



Published in final edited form as:

*Epidemiol Infect.* 2014 January ; 142(1): 163–171. doi:10.1017/S0950268813000800.

## Surveillance during an era of rapidly changing poliovirus epidemiology in India: the role of one vs. two stool specimens in poliovirus detection, 2000–2010

C. V. CARDEMIL<sup>1,2,\*</sup>, M. RATHEE<sup>3</sup>, H. GARY<sup>1</sup>, K. WANNEMUEHLER<sup>1</sup>, A. ANAND<sup>1</sup>, O. MACH<sup>1</sup>, S. BAHL<sup>3</sup>, S. WASSILAK<sup>1</sup>, S.Y. CHU<sup>1</sup>, A. KHERA<sup>4</sup>, H.S. JAFARI<sup>3</sup>, and M.A. PALLANSCH<sup>5</sup>

<sup>1</sup>Global Immunization Division, Center for Global Health, Centers for Disease Control and Prevention, Atlanta, GA, USA

<sup>2</sup>Epidemic Intelligence Service, Centers for Disease Control and Prevention, Atlanta, GA, USA

<sup>3</sup>World Health Organization – India, National Polio Surveillance Project, New Delhi, India

<sup>4</sup>Ministry of Health and Family Welfare, Government of India, New Delhi, India

<sup>5</sup>Division of Viral Diseases, National Center for Immunization and Respiratory Diseases, Centers for Disease Control and Prevention, Atlanta, GA, USA

### SUMMARY

Since 2004, efforts to improve poliovirus detection have significantly increased the volume of specimen testing from acute flaccid paralysis (AFP) patients in India. One option to decrease collection and testing burden would be collecting only a single stool specimen instead of two. We investigated stool specimen sensitivity for poliovirus detection in India to estimate the contribution of the second specimen. We reviewed poliovirus isolation data for 303984 children aged <15 years with AFP during 2000–2010. Using maximum-likelihood estimation, we determined specimen sensitivity of each stool specimen, combined sensitivity of both specimens, and sensitivity added by the second specimen. Of 5184 AFP patients with poliovirus isolates, 382 (7.4%) were identified only by the second specimen. Sensitivity was 91.4% for the first specimen and 84.5% for the second specimen; the second specimen added 7.3% sensitivity, giving a combined sensitivity of 98.7%. Combined sensitivity declined, and added sensitivity increased, as the time from paralysis onset to stool collection increased ( $P = 0.032$ ). The sensitivity added by the second specimen is important to detect the last chains of poliovirus transmission and to achieve certification of polio eradication. For sensitive surveillance, two stool specimens should continue to be collected from each AFP patient in India.

### Keywords

Infectious disease epidemiology; laboratory tests; polio; surveillance; surveillance system

\* Author for correspondence: Dr C. V. Cardemil, Centers for Disease Control and Prevention, 1600 Clifton Road, Atlanta, GA 30333, USA. (ccardemil@cdc.gov).

DECLARATION OF INTEREST

None.

## INTRODUCTION

Acute flaccid paralysis (AFP) surveillance forms the basis for detection of poliovirus cases globally. In the Global Polio Eradication Initiative (GPEI), AFP is defined as rapid progression of weakness with loss of voluntary movement and loss of muscle tone in any part of the body in a patient aged <15 years; or paralysis in a person of any age in whom polio is suspected [1]. AFP surveillance was started in India in 1997, and includes case investigations of all persons with suspected AFP, with collection of two stool specimens for poliovirus isolation [2]. In 2004–2005, in an effort to increase the sensitivity of the AFP surveillance system, the AFP case definition was broadened to include transient weakness and facial paralysis. To further enhance sensitivity, the number of AFP case-reporting sites nationally increased from 21403 in 2004 to 36629 in 2012 in an effort to include health facilities serving migrant and high-risk populations [2]. As a likely consequence of changes to increase sensitivity, the non-polio AFP rate in India increased from 2.0 to 13.5 cases/100000 population aged <15 years from 2000 to 2011, and is substantially higher than the World Health Organization (WHO) target rate of at least two non-polio AFP cases/100000 population aged <15 years [2, 3]. However, increased reporting of AFP cases requires higher resources for both case investigations and laboratory testing [4, 5]. While AFP surveillance accounted for <10% of the overall cost of the polio eradication programme in India in 2011 [6], AFP surveillance costs are not expected to decrease, even as India was removed from the WHO list of polio-endemic countries in January 2011, because of the need for a continued strong surveillance system [7].

Collection of two stool specimens is recommended by WHO because poliovirus is shed intermittently in stool, so testing only one sample could potentially miss an opportunity to detect the virus [8]. Nonetheless, collecting one stool specimen instead of two has been proposed as one way to reduce the case investigation and laboratory testing burden, and cost of the AFP surveillance system in India and other countries. In November 2010, the India Expert Advisory Group considered the role of the second stool sample for polio diagnosis in the context of the decline in reported polio cases from 266 in 2000 to 42 in 2010 [9]. Our study aimed to assess: (1) changes in stool collection and processing performance indicators, (2) the sensitivity of stool specimens in polio diagnosis, and (3) the number of polio cases identified only by the second specimen for AFP cases reported during 2000–2010. Using these results, we provide recommendations for maintaining high quality, sensitive AFP surveillance while taking into account limited programmatic resources.

## METHODS

We restricted the analysis to AFP cases in children aged <15 years reported to the National Polio Surveillance Project – India from 2000 to 2010. Stool specimens were tested at one of eight national laboratories in the country using cell culture on two poliovirus-sensitive cell lines according to WHO standards, followed by antigenic and/or molecular characterization of isolates [10]. The first and second specimens from each individual were analysed by the same methodology. Stool collection and processing performance indicators were assessed using WHO criteria [1]. Two stool specimens should be collected from each AFP case-patient within 14 days of paralysis onset and at least 24 h apart; each specimen must be of

adequate volume (8–10 g), and arrive at a WHO-accredited laboratory in good condition (i.e. no desiccation, no leakage, with adequate documentation and evidence that the cold chain was maintained) [1]. Polio cases were considered to be confirmed if wild poliovirus (WPV) type 1, WPV type 3, or vaccine-derived poliovirus (VDPV) was isolated in either stool specimen.

Polio cases with two stool results reported were further analysed to estimate single specimen, combined specimen, and added sensitivity. Methods for calculating the specimen sensitivity of each individual stool, the combined sensitivity of both first and second stool specimens (i.e. person sensitivity), and the added sensitivity of the second specimen have been described previously [11] (Fig. 1). In brief, this methodology derives maximum-likelihood estimates for the specimen sensitivities for each stool, and the approximate variances of the estimators. Each stool sample was considered to be independent of the other; therefore, each stool sample serves as the gold standard estimate for the other specimen.

Sensitivity estimates for confirmed polio cases were calculated for the entire period during 2000–2010 and by various factors, including stool adequacy, stool condition on arrival at the laboratory, and time interval of stool collection from onset of paralysis, patient age, and year. The Wald  $\chi^2$  test was used to test for equality of mean specimen sensitivities across categories. Results were divided into three time categories: 2000–2004 (before the AFP case definition change), 2005–2009 (after the AFP case definition change), and 2010 (a year of particularly low reported polio cases). The number of polio cases identified only by the second stool specimen was determined for the overall period 2000–2010, and by the same factors as in the sensitivity analyses. The Cochran–Armitage trend test was used to test for trends in proportions across ordinal categories. Data were analysed in SAS v. 9.2 (SAS Institute Inc., USA) and R v. 2.12.1 (R Foundation, Austria).

## RESULTS

### AFP cases, laboratory samples processed and stool surveillance indicators

From 2000 to 2010, 303984 children aged <15 years with AFP were identified. Of these, 290 763 (95.7%) had two stool samples with poliovirus isolation results reported, 2537 (0.8%) were missing one poliovirus isolation result, and 10684 (3.5%) were missing both poliovirus isolation results. The median age of children with AFP was 37 months (interquartile range 22–71), 207 774 (68%) AFP cases were children aged <5 years, and 108998 (41%) were females.

During 2000–2010, reported AFP cases increased from 8095 to 55616 and stool specimens processed by the laboratory increased from 15761 to 108 207 (Fig. 2). Confirmed polio cases fluctuated during 2000–2010, with peaks in 2002 (1603 cases), 2007 (877 cases), and 2009 (763 cases), and troughs in 2005 (66 cases) and 2010 (47 cases) (Fig. 2).

Indicators of stool adequacy, timeliness of specimen collection, condition on arrival at the laboratory, and time interval from collection of first and second stool specimens were above

the WHO target level of 80% for the entire period of 2000–2010 (Table 1), with little fluctuation from year to year.

The mean interval from onset of paralysis to arrival of the second stool specimen at the laboratory was 14.7 days during 2000–2004, 13.0 days during 2005–2009, and 12.2 days during 2010 (Fig. 3). The mean time interval from onset of paralysis to collection of the first stool sample accounted for the largest percentage of the total time interval in all categories, and decreased by 14% from 9.2 days during 2000–2004 to 7.9 days during 2010. The mean time interval from collection of the second stool specimen to its being sent to the laboratory decreased by 72% from 3.2 days during 2000–2004 to 1.2 days during 2010.

### Stool specimen sensitivity, combined sensitivity, and added sensitivity

Estimates of stool specimen sensitivity were calculated for the 5184 polio-confirmed cases with WPV type 1, WPV type 3, or VDPV isolated from at least one stool specimen (Table 2).

During 2000–2010, the specimen sensitivity of the first stool (91.4%) was significantly higher than the specimen sensitivity of the second stool (84.5%) ( $P < 0.0001$ ), combined sensitivity was 98.7%, and the added sensitivity of the second stool specimen was 7.3% (Table 3). There were no differences in sensitivity estimates by time period category ( $P = 0.960$ ). Specimen sensitivity was significantly higher when both stool specimens were adequate than when either of the specimens were not adequate ( $P = 0.037$ ). Of 350 specimens where both specimens were not adequate, 316 (90.3%) were inadequate due to poor timeliness (both stools not collected within 14 days after the onset of paralysis) and 30 (8.6%) were inadequate due to poor condition at the time of arrival at the laboratory.

Of the factors analysed affecting adequacy (stool condition, interval from onset of paralysis to collection of stool specimen 1, interval from collection of stool specimen 1 to collection of stool specimen 2), only interval from onset of paralysis to collection of stool specimen 1 significantly affected sensitivity ( $P = 0.03$ ) (Table 3). Specimens collected in the first week after the onset of paralysis had the highest specimen sensitivity (stool 1, 92.9%; stool 2, 85.8%) and combined sensitivity (99.0%), and lowest added sensitivity (6.1%) compared to later weeks. When categorized by the WHO timeliness guideline, specimens collected  $\leq 14$  days after the onset of paralysis had significantly higher specimen sensitivity (stool 1, 91.9%; stool 2, 85.4%) than specimens collected after the first 14 days (stool 1, 82.8%; stool 2, 72.9%;  $P = 0.025$ ). Specimens collected  $\leq 7$  days after the onset of paralysis also had higher specimen and combined sensitivity, and lower added sensitivity, than those collected  $> 7$  days, although this difference was not statistically significant ( $P = 0.071$ ).

Of children aged 60–180 months, specimen sensitivity was lower and added sensitivity was higher compared to specimen sensitivity and added sensitivity in children aged 0–59 months; however, the difference in mean specimen sensitivity between age groups was not statistically significant ( $P = 0.069$ ). A higher percentage of stools was collected after 14 days from children aged 60–180 months (21/160, 13%) than from children aged 0–59 months (298/5024, 6%).

### Cases identified by second stool specimen only

Of 5184 confirmed polio cases identified from 2000 to 2010, 382 (7.4%) were identified only by the second stool sample (Table 4). The percentage of cases identified only by the second stool sample was higher if both stools were inadequate ( $P=0.0001$ ), as the stool collection interval increased ( $P<0.0001$ ), and as patient age in months increased ( $P=0.0004$ ). Of the 382 confirmed polio cases identified by the second stool specimen only, 38 (10%) were from 33 districts and nine states that had previously been polio-free for at least 6 months (i.e. not geographically related to an ongoing outbreak).

## DISCUSSION

The second stool specimen contributed an additional 7.3% sensitivity overall, resulting in a combined sensitivity of 98.7% for both specimens. Both the first and second specimens were most sensitive if collected in the first week after the onset of paralysis. The second specimen detected an additional 382 polio cases that would not otherwise have been identified, which represents 7.2% of all polio-confirmed cases reported during 2000–2010; 38 (10%) were not geographically related to an ongoing outbreak. These results highlight the substantial contribution of the second stool specimen in India in identifying polio cases.

The first stool specimen had higher specimen sensitivity (91.4%) than the second stool specimen (84.5%). Reasons for lower specimen sensitivity for the second specimen are unclear, but could be related to the later time of second stool collection after illness onset [11, 12], viral excretion patterns, or to factors related to second stool specimen collection. Nonetheless, the second specimen increased the sensitivity of detection of confirmed polio cases as indicated by added sensitivity and the absolute number of cases identified by the second specimen only.

The lower specimen sensitivities when specimens were inadequate and in older children are due in part to the later collection time for some of these samples. This has greater implications currently as polio is historically a disease of young childhood; now, however, cases are increasingly occurring in older children and even in adults [13, 14]. As a result, healthcare providers might not recognize polio as the cause of AFP in older age groups as quickly as they do in younger children, which could lead to delayed stool collection times. Alternatively, older children who have partial immunity might have transient, asymptomatic infection with intermittent viral shedding resulting in discordant stool sample results; these discordant results would in turn decrease sensitivity.

The percentage of cases identified only by the second sample (7.2%) is similar to findings of previous studies in Latin America (8%) [4] and from the USA (10%) [8] but is lower than estimates reported previously from India (21%) [15] and the Western Pacific region (31%) [16]. The lower estimate in our analysis compared to a previous analysis of India's AFP surveillance system [15] could be due to improvements in the collection, storage and transport of specimens, as well as improved laboratory performance. Indeed, by 2000, both specimen sensitivity and the percentage of cases identified by the second stool sample (12%) approached comparable levels to the current analysis from 2000 to 2010. The fact that we did not find an increase in specimen sensitivity after 2000 suggests there may be a point at

which it is difficult to further increase specimen sensitivity despite improvements in field and laboratory performance.

The findings in this paper are subject to at least two limitations. First, the laboratories testing specimens changed during the analysis period and the prevalence of positive specimens going to specific laboratories also changed, making it difficult to assess laboratory-specific data over time. Second, for this analysis we must assume that isolation of poliovirus from one stool specimen is independent of the other. However, both samples are collected from the same AFP case-patient, often by the same health worker, and are usually transported together to the laboratory. Factors affecting specimen quality, including handling, packaging, and temperature, will affect both specimens equally, and may increase or decrease the laboratory's ability to isolate poliovirus. This lack of complete independence could result in overestimating or underestimating sensitivity. Nonetheless, the contribution of the second specimen is still marked, as this limitation does not affect the number of cases that would not have been detected without the second specimen.

India's AFP surveillance system is a 'best-case' scenario compared to other systems globally and caution should be exercised when generalizing the results to other areas with less highly functioning surveillance systems. On the other hand, some results from this study have important implications for other settings. It is a struggle to collect one stool specimen in some countries, and many specimens reach the laboratory in inadequate condition, or are collected too late after the onset of paralysis [17]. In these settings, specimen and combined sensitivity are likely to be lower because of later collection times, and the second stool sample would have greater impact on the sensitivity of AFP surveillance. But even in a well-functioning AFP surveillance system, the second stool sample detected cases that otherwise would have been missed, and added sensitivity to detection.

In addition to decreasing the number of stool specimens collected, other options exist for potentially reducing the burden of collecting and testing on the surveillance system and the laboratory. Returning to the AFP case definition used in India prior to 2005, or maintaining environmental sampling at current capacity rather than expanding sites, could achieve this goal. However, these scenarios will necessarily decrease sensitivity, to some extent. In principle, efforts could be reduced in the laboratory by changing from cell culture-based detection of poliovirus to newer molecular methods. Despite advances in these methods, cell culture in RD cells remains more sensitive than real-time PCR methods for detection of poliovirus from stool specimens [18]. While these methods require less personnel time, the cost of reagents is significantly higher if they are to be applied routinely. To date, the GPEI has not been willing to trade reduced sensitivity for a faster, but more expensive, laboratory result.

In certain circumstances, ending collection and testing of the second stool specimen might be appropriate. After examination of data from the Americas, it was suggested that in endemic countries with high laboratory proficiency, collection of the second stool sample may not be necessary [4, 19, 20]. As a result, a recommendation to collect only one stool sample was made by the Pan American Health Organization after certification of polio eradication in the Americas. For India, consideration could be given to ending the collection



and testing of the second stool specimen in the post-eradication era (following national certification 3 years after the last case is reported), if the AFP surveillance system continues to be strong. At the present time, however, when identification of every case is critical and maximum sensitivity is required to mitigate the risk of failure to detect poliovirus importations, our findings support continuing the collection of two stool specimens for AFP surveillance in India.

## ACKNOWLEDGEMENTS

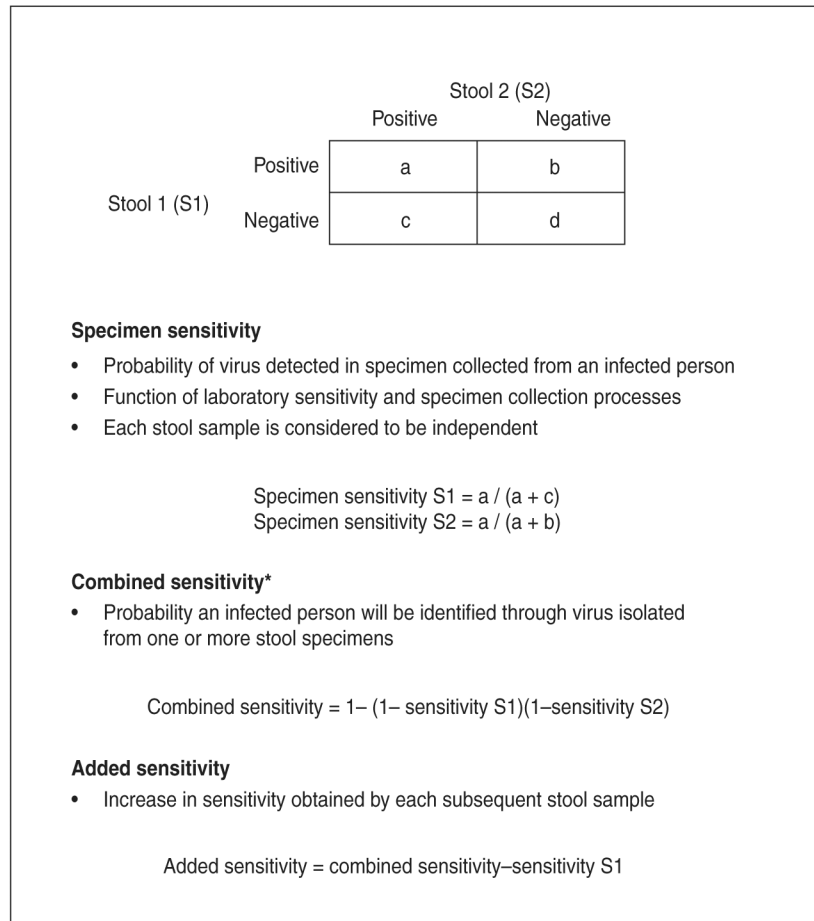
The authors acknowledge the World Health Organization – India for the contribution in coordinating, compiling and supporting data analysis for this study. We also thank the Government of India for making the data available for the study. This research was conducted as part of usual CDC activities and received no specific grant from any funding agency, commercial or not-for-profit sectors.

## REFERENCES

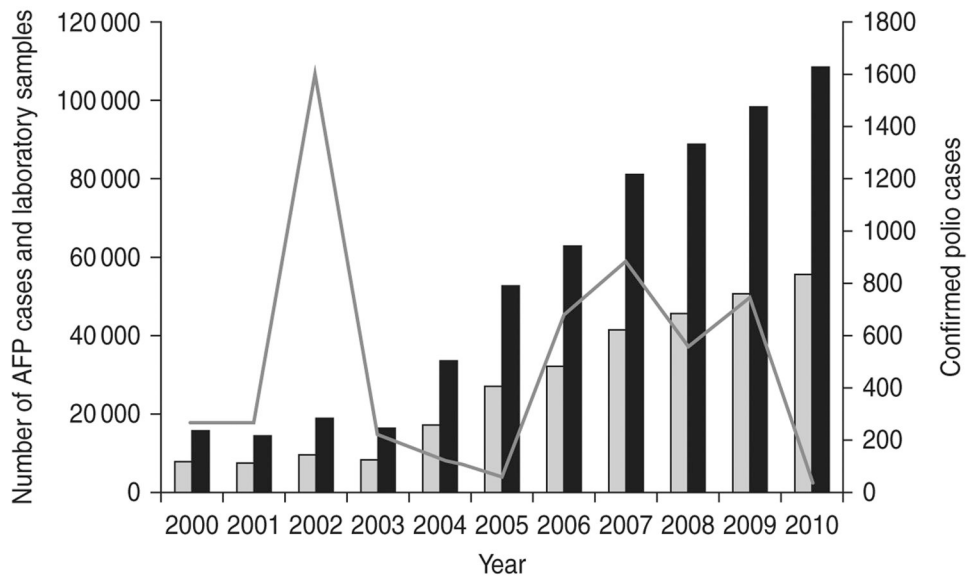
1. Government of India. Field Guide: Surveillance of Acute Flaccid Paralysis, 3rd edn. Child Health Division, Department of Family Welfare, Ministry of Health & Family Welfare, New Delhi 9 2005.
2. National Polio Surveillance Project. Components of AFP surveillance (<http://www.npsindia.org/Surveillance%20Strategy.asp>). Accessed 20 October 2012.
3. Centers for Disease Control and Prevention. Progress toward poliomyelitis eradication – India, January 2010–September 2011. *Morbidity and Mortality Weekly Report* 2011; 60: 1482–1486. [PubMed: 22048729]
4. Pinheiro FP, et al. Eradication of wild poliovirus from the Americas: wild poliovirus surveillance – laboratory issues. *Journal of Infectious Diseases* 1997; 175 (Suppl. 1): S43–49. [PubMed: 9203691]
5. National Polio Surveillance Project. Laboratory budget analysis, 11 2011.
6. World Health Organization. Global poliomyelitis eradication initiative: summary of external resource requirements by major category of activity, 2011–2012.
7. Independent Monitoring Board of the Global Polio Eradication Initiative. Report, 2 2012 ([www.polioeradication.org/portals/0/document/aboutus/governance/imb/5imbmeeting/imbreport\\_january2012.pdf](http://www.polioeradication.org/portals/0/document/aboutus/governance/imb/5imbmeeting/imbreport_january2012.pdf)). Accessed 20 October 2012.
8. Lennette EH. Problems of the viral diagnostic laboratory with respect to poliomyelitis. *Poliomyelitis: papers and discussions presented at the Fourth International Poliomyelitis Conference Philadelphia: J. B. Lippincott, 1958, pp. 377–386.*
9. India Expert Advisory Group for Polio Eradication. Conclusions and recommendations, 22nd Meeting of the India Expert Advisory Group for Polio Eradication, New Delhi, India, 1–2 11 2010.
10. World Health Organization Polio Laboratory Manual, 4th edn. Geneva, Switzerland, 2004 (<http://www.who.int/vaccines/en/poliolab/WHO-Polio-Manual-9.pdf>). Accessed 4 February 2013.
11. Gary HE Jr., Sanders R, Pallansch MA. A theoretical framework for evaluating the sensitivity of surveillance for detecting wild poliovirus: I. Factors affecting the detection sensitivity in a person with acute flaccid paralysis. *Journal of Infectious Diseases* 1997; 175 (Suppl. 1): S135–140. [PubMed: 9203705]
12. Alexander JP Jr., Gary HE Jr., Pallansch MA. Duration of poliovirus excretion and its implications for acute flaccid paralysis surveillance: a review of the literature. *Journal of Infectious Diseases* 1997; 175 (Suppl. 1): S176–182. [PubMed: 9203713]
13. Centers for Disease Control and Prevention. Outbreaks following wild poliovirus importations – Europe, Africa, and Asia, January 2009–September 2010. *Morbidity and Mortality Weekly Report* 2010; 59: 1393–1399. [PubMed: 21048560]
14. Centers for Disease Control and Prevention. Notes from the field: poliomyelitis outbreak – Republic of the Congo, September 2010–February 2011. *Morbidity and Mortality Weekly Report* 2011; 60: 312–313. [PubMed: 21412213]

15. Kohler KA, et al. Contribution of second stool specimen to increased sensitivity of poliovirus detection in India, 1998–2000. *Epidemiology and Infection* 2003; 131: 711–811. [PubMed: 12948371]
16. Yoneyama T, et al. Necessity of two-stool sample test for sensitive detection of poliovirus. *Japan Journal of Infectious Diseases* 2001; 54: 250–251.
17. World Health Organization. Performance of acute flaccid paralysis (AFP) surveillance and incidence of poliomyelitis, 2011. *Weekly Epidemiological Record* 2011; 86: 575–579. [PubMed: 22180898]
18. World Health Organization. Summary of discussions and recommendations of the 18th Informal Consultation of the Global Polio Laboratory Network, 28–29 6 2012 ([http://www.polioeradication.org/Portals/0/Document/Resources/GPLN\\_publications/GPLN\\_Meeting\\_recommendations\\_2012.pdf](http://www.polioeradication.org/Portals/0/Document/Resources/GPLN_publications/GPLN_Meeting_recommendations_2012.pdf)). Accessed 4 February 2013.
19. Silveira CM, et al. Polio diagnosis: one or two samples? *EPI Newsletter: Expanded Program on Immunization in the Americas* 1995; XVII: 1–2.
20. de Quadros CA, et al. Eradication of wild poliovirus from the Americas: acute flaccid paralysis surveillance, 1988–1995. *Journal of Infectious Diseases* 1997; 175 (Suppl. 1): S37–42. [PubMed: 9203690]

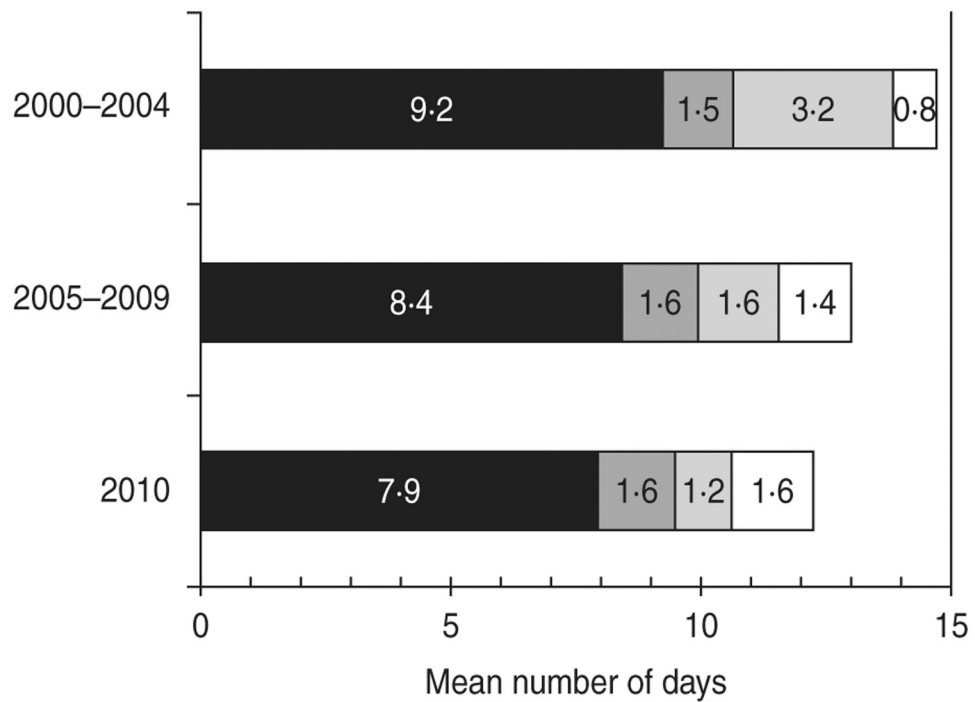




**Fig. 1.** Sensitivity definitions and calculations (Gary *et al.* [11]). \* Also known as person sensitivity.



**Fig. 2.** Acute flaccid paralysis (AFP) cases reported, confirmed polio cases reported, and stool specimens processed, India, 2000–2010. Confirmed polio cases include wild poliovirus (WPV) type 1, WPV type 3, and vaccine-derived poliovirus. ■, AFP cases; ■, laboratory samples processed; —, confirmed polio cases.



**Fig. 3.** Mean time interval from onset of paralysis and arrival of second specimen in laboratories, by components, India, 2000–2010. ■, Time from paralysis to collection of first stool sample (S1); ■, time from S1 collection to collection of second stool sample (S2); ■, time from S2 collection to sending S2 to the laboratory; □, time from sending S2 to arrival at the laboratory.

**Table 1.**

Indicators of stool specimen adequacy, timeliness, condition, and collection for acute flaccid paralysis (AFP) cases reported, India, 2000–2010\*

	<i>N</i>	%
<b>Stool adequacy<sup>†</sup></b>		
Both stools adequate	248829	85.5
Either inadequate	5706	1.8
Both inadequate	36515	12.6
<b>Time of stool specimen collection from onset of paralysis</b>		
Both stools ≤4 days	250651	86.1
Only stool 1 ≤4 days	4766	1.6
Both stools >14 days	35679	12.3
<b>Stool condition<sup>‡</sup></b>		
Both stools good	288892	99.3
Either poor	1249	0.4
Both poor	932	0.3
<b>Interval from collection of specimen 1 to collection of specimen 2</b>		
24–48 h	266149	91.3
>48 h	25202	8.7

\* AFP cases with missing data excluded from analysis; for all categories, <5% of all AFP cases in the database.

<sup>†</sup> Stool adequacy is defined as two stool specimens collected from each AFP case within 14 days of paralysis onset and at least 24 h apart; each specimen must be of adequate volume (8–10 g), and arrive at a WHO-accredited laboratory in good condition.

<sup>‡</sup> A stool specimen in good condition is defined as a specimen that arrives in the laboratory with neither desiccation nor leakage, with adequate documentation and evidence that the cold chain was maintained.

Non-polio acute flaccid paralysis (AFP) and polio-confirmed cases analysed for sensitivity, by outcome of testing

**Table 2.**

	Outcome of testing		Frequency				Total
	Specimen 1	Specimen 2	2000-2004	2005-2009	2010	2010	
Non-polio AFP cases	-	-	42 818	177 217	50061	270 096	
Polio-confirmed cases	+	+	1898	2123	35	4056	
	+	-	333	408	5	746	
	-	+	175	203	4	382	
Total polio-confirmed cases with two stool results			2406	2734	44	5184	

+, Indicates isolation of wild poliovirus (WPV) type 1, WPV type 3 or vaccine-derived poliovirus; -, indicates negative for all polioviruses

Sensitivity of first and second poliovirus specimens from children with acute flaccid paralysis (AFP) aged <15 years with two specimen results, India, 2000–2010

Table 3.

	N*	Specimen sensitivity stool 1, %	Specimen sensitivity stool 2, %	Combined sensitivity of stools 1 and 2, %	Added sensitivity of stool 2, %	P value <sup>†</sup>
<b>All polio-confirmed AFP cases, 2000–2010</b>						
	5184	91.4 (90.5–92.2)	84.5 (83.4–85.5)	98.7 (98.1–99.2)	7.3 (5.9–8.6)	<0.0001
<b>Time period</b>						
2000–2004	2406	91.6 (88.7–94.4)	85.1 (80.2–89.9)	98.7 (97.9–99.6)	7.2 (5.1–9.3)	0.960
2005–2009	2734	91.3 (88.7–93.9)	83.9 (79.2–88.5)	98.6 (97.8–99.4)	7.3 (5.5–9.2)	
2010	44	89.7 (64.6–100.0)	87.5 (57.2–100.0)	98.7 (92.6–100.0)	9.0 (0.0–28.3)	
<b>Stool adequacy<sup>‡</sup></b>						
Both adequate	4747	92.0 (90.0–94.0)	85.3 (81.7–88.8)	98.8 (98.3–99.4)	6.8 (5.4–8.3)	0.037
Neither adequate	350	84.2 (77.5–90.9)	75.2 (65.2–85.2)	96.1 (93.0–99.2)	11.9 (8.0–15.8)	
<b>Stool condition<sup>§</sup></b>						
Both good	5122	91.4 (89.4–93.3)	84.4 (81.1–87.8)	98.7 (98.1–99.2)	7.3 (5.9–8.7)	0.794
Both poor	30	89.7 (33.4–100.0)	96.3 (75.3–100.0)	99.6 (95.4–100.0)	10.0 (0.0–62.4)	
<b>Interval from onset of paralysis to collection of first stool specimen, days</b>						
≤7	3628	92.9 (90.7–95.1)	85.8 (81.6–90.0)	99.0 (98.4–99.6)	6.1 (4.4–7.7)	0.032
8–14	1237	88.8 (84.5–93.0)	83.2 (77.0–89.4)	98.1 (96.8–99.5)	9.4 (6.4–12.3)	
15–21	163	84.4 (73.0–95.8)	76.6 (60.4–92.8)	96.3 (91.4–100.0)	12.0 (5.1–18.8)	
22–60	156	81.7 (71.0–92.4)	69.9 (53.2–86.0)	94.4 (89.4–99.5)	12.7 (5.6–19.9)	
<b>Interval from collection of specimen 1 to collection of specimen 2, hours</b>						
24–48	4695	91.5 (89.5–93.6)	85.2 (81.7–88.6)	98.7 (98.2–99.3)	7.2 (5.7–8.7)	0.312
>48	489	90.0 (84.4–95.6)	77.8 (66.3–89.3)	97.8 (95.5–100.0)	7.8 (4.4–11.2)	
<b>Patient age, months</b>						
0–59	5024	91.6 (89.7–93.6)	84.9 (81.5–88.3)	98.7 (98.2–99.3)	7.1 (5.7–8.5)	0.069
60–180	160	82.7 (72.5–92.8)	70.5 (54.5–86.5)	94.9 (89.5–100.0)	12.2 (9.8–14.6)	
<b>Serotype</b>						
Wild poliovirus type 1	2990	91.3 (88.7–93.8)	84.5 (80.2–88.9)	98.6 (97.9–99.4)	7.4 (6.6–8.1)	0.982
Wild poliovirus type 3	2175	91.4 (88.4–94.4)	84.3 (79.1–89.5)	98.6 (97.8–99.5)	7.3 (6.4–8.1)	

	<i>N</i> <sup>*</sup>	Specimen sensitivity stool 1, %	Specimen sensitivity stool 2, %	Combined sensitivity of stools 1 and 2, %	Added sensitivity of stool 2, %	<i>P</i> value <sup>†</sup>
Vaccine-derived poliovirus (VDPV)	22	94.4 (83.8–100.0)	81.0 (64.2–97.8)	98.9 (91.4–99.8)	4.5 (0.0–18.9)	//

<sup>\*</sup>The category denominators for stool adequacy and stool condition do not add up to the overall denominator ( $n = 5184$ ) due to exclusion of certain subcategories due to small sample size (one stool adequate,  $n = 87$ ; one stool in good condition,  $n = 32$ ). Because serotypes are not mutually exclusive [of 5184 polio-confirmed cases nine were dually infected with wild poliovirus (WPV) types 1 and 3], it was necessary to treat WPV type 1 and type 3 cases independently for sensitivity analyses; this analysis assumes that one serotype does not influence the behaviour of the other in dually infected persons.

<sup>†</sup>*P* value represents the Wald  $\chi^2$  test for equality of mean specimen sensitivities across categories. For the interval from onset of paralysis to collection of first stool specimen, the *P* value represents the linear trend for mean specimen sensitivity across time collection intervals.

<sup>‡</sup>Stool adequacy is defined as two stool specimens collected from each AFP case within 14 days of paralysis onset and at least 24 h apart; each specimen must be of adequate volume (8–10 g), and arrive at a WHO-accredited laboratory in good condition.

<sup>§</sup> stool specimen in good condition is defined as a specimen that arrives at the laboratory with neither desiccation nor leakage, with adequate documentation and evidence that the cold chain was maintained.

// VDPV represents a separate category that is not directly comparable to other serotypes, so no statistical test was applied.



**Table 4.**

Percentage of polio-confirmed acute flaccid paralysis cases identified only by the second stool specimen, India, 2000–2010

	Total polio cases identified	Cases identified only by second stool (%)	<i>P</i> value *
<b>Overall</b>	5184	382 (7.4)	
<b>Stool condition</b>			
Both good	5122	379 (7.4)	0.5421
Both poor	30	3 (10.0)	
<b>Stool adequacy</b>			
Both adequate	4747	328 (6.9)	<0.0001
Both inadequate	350	45 (12.9)	
<b>Interval from paralysis onset to collection of first stool specimen (days)</b>			
<8	3628	222 (6.1)	<0.0001
8–14	1237	118 (9.5)	
15–21	163	20 (12.4)	
22–60	156	22 (13.9)	
<b>Patient age, months</b>			
0–11	1586	104 (6.6)	0.0004
12–23	2105	141 (6.7)	
24–35	847	72 (8.5)	
36–59	475	43 (9.1)	
60–180	171	22 (12.9)	
<b>Time period category</b>			
2000–2004	2406	175 (7.3)	0.7527
2005–2009	2734	203 (7.4)	
2010	44	4 (9.1)	

\* *P* value represents trend test for proportions across ordinal categories.