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Arthritis Caused by MRSA CC398 in Patient without Animal Contact, Japan

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Clonal complex 398 methicillin-resistant *Staphylococcus aureus* (MRSA) is a typical lineage of livestock-associated MRSA. We report a case of intractable arthritis of the shoulder joint caused by a multidrug-resistant Panton-Valentine leukocidin–positive livestock-associated MRSA clonal complex 398 sequence type 1232 clone in a patient in Japan who had no animal contact.

n the past decade, methicillin-resistant *Staphylo*-Lcoccus aureus (MRSA) has been detected in livestock, including swine, poultry, and veal calves (1,2). In general, the virulence of animal-derived livestock-associated (LA-MRSA) strains is considered to be lower than that of community-acquired MRSA lineages (3). However, LA-MRSA strains can effectively colonize and infect humans, with subsequent transmission in both community and hospital settings. Human colonization with LA-MRSA sequence type (ST) 398 was first recognized among swine farmers in France and the Netherlands in the early 2000s (4). According to Larsen et al., clonal complex (CC) 398 MRSA accounted for 21% of MRSA isolated from skin and soft tissue infections in Denmark during 2010-2015 (5). However, ST398 MRSA has not been isolated in patients in Japan. We report a case of intractable arthritis of the shoulder joint caused by a multidrug-resistant Panton-Valentine leukocidin (PVL)-positive LA-MRSA CC398 (ST1232) clone in a patient in Japan who had no animal contact.

We performed MRSA identification, staphylococcal cassette chromosome (SCC) *mec* typing, *spa* typing, multilocus sequence typing (MLST), MIC determination, and PCR assays for detecting virulence factors and antimicrobial resistance genes, as described previously (1,6). The study protocol was approved by the Tokyo University of Pharmacy and Life Sciences Ethics Committee (approval no. 12–09).

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The patient, a 74-year-old man who lives in Tokyo, had received dialysis treatments 3 times a week since April 2018. He reported no overseas travel or animal contact. In September 2018, he felt pain in his right shoulder joint and was admitted to the Tokyo Medical University Hachioji Medical Center. At admission, his leukocyte count was 18,600 cells/ μ L and his C-reactive protein level was 7.0 mg/dL; we began treatment with cefazolin immediately (day 0). We isolated MRSA from venous blood and joint fluid, and we switched the antimicrobial agent to vancomycin on day 1.

Molecular epidemiologic analysis showed that the MRSA THI2018-120 strain we isolated is classified into SCCmec type V and spa type t034. Moreover, MLST analysis revealed that the strain was ST1232, a singlelocus variant of ST398 that belongs to CC398. When we determined antimicrobial susceptibilities, the THI2018-120 strain exhibited multidrug resistance to oxacillin, gentamicin, clarithromycin, clindamycin, and tetracycline (Table). We detected resistance genes for aminoglycoside (aacA-aphD) and tetracycline (tet[K]). However, we did not find known macrolide resistance genes, including ermA, ermB, ermC, ermT, mphC, and msrA/B, or clindamycin resistance genes lnuA, lnuB, lnuC, and lnuD (1,6). We conducted experiments to detect virulence factors, which detected the lukS/F-PV genes (Table). In addition, the THI2018-120 strain carried clfA, *clfB*, and *fnbA*, which are microbial surface components recognizing adhesive matrix molecules (6).

On day 18, we performed surgical debridement. On day 29, the patient had a drug reaction to vancomycin, so we switched the antimicrobial agent to daptomycin. On day 80, we added oral rifampin to the patient's regimen to treat prolonged chronic osteomyelitis. The patient's symptoms improved, and we switched from daptomycin to oral levofloxacin on day 108. On day 119, the patient was discharged when we no longer detected MRSA in pus from drained and nonopen lesions.

 Table. Antimicrobial drug resistance for clonal complex 398

 sequence type 1232 staphylococcal cassette chromosome mec

 type V methicillin-resistant Staphylococcus aureus strain isolated

 from a patient in Japan*

| nom a paton no apan | | |
|---------------------|-----------------|----------------|
| Antimicrobial drug | MIC, μg/mL | Susceptibility |
| Ampicillin | 4 | ND |
| Oxacillin | 8 | R |
| Fosfomycin | 0.5 | S |
| Gentamicin | 32 | R |
| Levofloxacin | 0.25 | S |
| Clarithromycin | 64 | R |
| Clindamycin | <u>></u> 256 | R |
| Tetracycline | 128 | R |
| Vancomycin | 1 | S |
| Daptomycin | 0.5 | S |

*Antimicrobial resistance genes included *mecA*, *aacA-aphD*, and *tet(K)*. Virulence factors included *lukS/F-PV*, *clfA*, *clfB*, and *fnbA*. ND, breakpoint is not defined by Clinical and Laboratory Standards Institute criteria; R, resistant; S, susceptible.

Human infections with the PVL-positive ST1232 MRSA strain are rare but were reported in New Zealand during 2011–2013 (1). In 2015, a fatal infection caused by a PVL-positive ST398 MRSA was reported in a patient who was infected in China but developed symptoms in Japan (7).

As mentioned, the virulence of animal-derived ST398 MRSA strains is considered to be lower than that of community-acquired MRSA (3). However, we presume that PVL production enhanced the severity of this case. Recent surveillance data suggest that not all cases of MRSA CC398 occurring among humans are related to animals (8). Our patient had no connection to animals, which suggests that this strain might be more common in Japan than previously thought. Further investigation, including whole-genome analysis of this isolate, could provide accurate phylogeny with higher resolution. In addition, a more robust estimation of this strain's virulence might elucidate the actual transmission route in this patient.

We previously reported the increased prevalence of the PVL-positive USA300 and USA300-LV clones in Japan, which were disseminated from North and Latin America (6). We also reported a case of septic arthritis by a PVL-positive ST772 Bengal-Bay clone, which is a predominant clone in India (9). Those data suggest that diverse and highly pathogenic PVL-positive MRSA clones have been entering Japan from abroad. We hypothesize that the PVL-positive ST1232 MRSA strain in this case also was transmitted from abroad. The transmission route of antimicrobial-resistant strains might be not only by humans but also by imported edible meat (10). From a One Health perspective, increased monitoring of imported livestock products is needed to prevent antimicrobial-resistant strains entering from other countries.

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Detection of Rocio Virus SPH 34675 during Dengue Epidemics, Brazil, 2011–2013

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Recent seroprevalence studies in animals detected Rocio virus in regions of Brazil, indicating risk for reemergence of this pathogen. We identified Rocio virus RNA in samples from 2 human patients for whom dengue fever was clinically suspected but ruled out by laboratory findings. Testing for infrequent flavivirus infections should expedite diagnoses.

Brazil has been affected by outbreaks caused by viruses of the genus *Flavivirus*, such as dengue (DENV), Zika, and yellow fever viruses, along with coinfections with other arboviruses (1). The amino acid sequences of polyproteins from viruses of this genus are very similar, which has limited the development of detection methods, often resulting in cross-reactions within serocomplexes during serologic testing (2). Therefore, tracking in areas where mosquito-specific flaviviruses co-circulate may have led to underestimated infections because of the detection and the hierarchy of disease based on medical importance.

Rocio virus (ROCV) is a potentially emerging neurotropic flavivirus in Brazil; however, because relatively little is known about the biology of this virus, technologies for its detection are limited (3–5). In 1975, ROCV was found to be related to the causative agent of a fatal outbreak of human encephalitis in Brazil; the case-fatality rate was 13%, and neurologic sequelae affected 20% of patients (5). The unexpected outbreak ended in 1980, but little documentation exists with regard to circulation of ROCV in Brazil.

To determine the extent of ROCV circulation in different areas of Brazil, we screened 647 serum samples collected during an outbreak of dengue fever during 2011–2013. The samples came from patients in care units of the public health system, which offer 24-hour outpatient urgent care, and emergency services in the city of Goiânia, central Brazil. The samples were from patients of all age groups and sexes