



HHS Public Access

Author manuscript

Thyroid. Author manuscript; available in PMC 2020 May 01.

Published in final edited form as:

Thyroid. 2019 May ; 29(5): 631–641. doi:10.1089/thy.2018.0417.

Pre and Postnatal Polybrominated Diphenyl Ether Concentrations in Relation to Thyroid Parameters Measured During Early Childhood

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Abstract

Background: Penta-brominated diphenyl ethers (PentaBDEs) are endocrine disrupting chemicals that structurally resemble thyroid hormones and were widely used as flame retardants in household consumer products from 1975–2004. Polybrominated diphenyl ethers (PBDEs) cross the placenta and evidence suggests that for many children, body burdens may peak during toddler years. We aimed to understand the impact of exposure timing by examining both pre- and postnatal exposure to BDE-47, the predominant PentaBDE congener detected in humans, in relation to thyroid hormone parameters measured during early childhood.

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Disclosure: The authors report no conflicts of interest in this work.

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Methods: The Columbia Center for Children’s Environmental Health Mothers and Newborns Study is a prospective birth cohort of African American and Dominican maternal-child pairs. Pregnant women were recruited from two prenatal clinics in Northern Manhattan and the South Bronx between 1998 and 2006. Participants included 158 children with 1) plasma PBDE concentrations measured at birth and toddler years (age 2-3), and 2) serum thyroid parameters measured at 3 and/or 5 years. Outcomes included concentrations of serum thyroid stimulating hormone (TSH), free thyroxine (fT₄) and total thyroxine (T₄).

Results: Children with high exposure to BDE-47 during the prenatal period (–17%, 95% CI – 29, –2) or toddler age (-19%, 95% CI: –31, –5) had significantly lower geometric mean TSH levels compared to children with low BDE-47 exposure throughout early life. Associations with T₄ were also inverse, however, they did not reach statistical significance at the p=0.05 level. Sex-stratified models suggest associations with postnatal exposure may be stronger among boys compared to girls.

Conclusions: The thyroid regulatory system may be sensitive to BDE-47 during prenatal and postnatal periods.

Keywords

flame retardants; organohalogen; endocrine disruption; prenatal; childhood; thyroid hormone

Introduction

Endocrine disrupting chemicals (EDCs) contribute substantially to human morbidity and are estimated to result in hundreds of billions in costs per year (1). EDCs are defined by their ability to cause changes in endocrine function and consist of several classes of chemicals with varying structures, actions and endocrine targets (2). Many EDCs were introduced into United States commerce beginning in the 1970s to augment food packaging, personal care products and other household items (3). For example, following reports indicating improperly extinguished cigarettes were the leading cause of household fires (4), polybrominated diphenyl ethers (PBDEs) began to be used as flame retardants in electronics and household furnishings in 1975 (5). PBDEs were used as three technical mixtures known as PentaBDE, OctaBDE, and DecaBDE, each of which is comprised of several congeners. PentaBDE, which is estimated to make up 90% of the human body burden, was primarily applied to products containing polyurethane foam, including couches, car seats, carpet padding and other upholstered items (6). PentaBDE was often used in large volumes; for example, reports indicate it comprised up to 3% by weight of the polyurethane foam contained within a couch (7). During manufacturing, PBDEs are not chemically bonded to base polymers and thus have a propensity to migrate into the indoor environment and settle in house dust (8). Human exposure occurs primarily through incidental ingestion of dust, placing young children at risk for elevated exposure due to their frequent hand to mouth behavior and often close proximity to the floor (5, 9–11). Owing to their lipophilic properties, PBDEs have long half-lives (PentaBDE congeners: 1.6- 6.5 years) (12) and are known to penetrate the fat-soluble placenta, as well as partition into breastmilk (13, 14). It is estimated that over 46,000 tons of PentaBDE were used in North America until its phase-out in 2004, leading to nearly ubiquitous exposure and body burdens that are the highest in the

world (15, 16). Despite their phase-out, PBDE exposure continues owing to their resistance to environmental degradation and ongoing release from consumer products that are infrequently replaced (15).

PBDE congeners consist of a diphenyl ether backbone around which varying numbers of bromine atoms are attached (5). This molecular structure closely resembles that of the halogenated (iodine) thyroid hormones triiodothyronine (T_3) and thyroxine (T_4) (17), supporting the putative interaction of PBDEs with thyroid hormone transport proteins, receptors and/or degradation enzymes (18). Thyroid hormones bind to receptors in nearly every organ in the human body and play critical roles in the regulation of growth, metabolism and brain development (19). *In vitro* research indicates hydroxylated metabolites of BDE-47, the predominant congener detected in humans, markedly inhibit the capacity of T_3 to bind with receptors (20) and evidence from animal research suggests PBDE exposure alters thyroid hormone homeostasis (18), as well as other thyroid- dependent processes. For example, research conducted in *Xenopus laevis* has demonstrated BDE-47 exposure arrests thyroid-dependent metamorphosis of tadpole into froglet and disrupts thyroid hormone-related gene expression in the brain (21, 22). Likewise, research conducted in avian and murine models has consistently demonstrated associations between prenatal exposure to PentaBDE congeners, including BDE-47, with reductions in circulating levels of T_4 (reviewed by: (18, 23)).

Widespread research supports classification of PBDEs as developmental neurotoxicants (24–27) (11, 24–26), with disruption of thyroid hormones as a leading putative mechanism underlying observed relationships (18, 28). However, despite convincing evidence from *in vitro* and animal research, results from studies investigating PBDEs in relation to thyroid hormone function in humans include a mix of negative, positive and null associations (29–34). Notably, previous studies have measured thyroid hormone parameters in maternal blood (29, 30, 34) collected during pregnancy or parturition, cord blood (29, 32, 34), or infant blood (31, 32) collected within hours to weeks of birth; periods when transient, yet substantial endocrine system changes occur, including profound alterations to the thyroid regulatory system (35). It is plausible that inconsistencies across previous human studies are partially attributable to misclassification introduced by the timing of thyroid hormone measurement. In the present study, we addressed this limitation by examining plasma PBDE concentrations in samples collected at birth and during toddler years (age 2-3) in relation to TSH, free T_4 and total T_4 levels measured in serum samples collected during early childhood (3-5 years). Based on results from animal research, we hypothesized prenatal exposure to BDE-47 would be associated with lower T_4 levels at birth (reviewed by: (18, 23)). We further hypothesize these associations will act through a programming pathway, leading to effects that persist throughout childhood. Owing to the typically higher exposure of young children compared to fetuses (via direct interaction with the external environment), we also hypothesize that children with low prenatal exposure, but high postnatal exposure will show evidence of a dysregulated thyroid regulatory system during childhood. This is the largest prospective study to examine both pre- and postnatal exposure to PBDEs in relation to thyroid endpoints.

Subjects and Methods

Study Sample

The sample includes a subset of participants enrolled in the Columbia Center for Children's Environmental Health (CCCEH) Mothers and Newborns birth cohort, which recruited African American and Dominican women from New York City between 1998 and 2006. Women were ineligible for study participation if they were outside the ages of 18-35 years, initiated prenatal care after the 20th week of pregnancy, had a multiple pregnancy, used tobacco products or illicit drugs, had diabetes, had hypertension, or were HIV positive. Women were considered fully enrolled if a maternal or umbilical cord blood sample was collected at the child's delivery. During pregnancy and at each postnatal study visit a bilingual research worker conducted a structured interview to collect information about sociodemographic and lifestyle factors. At delivery, study staff collected umbilical cord blood; at ages 2, 3, and 5-year follow-up visits a pediatric phlebotomist collected child venous blood. All samples were transported to the CCCEH laboratory immediately following collection, where the buffy coat, packed red blood cells, and plasma were separated and frozen at -70°C . Additional details describing the cohort design, recruitment and follow-up have been previously published (36–38).

At delivery, 727 mothers remained eligible and were fully enrolled in the cohort; at 2-year, 3-year and 5-year follow-up visits, 566 (78%), 562 (77%), and 551 (76%) maternal-child pairs remained in the study, respectively. At these postnatal follow-up visits, blood was collected from 92-98% of children. We measured PBDE concentrations in all available stored cord plasma samples and in early childhood samples among children with follow-up data (cord plasma $n=327$, 2-year $n=43$, 3-year $n=102$, 2-year and 3-year = 13). We measured thyroid hormone parameters in 185 of the 327 children with a measure of PBDE concentrations at age 3 years ($n=112$), 5 years ($n=35$) or both 3 and 5 years ($n=38$) (Figure 1). The study protocol was approved by the Institutional Review Board of Columbia University Medical Center. It was determined at the Centers for Disease Control and Prevention (CDC) that the agency was not engaged in human subjects' research. Before each study visit, mothers gave written informed consent for herself and for her child.

PBDE Analysis

The CDC's Persistent Organic Pollutants Biomonitoring Laboratory measured PBDE concentrations in umbilical cord and venous plasma samples. Detailed analytic methods are available elsewhere (39, 40) and information pertaining to analysis of PBDEs in this cohort has been previously described (9, 41). Briefly, samples were fortified with internal standards followed by automated liquid-liquid extraction using a Gilson 215 liquid handler (Gilson Inc., Middleton, WI). Final analytical determinations were made by gas chromatography isotope dilution high-resolution mass spectrometry using a DFS instrument (Thermo Fisher Scientific, Bremen, Germany). Each analytical batch was comprised of method blanks ($n=3$), quality control samples ($n=3$) and study samples ($n=26$). All reported data were subtracted from the median concentration detected in method blank samples. Co-extracted lipids were removed using a silica: silica/sulfuric acid column with automation on a Rapid Trace SPE work station (Biotage, Uppsala, Sweden) and total cholesterol and triglycerides

were determined on a Roche Hitachi 912 Chemistry Analyzer (GMI Inc, Ramsey, MN). We estimated total cord blood lipid levels, including unmeasured free cholesterol and phospholipids, by summation of individual lipid components using an umbilical cord blood-specific formula [total cord blood lipids = $2.657 \times$ total cord blood cholesterol + cord blood triglycerides + 0.268, in g lipids/L plasma] (A Sjodin, personal communication, November 2016). We estimated child total blood lipids using the short formula developed by Phillips et al. (42). We examined PBDEs as a lipid standardized variable in all models (ng/g lipid).

Thyroid Hormone Analysis

Thyroid stimulating hormone (TSH), freeT₄ and total T₄ were measured in child serum samples by the Clinical and Epidemiologic Research Laboratory at Boston Children's Hospital. All analyses were performed by automated immunoassay using a competitive electrochemiluminescence detection system (Roche Diagnostics, Indianapolis, IN). The lowest detection limits were 0.005 µIU/mL, 0.26 pmol/L, and 5.4 nmol/L for TSH, free T₄, and total T₄, respectively. Day-to-day imprecision values ranged from 1.8%-5.4% for 0.09-3.96 µIU/mL of TSH, 3.5%-6.6% for 8.75-50.70 pmol/L of free T₄, and 3.0%-6.9% for 33.4-237 nmol/L of total T₄. We measured maternal iodide concentrations, which is an essential substrate for thyroid hormone biosynthesis (43), in maternal spot urine samples collected during the third trimester. Before performing statistical analyses, we adjusted iodide for specific gravity to control for variation in urinary dilution.

Statistical Analysis

We focused on BDE-47 (percent detect: 80%), which was the only congener for which cord plasma concentrations were detectable in more than 50% of samples; concentrations in child plasma (age 2-3 years) were detectable in 99% of samples. Consistent with other studies, in our samples cord plasma BDE-47 concentrations were moderately to highly correlated with the other primary congeners that comprise the Penta-BDE formulation (Spearman's rho: BDE-99: 0.83, BDE-100: 0.76, BDE-153: 0.47; all $p < 0.01$). As previously described (41), we used a distribution-based approach to multiply impute values for plasma BDE-47 concentrations below the sample-specific limit of detection (LOD), which is determined by the sample's volume and lipid content.

We performed latent class growth analysis (LCGA) using the SAS Proc Traj procedure (44) to estimate trajectories of BDE-47 concentration between birth and 3 years. LCGA is a group-based modeling technique that empirically clusters individuals with a shared temporal pattern of change for a given characteristic (i.e. change in PBDE concentration over early life) (45). Before estimating trajectories, we log₁₀-transformed continuous BDE-47 concentrations (ng/g lipid) to better estimate a normal distribution and replaced non-detected concentrations with the sample-specific mean value across the 10 imputed datasets. We iteratively tested models with varying numbers of groups (2-5) and shapes (linear-cubic) and determined the optimal number of trajectories based on: 1) visual confirmation of distinct trajectories, each of which comprised >10% of the data, 2) evaluation of the Bayesian Information Criterion, and 3) evaluation of the average posterior probability of group membership. Additional details describing LCGA model fitting are provided in the Supplemental Material (Table S1).

We used multivariable linear regression to examine associations between trajectories of BDE-47, treated as a categorical variable, and thyroid hormone parameters collected between 3 years and 5 years. We used the generalized estimating equations (GEE) approach with an exchangeable working correlation to account for repeated thyroid measures within a child over time. We selected an exchangeable working correlation based on evaluation of the empirical correlation matrix, which did not indicate an autoregressive relationship, as well as evaluation of the quasi-likelihood information criterion (QIC). In all models, we expressed TSH, free T₄, and total T₄ as continuous variables and log₁₀-transformed TSH to better approximate a normal distribution. Models including an interaction term between age and BDE-47 trajectory did not indicate that the association between PBDEs and thyroid parameters significantly varies by age at blood collection. We further examined separate models for thyroid hormone parameters measured at ages 3 and 5 years in sensitivity analyses.

Intra-individual thyroid hormone concentrations decrease with age (46); therefore, we *a priori* included exact age at blood draw as a time-varying covariate. We constructed Directed Acyclic Graphs (DAG) based on substantive knowledge and previously published research to identify the minimal set of covariates sufficient to estimate the unconfounded effect of PBDEs on thyroid hormone parameters, which included only race/ethnicity (Figure S2). The set of potential confounders we considered included: sex, race/ethnicity (African American/Dominican), date of birth, gestational age (in weeks), birth weight (in grams), prenatal environmental tobacco smoke exposure (yes/no as previously described (47)), breastfeeding history (<12 weeks/ ≥12 weeks), parity (nulliparous/multiparous), relationship status (unmarried/married or with the same partner for 7 or more years), maternal age (in years), material hardship (none/unable to afford food, clothing or housing), maternal education (less than high school/high school or equivalent), and maternal employment (employed/not employed). All variables relating to the mother or household were collected during the prenatal period and variables relating to the delivery were extracted from hospital medical records. Information on breastfeeding history was collected at 3-, 6-, 12-, 24- and 36-month follow-up visits. In sensitivity analyses, we further evaluated the influence of covariate selection by examining *a priori* (age at blood draw-only) and fully-adjusted models.

Given sex differences in the incidence of many thyroid-related diseases (48, 49), we explored potential effect modification by child sex using cross product terms and sex-stratified models. We examined the influence of maternal iodide status during pregnancy by stratifying participants by the pregnancy-specific threshold for population iodine sufficiency (< 150 µg/L) and examining models within each stratum (50). Finally, to compare the results with findings from other cohort studies, we used the GEE approach to examine associations between plasma BDE-47 concentrations treated as a continuous, log₁₀-transformed variable measured in cord blood or age 3-year blood in relation to repeated thyroid hormone parameters. We performed statistical analyses using SAS v9.4 (SAS Institute) or RStudio v0.99.891 and constructed DAGs using DAGitty v2.3(51).

Results

Table 1 presents characteristics of the study population. All maternal-child pairs are African American (45%) or Dominican (55%) and at delivery 37% of mothers had less than a high school education, 76% were not in a stable relationship, and 36% reported experiencing material hardship. Sociodemographic and lifestyle characteristics were similarly distributed between children included in the analysis and those excluded due to missing PBDE or thyroid hormone data, with the following exceptions: the excluded sample had a higher proportion of Dominican participants (68% versus 55%, $p<0.01$), fewer nulliparous mothers (42% versus 51%, $p=0.04$), and on average newborns had lower birthweights (mean difference: 119 grams, $p<0.01$). The difference in birthweight may reflect racial/ethnic variation between the included and excluded samples, as African American newborns weighed on average 148 grams less than Dominican infants ($p<0.01$). Among included children, BDE-47 concentrations (birth, age 2, age 3) and thyroid hormone parameters (age 3, age 5) were not significantly associated with parity or birthweight. At ages 2 and 3 years, PBDE concentrations were significantly higher among children included in the analysis compared to children excluded ($p=0.05$), which likely reflects that included children were more likely to be born earlier during the enrollment period (i.e. prior to the 2004 phase-out of BDE-47) to allow time to age into later blood draws (9).

We detected BDE-47 in 80% of cord plasma samples and 99% of toddler plasma samples. The lower detection frequency in cord blood is consistent with results from an independent New York City-based cohort (BDE-47: 81%) and a Baltimore-based cohort (BDE-47: 90%), which both measured PBDE concentrations in cord blood (52, 53). As expected, plasma BDE-47 concentrations were significantly lower at birth (14.2 ± 0.9 , $n=327$) compared to toddler years (age 2: $GM\pm GSE$: 37.8 ± 5.8 , $n=56$, paired t-test using \log_{10} -transformed BDE-47 in ng/g lipids: $t=-4.07$, $p=0.0002$; age 3: $GM\pm GSE$: 32.0 ± 3.1 , $n=115$, paired t-test using \log_{10} -transformed BDE-47 in ng/g lipid: $t=-6.90$, $p<0.0001$). BDE-47 in cord plasma correlated poorly with BDE-47 in child plasma measured at age 2 years ($p=-0.03$, $p=0.82$) or 3-years ($p=0.09$, $p=0.36$), however, BDE-47 measured at age 2 years was strongly correlated with BDE-47 measured at age 3 years ($p=-0.79$, $p<0.01$). Exposure percentiles for each group are provided in Table S3. As illustrated by Figure 2a, the best fitting LCGA model revealed three trajectories of BDE-47 exposure characterized by 1) “persistent low” (34%), 2) “high-decreasing” (28%), and 3) “low-increasing” (38%) plasma concentrations across early childhood.

In models examining BDE-47 trajectories in relation to serum thyroid parameters measured between 3 and 5 years, children assigned to the ‘high-decreasing’ or ‘low-increasing’ trajectory had 17% (95% CI: -29, -2) and 19% (95% CI: -31, -5) lower geometric mean TSH levels compared to children assigned to the ‘persistent low’ trajectory, respectively. Associations between each of these trajectories and T_4 levels (free and total) were also inverse, however, they did not reach statistical significance at the $p=0.05$ level (Table 2, which presents estimates from GEE models, and Figure 2b, which plots adjusted mean thyroid parameter concentrations stratified by BDE-47 trajectory). Results from fully-adjusted models, as well as models examining thyroid hormones at ages 3 and 5 years separately did not substantially deviate from these results (see Supplemental Material,

Tables S4 and S5). Models examining cord plasma BDE-47 as a continuous variable are presented in Figure 3 and Table S6.

We observed no significant sex differences in the proportion of children assigned to each trajectory. Likewise, thyroid hormone parameters did not significantly differ between girls and boys at age 3 or 5 years (Supplemental Material, Table S7). In sex-stratified models (see Supplemental Material, Figure S8 and Table S9), the inverse association observed between the ‘low-increasing’ BDE-47 trajectory (versus ‘persistent low’ trajectory) and childhood thyroid parameters (TSH and free T₄) was augmented among boys (percent change TSH: -30, 95% CI: -45, -11, p-interaction: 0.12; unit change free T₄: -1.18, 95% CI: -2.18, -0.20, p-interaction: 0.21) and attenuated among girls (percent change TSH: -8, 95% CI: -26, 13.3; unit (nmol/L) change free T₄: -0.19, 95% CI: -1.19, 0.82). While the interaction terms did not reach statistical significance at the p=0.05 level, these findings suggest sex may modify the association between postnatal BDE-47 exposure and thyroid hormone parameters. Given our relatively small sample size for investigating interactions, it will be important that these findings are replicated by other research groups.

Specific-gravity adjusted urinary iodide concentrations among the 115 mothers with an available urine sample ranged from 45.4 to 425.9 µg/L; 27% of mothers had a concentration below the pregnancy-specific threshold for population iodine sufficiency (150 µg/L). In age, ethnicity and specific-gravity adjusted models, we detected no significant interaction between BDE-47 and maternal urinary iodine status, treated as a continuous or categorical variable (<150 v.s. 150 µg/L) for any thyroid hormone parameter.

Discussion

Compared to children with low cord plasma BDE-47 concentrations (GM±GSD: 5.8±0.4 ng/g lipid) that remained low throughout early childhood (GM±GSD: 13.8±1.2 ng/g lipid), children with high prenatal exposure (GM±GSD: 66.6±6.1 ng/g lipid) that decreased after birth had significantly lower circulating TSH levels measured between the ages of 3 and 5 years. TSH is a key effector and stimulus of the hypothalamic-pituitary-thyroid (HPT) axis, which maintains circulating thyroid hormone levels around an intra-individual set point. Briefly, low levels of circulating T₃ and T₄ stimulate the pituitary gland to release TSH, which in turn stimulates the thyroid gland to produce and secrete T₃ and T₄ (43, 54). Evidence from animal models and human clinical studies suggests the set point around which this negative feedback mechanism responds may be partially determined during gestation (55–58). Our findings suggest prenatal exposure to PBDEs may program a ‘reactive HPT axis’ phenotype such that less TSH is required to stimulate production and release of adequate T₄.

Children with low cord plasma BDE-47 concentrations (GM±GSD: 13.8±1.2 ng/g lipid) that increased during toddler years (GM±GSD: 106.9±9.2 ng/g lipid) also had significantly lower TSH concentrations between age 3-5 years compared to children with low cord plasma BDE-47 concentrations (GM±GSD: 5.8±0.4 ng/g lipid) that remained low during toddler years (GM±GSD: 13.8±1.3 ng/g lipid). Interestingly, the magnitude of this association was stronger among boys, despite our finding of no significant difference in BDE-47

concentrations or TSH levels between girls and boys. Boys with high exposure during toddler years also showed significantly lower free and total T₄ levels compared to boys with persistent low BDE-47 exposure. Our finding of depressed T₄ levels is consistent with research conducted on murine models, which have consistently found PBDE exposure to be associated with reduced serum T₄ levels (59). Putative mechanisms underlying this finding include PBDE interference with thyroid hormone transport and metabolism. For example, research conducted in mice suggests PBDEs induce upregulation of thyroid hormone metabolizing enzymes, resulting in enhanced clearance of T₄ (60) and both animal and *in vitro* studies suggest PBDEs or their hydroxylated metabolites may bind and displace T₄ from protein transporters thereby disrupting circulating levels (18).

Several birth cohort studies (n>100) have investigated cross sectional associations between prenatal exposure to PBDEs and thyroid hormone parameters measured in maternal or cord blood collected during pregnancy, delivery, or early postnatal life with a combination of positive, negative and null findings (Figure 3, Table S10) (29–34). Results from these studies are inconsistent and difficult to compare to our findings given variation in the measurement of thyroid hormones during developmental periods when normal fluctuations in HPT axis homeostasis occur. For example, an estrogen-induced elevation of thyroid binding globulin and placental production of chorionic gonadotrophin triggers maternal T₄ levels to increase sharply and TSH levels to fall during the first trimester of pregnancy, and during delivery a stress and cold-evoked surge in TSH occurs in the newborn, followed by a reflexive increase in T₄ over the next 24-48 hours (35, 43, 61).

Similar to studies focused on pregnancy and infancy, results from research investigating cross-sectional associations between PBDEs and thyroid dysregulation during childhood are inconsistent and difficult to compare due to differences in study design and variation in both the distribution of PBDE concentrations and congeners detected. Specifically, of the eight studies we identified examining postnatal exposure to PBDEs and thyroid parameters, three focused on special populations with unusually high exposure levels (i.e. children living and working near electronics recycling facilities in China) (62–64), two detected unusually low PBDE concentrations for unexplained reasons (65, 66), and one was conducted among a small sample (n<30) of older children (ages 14-18 years) (67). Among 80 children admitted to a hospital for a non-endocrine related disease between the ages of 1 and 5 years, Jacobson et al. detected a significant positive association between BDE-47 (ng/g lipid) and TSH, but no significant associations with total or free T₄ or T₃, reverse T₃, or T₃ uptake (68). A number of factors could underlie the differences in our findings, including study design (cross sectional versus prospective), variation in age at time of thyroid parameter measurement (1-5 years versus 3-5 years), or differences in the source populations (i.e. general population recruitment versus hospital-based recruitment).

Only one other study has prospectively examined associations between both prenatal and postnatal BDE-47 concentrations and thyroid hormone levels measured during early childhood. Using multiple informant models, Vuong et al. detected significant inverse associations between maternal log₁₀-BDE-47 concentrations measured during pregnancy and ln-TSH measured at age 3 years (β : -0.20, 95% CI: -0.38, -0.03) among 158 maternal-child pairs living in Cincinnati, Ohio (69). Also consistent with our finding of a stronger

association between the early prenatal high BDE-47 trajectory and TSH among girls, in sex-stratified models, albeit not significantly different from boys, Vuong et al. found that inverse associations between prenatal BDE-47 and ln-TSH were only statistically significant among girls. While we did not detect evidence of a sex-specific effect between prenatal BDE-47 and free T₄, Vuong et al. found a significant inverse association only among boys (69). Additionally, in contrast to our observation of inverse associations between postnatal BDE-47 and ln-TSH, the researchers detected significant positive associations between serum BDE-47 measured at age 2 years (n=71) (but not 1 (n=77) or 3 (n=71) years), and both ln-TSH and total T₄ measured at age 3 years (69); results of sex interactions with postnatal exposure were not reported.

Despite evidence indicating: 1) sex-specific associations between PBDEs and thyroid gland function in birds (70), 2), interactions with sex hormone receptors in fish and rodents (71–73), and 3) altered sex hormone levels in children (74) and pregnant women (75), few studies have investigated sexually-dimorphic effects of PBDEs on thyroid hormone disruption. Specific mechanisms underlying our observation of stronger associations with postnatal exposure among boys compared to girls are unknown, however, the high degree of overlap between the HPT axis and the hypothalamic-pituitary-gonadal axis, which regulates circulating sex hormone levels, suggests disruption in one system may have downstream consequences for the other (76). For example, evidence suggests hypogonadism in hypothyroid men reflects a hypothyroidism-induced blunted pituitary response to gonadotropin-releasing hormone secreted by the hypothalamus (77). Further, male reproductive organs, including the pre-pubertal testis, are thyroid-responsive tissues (78) and animal studies have demonstrated that experimentally induced hypothyroidism results in testicular damage, decreased testosterone concentrations, and arrest of sexual maturity (79, 80). Research designed to investigate the effect of PBDEs on overlapping pathways central to both thyroid and reproductive hormone homeostasis is needed to more fully understand the mechanisms underlying our sex-specific findings.

In addition to the longitudinal design, the present study has several strengths. First, PBDE concentrations were comparable to other geographically and temporally similar birth cohorts and reflect general population exposure (52, 53). Additionally, our relatively large sample size allowed us to examine many potential confounders, as well as explore effect modification by child sex and maternal iodide status during pregnancy. Unfortunately, we were not able to evaluate selenium, which is known to be an important determinant of thyroid status (81). Additional limitations include our lack of T₃ levels, thyroid binding protein levels, and PBDE metabolite data, which structurally resemble endogenous thyroid hormones more closely than parent congeners (18). Finally, we analyzed thyroid parameters by immunoassay, which may be affected by variation in serum thyroid binding protein levels (43).

Overall, our findings suggest the thyroid regulatory system may be sensitive to disruption by PBDEs during both the prenatal and postnatal period. Pregnant women and young children should minimize exposure to these endocrine disrupting chemicals. While research on PBDE exposure intervention studies is limited, results from observational research suggest several behavioral modifications for reducing contact with dust may be effective in limiting

PBDE exposure, including wet or damp mopping the home (9, 41), wiping down plastic toys (82), vacuuming with a HEPA filter and/or wearing a dust mask while vacuuming, washing hands frequently (83), avoiding hand to mouth behaviors (82) (i.e. thumb sucking, nail biting), and purchasing flame retardant free furniture and furnishings, which can be readily identified in the United States by examining product tags.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments:

We thank Dr. Lori Hoepner for assistance with data management and Pat Vuguin for review of manuscript drafts.

Funding: This work was supported by National Institutes of Health [grant numbers R01ES021806, R01ES013543, P50ES009600, T32ES023772 and T32ES007322]; and Environmental Protection Agency FP-91779001.

References

1. Trasande L, Zoeller RT, Hass U, Kortenkamp A, Grandjean P, Myers JP, DiGangi J, Bellanger M, Hauser R, Legler J, Skakkebaek NE, Heindel JJ 2015 Estimating burden and disease costs of exposure to endocrine-disrupting chemicals in the European union. *J Clinical Endocrinol Metab* 100:1245–1255. [PubMed: 25742516]
2. Zoeller RT, Brown TR, Doan LL, Gore AC, Skakkebaek NE, Soto AM, Woodruff TJ, Vom Saal FS 2012 Endocrine-disrupting chemicals and public health protection: a statement of principles from The Endocrine Society. *Endocrinology* 153:4097–4110. [PubMed: 22733974]
3. Bergman A, Heindel JJ, Jobling S, Kidd KA, Zoeller RT 2013 State of the Science of Endocrine Disrupting Chemicals 2012 United Nations Environment Programme and World Health Organization, Geneva, Switzerland.
4. Callahan P, Roe S, Hawthorne M 2012 Playing with Fire Chicago Tribune, Chicago, IL.
5. EPA 2010 An Exposure Assessment of Polybrominated Diphenyl Ethers. National Center for Environmental Assessment Environmental Protection Agency, Washington, DC.
6. Talsness CE 2008 Overview of toxicological aspects of polybrominated diphenyl ethers: a flame-retardant additive in several consumer products. *Environmental Research* 108:158–167. [PubMed: 18949835]
7. Cobb, D; Analysis of flame retardant chemicals added to foams, fabric, batting, loose fill and barriers.. Memorandum to Dale R Ray, Project Manager, Upholstered Furniture, Consumer Products Safety Commission. 2005.
8. Zhang X, Diamond ML, Robson M, Harrad S 2011 Sources, emissions, and fate of polybrominated diphenyl ethers and polychlorinated biphenyls indoors in Toronto, Canada. *Environmental Science & Technology* 45:3268–3274. [PubMed: 21413794]
9. Cowell WJ, Sjodin A, Jones R, Wang Y, Wang S, Herbstman JB 2018 Temporal trends and developmental patterns of plasma polybrominated diphenyl ether concentrations over a 15-year period between 1998 and 2013 *J Expo Sci Environ Epidemiol* 29(1):49–60. [PubMed: 29618764]
10. Toms LM, Sjodin A, Harden F, Hobson P, Jones R, Edenfield E, Mueller JF 2009 Serum polybrominated diphenyl ether (PBDE) levels are higher in children (2-5 years of age) than in infants and adults. *Environmental Health Perspectives* 117:1461–1465. [PubMed: 19750114]
11. Vuong AM, Braun JM, Yolton K, Xie C, Webster GM, Sjodin A, Dietrich KN, Lanphear BP, Chen A 2017 Prenatal and postnatal polybrominated diphenyl ether exposure and visual spatial abilities in children. *Environmental Research* 153:83–92. [PubMed: 27915227]
12. Geyer H, Schramm K, Darnerud P, Aune M, Feicht E, Fried K 2004 Terminal elimination half-lives of the brominated flame retardants TBBPA, HBCD, and lower brominated PBDEs in humans. *Organohalogen Comp* 66:5.

13. Adgent MA, Hoffman K, Goldman BD, Sjodin A, Daniels JL 2014 Brominated flame retardants in breast milk and behavioural and cognitive development at 36 months. *Paediatric and Perinatal Epidemiology* 28:48–57. [PubMed: 24313667]
14. Leonetti C, Butt CM, Hoffman K, Hammel SC, Miranda ML, Stapleton HM 2016 Brominated flame retardants in placental tissues: associations with infant sex and thyroid hormone endpoints. *Environmental Health* 15:113. [PubMed: 27884139]
15. Abbasi G, Buser AM, Soehl A, Murray MW, Diamond ML 2015 Stocks and flows of PBDEs in products from use to waste in the U.S. and Canada from 1970 to 2020. *Environmental Science & Technology* 49:1521–1528. [PubMed: 25548829]
16. Fromme H, Becher G, Hilger B, Volkel W 2016 Brominated flame retardants - exposure and risk assessment for the general population. *Int J Hyg Environ Health* 219:1–23. [PubMed: 26412400]
17. Mary EG, Zoeller RT 2010 Thyroid Hormones' Impact on the Developing Brain: Possible Mechanisms of Neurotoxicity. In: Harry GJ, Tilson HA (eds) *Neurotoxicology*, 3rd edition. Informa Healthcare, New York, NY, pp 79–111.rd
18. Costa LG, de Laat R, Tagliaferri S, Pellacani C 2014 A mechanistic view of polybrominated diphenyl ether (PBDE) developmental neurotoxicity. *Toxicology Letters* 230:282–294. [PubMed: 24270005]
19. Williams GR 2008 Neurodevelopmental and neurophysiological actions of thyroid hormone. *Journal of Neuroendocrinology* 20:784–794. [PubMed: 18601701]
20. Kitamura S, Shinohara S, Iwase E, Sugihara K, Uramaru N, Shigematsu H, Fujimoto N, Ohta S 2008 Affinity for thyroid hormone and estrogen receptors of hydroxylated polybrominated diphenyl ethers. *J Health Sci* 54(5).
21. Balch GC, Velez-Espino LA, Sweet C, Alaee M, Metcalfe CD 2006 Inhibition of metamorphosis in tadpoles of *Xenopus laevis* exposed to polybrominated diphenyl ethers (PBDEs). *Chemosphere* 64:328–338. [PubMed: 16455129]
22. Yost AT, Thornton LM, Venables BJ, Sellin Jeffries MK 2016 Dietary exposure to polybrominated diphenyl ether 47 (BDE-47) inhibits development and alters thyroid hormone-related gene expression in the brain of *Xenopus laevis* tadpoles. *Environmental toxicology and pharmacology* 48:237–244. [PubMed: 27838513]
23. Birnbaum LS, Staskal DF 2004 Brominated flame retardants: cause for concern? *Environmental Health Perspectives* 112(1):9–17. [PubMed: 14698924]
24. Lam J, Lanphear BP, Bellinger DC, Axelrad DA, McPartland J, Sutton P, Davidson L, Daniels N, Sen S, Woodruff TJ 2017 Developmental PBDE exposure and IQ/ADHD in childhood: a systematic review and meta-analysis. *Environmental Health Perspectives* 125(8).
25. Linares V, Belles M, Domingo JL 2015 Human exposure to PBDE and critical evaluation of health hazards. *Arch Toxicol* 89:335–356. [PubMed: 25637414]
26. Roth N, Wilks MF 2014 Neurodevelopmental and neurobehavioural effects of polybrominated and perfluorinated chemicals: a systematic review of the epidemiological literature using a quality assessment scheme. *Toxicology Letters* 230:271–281. [PubMed: 24583043]
27. Vuong AM, Yolton K, Dietrich KN, Braun JM, Lanphear BP, Chen A 2017 Exposure to polybrominated diphenyl ethers (PBDEs) and child behavior: Current findings and future directions. *Hormones and Behavior*
28. Mughal B, Fini J, Demeneix B 2018 Thyroid-disrupting chemicals and brain development: an update. *Endocr Connect* 7:R160–R186. [PubMed: 29572405]
29. Abdelouahab N, Langlois MF, Lavoie L, Corbin F, Pasquier JC, Takser L 2013 Maternal and cord-blood thyroid hormone levels and exposure to polybrominated diphenyl ethers and polychlorinated biphenyls during early pregnancy. *American Journal of Epidemiology* 178:701–713. [PubMed: 23924579]
30. Chevrier J, Harley KG, Bradman A, Gharbi M, Sjodin A, Eskenazi B 2010 Polybrominated diphenyl ether (PBDE) flame retardants and thyroid hormone during pregnancy. *Environmental Health Perspectives* 118(10):1444–1449. [PubMed: 20562054]
31. Chevrier J, Harley KG, Bradman A, Sjodin A, Eskenazi B 2011 Prenatal exposure to polybrominated diphenyl ether flame retardants and neonatal thyroid-stimulating hormone levels

- in the CHAMACOS study. *American Journal of Epidemiology* 174:1166–1174. [PubMed: 21984658]
32. Herbstman JB, Sjodin A, Apelberg BJ, Witter FR, Halden RU, Patterson DG, Panny SR, Needham LL, Goldman LR 2008 Birth delivery mode modifies the associations between prenatal polychlorinated biphenyl (PCB) and polybrominated diphenyl ether (PBDE) and neonatal thyroid hormone levels. *Environmental Health Perspectives* 116(10):1376–1382. [PubMed: 18941581]
 33. Stapleton HM, Eagle S, Anthopolos R, Wolkin A, Miranda ML 2011 Associations between polybrominated diphenyl ether (PBDE) flame retardants, phenolic metabolites, and thyroid hormones during pregnancy. *Environmental Health Perspectives* 119(10):1454–1459. [PubMed: 21715241]
 34. Vuong AM, Webster GM, Romano ME, Braun JM, Zoeller RT, Hoofnagle AN, Sjodin A, Yolton K, Lanphear BP, Chen A 2015 Maternal Polybrominated Diphenyl Ether (PBDE) Exposure and Thyroid Hormones in Maternal and Cord Sera: The HOME Study, Cincinnati, USA. *Environmental Health Perspectives* 123(10):1079–1085. [PubMed: 25893858]
 35. Glinoe D 1997 The regulation of thyroid function in pregnancy: pathways of endocrine adaptation from physiology to pathology. *Endocr Rev* 18:404–433. [PubMed: 9183570]
 36. Perera FP, Rauh V, Whyatt RM, Tsai WY, Tang D, Diaz D, Hoepner L, Barr D, Tu YH, Camann D, Kinney P 2006 Effect of prenatal exposure to airborne polycyclic aromatic hydrocarbons on neurodevelopment in the first 3 years of life among inner-city children. *Environmental Health Perspectives* 114(8):1287–1292. [PubMed: 16882541]
 37. Rauh VA, Garfinkel R, Perera FP, Andrews HF, Hoepner L, Barr DB, Whitehead R, Tang D, Whyatt RW 2006 Impact of prenatal chlorpyrifos exposure on neurodevelopment in the first 3 years of life among inner-city children. *Pediatrics* 118:e1845–1859. [PubMed: 17116700]
 38. Whyatt RM, Camann DE, Kinney PL, Reyes A, Ramirez J, Dietrich J, Diaz D, Holmes D, Perera FP 2002 Residential pesticide use during pregnancy among a cohort of urban minority women. *Environmental Health Perspectives* 110(5):507–514. [PubMed: 12003754]
 39. Jones R, Edenfield E, Anderson S, Zhang Y, Sjodin A 2012 Semi-automated extraction and cleanup method for measuring persistent organic pollutants in human serum. *Organohalogen Comp* 74:97–98.
 40. Sjodin A, Jones RS, Lapeza CR, Focant JF, McGahee EE, Patterson DG, 2004 Semiautomated high-throughput extraction and cleanup method for the measurement of polybrominated diphenyl ethers, polybrominated biphenyls, and polychlorinated biphenyls in human serum. *Analytical Chemistry* 76:1921–1927. [PubMed: 15053652]
 41. Cowell WJ, Sjodin A, Jones R, Wang Y, Wang S, Herbstman JB 2018 Determinants of prenatal exposure to polybrominated diphenyl ethers (PBDEs) among urban, minority infants born between 1998–2006. *Environmental Pollution* 223:774–781.
 42. Phillips DL, Pirkle JL, Burse VW, Bernert JT Jr., Henderson LO, Needham 1989 Chlorinated hydrocarbon levels in human serum: effects of fasting and feeding. *Arch Environ Contam Toxicol* 18:495–500. [PubMed: 2505694]
 43. Werner & Ingbar's the thyroid: a fundamental and clinical text 2013 10th ed. Wolters Kluwer, Lippincott Williams & Wilkins Health, Philadelphia, PA.
 44. Jones B, Nagin D, KA R 2001 A SAS procedure based on mixture models for estimating developmental trajectories. *Sociological Methodology Research* 29:374–393.
 45. Nagin D 2005 Group-based modeling of development Harvard University Press, Cambridge, MA.
 46. Elmlinger MW, Kuhnel W, Lambrecht HG, Ranke MB 2001 Reference intervals from birth to adulthood for serum thyroxine (T4), triiodothyronine (T3), free T3, free T4, thyroxine binding globulin (TBG) and thyrotropin (TSH). *Clinical chemistry and laboratory medicine* 39:973–979.
 47. Rauh VA, Whyatt RM, Garfinkel R, Andrews H, Hoepner L, Reyes A, Diaz D, Camann D, Perera FP 2004 Developmental effects of exposure to environmental tobacco smoke and material hardship among inner-city children. *Neurotoxicology and Teratology* 26:373–385. [PubMed: 15113599]
 48. Hollowell JG, Staehling NW, Flanders WD, Hannon WH, Gunter EW, Spencer CA, Braverman LE 2002 Serum TSH, T(4), and thyroid antibodies in the United States population (1988 to 1994): National Health and Nutrition Examination Survey (NHANES III). *J Clin Endocrinol Metab* 87:489–499.

49. Vanderpump MP, Tunbridge WM, French JM, Appleton D, Bates D, Clark F, Grimley Evans J, Hasan DM, Rodgers H, Tunbridge F, et al. 1995 The incidence of thyroid disorders in the community: a twenty-year follow-up of the Whickham Survey. *Clin Endocrinol* 43:55–68.
50. World Health Organization 2007 Assessment of the iodine deficiency disorders and monitoring their elimination Geneva, Switzerland.
51. Textor J, Hardt J, Knuppel S 2011 DAGitty: a graphical tool for analyzing causal diagrams. *Epidemiology* 22:745.
52. Herbstman JB, Sjodin A, Apelberg BJ, Witter FR, Patterson DG, Halden RU, Jones RS, Park A, Zhang Y, Heidler J, Needham LL, Goldman LR 2007 Determinants of prenatal exposure to polychlorinated biphenyls (PCBs) and polybrominated diphenyl ethers (PBDEs) in an urban population. *Environmental Health Perspectives* 115(12):1794–1800. [PubMed: 18087602]
53. Herbstman JB, Sjodin A, Kurzon M, Lederman SA, Jones RS, Rauh V, Needham LL, Tang D, Niedzwiecki M, Wang RY, Perera F 2010 Prenatal exposure to PBDEs and neurodevelopment. *Environmental Health Perspectives* 118(5):712–719. [PubMed: 20056561]
54. Andersen S, Pedersen KM, Bruun NH, Laurberg P 2002 Narrow individual variations in serum T(4) and T(3) in normal subjects: a clue to the understanding of subclinical thyroid disease. *J Clin Endocrin Metab* 87:1068–1072.
55. Azizi F, Vagenakis AG, Bollinger J, Reichlin S, Braverman LE, Ingbar SH 1974 Persistent abnormalities in pituitary function following neonatal thyrotoxicosis in the rat. *Endocrinology* 94:1681–1688. [PubMed: 4208653]
56. Bagattini B, Cosmo CD, Montanelli L, Piaggi P, Ciampi M, Agretti P, Marco GD, Vitti P, Tonacchera M 2014 The different requirement of L-T4 therapy in congenital athyreosis compared with adult-acquired hypothyroidism suggests a persisting thyroid hormone resistance at the hypothalamic-pituitary level. *European Journal of Endocrinology* 171:615–621. [PubMed: 25305309]
57. Cavaliere H, Medeiros-Neto GA, Rosner W, Kourides IA 1985 Persistent pituitary resistance to thyroid hormone in congenital versus later-onset hypothyroidism. *J Endocrinol Invest* 8:527–532. [PubMed: 3938790]
58. Walker P, Courtin F 1985 Transient neonatal hyperthyroidism results in hypothyroidism in the adult rat. *Endocrinology* 116:2246–2250. [PubMed: 3996311]
59. Zhou T, Ross DG, DeVito MJ, Crofton KM 2001 Effects of short-term in vivo exposure to polybrominated diphenyl ethers on thyroid hormones and hepatic enzyme activities in weanling rats. *Toxicological Sciences* 61:76–82. [PubMed: 11294977]
60. Szabo DT, Richardson VM, Ross DG, Diliberto JJ, Kodavanti PR, Birnbaum LS 2009 Effects of perinatal PBDE exposure on hepatic phase I, phase II, phase III, and deiodinase 1 gene expression involved in thyroid hormone metabolism in male rat pups. *Toxicological Sciences* 107:27–39. [PubMed: 18978342]
61. Fisher DA, Klein AH 1981 Thyroid development and disorders of thyroid function in the newborn. *NEJM* 304:702–712. [PubMed: 6258072]
62. Han G, Ding G, Lou X, Wang X, Han J, Shen H, Zhou Y, Du L 2011 Correlations of PCBs, DIOXIN, and PBDE with TSH in children's blood in areas of computer E-waste recycling. *Biomed Environ Sci* 24:112–116. [PubMed: 21565681]
63. Xu P, Lou X, Ding G, Shen H, Wu L, Chen Z, Han J, Han G, Wang X 2014 Association of PCB, PBDE and PCDD/F body burdens with hormone levels for children in an e- waste dismantling area of Zhejiang Province, China. *The Science of the Total Environment* 499:55–61. [PubMed: 25173862]
64. Xu X, Liu J, Zeng X, Lu F, Chen A, Huo X 2014 Elevated serum polybrominated diphenyl ethers and alteration of thyroid hormones in children from Guiyu, China. *PLoS One* 9:e113699. [PubMed: 25415336]
65. Gascon M, Vrijheid M, Martinez D, Forn J, Grimalt JO, Torrent M, Sunyer J 2011 Effects of pre and postnatal exposure to low levels of polybromodiphenyl ethers on neurodevelopment and thyroid hormone levels at 4 years of age. *Environment International* 37:605–611. [PubMed: 21237513]

66. Kicinski M, Viaene MK, Den Hond E, Schoeters G, Covaci A, Dirtu AC, Nelen V, Bruckers L, Croes K, Sioen I, Baeyens W, Van Larebeke N, Nawrot TS 2012 Neurobehavioral function and low-level exposure to brominated flame retardants in adolescents: a cross-sectional study. *Environmental Health* 11:86. [PubMed: 23151181]
67. Leijs MM, ten Tusscher GW, Olie K, van Teunenbroek T, van Aalderen WM, de Voogt P, Vulsma T, Bartonova A, Kraymer von Krauss M, Mosoiu C, Riojas-Rodriguez H, Calamandrei G, Koppe JG 2012 Thyroid hormone metabolism and environmental chemical exposure. *Environmental Health* 11 Suppl 1:S10. [PubMed: 22759492]
68. Jacobson MH, Barr DB, Marcus M, Muir AB, Lyles RH, Howards PP, Pardo L, Darrow LA 2016 Serum polybrominated diphenyl ether concentrations and thyroid function in young children. *Environmental Research* 149:222–230. [PubMed: 27228485]
69. Vuong AM, Braun JM, Webster GM, Thomas Zoeller R, Hoofnagle AN, Sjodin A, Yolton K, Lanphear BP, Chen A 2018 Polybrominated diphenyl ether (PBDE) exposures and thyroid hormones in children at age 3 years. *Environment International* 117:339–347. [PubMed: 29787984]
70. Fernie KJ, Marteinson SC 2016 Sex-specific changes in thyroid gland function and circulating thyroid hormones in nestling American kestrels (*Falco sparverius*) following embryonic exposure to polybrominated diphenyl ethers by maternal transfer. *Environ Toxicol Chem* 35:2084–2091. [PubMed: 26757407]
71. Lefevre PL, Wade M, Goodyer C, Hales BF, Robaire B 2016 A mixture reflecting polybrominated diphenyl ether (PBDE) profiles detected in human follicular fluid significantly affects steroidogenesis and induces oxidative stress in a female human granulosa cell line. *Endocrinology* 157:2698–2711. [PubMed: 27219277]
72. Noyes P, Stapleton H 2014 PBDE flame retardants: toxicokinetics and thyroid hormone endocrine disruption in fish. *Endocrine Disruptors* 2(1).
73. Sarkar D, Chowdhury JP, Singh SK 2016 Effect of polybrominated diphenyl ether (BDE-209) on testicular steroidogenesis and spermatogenesis through altered thyroid status in adult mice. *Gen Comp Endocrinol* 239:50–61. [PubMed: 26602377]
74. Eskenazi B, Rauch SA, Tenerelli R, Huen K, Holland NT, Lustig RH, Kogut K, Bradman A, Sjodin A, Harley KG 2017 In utero and childhood DDT, DDE, PBDE and PCBs exposure and sex hormones in adolescent boys: The CHAMACOS study. *Int J Hyg Environ Health* 220:364–372. [PubMed: 27876543]
75. Gao Y, Chen L, Wang C, Zhou Y, Wang Y, Zhang Y, Hu Y, Ji L, Shi R, Cui C, Ding G, Jin J, Tian Y 2016 Exposure to polybrominated diphenyl ethers and female reproductive function: A study in the production area of Shandong, China. *The Science of the Total Environment* 572:9–15. [PubMed: 27485910]
76. Krassas GE, Pontikides N 2004 Male reproductive function in relation with thyroid alterations. *Best Pract Res Clin Endocrinol Metab* 18:183–195. [PubMed: 15157835]
77. Velazquez EM, Bellabarba Arata G 1997 Effects of thyroid status on pituitary gonadotropin and testicular reserve in men. *Arch Androl* 38:85–92. [PubMed: 9017126]
78. Jannini EA, Ulisse S, D'Armiento M 1995 Thyroid hormone and male gonadal function. *Endocr Rev* 16:443–459. [PubMed: 8521789]
79. Asker ME, Hassan WA, El-Kashlan AM 2015 Experimentally induced hyperthyroidism influences oxidant and antioxidant status and impairs male gonadal functions in adult rats. *Andrologia* 47:644–654. [PubMed: 25220112]
80. Chandrasekhar Y, Holland MK, D'Occhio MJ, Setchell BP 1985 Spermatogenesis, seminal characteristics and reproductive hormone levels in mature rams with induced hypothyroidism and hyperthyroidism. *J Endocrinol* 105:39–46. [PubMed: 3921644]
81. Arthur JR, Beckett GJ, Mitchell JH 1999 The interactions between selenium and iodine deficiencies in man and animals. *Nutr Res Rev* 12:55–73. [PubMed: 19087446]
82. Hoffman K, Webster TF, Sjodin A, Stapleton HM 2017 Toddler's behavior and its impacts on exposure to polybrominated diphenyl ethers. *J Expo Sci Environ Epidemiol* 27:193–197. [PubMed: 26956938]

83. Watkins DJ, McClean MD, Fraser AJ, Weinberg J, Stapleton HM, Sjodin A, Webster TF 2011 Exposure to PBDEs in the office environment: evaluating the relationships between dust, handwipes, and serum. *Environmental Health Perspectives* 119(9):1247–1252. [PubMed: 21715243]

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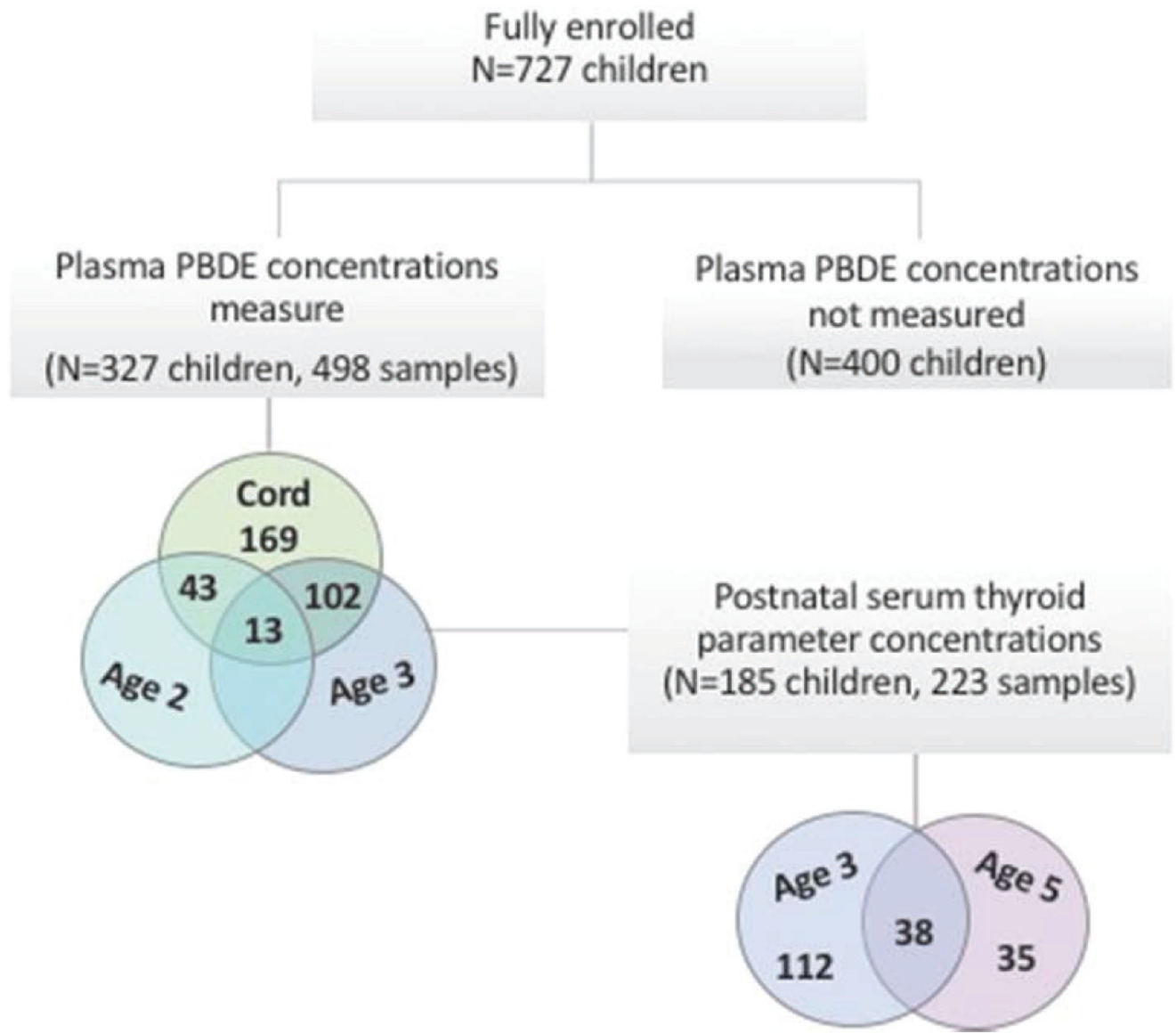


Figure 1. Flow diagram of participant selection from the Columbia Center for Children's Environmental Health Mothers and Newborns birth cohort study.

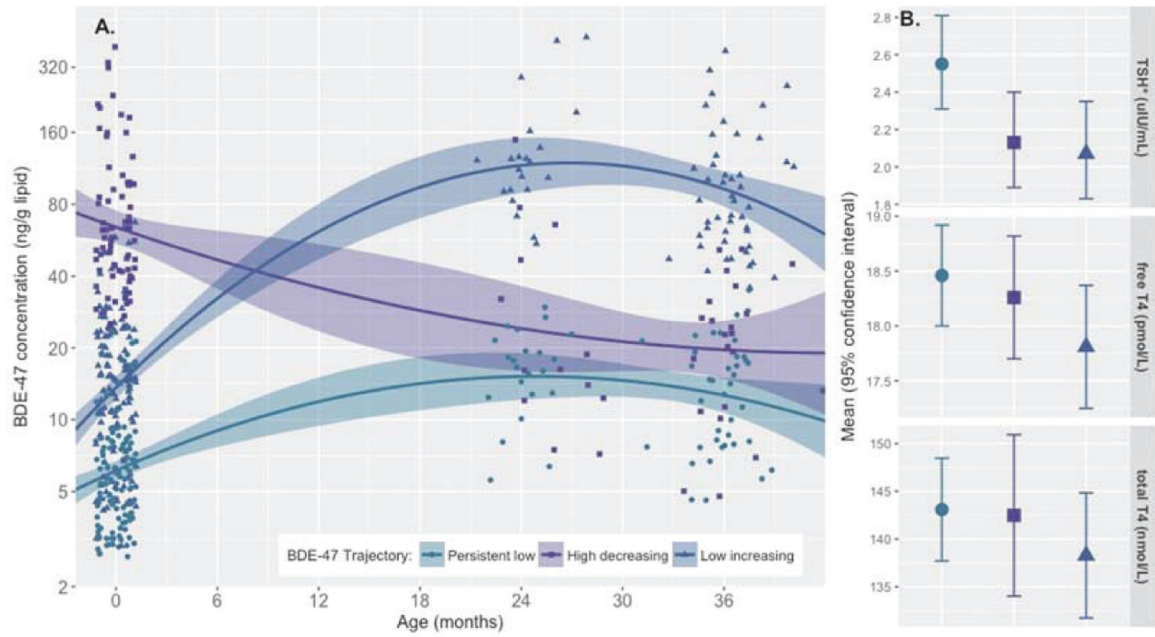


Figure 2.

A. Trajectories of BDE-47 concentration from birth through age 3-years. B. Age and ethnicity adjusted mean thyroid parameter levels by BDE-47 trajectory. *TSH is geometric mean.

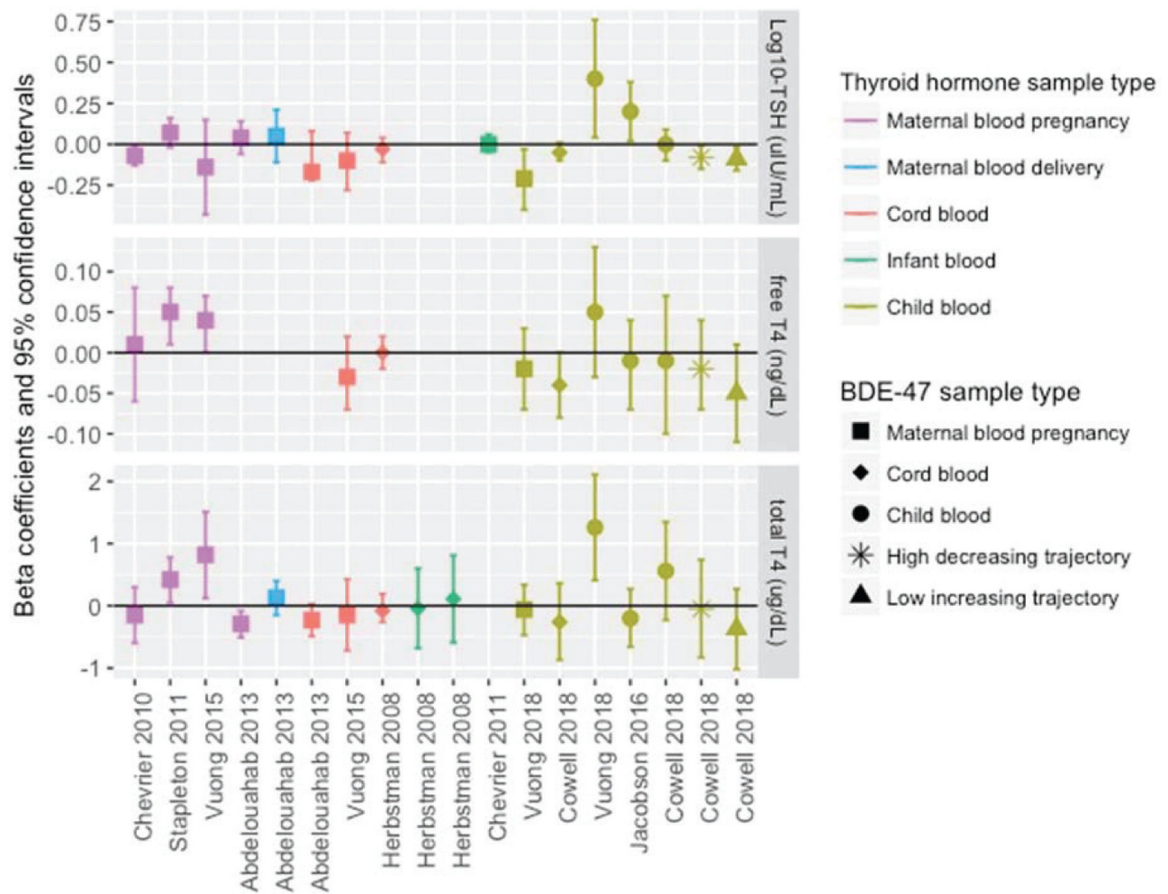


Figure 3.

Results from multiple linear regression models examining associations between \log_{10} BDE-47 (ng/g lipid) and thyroid hormone parameters (\log_{10} TSH: μ IU/mL; free T₄: ng/dL; total T₄: μ g/dL) measured as continuous variables reported by 7 North American birth cohort studies.

Herbstman 2008 and Stapleton 2011 applied a natural-log transformation to BDE-47 and TSH rather than a \log_{10} -transformation. Stapleton 2011 additionally natural-log transformed free T₄. To facilitate comparison of our results to others, we re-analyzed final models expressing free T₄ and total T₄ in units of ng/dL and μ g/dL, respectively. Abdelouahab 2013 modeled free T₄ measured in cord blood on a pmol/L basis, therefore, we excluded these results from the figure to accommodate the y-axis scale. Supplemental Material Table 8 presents summary data for BDE-47 and thyroid hormone parameters measured by each study.

Table 1.

Participant characteristics, BDE-47 concentrations and thyroid parameter levels among maternal-child pairs included in the analysis (n=185)

	N (%) or mean±SD			
	Included	N sample	Excluded	N sample
African American *	83 (45)	185	171 (32)	542
Dominican *	102 (55)	185	371 (68)	542
Nulliparous *	95 (51)	185	229 (42)	538
Maternal age (years)	24.9±4.8	185	25.3±5.0	542
< High school education	68 (37)	185	190 (36)	532
Stable relationship	44 (24)	185	150 (28)	538
Employed	117 (63)	185	282 (52)	539
Material hardship	66 (36)	185	220 (42)	530
Male	85 (46)	185	266 (49)	542
Prenatal ETS exposure	70 (38)	185	184 (34)	540
Gestational age (weeks)	39.3±1.2	185	38.5±6.3	542
Birthweight (kg) *	3.5±0.5	185	3.3±0.6	535
Breastfed 12 weeks	64 (35)	185	173 (34)	502
BDE-47 (ng/g lipid) ^a				
Prenatal	14.2±1.2	185	14.1±1.2	142
2 years *	43.9±7.4	45	20.4±6.2	11
3 years *	38.0±5.3	60	26.6±3.6	55
TSH (μIU/mL) ^a				
3 years	2.4±0.1	150	2.3±0.10	125
5 years	2.1±0.1	73	2.3±0.2	36
Free T ₄ (pmol/L)				
3 years	18.3±2.2	150	18.3±2.5	125
5 years	18.1±2.2	73	18.2±1.8	36
Total T ₄ (nmol/L)				
3 years	140.6±25.3	150	140.8±26.8	125
5 years	146.4±31.8	73	148.9±31.3	36

Notes:

* Included and excluded significantly different at p=0.05

Abbreviations: BDE, brominated diphenyl ether; ETS, environmental tobacco smoke; T₄, thyroxine; TSH, thyroid stimulating hormone

^a geometric mean

Table 2.

Age- and ethnicity-adjusted associations (β , 95% CI) between BDE-47 trajectories (ng/g lipid) and serum thyroid parameters; n=185 children and 223 observations.

BDE-47 trajectory	Log₁₀TSH (μIU/mL)	Free T₄ (pmol/L)	Total T₄ (nmol/L)
Persistent low (34%)	Reference	Reference	Reference
High decreasing (28%)	-0.08 (-0.15, -0.01)	-0.20 (-0.92, 0.52)	-0.61 (-10.66, 9.44)
Low increasing (38%)	-0.09 (-0.16, -0.02)	-0.65 (-1.37, 0.07)	-4.80 (-13.11, 3.51)

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