



HHS Public Access

Author manuscript

Free Radic Biol Med. Author manuscript; available in PMC 2019 August 20.

Published in final edited form as:

Free Radic Biol Med. 2018 August 20; 124: 484–492. doi:10.1016/j.freeradbiomed.2018.04.579.

Metal exposure and oxidative stress markers in pregnant Navajo Birth Cohort Study participants

Erica J. Dashner-Titus^{a,1}, Joseph. Hoover^{b,1}, Li Luo^{c,d}, Ji-Hyun Lee^{c,d}, Ruofei Du^{c,d}, Ke Jian Liu^a, Maret G Traber^e, Emily Ho^{e,f}, Johnnye Lewis^b, and Laurie G. Hudson^{a,*}

^aDepartment of Pharmaceutical Sciences, College of Pharmacy, University of New Mexico, 1 University of New Mexico, Albuquerque, NM

^bCommunity Environmental Health Program, College Of Pharmacy, University of New Mexico, 1 University of New Mexico, Albuquerque, NM 87131

^cBiostatistics Shared Resource, The UNM Comprehensive Cancer Center, Albuquerque, NM, 87131, USA

^dUNM METALS Biostatistics and Data Management (BDM) Core (Luo, Senior author for BDM team)

^eLinus Pauling Institute, Oregon State University, 307 Linus Pauling Science Center, Corvallis, OR 97331

^fMoore Family Center for Whole Grain Foods, Nutrition & Preventive Health, School of Biological & Population Health Sciences, College of Public Health & Human Sciences, 211 Milam Hall, Oregon State University, Corvallis, OR 97331

Abstract

Contamination of soil and water by waste from abandoned uranium mines has led to chronic exposures to metal mixtures in Native American communities. Our previous work demonstrated that community exposures to mine waste increase the likelihood of developing cardiovascular disease, as well as the likelihood of developing multiple chronic diseases including diabetes, hypertension and kidney disease. Exposure to various environmental metals is associated with elevated oxidative stress, which is considered a contributor to these and other chronic disease states. The purpose of the current research was to assess potential associations between exposure to uranium and arsenic and evidence for increased oxidative stress as measured by urinary F₂-isoprostanes in pregnant women enrolled in the Navajo Birth Cohort Study. The current study also included an analysis of zinc as a potential mediator of oxidative stress in the study population. Urinary arsenic and uranium, serum zinc and urinary F₂-isoprostanes were measured for each study participant at enrollment. Study participants were pregnant women with median age of 26.8; 18.9 % were enrolled in the 1st trimester, 44.7% were enrolled in the 2nd trimester, and 36.4%

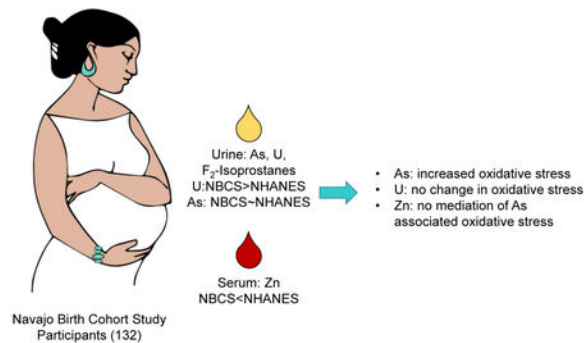
*Corresponding author.

¹Authors contributed equally to the work and are listed alphabetically

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

were enrolled in the 3rd trimester. Median urinary metal levels were 5.5 and 0.016 $\mu\text{g/g}$ creatinine for arsenic and uranium, respectively. Multivariable regression analysis indicated a significant association between arsenic exposure and the lipid peroxidation product 8-iso-prostaglandin $\text{F}_{2\alpha}$, controlling for zinc and trimester. No associations were detected with uranium despite evidence that levels were in the Navajo Birth Cohort samples were 2.3 times the median reported for women in the National Health and Nutrition Examination Survey (2011-12). Zinc was not found to have any causal mediation of the effects of the other metals on oxidative stress. The current work is consistent with other studies that have detected an association between arsenic and elevated oxidative stress. In contrast to arsenic, uranium did not appear to increase oxidative stress response in this study population. These findings are relevant to assessing the potential human impact of chronic exposure to mixed metal waste from abandoned uranium mines.

Graphical abstract



Keywords

Arsenic; uranium; zinc; oxidative stress; isoprostanes; Navajo Birth Cohort Study; AI/AN

INTRODUCTION

There are more than 4,000 abandoned uranium mines (AUMs⁹) in the Western United States [1] with more than 500 located on the Navajo Nation [2, 3]. These abandoned mine sites are often located in close proximity to Native American communities leading to community concerns about health impacts due to mine waste exposures. People may be exposed to AUM waste containing uranium, arsenic, and other co-occurring metals via air, soil or ground water [4-6]. On the Navajo Nation for example, more than 30% of the population lacks access to regulated public drinking water and must rely on unregulated water supplies or other sources for drinking water. A previous analysis of more than 500 unregulated water sources indicated that 15.1% and 12.8% of these unregulated sources exceeded national drinking water maximum contaminant levels (MCLs) for arsenic and uranium, respectively [7], providing potential exposure sources for arsenic and/or uranium. Navajo community members have expressed particular concern about uranium exposures [9]. Exposure to metals found in abandoned mine waste is associated with chronic disease occurrence among the Navajo and other Native American tribes. A survey of over 1300 people living on the Navajo Nation discovered that Navajo community members with ongoing community-level

exposures to uranium mine waste have an increased likelihood of several chronic conditions such as cardiovascular disease, diabetes, and kidney disease [8, 9]. These findings suggest that chronic exposures to AUM waste affect human health.

Numerous chronic diseases, including cardiovascular and renal disease, are associated with elevated oxidative damage due to excess generation of reactive oxygen species [10-13]. Experimental studies in cells and animal models link both arsenic [14-18] and uranium [19-23] to generation of oxidative stress. In contrast, studies to investigate the association between arsenic exposure and biomarkers of oxidative stress in human populations have not yielded consistent findings [24-34] due in part to different study populations and detection methods for oxidative stress. The lack of studies on uranium exposure and direct biomarkers of oxidative stress is a gap in knowledge that limits our ability to understand the relationships between different metal exposures, oxidative stress and associated diseases in humans.

An ongoing cohort study on the Navajo Nation, the Navajo Birth Cohort Study (NBCS) [35], is examining the effects of uranium and other metal/metalloid exposures on birth outcomes and development. Currently there is limited information about the association between metal exposure and oxidative stress during pregnancy. In the NBCS, biomonitoring results indicate urine uranium concentrations exceed those in the US population, while serum zinc, a metal with antioxidant properties, is frequently below World Health Organization (WHO) recommended sufficiency concentrations [3]. This zinc deficiency has been observed in pregnant Navajo women for more than 35 years [36], but also occurs in men in the population. Therefore, samples from this study provide an opportunity to understand the effect of exposures to known metal inducers of oxidative damage on measures of oxidative stress, and test the hypothesis that zinc may have a potential modifying role in those responses.

Based on evidence from experimental models that arsenic and uranium exposures and zinc deficiency increase oxidative stress [14-23, 37-40], the goal of the current study was to investigate the association between metal exposure and oxidative stress biomarkers, and determine whether serum zinc moderates the effects of these exposures. The lipid peroxidation product 8-iso-prostaglandin $F_{2\alpha}$ (8-iso-PGF $_{2\alpha}$) was used as a urinary oxidative stress biomarker. Because 8-iso-PGF $_{2\alpha}$ is also generated through an enzymatic pathway by prostaglandin-endoperoxide synthases, the ratio of 8-iso-PGF $_{2\alpha}$ to prostaglandin $F_{2\alpha}$ (PGF $_{2\alpha}$) has been established to distinguish enzymatic versus chemical lipid peroxidation as a biomarker of oxidative stress and was included in this study [41, 42]. We measured urinary arsenic and uranium, serum zinc and urinary F_2 -isoprostanes for 132 participants from the parent NBCS study. This group was then stratified by serum zinc concentrations (low vs. high) while preserving the range of uranium and arsenic found in the population to allow assessment of the impact of zinc on metal-associated oxidative stress. Metal values in the study population were compared to national values obtained from the National Health and Nutrition Examination Survey (NHANES) [43]. The reported findings provide insights into oxidative stress and exposure to metals associated with AUMs.

MATERIALS AND METHODS

Study Overview

The study site, the Navajo Nation, is located in the Four Corners area of the southwestern United States. In 2013 the NBCS began recruiting pregnant women between 14 and 45 years of age who had lived on the Navajo Nation for at least 5 years, were willing to deliver at a participating hospital, and have their child followed up for one year postnatally. At the time of enrollment, a blood and urine sample was collected from the participant. Socioeconomic, demographic, and lifestyle information was also collected in the home shortly after enrollment. A subset of the maternal enrollment urine samples was selected for oxidative stress analysis, which forms the basis of this investigation.

Study Population

The NBCS was initiated to address Navajo community concerns about how chronic environmental exposure to uranium mine waste affects human health. The research team led by the University of New Mexico (UNM) Health Sciences Center Community Environmental Health Program included partnerships with Navajo Nation Department of Health, Navajo Area Indian Health Service, Southwest Research Information Center and the US Centers for Disease Control and Prevention Agency for Toxic Substances and Disease Registry (CDC/ATSDR) National Center for Environmental Health (NCEH). Trained Indian Health Service staff recruited pregnant women during prenatal visits at one of six participating Indian Health Service or Public Law 638 hospitals on the Navajo Nation. The enrollment protocol prioritized recruitment of women during their 1st trimester but an open enrollment process was used, allowing women to enroll at any time during their pregnancy. For the present study, urine samples from NBCS participants were stratified by serum zinc concentration above and below the WHO level of sufficiency of 70 µg/dL. Additionally, we limited the sample population to individuals who also had their enrollment urine sample analyzed for uranium, total arsenic, arsenous (III) acid (arsenite, AsIII), and dimethylarsinic acid (DMA). Then we randomly selected 66 urine samples from each zinc group for inclusion in the present study out of 204 women enrolled in the NBCS at that time. Written informed consent was obtained from all study participants and the study protocol approved by the University of New Mexico Institutional Review Board (HRPO 11-310) and the Navajo Nation Human Research Review Board (NRR 11.323).

Sample Collection and Preparation

Trained hospital staff collected biospecimen samples using pre-screened metal-free collection cups, transfer pipettes, and Nalgene cryo-vials provided by CDC NCEH Division of Laboratory Sciences (DLS). During a prenatal hospital appointment trained hospital staff collected spot urine samples in sterile 50 mL urine collection cup. After collection, a laboratory staff member transferred a 1.8 mL aliquot of urine to separate 2.0 mL Nalgene cryo-vials for multi-element metals, total arsenic, speciated arsenic, and creatinine analysis. Hospital laboratory staff also collected peripheral blood via venipuncture and then allowed the blood to clot at room temperature for 30 to 40 minutes. Once clotted laboratory staff centrifuged the blood tube at 2,400 revolutions per minute for 15 minutes to separate the serum and then transferred 1.8 mL aliquot of serum to a 2.0 mL Nalgene cryo-vial. After

processing urine and serum samples, hospital staff placed all cryo-vials in a -80°C freezer for storage and transferred on dry-ice to freezer storage facilities at UNM. Chain of Custody forms were completed, reviewed, and validated at each stage of collection, storage and analysis.

Metal Biomonitoring Analysis

UNM staff shipped samples on dry ice to CDC DLS for analysis. NCEH laboratory staff prepared urine samples for uranium and other metals analyses using NCEH Method 3018.3; samples analyzed for total arsenic were prepared using Method 3018A.2 [44]; and samples analyzed for speciated arsenic were prepared using Method 3000.11 [45]. Chemical concentrations in urine and serum were measured using Inductively Coupled Plasma – Dynamic Reaction Cell – Mass Spectrometry (ICP-DRC-MS) [46]. Arsenic species concentrations were determined in separate aliquots using High Performance Liquid Chromatography (HPLC) and an anion exchange column to separate species prior to ICP-DRC-MS [47]. Urinary creatinine was determined using Roche/Hitachi Modular P Chemistry Analyzer [48].

Measurement of Urinary Isoprostanes by HPLC-Tandem Mass Spectrometry

Analysis of urinary F₂-isoprostanes and prostaglandin F_{2 α} in urine was performed by the Linus Pauling Institute Oxidative and Nitrate Stress Core Laboratory by HPLC-tandem mass spectrometry as described previously [49]. Creatinine was quantified in each sample by the same laboratory. Briefly, urine was thawed at room temperature, mixed by inversion, and centrifuged (200x *g*, 5 min). Aliquots of the supernatant were mixed with methanol and internal standards followed by addition of 0.02 M bis-tris-HCl. Samples were pH adjusted to 6.0 \pm 0.05. Strata X-AW cartridges (100 mg/3 mL, Phenomenex, Torrance, CA) were each pre-conditioned with methanol, then water. Diluted urine samples were loaded, the cartridges rinsed with successively with methanol/water, acetonitrile and cartridges dried under a gentle vacuum (\sim 5 mm Hg, 30 sec). Cartridges were then eluted 3-times with 1 mL methanol, and the eluants from each cartridge pooled and collected into glass tubes. Samples were dried under nitrogen gas, reconstituted in 200 μL methanol containing 0.1% formic acid (v:v), and injected onto the LC-MS-MS. Analytes were detected and quantified using SRM: F₂-isoprostanes/prostaglandin F_{2 α} , *m/z* 353 to 193; 8-iso-PGF_{2 α} -d₄ internal standard, *m/z* 357 to 197. Samples were analyzed by HPLC-tandem mass spectrometry against standard curves for authentic standards [49].

Statistical Methods

Summary statistics including median (interquartile range) for continuous variables and frequency (%) for categorical variables were used to describe the demographics, environmental characteristics, chemical exposures of urinary uranium, urinary total arsenic, DMA, AsIII, and serum zinc of the participants in the NBCS, overall and by zinc groups (> 70 vs < 70 $\mu\text{g}/\text{dL}$ for high vs low categorical membership).

The urine chemical measurements were corrected for urine creatinine, and values below limit of detection (LOD) were replaced by the LOD value divided by 2. Wilcoxon rank-sum tests were used to compare continuous variables between zinc groups. Chi-squared tests and

Fisher Exact tests for small expected numbers were performed to compare categorical variables between zinc groups. Pearson correlation coefficients along with the corresponding 95% confidence intervals (CIs) and scatterplots were used to summarize the correlation between log-transformed chemicals.

We compared the chemical exposures in the NBCS study to the national levels measured in women surveyed in the NHANES. The chemical exposure data were extracted from the NHANES year 2011-2012 database for women to represent the national population. Summary statistics of NHANES were calculated following the analysis guideline to account for the complex design features of NHANES including stratification, cluster sampling, and weighting [50].

Geometric mean along with 95% CIs were summarized for the chemical exposures in NBCS and the NHANES study. Statistical significance was determined using two-sample Welch's t-test comparing the mean (standard error) of the log transformed analytes, which took into account the NHANES design features.

The oxidative stress biomarkers as the primary outcomes include 8-iso-PGF_{2α}, PGF_{2α}, and the ratio of 8-iso-PGF_{2α} to PGF_{2α} to distinguish enzymatic versus chemical lipid peroxidation [41, 42]. Descriptive statistics were summarized for those variables by trimester at enrollment (1st, 2nd, 3rd). The measurements were corrected for urine creatinine and variables with skewed distributions were log transformed.

Univariable linear regression analyses were used to examine the association between each demographic variable and chemical exposure and the oxidative stress outcome (prostaglandin ratio or 8-iso-PGF_{2α}). Multiple linear regression models for the oxidative stress biomarkers were used to evaluate the impact of chemical exposures while adjusting for other potential confounding covariates. Covariates evaluated in the analysis included: age at interview, pre-pregnancy body mass index (BMI), trimester at enrollment, educational attainment (above or below high school), household income (above or below \$20,000), employment status (currently employed or not), alcohol in the past year (yes or no), tobacco smoking (never, current or former smoker), vitamin intake (yes or no), and use of wood or coal for home heating (yes or no). Linear models with a backwards variable selection method based on the Akaike information criterion (AIC) measure were used for assessing the effects of variables along with their interactions on the oxidative stress biomarker variables. We also performed multiple linear regression stratified by zinc groups to describe the different effects of urinary total arsenic on 8-iso-PGF_{2α} moderated by high vs low zinc.

To assess the potential for zinc to mediate the effect of each metal (uranium, total arsenic, AsIII and DMA) on oxidative stress biomarkers (prostaglandin ratio and 8-iso-PGF_{2α}), we used a quasi-Bayesian Monte Carlo causal mediation analysis [51, 52]. Covariates including trimester, education, household income, and tobacco were evaluated and adjusted for in the mediation analyses. We also then stratified this analysis by trimester to assess any confounding effects of changes in zinc across trimesters.

No adjustments for the multiple comparison tests were considered due to the discovery nature of this study. All analyses were conducted using the SAS 9.4 and R 3.4.1.

RESULTS

Selected Study Population Demographic Characteristics

Selected demographic characteristics for the subjects in this study are shown in Table 1. Study participants were drawn from the NBCS and all 132 participants included in this study were pregnant women between the ages of 16 to 42 with a median age of 26.8 years old. Analysis of survey information indicated that 38.6% of women had education beyond high school, 57.6% were unemployed and 43.2% had an annual household income below \$20,000 at the time of study enrollment. The majority of women (59.8%) were taking vitamin supplements and were overweight or obese based on pre-pregnancy BMI (52.2%). Current cigarette smoking is negligible among the NBCS pregnant women (<1%) which is consistent with low tobacco usage overall in the Navajo population as reported previously by Redwood et al. [53]. However, other exposures such as wood or coal heating or use of ceremonial tobacco were noted as potential contributors to oxidative stress. The survey questions used to generate Table 1 are provided in Supplemental Table 1.

Metal Biomonitoring Characteristics of the Study Population

Urinary total arsenic, AsIII, DMA, uranium and serum zinc, were measured by ICP-DRC-MS as described in Materials and Methods (Table 2). Values for the same metals in women surveyed for the NHANES [54] are shown in Table 2. Of the 132 individuals in the sample population, urinary metals were detected among 132 (100%) for urinary total arsenic, 105 (79.5%) for urinary DMA, 88 (66.7%) for urinary AsIII, and 128 for urinary uranium (97.0%) (Table 2). Serum zinc was detected in 132 (100%) individuals from the sample population (Table 2). Total arsenic concentrations had a weak to moderate correlation with AsIII levels within the population (Supplemental Figure 1A), however, as expected, total arsenic correlated strongly with the metabolite DMA (Supplemental Figure 1B).

The median concentration for serum zinc was 67 µg/dL and median concentrations for urinary total arsenic, DMA, AsIII, and uranium were 5.5 µg/g, 4.3 µg/g, 0.41 µg/g, and 0.016 µg/g creatinine respectively. To provide context for the biomonitoring values, information for women was extracted from the NHANES 2011-12. Compared to the median for women included in the NHANES 2011-12 survey, urinary uranium was 2.3 times greater for NBCS samples (Table 2) with 88.6% and 15.9% of the sample population exceeding the NHANES 50th and 95th percentiles, respectively (Supplemental Table 2). Urine uranium concentrations for NBCS participants were greater than observed values from the NHANES study (p-value <0.001).

Urinary concentrations of total arsenic were lower for NBCS participants compared to NHANES values (p-value <0.001) (supplemental Table 2). Only 40.2% and 0.8% of NBCS participants had total arsenic urine concentrations exceeding the NHANES 50th and 95th percentiles, respectively. Serum concentrations of zinc were also lower in NBCS participants when compared to NHANES (p-value<0.001) (supplemental Table 2). No statistically significant difference was observed for DMA. Concentrations of urinary AsIII and serum zinc decreased from the 1st to 3rd trimester while concentrations of urinary uranium, total arsenic, and DMA remained stable (Table 3).

We observed no significant difference in urinary uranium or arsenic concentrations between the low versus high zinc groups suggesting that zinc status did not influence urinary metal levels (Table 2). The median concentration for the high zinc group was 78 µg/dL, which is similar to the NHANES values of 80 µg/dL and above the WHO sufficiency standard of 70 µg/dL. The median for the low zinc group was 48 µg/dL and well below the NHANES values and zinc sufficiency standard. The high and low zinc groups did not differ in the demographic characteristics with the exception of smoker classification. There were more Never Smokers and fewer Former Smokers in the low zinc group. Additionally, the high zinc group included more women who enrolled in the NBCS during the 1st trimester and low zinc group had more women enroll during the 3rd trimester (Table 1).

Characteristics of Oxidative Stress Biomarkers in the Study Population

Table 4 provides the summary statistics for oxidative stress biomarkers as measured by HPLC-Tandem Mass Spectrometry. The biomarker 8-iso-PGF_{2α} is a well-regarded marker for detection of chemical lipid peroxidation in human studies but does not take into account enzymatic contributions to total 8-iso-PGF_{2α}. The use of a prostaglandin ratio (8-iso-PGF_{2α} to PGF_{2α}) has been described to overcome this issue [41, 42]. Of the oxidative stress biomarkers measured in this study, PGF_{2α} and the prostaglandin ratio, but not 8-iso-PGF_{2α}, were significantly different based on pregnancy trimester (Table 4). When only zinc is considered, comparison between the low and high zinc status subsets revealed no difference in the oxidative stress biomarkers 8-iso-PGF_{2α} or the prostaglandin ratio (Figure 1). The urinary 8-iso-PGF_{2α} levels measured in this study are comparable to previous reports in pregnant populations [55, 56]. Based on findings that the isoprostane metabolites dinorF1 and F2 values were subject to variability between different batch analyses (data not shown), only 8-iso-PGF_{2α}, PGF_{2α} and the prostaglandin ratio were included for the regression models.

Ceremonial tobacco use was associated significantly with higher levels 8-iso-PGF_{2α} and prostaglandin ratio oxidative stress markers (Supplemental Tables 3 and 4). In addition, significantly lower 8-iso-PGF_{2α} levels were observed in populations with a household income above \$20,000 and education above high school (Supplemental Table 3). The prostaglandin ratio was significantly affected by trimester stage (Supplemental Table 4).

Associations between metals and oxidative stress biomarkers

In a univariable analysis of the linear regression model there were no significant associations with environmental metals and 8-iso-PGF_{2α} (Table 5). Of the potential confounders, low income and ceremonial tobacco use had a significant association with urinary 8-iso-PGF_{2α} in the univariable analysis (Table 5). A multivariable analysis revealed significant main effects and interaction effects between total arsenic and low zinc on the 8-iso-PGF_{2α} outcome (Table 5).

The findings differed when using the prostaglandin ratio as the oxidative stress biomarker. In the univariable analysis there was a positive association with the arsenic metabolite DMA, but not other metals (Table 6) and a marginal positive association between total arsenic and the prostaglandin ratio was observed in the multivariable analysis. Notably, different sets of

confounding variables were identified for each of the oxidative stress biomarkers. For example, pregnancy trimester was identified as a confounding variable for the prostaglandin ratio but not for 8-iso-PGF_{2α} alone (Table 6). This can be accounted for by the lack of significant difference of 8-iso-PGF_{2α} by trimester whereas PGF_{2α} was significantly increased as pregnancy progressed from 1st to 3rd trimester (Table 4).

In order to better understand the arsenic and zinc interaction term from Table 5, we ran the stratified multivariable analysis by serum zinc group. We observed that the association between total arsenic and oxidative stress was modified by zinc group. Specifically, arsenic was positively associated with increased 8-iso-PGF_{2α} for the high serum zinc group, but not for the low serum zinc group (Table 7). In terms of the mediation effect of zinc on the association of metal/metalloids with oxidative stress, the causal mediation analysis (Materials and Methods) yielded no significant results of zinc mediation.

DISCUSSION

The Navajo Nation was a site of extensive uranium mining for more than 40 years [2, 35] and even in the absence of current active mining, potential exposures persist through proximity to AUMs, and water and soil contamination [57]. Uranium and arsenic were the focus of this study based on the prevalence of these metals in AUM waste, elevated levels in many water sources on Navajo land [7] and biomonitoring evidence indicating elevated uranium exposures in individuals [9]. Both metals have been reported to induce oxidative stress in experimental models [14-23], although the specific mechanisms that might be pertinent to a human population at environmental levels are currently unknown. Numerous studies link exposures to uranium or arsenic or mixed metals to a multitude of adverse health effects such as renal, cardiovascular and immune diseases [8, 9, 58-63]. Although oxidative stress is viewed as a mechanism underlying many metal-associated adverse health effects, there is limited knowledge regarding the potential association between uranium exposure and oxidative stress in human populations. Understanding potential roles of environmental metal exposure and oxidative stress during pregnancy is important because oxidative stress has been implicated in various pregnancy disorders, most notably pre-eclampsia and preterm labor [64].

The median urinary uranium levels in this study population were more than double those reported in the general US population based on NHANES data, yet no significant association between uranium and the oxidative stress biomarkers was detected. This finding is consistent with experimental studies where concentrations of uranium at or above 100 μM were required to elicit a measureable oxidative stress response [19-23] and suggests that the level of uranium exposure in these NBCS participants is below the threshold for detectable increase in oxidative stress.

We identified a significant positive association between urinary total arsenic and the oxidative stress biomarker 8-iso PGF_{2α} in a multivariable analysis. This finding is in agreement with several other population studies investigating arsenic exposure and oxidative stress, but results vary across studies. Positive associations between arsenic exposure and the urinary oxidative stress markers of DNA damage (8-oxodG or 8-OHdG) [26, 30, 32-34] or

8-iso-PGF_{2α} [34] have been reported. A study in children found a positive correlation between arsenic exposure and salivary 8-OHdG, but not urinary 8-OHdG [24] and a dose relationship study based on different arsenic exposures failed to identify a significant association between arsenic and protein carbonyl or 8-oxo-2'-deoxyguanosine levels [28]. Another study found the strongest association between 8-oxodG and percent monomethylarsonic acid (MMA) compared to a weak association with urinary inorganic arsenic [29]. Those authors proposed that the differences from other reported findings may have been due to differences in arsenic metabolism in their indigenous study population [29]. Despite differences in study populations in terms of age, ethnicity, gender, pregnancy status and other factors, as well as lack of uniformity in specific oxidative stress biomarkers selected overall, our findings in the NBCS participants and those reported in the literature provide evidence of arsenic-associated elevation of oxidative stress in humans.

In this study we hypothesized that zinc status would have an impact on metal-associated oxidative stress based on the antioxidant properties of zinc [65, 66]. However, we did not obtain definitive evidence that zinc mediated the arsenic effect with regard to the oxidative stress biomarker 8-iso-PGF_{2α}. One possible explanation is the impact of pregnancy on zinc status. Decreased serum zinc is common in pregnancy with reported mean values of approximately 58-60 μg/dL at term [66-69]. During pregnancy, women with serum zinc values <56 μg/dL are considered zinc deficient [70] and the mean serum zinc value in the NBCS study participants was below this definition of zinc sufficiency. Multivariable analysis using the standard biomarker 8-iso-PGF_{2α} revealed a positive association between low serum zinc and elevated oxidative stress as has been reported previously in adults [66, 71].

The biomarker 8-iso-PGF_{2α} is considered the best fatty acid indicator of oxidative stress *in vivo* [10, 72] and is widely used in human studies. One caveat is that cyclooxygenases 1 & 2 can contribute to the formation of 8-iso-PGF_{2α} independent of oxidative stress stimuli. Recent studies in humans and animal models suggest that the prostaglandin ratio of 8-iso-PGF_{2α}/PGF_{2α} is a better indicator of oxidative stress because it accounts for changes in biosynthesis pathways [41, 42]. In this study we compared 8-iso-PGF_{2α} or the prostaglandin ratio as the oxidative stress biomarker variables and noted differences based on the biomarker used. For example, in the linear regression models pregnancy trimester was significantly associated with oxidative stress when the prostaglandin ratio was used as the biomarker variable. This association was absent when 8-iso-PGF_{2α} alone was the biomarker variable. We found that PGF_{2α}, but not 8-iso-PGF_{2α}, levels increased as pregnancy advanced which is consistent with other findings that oxidative stress increases with pregnancy [55, 56]. This comparison provides an example of the potential value of incorporating the prostaglandin ratio into analyses of F₂-isoprostanes as biomarkers of oxidative stress in addition to the more standard use of 8-iso-PGF_{2α}. Use of both markers would allow for comparison of findings with established literature and expand understanding of the benefits of the ratio especially under conditions or in populations where endogenous contributions to total production of 8-iso-PGF_{2α} may be an important factor.

Very few demographic characteristics were associated with either oxidative stress biomarker, but ceremonial tobacco use was significant in the regression models. Ceremonial tobacco use is associated with traditional practices and may reflect complex exposures from the local

plant sources or other aspects of cultural ceremonies incorporating this product. Although we do not know how or why intermittent ceremonial tobacco use might be associated with systemic oxidative stress, this example highlights the importance of taking into consideration culture and local practices in studies of indigenous populations.

Challenges in this study include the relative paucity of data in the Navajo population, limited published information on metals and oxidative stress in pregnant women, and the NHANES sample not being representative of many aspects of our study group including a sufficient numbers of Native Americans for comparison. While NHANES values are representative of the overall US population, the survey is not designed to assess pregnant women specifically. During the 2011-12 NHANES cycle for example, less than 5% of participating women had a clinically confirmed pregnancy. Pregnancy results in changes to urinary excretion of metals and decreases in serum zinc concentrations. For example, previous work has demonstrated that the prevalence of arsenic species changes throughout pregnancy due to greater methylation [73]. Additionally, previous work has indicated that pregnant women have lower serum zinc concentrations than the general population because increased copper absorption interferes with zinc absorption [74]. No information about changes in uranium excretion throughout pregnancy is available. Therefore, the representative NHANES values may not fully represent urinary or serum concentrations of pregnant women in the United States but it remains the only large sample data set using consistent, high quality, and comparable analytical methods to use for comparison of population biomonitoring data. This study was focused on metals (arsenic and uranium) known to exceed the MCL in local water sources. Future studies may include additional metals in relation to biomarkers of oxidative stress.

In conclusion, we find evidence for elevated urinary uranium in women enrolled in the NBCS when compared to NHANES values, but no corresponding increase in oxidative stress measures associated with uranium. In contrast, arsenic is associated with increased levels of urinary 8-iso-PGF_{2α} as has been reported in other populations. Despite established antioxidant properties of zinc, zinc was not found to have causal mediation of the effects of the other metals on oxidative stress. Direct comparison of the prostaglandin ratio versus 8-iso-PGF_{2α} alone as biomarkers of oxidative stress reveals the utility of the prostaglandin ratio for studies of populations with known underlying factors, such as pregnancy, that may affect oxidative stress measurements.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

ACKNOWLEDGEMENTS

This work was supported by the National Institutes of Health [Competing Revision to 1R01ES021100 NIH Revision Awards for Creating Virtual Consortium for Translational/Transdisciplinary Environmental Research (ViCTER)(R01); Navajo Birth Cohort Study: CDC/NCEH/ATSDR 5 U01 TS000135-05 and 1P50ES026102 UNM Center for Native Environmental Health Equity NIEHS/NIMHD P50ES026102 & USEPA (#83615701). Additional support for Shared Resources was provided by 2P30 CA118100 UNM Comprehensive Cancer Center NCI and training support for Drs. Hoover and Dashner by 5K12GM088021-08 Academic Science Education and Research Training (ASERT) Program and support from the Oregon Agricultural Experimental Research Station. We also acknowledge the expertise of Dr. Jaewoo Choi and Scott Leonard of the Linus Pauling Institute for the analysis of urinary isoprostanes.

This material was developed in part under Assistance Agreement No. 83615701 awarded by the U.S. Environmental Protection Agency to the University of New Mexico Health Sciences Center. It has not been formally reviewed by EPA. The views expressed are solely those of the authors and do not necessarily reflect those of the Agency. EPA does not endorse any products or commercial services mentioned in this publication.

A portion of the research reported in this publication was supported by the National Institute of Environmental Health Sciences of the National Institutes of Health under Award Number P50ES026102. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

We would like to acknowledge all of our indigenous partners we have had the pleasure of working with over the last three decades and from whom we have learned about the lasting impact of the abandoned mines, with particular acknowledgement of those from Tachee/Blue Gap and Red Water Pond Road communities on Navajo, the 1304 families in our original DiNEH Project, and the more than 750 families currently in the Navajo Birth Cohort Study. We also acknowledge the partnership of our research partners from Southwest Research and Information Center and their Navajo Home Environmental Assessment staff, Navajo Nation Department of Health including Mae-Gilene Begay and the Community Health and Environmental Research Specialists, Navajo Nation Environmental Protection Agency, and faculty and staff from the UNM METALS team including Dr. David Begay of CEHP for his support in understanding Navajo culture and tradition. Finally, all Navajo Nation research is reviewed and monitored by the Navajo Nation Human Research Review Board.

Abbreviations⁹

8-iso-PGF_{2α}	8-iso prostaglandin F _{2α}
AsIII	arsenite
AUM	abandoned uranium mine
ATSDR	Agency for Toxic Substances and Disease Registry
BMI	Body Mass Index
CDC	Center for Disease Control and Prevention
CI	confidence interval
DLS	Division of Laboratory Sciences
DMA	dimethylarsinic acid
HPLC	High Performance Liquid Chromatography
ICP-DRC-MS	Inductively Coupled Plasma – Dynamic Reaction Cell – Mass Spectrometry
IQR	interquartile range
LOD	Limit of Detection
MCL	Maximum Contaminant Level
NBCS	Navajo Birth Cohort Study
NCEH	National Center for Environmental Health
NHANES	National Health and Nutrition Examination Survey
PGF_{2α}	prostaglandin F _{2α}

UNM University of New Mexico

WHO World Health Organization

REFERENCES

1. Agency, U.S.E.P., Technical Report on Technologically Enhanced Naturally Occurring Radioactive Materials from Uranium Mining, R.P.D. Office of Radiation and Indoor Air, Editor. 2008: Washington DC.
2. Agency, U.S.E.P., Health and Environmental Impacts of Uranium Contamination in the Navajo Nation Five-Year Plan. 2016.
3. Lewis J, et al., Environmental exposures to metals in Native communities and implications for child development: basis for the Navajo birth cohort study. *J Soc Work Disabil Rehabil*, 2015 14(3-4): p. 245–69. [PubMed: 26151586]
4. Blake JM, et al., Elevated Concentrations of U and Co-occurring Metals in Abandoned Mine Wastes in a Northeastern Arizona Native American Community. *Environ Sci Technol*, 2015 49(14): p. 8506–14. [PubMed: 26158204]
5. Corlin L, et al., Health Effects and Environmental Justice Concerns of Exposure to Uranium in Drinking Water. *Curr Environ Health Rep*, 2016 3(4): p. 434–442. [PubMed: 27815781]
6. Orescanin V, et al., Characterization and treatment of water used for human consumption from six sources located in the Cameron/Tuba City abandoned uranium mining area. *J Environ Sci Health A Tox Hazard Subst Environ Eng*, 2011 46(6): p. 627–35. [PubMed: 21547818]
7. Hoover J, et al., Elevated Arsenic and Uranium Concentrations in Unregulated Water Sources on the Navajo Nation, USA. *Expo Health*, 2017 9(2): p. 113–124. [PubMed: 28553666]
8. Hund L, et al., A Bayesian framework for estimating disease risk due to exposure to uranium mine and mill waste on the Navajo Nation. *Journal of the Royal Statistical Society*, 2015 178(4): p. 1069–1091.
9. Lewis J, Hoover J, and MacKenzie D, Mining and Environmental Health Disparities in Native American Communities. *Curr Environ Health Rep*, 2017 4(2): p. 130–141. [PubMed: 28447316]
10. Il'yasova D, Scarbrough P, and Spasojevic I, Urinary biomarkers of oxidative status. *Clin Chim Acta*, 2012 413(19-20): p. 1446–53. [PubMed: 22683781]
11. Friehoff J, et al., Clinical Relevance of Biomarkers of Oxidative Stress. *Antioxid Redox Signal*, 2015 23(14): p. 1144–70. [PubMed: 26415143]
12. Deeb RS and Hajjar DP, Repair Mechanisms in Oxidant-Driven Chronic Inflammatory Disease. *Am J Pathol*, 2016 186(7): p. 1736–1749. [PubMed: 27171899]
13. Ratliff BB, et al., Oxidant Mechanisms in Renal Injury and Disease. *Antioxid Redox Signal*, 2016 25(3): p. 119–46. [PubMed: 26906267]
14. Valko M, et al., Redox- and non-redox-metal-induced formation of free radicals and their role in human disease. *Arch Toxicol*, 2016 90(1): p. 1–37. [PubMed: 26343967]
15. Flora SJ, Arsenic-induced oxidative stress and its reversibility. *Free Radic Biol Med*, 2011 51(2): p. 257–81. [PubMed: 21554949]
16. Jomova K, et al., Arsenic: toxicity, oxidative stress and human disease. *J Appl Toxicol*, 2011 31(2): p. 95–107. [PubMed: 21321970]
17. Cooper KL, Liu KJ, and Hudson LG, Enhanced ROS production and redox signaling with combined arsenite and UVA exposure: contribution of NADPH oxidase. *Free Radic Biol Med*, 2009 47(4): p. 381–8. [PubMed: 19414066]
18. Shi H, Hudson LG, and Liu KJ, Oxidative stress and apoptosis in metal ion-induced carcinogenesis. *Free Radic Biol Med*, 2004 37(5): p. 582–93. [PubMed: 15288116]
19. Periyakaruppan A, et al., Uranium induces oxidative stress in lung epithelial cells. *Arch Toxicol*, 2007 81(6): p. 389–95. [PubMed: 17124605]
20. Gagnaire B, et al., Effects of depleted uranium on oxidative stress, detoxification, and defence parameters of zebrafish *Danio rerio*. *Arch Environ Contam Toxicol*, 2013 64(1): p. 140–50. [PubMed: 23052361]

21. Garmash SA, et al., Pro-oxidative, genotoxic and cytotoxic properties of uranyl ions. *J Environ Radioact*, 2014 127: p. 163–70. [PubMed: 23312590]
22. Zheng J, et al., Hydrogen sulfide (H₂S) attenuates uranium-induced acute nephrotoxicity through oxidative stress and inflammatory response via Nrf2-NF- κ B pathways. *Chem Biol Interact*, 2015 242: p. 353–62. [PubMed: 26523793]
23. Hao Y, et al., Ghrelin protects against depleted uranium-induced apoptosis of MC3T3-E1 cells through oxidative stress-mediated p38-mitogen-activated protein kinase pathway. *Toxicol Appl Pharmacol*, 2016 290: p. 116–25. [PubMed: 26529667]
24. Hinhumpatch P, et al., Oxidative DNA damage and repair in children exposed to low levels of arsenic in utero and during early childhood: application of salivary and urinary biomarkers. *Toxicol Appl Pharmacol*, 2013 273(3): p. 569–79. [PubMed: 24128852]
25. Pei Q, et al., Oxidative DNA damage of peripheral blood polymorphonuclear leukocytes, selectively induced by chronic arsenic exposure, is associated with extent of arsenic-related skin lesions. *Toxicol Appl Pharmacol*, 2013 266(1): p. 143–9. [PubMed: 23142755]
26. Xu Y, et al., Association of oxidative stress with arsenic methylation in chronic arsenic-exposed children and adults. *Toxicol Appl Pharmacol*, 2008 232(1): p. 142–9. [PubMed: 18640141]
27. De Vizcaya-Ruiz A, et al., Biomarkers of oxidative stress and damage in human populations exposed to arsenic. *Mutat Res*, 2009 674(1-2): p. 85–92. [PubMed: 18984063]
28. Harper KN, et al., A dose-response study of arsenic exposure and markers of oxidative damage in Bangladesh. *J Occup Environ Med*, 2014 56(6): p. 652–8. [PubMed: 24854259]
29. Engstrom KS, et al., Low 8-oxo-7,8-dihydro-2'-deoxyguanosine levels and influence of genetic background in an Andean population exposed to high levels of arsenic. *Mutat Res*, 2010 683(1-2): p. 98–105. [PubMed: 19896490]
30. Engstrom KS, et al., Chronic exposure to cadmium and arsenic strongly influences concentrations of 8-oxo-7,8-dihydro-2'-deoxyguanosine in urine. *Free Radic Biol Med*, 2010 48(9): p. 1211–7. [PubMed: 20153423]
31. Burgess JL, et al., Environmental Arsenic Exposure and Urinary 8-OHdG in Arizona and Sonora. *Clin Toxicol (Phila)*, 2007 45(5): p. 490–8. [PubMed: 17503254]
32. Lu S, et al., Trace elements are associated with urinary 8-hydroxy-2'-deoxyguanosine level: a case study of college students in Guangzhou, China. *Environ Sci Pollut Res Int*, 2016 23(9): p. 8484–91. [PubMed: 26782679]
33. Kubota R, et al., Urinary 8-hydroxy-2'-deoxyguanosine in inhabitants chronically exposed to arsenic in groundwater in Cambodia. *J Environ Monit*, 2006 8(2): p. 293–9. [PubMed: 16470262]
34. Wang T, et al., The effects of heavy metals and their interactions with polycyclic aromatic hydrocarbons on the oxidative stress among coke-oven workers. *Environ Res*, 2015 140: p. 405–13. [PubMed: 25956561]
35. Hunter CM, et al., The Navajo Birth Cohort Study. *J Environ Health*, 2015 78(2): p. 42–5. [PubMed: 26502566]
36. Butte NF, Calloway DH, and Van Duzen JL, Nutritional assessment of pregnant and lactating Navajo women. *Am J Clin Nutr*, 1981 34(10): p. 2216–28. [PubMed: 7293950]
37. Bruno RS, et al., Dietary zinc restriction in rats alters antioxidant status and increases plasma F₂ isoprostanes. *J Nutr Biochem*, 2007 18(8): p. 509–18. [PubMed: 17142032]
38. Kloubert V and Rink L, Zinc as a micronutrient and its preventive role of oxidative damage in cells. *Food Funct*, 2015 6(10): p. 3195–204. [PubMed: 26286461]
39. Oteiza PI, Zinc and the modulation of redox homeostasis. *Free Radic Biol Med*, 2012 53(9): p. 1748–59. [PubMed: 22960578]
40. Prasad AS, Zinc is an Antioxidant and Anti-Inflammatory Agent: Its Role in Human Health. *Front Nutr*, 2014 1: p. 14. [PubMed: 25988117]
41. van't Erve TJ, et al., Reinterpreting the best biomarker of oxidative stress: The 8-iso-PGF(2 α)/PGF(2 α) ratio distinguishes chemical from enzymatic lipid peroxidation. *Free Radic Biol Med*, 2015 83: p. 245–51. [PubMed: 25772010]
42. Van't Erve TJ, et al., Reinterpreting the best biomarker of oxidative stress: The 8-iso-prostaglandin F₂ α /prostaglandin F₂ α ratio shows complex origins of lipid peroxidation biomarkers in animal models. *Free Radic Biol Med*, 2016 95: p. 65–73. [PubMed: 26964509]

43. Prevention, C.f.D.C.a., National Health and Nutrition Examination Survey Data, N.C.f.H. Statistics, Editor. 2011-2012: Hyattsville, MD.
44. Prevention, C.f.D.C.a., Laboratory Procedure Manual, Multi-Element ICP-DRC-MS Renamed from “Inductively Coupled Plasma-Mass Spectrometry (ICP-DRC-MS)” Method No: 3018.3 (15 element panel) and 3018A.2 (total arsenic). 2012.
45. Prevention, C.f.D.C.a., National Health and Nutrition Examination Survey, 2011-2012 Data Documentation, Codebook, and Frequencies, Arsenics - Total & Speciated - Urine (UAS_G).
46. Prevention, C.f.D.C.a., Laboratory Procedure Manual, Serum Multi-Element ICP-DRC-MS Method No: ICPDRCMS-3006.7.
47. Prevention, C.f.D.C.a., Laboratory Procedure Manual, Urine arsenic speciation HPLCICPDRCMS (Renamed from High Performance Liquid Chromatography Inductively Coupled Plasma Dynamic Reaction Cell Mass Spectrometry (HPLC-ICP-DRC-MS)) Method No: 3000.11 (Formerly 0161A/01-OD). 2011.
48. Prevention, C.f.D.C.a., Laboratory Procedure Manual, Urinary Creatinine
49. Taylor AW, Bruno RS, and Traber MG, Women and smokers have elevated urinary F(2)-isoprostane metabolites: a novel extraction and LC-MS methodology. *Lipids*, 2008 43(10): p. 925–36. [PubMed: 18751751]
50. Prevention, C.f.D.C.a., National Health and Nutrition Examination Survey: Analytic Guidelines, 2011-2012.
51. Imai K, Keele L, and Yamamoto T, Identification, Inference and Sensitivity Analysis for Causal Mediation Effects. *Statistical Science*, 2010 25(1): p. 51–71.
52. Imai K, Keele L, and Tingley D, A general approach to causal mediation analysis. *Psychol Methods*, 2010 15(4): p. 309–34. [PubMed: 20954780]
53. Redwood D, et al., Differences in cigarette and smokeless tobacco use among American Indian and Alaska Native people living in Alaska and the Southwest United States. *Nicotine Tob Res*, 2010 12(7): p. 791–6. [PubMed: 20525781]
54. Prevention, C.f.D.C.a., Fourth Report on Human Exposure to Environmental Chemicals, Updated Tables, U.S.D.o.H.a.H. Services, Editor. 2017: Atlanta, GA.
55. Ishihara O, et al., Isoprostanes, prostaglandins and tocopherols in pre-eclampsia, normal pregnancy and non-pregnancy. *Free Radic Res*, 2004 38(9): p. 913–8. [PubMed: 15621708]
56. Palm M, et al., F(2)-isoprostanes, tocopherols and normal pregnancy. *Free Radic Res*, 2009 43(6): p. 546–52. [PubMed: 19384749]
57. Agency, U.S.E.P., Addressing uranium contamination on the Navajo Nation., P.S.S. Program, Editor. 2014: San Francisco, CA.
58. Okaneku J, et al., Urine uranium concentrations and renal function in residents of the United States--2001 to 2010. *Clin Toxicol (Phila)*, 2015 53(10): p. 931–4. [PubMed: 26468995]
59. Brugge D and Buchner V, Health effects of uranium: new research findings. *Rev Environ Health*, 2011 26(4): p. 231–49. [PubMed: 22435323]
60. Attreed SE, Navas-Acien A, and Heaney CD, Arsenic and Immune Response to Infection During Pregnancy and Early Life. *Curr Environ Health Rep*, 2017 4(2): p. 229–243. [PubMed: 28488132]
61. Sanchez TR, Perzanowski M, and Graziano JH, Inorganic arsenic and respiratory health, from early life exposure to sex-specific effects: A systematic review. *Environ Res*, 2016 147: p. 537–55. [PubMed: 26891939]
62. Cosselman KE, Navas-Acien A, and Kaufman JD, Environmental factors in cardiovascular disease. *Nat Rev Cardiol*, 2015 12(11): p. 627–42. [PubMed: 26461967]
63. Bjorklund G, et al., Recent aspects of uranium toxicology in medical geology. *Environ Res*, 2017 156: p. 526–533. [PubMed: 28431380]
64. Duhig K, Chappell LC, and Shennan AH, Oxidative stress in pregnancy and reproduction. *Obstet Med*, 2016 9(3): p. 113–6. [PubMed: 27630746]
65. Maret W, Zinc biochemistry: from a single zinc enzyme to a key element of life. *Adv Nutr*, 2013 4(1): p. 82–91. [PubMed: 23319127]

66. Prasad AS, et al., Zinc supplementation decreases incidence of infections in the elderly: effect of zinc on generation of cytokines and oxidative stress. *Am J Clin Nutr*, 2007 85(3): p. 837–44. [PubMed: 17344507]
67. Izquierdo Alvarez S, et al., Updating of normal levels of copper, zinc and selenium in serum of pregnant women. *J Trace Elem Med Biol*, 2007 21 Suppl 1: p. 49–52. [PubMed: 18039497]
68. Choi R, et al., A Prospective Study of Serum Trace Elements in Healthy Korean Pregnant Women. *Nutrients*, 2016 8(11).
69. Wrzesniak M, et al., The Influence of Tobacco Smoke on Protein and Metal Levels in the Serum of Women during Pregnancy. *PLoS One*, 2016 11(8): p. e0161342. [PubMed: 27548057]
70. International Zinc Nutrition Consultative, G., et al., International Zinc Nutrition Consultative Group (IZiNCG) technical document #1. Assessment of the risk of zinc deficiency in populations and options for its control. *Food Nutr Bull*, 2004 25(1 Suppl 2): p. S99–203. [PubMed: 18046856]
71. Tomey KM, et al., Dietary fat subgroups, zinc, and vegetable components are related to urine F2-isoprostane concentration, a measure of oxidative stress, in midlife women. *J Nutr*, 2007 137(11): p. 2412–9. [PubMed: 17951478]
72. Dorjgochoo T, et al., Major metabolite of F2-isoprostane in urine may be a more sensitive biomarker of oxidative stress than isoprostane itself. *Am J Clin Nutr*, 2012 96(2): p. 405–14. [PubMed: 22760572]
73. Hopenhayn C, et al., Profile of urinary arsenic metabolites during pregnancy. *Environ Health Perspect*, 2003 111(16): p. 1888–91. [PubMed: 14644662]
74. Gibson RS, The role of diet- and host-related factors in nutrient bioavailability and thus in nutrient-based dietary requirement estimates. *Food Nutr Bull*, 2007 28(1 Suppl International): p. S77–100. [PubMed: 17521121]

Highlights

- Urinary uranium, but not arsenic, is elevated in the study participants compared to NHANES.
- Arsenic is associated with increased levels of urinary 8-iso-prostaglandin F_{2α}.
- Uranium is not associated with elevated urinary F₂-isoprostanes.
- Zinc was not found to have any causal mediation of the effects of the other metals on oxidative stress.

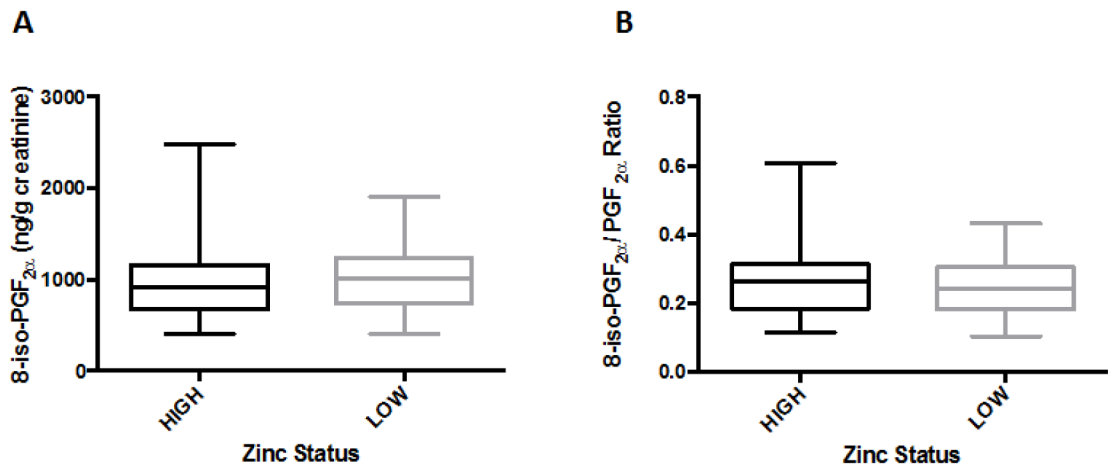


Figure 1. No difference in oxidative stress biomarkers by zinc status.

Participant results were separated by zinc status and analyzed for changes in the oxidative stress markers **A.** 8-iso-PGF_{2α} and **B.** Prostaglandin ratio (8-iso-PGF_{2α} / PGF_{2α})

Table 1.
Selected demographic characteristics for NBCS participants

Characteristic		Total (n = 132)	Zinc Group		p
			High (n = 66)	Low (n = 66)	
Median Age [IQR]		26.80 [22.33 - 31.2]	26.35 [22.0 - 30.38]	27.15 [22.5 - 31.2]	0.664 ^a
Trimester stage (%)	1st	25 (18.9)	25 (37.9)	0 (0.0)	<0.001 ^b
	2nd	59 (44.7)	32 (48.5)	27 (40.9)	
	3rd	48 (36.4)	9 (13.6)	39 (59.1)	
Pre pregnancy BMI (%)	Normal ^c	28 (21.2)	13 (19.7)	15 (22.7)	0.536 ^b
	Overweight ^c	37 (28.0)	13 (19.7)	24 (36.4)	
	Obese ^c	32 (24.2)	15 (22.7)	17 (25.8)	
	Information unavailable	35 (26.5)	25 (37.9)	10 (15.2)	
Vitamin usage (%)	No	39 (29.5)	22 (33.3)	17 (25.8)	0.241 ^b
	Yes	79 (59.9)	34 (51.5)	45 (68.2)	
	No Response	14 (10.6)	10 (15.2)	4 (6.1)	
Education above high school (%)	No	65 (49.2)	28 (42.4)	37 (56.1)	0.385 ^b
	Yes	51 (38.6)	27 (40.9)	24 (36.4)	
	No Response	16 (12.1)	11 (16.7)	5 (7.6)	
Annual household income <\$20,000 (%)	No	41 (31.1)	23 (34.8)	18 (27.3)	0.182 ^b
	Yes	57 (43.2)	23 (34.8)	34 (51.5)	
	No Response	34 (25.8)	20 (30.3)	14 (21.2)	
Currently unemployed (%)	No	42 (31.8)	24 (36.4)	18 (27.3)	0.17 ^b
	Yes	76 (57.6)	32 (48.5)	44 (66.7)	
	No Response	14 (10.6)	10 (15.2)	4 (6.1)	
Alcohol usage in the past year (%)	No	84 (63.6)	42 (63.6)	42 (63.6)	0.506 ^b
	Yes	34 (25.8)	14 (21.2)	20 (30.3)	
	No Response	14 (10.6)	10 (15.2)	4 (6.1)	
Cigarette usage (%)	Never Smoked	89 (67.4)	38 (57.6)	51 (77.3)	0.018 ^d
	Current Smoker	1 (0.8)	0 (0.0)	1 (1.5)	
	Former Smoker	16 (12.1)	12 (18.2)	4 (6.1)	
	No Response	26 (19.7)	16 (24.2)	10 (15.2)	
Ceremonial tobacco usage (%)	No	73 (55.3)	38 (57.6)	35 (53.0)	0.278 ^b
	Yes	45 (34.1)	18 (27.3)	27 (40.9)	
	No Response	14 (10.6)	10 (15.2)	4 (6.1)	
Wood used for home heating (%)	No	70 (53.0)	29 (43.9)	41 (62.1)	0.163 ^b
	Yes	48 (36.4)	27 (40.9)	21 (31.8)	
	No Response	14 (10.6)	10 (15.2)	4 (6.1)	

Characteristic		Total (n = 132)	Zinc Group		p
			High (n = 66)	Low (n = 66)	
Coal used for home heating (%)	No	28 (21.2)	11 (16.7)	17 (25.8)	0.438 ^b
	Yes	90 (68.2)	45 (68.2)	45 (68.2)	
	No Response	14 (10.6)	10 (15.2)	4 (6.1)	

Abbreviations: BMI, body mass index; IQR, interquartile range

^aWilcoxon rank sum test was used to calculate the P-value comparing low vs. high zinc groups.

^bChi-square tests were used to calculate the P-value comparing low vs. high zinc groups.

^cBMI defined as Normal (18.5 - 24.9 kg/m²), Overweight (25 - 29.9 kg/m²), and Obese (>30 kg/m²)

^dFisher Exact tests were used to calculate the P-value comparing low vs. high zinc groups.

Table 2.

Summary of metal levels in the NBCS population

Metals	NHANES ^a Median [IQR]	NBCS Median [IQR]	% Below LOD	Zinc Group	
				High [IQR]	Low [IQR]
Uranium (µg/g creatinine)	0.007 [0.005 -0.013]	0.016 [0.0098 - 0.025]	3.0	0.013 [0.0094 - 0.021]	0.017 [0.011 - 0.027]
Total Arsenic (µg/g creatinine)	6.6 [4.2-15.0]	5.5 [4.2-8.2]	0.0	5.1 [4.1 - 7.9]	5.9 [4.8 - 8.6]
DMA (µg/g creatinine)	4.1 [2.7-6.7]	4.3 [2.9-5.8]	20.5	3.9 [2.6-5.2]	4.6 [3.2-6.2]
AsIII (µg/g creatinine)	<LOD [< LOD - <LOD]	0.41 [0.019-0.58]	33.3	0.43 [0.24 - 0.58]	0.37 [0.16-0.58]
Zinc (µg/dL)	80 [72 -88]	67 [48.0 - 78.0]	0.0	78 [74.0 - 85.0]	48 [44 -51]

Abbreviations: NBCS, Navajo Birth Cohort Study; NHANES, National Health and Nutrition Examination Survey; LOD, limit of detection; IQR, interquartile range; DMA, dimethylarsinic acid; AsIII, arsenite

^aSummary statistics were based on 2011-2012 NHANES dataset restricted to women; Urine chemical measurements were corrected for creatinine.

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Table 3.

Distribution of metals by trimester.

Metal	Trimester	n	Median [IQR]	<i>p</i> ^a
Uranium (µg/g creatinine)	1st	25	0.014 [0.010 - 0.021]	0.76
	2nd	59	0.016 [0.009 - 0.024]	
	3rd	48	0.016 [0.011 - 0.026]	
Total Arsenic (µg/g creatinine)	1st	25	4.992 [4.097 - 7.910]	0.55
	2nd	59	5.573 [4.293 - 8.287]	
	3rd	48	5.723 [4.565 - 8.632]	
DMA (µg/g creatinine)	1st	25	3.460 [2.629 - 5.806]	0.357
	2nd	59	4.572 [2.802 - 5.894]	
	3rd	48	4.307 [3.177 - 5.750]	
AsIII (µg/g creatinine)	1st	25	0.535 [0.379 - 0.892]	0.0017
	2nd	59	0.442 [0.257 - 0.574]	
	3rd	48	0.259 [0.137-0.482]	
Zinc (µg/dL)	1st	25	77 [74.0 - 82.9]	1.78E-11
	2nd	59	72 [49.65 - 82.0]	
	3rd	48	49.95 [44.075 - 52.851]	

Abbreviations: IQR, interquartile range; DMA, dimethylarsinic acid; AsIII, arsenite

^aP-values were calculated using one-way ANOVA F test of the log transformed chemical exposures among three trimesters.

Table 4:

Summary statistics for oxidative stress outcome variables.

		Overall	First Trimester	Second Trimester	Third Trimester	
	Unit ^b	Median [IQR]	Median [IQR]	Median [IQR]	Median [IQR]	<i>p</i> ^c
8-isoPGF_{2a}	ng/g creatinine	935 [708 - 1201]	918 [674 - 1162]	881 [699 - 1219]	1009 [850 - 1171]	0.4633
prostaglandin-F_{2a}	ng/g creatinine	3897 [2952 - 5373]	3225 [2571 - 4073]	3892 [2985 - 5058]	4929 [3572 - 6905]	0.0013
Prostaglandin Ratio^a		0.25 [0.18 - 0.31]	0.30 [0.27 - 0.32]	0.25 [0.19 - 0.30]	0.20 [0.16 - 0.29]	0.0036

Abbreviations: IQR, interquartile range

^aRatio of 8-isoPGF_{2a} to PGF_{2a}.^bMeasurements were corrected for urine creatinine.^cKruskal-Wallis tests were used to calculate the p-value comparing different trimesters.

Table 5.

Regression coefficients of metal exposure, income, and ceremonial tobacco use for oxidative stress biomarker 8-isoPGF_{2α}.

Variable ^a	Univariable			Multivariable ^d		
	Coefficient ^b	Standard Error ^b	p ^c	Coefficient	Standard Error	p ^c
Uranium	-0.044	0.044	0.32	-	-	-
DMA	0.097	0.066	0.15	-	-	-
Total Arsenic	0.066	0.069	0.34	0.341	0.133	0.012
Low Zinc	0.086	0.066	0.20	0.679	0.327	0.041
Total As/Zinc Group Interaction	-	-	-	-0.380	0.181	0.038
Annual Household Income <\$20,000	0.158	0.078	0.044	0.168	0.076	0.030
Ceremonial Tobacco Use	0.167	0.071	0.021	0.142	0.079	0.076

Abbreviations: DMA, dimethylarsinic acid

^aMetals were log transformed. Zinc was a binary variable. Outcome variable (8-isoPGF_{2α}) was log transformed.

^bThe estimates of the regression coefficients and the standard errors from the linear regression models.

^cP-values were calculated using t tests.

^dMultivariable model with backward selection using AIC criteria modeling the relationship between metals and oxidative stress biomarker 8-isoPGF_{2α}. R²=0.15.

Table 6.

Regression coefficients of metal exposure, education, ceremonial tobacco use and trimester for the prostaglandin ratio.

Variable ^a	Univariable			Multivariable ^d		
	Coefficient ^b	Standard Error ^b	p ^c	Coefficient	Standard Error	p ^c
Uranium	0.008	0.010	0.44	-0.055	0.043	0.21
Total Arsenic	0.031	0.015	0.043	0.19	0.096	0.053
DMA	0.021	0.008	0.01	-	-	-
Uranium/Total Arsenic Interaction	-	-	-	0.036	0.023	0.11
Low Zinc Group	-0.015	0.015	0.320	-	-	-
Education above high school	-0.024	0.017	0.157	-0.03	0.015	0.049
Ceremonial Tobacco Use	0.035	0.016	0.037	0.035	0.016	0.026
2nd Trimester	-0.057	0.020	0.004	-0.052	0.021	0.017
3rd Trimester	-0.079	0.020	0.00016	-0.089	0.022	0.00009

Abbreviations: DMA, dimethylarsinic acid

^aMetals were log transformed.

^bThe estimates of the regression coefficients and the standard errors from the linear regression models.

^cP-values were calculated using t tests.

^dFinal multivariable model with backward selection using AIC criteria modeling the relationship between chemicals and oxidative stress biomarker ratio. R²=0.23.

Table 7.

Regression coefficients of total arsenic and the oxidative stress biomarker 8-isoPGF_{2α}, stratified by zinc group.

Variable ^a	Low Zinc group			High Zinc group		
	Coefficient ^b	Standard Error ^b	p ^c	Coefficient	Standard Error	p ^c
Total Arsenic	-0.045	0.12	0.71	0.34	0.14	0.018
Annual Household Income <\$20,000	0.097	0.10	0.35	0.24	0.11	0.040
Ceremonial Tobacco Use	0.099	0.10	0.34	0.20	0.13	0.13

^aMetals were log transformed. Zinc was a binary variable. Outcome variable (8-isoPGF_{2α}) was log transformed.

^bThe estimates of the regression coefficients and the standard errors from the linear regression models.

^cP-values were calculated using t tests.