TOXICOLOGICAL PROFILE FOR CHLORINE

U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES
Public Health Service
Agency for Toxic Substances and Disease Registry

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DISCLAIMER

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UPDATE STATEMENT

A Toxicological Profile for Chlorine, Draft for Public Comment was released in October 2007. This edition supersedes any previously released draft or final profile.

Toxicological profiles are revised and republished as necessary. For information regarding the update status of previously released profiles, contact ATSDR at:

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FOREWORD

This toxicological profile is prepared in accordance with guidelines* developed by the Agency for Toxic Substances and Disease Registry (ATSDR) and the Environmental Protection Agency (EPA). The original guidelines were published in the *Federal Register* on April 17, 1987. Each profile will be revised and republished as necessary.

The ATSDR toxicological profile succinctly characterizes the toxicologic and adverse health effects information for these toxic substances described therein. Each peer-reviewed profile identifies and reviews the key literature that describes a substance's toxicologic properties. Other pertinent literature is also presented, but is described in less detail than the key studies. The profile is not intended to be an exhaustive document; however, more comprehensive sources of specialty information are referenced.

The focus of the profiles is on health and toxicologic information; therefore, each toxicological profile begins with a public health statement that describes, in nontechnical language, a substance's relevant toxicological properties. Following the public health statement is information concerning levels of significant human exposure and, where known, significant health effects. The adequacy of information to determine a substance's health effects is described in a health effects summary. Data needs that are of significance to protection of public health are identified by ATSDR.

Each profile includes the following:

- (A) The examination, summary, and interpretation of available toxicologic information and epidemiologic evaluations on a toxic substance to ascertain the levels of significant human exposure for the substance and the associated acute, subacute, and chronic health effects;
- (B) A determination of whether adequate information on the health effects of each substance is available or in the process of development to determine levels of exposure that present a significant risk to human health of acute, subacute, and chronic health effects; and
- (C) Where appropriate, identification of toxicologic testing needed to identify the types or levels of exposure that may present significant risk of adverse health effects in humans.

The principal audiences for the toxicological profiles are health professionals at the Federal, State, and local levels; interested private sector organizations and groups; and members of the public.

This profile reflects ATSDR's assessment of all relevant toxicologic testing and information that has been peer-reviewed. Staffs of the Centers for Disease Control and Prevention and other Federal scientists have also reviewed the profile. In addition, this profile has been peer-reviewed by a nongovernmental panel and was made available for public review. Final responsibility for the contents and views expressed in this toxicological profile resides with ATSDR.

Thomas R. Frieden, M.D., M.P.H.

Administrator

Agency for Toxic Substances and

Disease Registry

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*Legislative Background

The toxicological profiles are developed under the Comprehensive Environmental Response, Compensation, and Liability Act of 1980, as amended (CERCLA or Superfund). CERCLA section 104(i)(1) directs the Administrator of ATSDR to "... effectuate and implement the health related authorities" of the statute. This includes the preparation of toxicological profiles for hazardous substances most commonly found at facilities on the CERCLA National Priorities List and that pose the most significant potential threat to human health, as determined by ATSDR and the EPA. Section 104(i)(3) of CERCLA, as amended, directs the Administrator of ATSDR to prepare a toxicological profile for each substance on the list. In addition, ATSDR has the authority to prepare toxicological profiles for substances not found at sites on the National Priorities List, in an effort to "... establish and maintain inventory of literature, research, and studies on the health effects of toxic substances" under CERCLA Section 104(i)(1)(B), to respond to requests for consultation under section 104(i)(4), and as otherwise necessary to support the site-specific response actions conducted by ATSDR.

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QUICK REFERENCE FOR HEALTH CARE PROVIDERS

Toxicological Profiles are a unique compilation of toxicological information on a given hazardous substance. Each profile reflects a comprehensive and extensive evaluation, summary, and interpretation of available toxicologic and epidemiologic information on a substance. Health care providers treating patients potentially exposed to hazardous substances will find the following information helpful for fast answers to often-asked questions.

Primary Chapters/Sections of Interest

Chapter 1: Public Health Statement: The Public Health Statement can be a useful tool for educating patients about possible exposure to a hazardous substance. It explains a substance's relevant toxicologic properties in a nontechnical, question-and-answer format, and it includes a review of the general health effects observed following exposure.

- **Chapter 2: Relevance to Public Health**: The Relevance to Public Health Section evaluates, interprets, and assesses the significance of toxicity data to human health.
- **Chapter 3: Health Effects**: Specific health effects of a given hazardous compound are reported by type of health effect (death, systemic, immunologic, reproductive), by route of exposure, and by length of exposure (acute, intermediate, and chronic). In addition, both human and animal studies are reported in this section.

NOTE: Not all health effects reported in this section are necessarily observed in the clinical setting. Please refer to the Public Health Statement to identify general health effects observed following exposure.

Pediatrics: Four new sections have been added to each Toxicological Profile to address child health issues:

Section 1.6 How Can (Chemical X) Affect Children?

Section 1.7 How Can Families Reduce the Risk of Exposure to (Chemical X)?

Section 3.7 Children's Susceptibility

Section 6.6 Exposures of Children

Other Sections of Interest:

Section 3.8 Biomarkers of Exposure and Effect Section 3.11 Methods for Reducing Toxic Effects

ATSDR Information Center

Phone: 1-800-CDC-INFO (800-232-4636) or 1-888-232-6348 (TTY) Fax: (770) 488-4178

The following additional material can be ordered through the ATSDR Information Center:

Case Studies in Environmental Medicine: Taking an Exposure History—The importance of taking an exposure history and how to conduct one are described, and an example of a thorough exposure history is provided. Other case studies of interest include Reproductive and Developmental Hazards; Skin Lesions and Environmental Exposures; Cholinesterase-Inhibiting Pesticide Toxicity; and numerous chemical-specific case studies.

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Managing Hazardous Materials Incidents is a three-volume set of recommendations for on-scene (prehospital) and hospital medical management of patients exposed during a hazardous materials incident. Volumes I and II are planning guides to assist first responders and hospital emergency department personnel in planning for incidents that involve hazardous materials. Volume III—

Medical Management Guidelines for Acute Chemical Exposures—is a guide for health care professionals treating patients exposed to hazardous materials.

Fact Sheets (ToxFAQs) provide answers to frequently asked questions about toxic substances.

Other Agencies and Organizations

The National Center for Environmental Health (NCEH) focuses on preventing or controlling disease, injury, and disability related to the interactions between people and their environment outside the workplace. Contact: NCEH, Mailstop F-29, 4770 Buford Highway, NE, Atlanta, GA 30341-3724 • Phone: 770-488-7000 • FAX: 770-488-7015.

The National Institute for Occupational Safety and Health (NIOSH) conducts research on occupational diseases and injuries, responds to requests for assistance by investigating problems of health and safety in the workplace, recommends standards to the Occupational Safety and Health Administration (OSHA) and the Mine Safety and Health Administration (MSHA), and trains professionals in occupational safety and health. Contact: NIOSH, 200 Independence Avenue, SW, Washington, DC 20201 • Phone: 800-356-4674 or NIOSH Technical Information Branch, Robert A. Taft Laboratory, Mailstop C-19, 4676 Columbia Parkway, Cincinnati, OH 45226-1998 • Phone: 800-35-NIOSH.

The National Institute of Environmental Health Sciences (NIEHS) is the principal federal agency for biomedical research on the effects of chemical, physical, and biologic environmental agents on human health and well-being. Contact: NIEHS, PO Box 12233, 104 T.W. Alexander Drive, Research Triangle Park, NC 27709 • Phone: 919-541-3212.

Referrals

The Association of Occupational and Environmental Clinics (AOEC) has developed a network of clinics in the United States to provide expertise in occupational and environmental issues. Contact:

AOEC, 1010 Vermont Avenue, NW, #513, Washington, DC 20005 • Phone: 202-347-4976
• FAX: 202-347-4950 • e-mail: AOEC@AOEC.ORG • Web Page: http://www.aoec.org/.

The American College of Occupational and Environmental Medicine (ACOEM) is an association of physicians and other health care providers specializing in the field of occupational and environmental medicine. Contact: ACOEM, 25 Northwest Point Boulevard, Suite 700, Elk Grove Village, IL 60007-1030 • Phone: 847-818-1800 • FAX: 847-818-9266.

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THE PROFILE HAS UNDERGONE THE FOLLOWING ATSDR INTERNAL REVIEWS:

- 1. Health Effects Review. The Health Effects Review Committee examines the health effects chapter of each profile for consistency and accuracy in interpreting health effects and classifying end points.
- 2. Minimal Risk Level Review. The Minimal Risk Level Workgroup considers issues relevant to substance-specific Minimal Risk Levels (MRLs), reviews the health effects database of each profile, and makes recommendations for derivation of MRLs.
- 3. Data Needs Review. The Applied Toxicology Branch reviews data needs sections to assure consistency across profiles and adherence to instructions in the Guidance.
- 4. Green Border Review. Green Border review assures the consistency with ATSDR policy.

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PEER REVIEW

A peer review panel was assembled for chlorine. The panel consisted of the following members:

- 1. John Balmes, M.D., Professor in Residence, Department of Medicine, University of California, San Francisco, San Francisco, California;
- 2. Meryl Karol, Ph.D., Professor Emeritus, Associate Dean for Academic Affairs and Research, University of Pittsburgh, Pennsylvania; and
- 3. Dennis Shusterman, M.D., MPH, Professor Emeritus, Department of Medicine, University of California, San Francisco, California.

These experts collectively have knowledge of chlorine's physical and chemical properties, toxicokinetics, key health end points, mechanisms of action, human and animal exposure, and quantification of risk to humans. All reviewers were selected in conformity with the conditions for peer review specified in Section 104(I)(13) of the Comprehensive Environmental Response, Compensation, and Liability Act, as amended.

Scientists from the Agency for Toxic Substances and Disease Registry (ATSDR) have reviewed the peer reviewers' comments and determined which comments will be included in the profile. A listing of the peer reviewers' comments not incorporated in the profile, with a brief explanation of the rationale for their exclusion, exists as part of the administrative record for this compound.

The citation of the peer review panel should not be understood to imply its approval of the profile's final content. The responsibility for the content of this profile lies with the ATSDR.

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1. PUBLIC HEALTH STATEMENT

This public health statement tells you about chlorine and the effects of exposure to it.

The Environmental Protection Agency (EPA) identifies the most serious hazardous waste sites in the nation. These sites are then placed on the National Priorities List (NPL) and are targeted for long-term federal clean-up activities. Chlorine gas is too reactive to be detected in environmental media at hazardous waste sites. Any chlorine gas released at these sites would be quickly converted to other substances whose primary source may or may not have been chlorine.

When a substance is released either from a large area, such as an industrial plant, or from a container, such as a drum or bottle, it enters the environment. Such a release does not always lead to exposure. You can be exposed to a substance only when you come in contact with it. You may be exposed by breathing, eating, or drinking the substance, or by skin contact. Since chlorine is highly reactive, you are unlikely to be exposed directly to it unless there has been a large scale accidental release in the nearby vicinity.

If you are exposed to chlorine, many factors will determine whether you will be harmed. These factors include the dose (how much), the duration (how long), and how you come in contact with it. You must also consider any other chemicals you are exposed to and your age, sex, diet, family traits, lifestyle, and state of health.

The subject of this profile is molecular chlorine (Cl₂), which exists as a gas under normal environmental conditions or as a liquid when stored under pressure. Although molecular chlorine is used in some water disinfection processes, the resulting solution, commonly referred to as chlorinated water, does not actually contain molecular chlorine. Therefore, water disinfection and the chemicals present in chlorinated water, such as hypochlorite, are not the main focus of this document and are only discussed where relevant.

1.1 WHAT IS CHLORINE?

Chlorine is a gas with a very irritating odor	It is very unstable and quickly reacts with many substances to form other chemicals.
Used in manufacturing and water disinfection	Chlorine is an extremely important industrial chemical that is used in the production of thousands of products. It is also used for water disinfection, although the chlorine itself is quickly transformed into other chemicals at the beginning of the process.
Chlorine gas is not present in chlorinated water	A common misconception is that molecular chlorine (Cl ₂) is present in chlorinated water. During water chlorination, molecular chlorine gas may be added to the water at first; however, the chlorine is quickly transformed into other chemicals, which actually disinfect the water. Hypochlorous acid and hypochlorite anion are two of these chemicals that disinfect the water. The terms "free chlorine" and "aqueous chlorine" in drinking water usually refer to the amount of hypochlorous acid and hypochlorite in the water. It is important to recognize that these compounds are different from molecular chlorine.
Bleach is not chlorine	One of the important products that chlorine is used to make is bleach, and people sometimes confuse chlorine with bleach. Bleach contains a compound called sodium hypochlorite. If you mix acidic chemicals with bleach, chlorine can be formed and given off as a gas.

For more information on the sources, properties, and uses of chlorine, see Chapters 4 and 5.

1.2 WHAT HAPPENS TO CHLORINE WHEN IT ENTERS THE ENVIRONMENT?

Chlorine is very unstable in the environment	Chlorine is very unstable, and reacts with a variety of chemicals and water when it is released into the environment.
Rapidly broken down • Air	Chlorine is broken down by sunlight within a matter of several minutes.
• Water	Chlorine dissolves in water and is converted into chloride and hypochlorous acid.
Chlorine can travel from its source	If chlorine is spilled into water or onto soil or if it is released from a tank into the air, the chlorine will evaporate very quickly forming a greenish-yellow cloud that can be carried by the wind from the source.

For more information on chlorine in the environment, see Chapter 6.

1.3 HOW MIGHT I BE EXPOSED TO CHLORINE?

Most people are not expected to be exposed to chlorine	Because chlorine is so reactive, it is not normally detected in the environment except for very low levels in the air above seawater.
Accidental exposure to chlorine	You may be exposed through breathing, skin contact, and eye contact if an accident involving chlorine takes place nearby, such as a liquid chlorine spill, a leak from a chlorine tank, or a leak from a facility that produces or uses chlorine. You may also be exposed to chlorine if you mix household chemicals such as toilet cleaner with bleach. Hypochlorous acid is used to treat swimming pool water. You may be exposed to chlorine gas through the improper use of swimming pool chemicals.
Workplace air	People who work in places where chlorine is made or used may be exposed to low levels over a period of time. People may be exposed to high levels if a large amount of chlorine is released during an accident.

For more information on human exposure to chlorine, see Chapter 6.

1.4 HOW CAN CHLORINE ENTER AND LEAVE MY BODY?

Chlorine gas enters your body only when you breathe it in.	Chlorine gas can enter your body through your nose or your mouth. At low concentrations (less than 10 ppm), almost all of the chlorine is removed from the air in the upper part of the respiratory airways and only a very small amount may reach your lungs. If you drink hypochlorite solution, it may react with the acids in your stomach and possibly form chlorine gas.
Immediately reacts with other chemicals	Chlorine gas reacts with the water in the cells located in the surface of the respiratory airways and forms other compounds that produce irritation of the airways. Most of these compounds eventually are transformed into chloride ions, which are normal components of the body.

For more information on how chlorine enters and leaves the body, see Chapter 3.

1.5 HOW CAN CHLORINE AFFECT MY HEALTH?

This section looks at studies concerning potential health effects in animal and human studies.

The effect of chlorine on human health depends on how much chlorine is present, how you are exposed to it, and the length of exposure.

Short-term exposure to chlorine in air	The following effects have been observed in humans briefly exposed to chlorine: • mild nose irritation at 1–3 ppm • eye irritation at 5 ppm • throat irritation at 5–15 ppm • immediate chest pain, vomiting, changes in breathing rate, and cough at 30 ppm • lung injury (toxic pneumonitis) and pulmonary edema (fluid in the lungs) at 40–60 ppm • death after 30 minute exposure to 430 ppm • death after a few minute exposure to 1,000 ppm The concentrations listed above are approximate; the effects will depend also on exposure duration. In general, people who suffer from respiratory conditions such as allergies or hay fever, or who are heavy smokers, tend to experience more severe effects than healthy subjects or nonsmokers.
Long-term exposure to chlorine in air	No significant harmful health effects were observed in workers exposed for years to relatively low concentrations of chlorine (around 1 ppm). The tissues inside the nose were principally affected in animals exposed to chlorine for longer durations.
Short-term exposure to hypochlorite solution by ingestion	Drinking small amounts of hypochlorite solution (less than a cup) can produce irritation of the esophagus. Drinking concentrated hypochlorite solution can produce severe damage to the upper digestive tract and even death. These effects are most likely caused by the caustic nature of the hypochlorite solution and not from exposure to molecular chlorine.
Long-term exposure to hypochlorite solution by ingestion	There is no information on long-term ingestion of hypochlorite solution in humans. Animals that drank hypochlorite solution in water for up to 2 years did not show any significant health effects. The amount of hypochlorite solution in the water that the animals drank was much smaller than what is found in household bleach.
Skin exposure to hypochlorite solution	Spilling hypochlorite solution on the skin can produce irritation. The severity of the effects depends on the concentration of sodium hypochlorite in the bleach.

Further information on the health effects of chlorine in humans and animals can be found in Chapters 2 and 3.

1.6 HOW CAN CHLORINE AFFECT CHILDREN?

This section discusses potential health effects in humans from exposures during the period from conception to maturity at 18 years of age.

Children are likely to have similar effects as adults, but may be more sensitive than adults	Short-term exposures (minutes) to high concentrations of chlorine affect children in the same manner they affect adults (i.e., mucous membrane and respiratory tract irritation). We do not know what the effects could be in children following longer-term (weeks or longer), low-level exposure to chlorine gas, but this type of exposure occurs only in workers and is not relevant to children. We also do not know what the effects could be in children following longer-term, low-level exposure to hypochlorite solution.
Birth defects	We do not know whether exposure to chlorine gas during pregnancy can result in damage to unborn babies because there are no studies of pregnant women or pregnant animals exposed to chlorine gas.
	One study of rats exposed to hypochlorite solution during pregnancy found no evidence of birth defects or any other developmental alteration in the baby rats. The amount of chlorine that the rats consumed was many times higher than what people are normally exposed to through drinking water.

1.7 HOW CAN FAMILIES REDUCE THE RISK OF EXPOSURE TO CHLORINE?

Do not mix bleach with household cleaners	Chlorine gas can be released to the air when bleach is mixed with other cleaning solutions that contain an acid like some toilet cleaners. Mixing bleach with ammonia also produces very hazardous gases, such as chloramines.
Store household chemicals out of reach of young children	Always store household chemicals in their original labeled containers out of reach of young children to prevent accidental poisonings. Never store household chemicals in containers children would find attractive to eat or drink from, such as old soda bottles.
Follow instructions for swimming pool disinfection	Chlorine gas can also be released to the air when chemicals used to chlorinate swimming pools are mishandled. If you have a swimming pool at home, read the labels of the chlorination products carefully and do not let children play with these products.

1.8 IS THERE A MEDICAL TEST TO DETERMINE WHETHER I HAVE BEEN EXPOSED TO CHLORINE?

There are no medical tests available for	There are no medical tests to determine whether you have been exposed specifically to chlorine.
chlorine	Chlorine is transformed in the body into chloride ions, which are normal components of the body. An enormous amount of chlorine has to be inhaled or ingested in order to detect a significant increase in chloride ions in the blood. This has occurred in a few cases of ingestion of large amounts of hypochlorite solution and one of them was a fatal case.

1.9 WHAT RECOMMENDATIONS HAS THE FEDERAL GOVERNMENT MADE TO PROTECT HUMAN HEALTH?

The federal government develops regulations and recommendations to protect public health. Regulations can be enforced by law. The EPA, the Occupational Safety and Health Administration (OSHA), and the Food and Drug Administration (FDA) are some federal agencies that develop regulations for toxic substances. Recommendations provide valuable guidelines to protect public health, but cannot be enforced by law. The Agency for Toxic Substances and Disease Registry (ATSDR) and the National Institute for Occupational Safety and Health (NIOSH) are two federal organizations that develop recommendations for toxic substances.

Regulations and recommendations can be expressed as "not-to-exceed" levels, that is, levels of a toxic substance in air, water, soil, or food that do not exceed a critical value that is usually based on levels that affect animals; they are then adjusted to levels that will help protect humans. Sometimes these not-to-exceed levels differ among federal organizations because they used different exposure times (an 8-hour workday or a 24-hour day), different animal studies, or other factors.

Recommendations and regulations are also updated periodically as more information becomes available. For the most current information, check with the federal agency or organization that provides it.

Some regulations and recommendations for chlorine include the following:

Levels in air set by EPA	EPA established an environmental air limit of 0.5 ppm. Exposure to higher levels could result in discomfort and irritation. Dependent on the concentration, these effects may be reversible when exposure ends.
Levels in workplace air set by OSHA	OSHA set a legal limit of 1 ppm chlorine in air as a ceiling limit. At no time should a worker's exposure exceed this limit.
Levels in drinking water set by EPA	EPA established a maximum contaminant level (MCL) and maximum residual disinfectant level (MRDL) of 4 mg/L for free chlorine in drinking water.

1.10 WHERE CAN I GET MORE INFORMATION?

If you have any more questions or concerns, please contact your community or state health or environmental quality department, or contact ATSDR at the address and phone number below.

ATSDR can also tell you the location of occupational and environmental health clinics. These clinics specialize in recognizing, evaluating, and treating illnesses that result from exposure to hazardous substances.

Toxicological profiles are also available on-line at www.atsdr.cdc.gov and on CD-ROM. You may request a copy of the ATSDR ToxProfilesTM CD-ROM by calling the toll-free information and technical assistance number at 1-800-CDCINFO (1-800-232-4636), by e-mail at cdcinfo@cdc.gov, or by writing to:

Agency for Toxic Substances and Disease Registry Division of Toxicology and Environmental Medicine 1600 Clifton Road NE Mailstop F-62 Atlanta, GA 30333

Fax: 1-770-488-4178

CHLORINE 8 1. PUBLIC HEALTH STATEMENT

Organizations for-profit may request copies of final Toxicological Profiles from the following:

National Technical Information Service (NTIS) 5285 Port Royal Road Springfield, VA 22161 Phone: 1-800-553-6847 or 1-703-605-6000

Web site: http://www.ntis.gov/

2. RELEVANCE TO PUBLIC HEALTH

2.1 BACKGROUND AND ENVIRONMENTAL EXPOSURES TO CHLORINE IN THE UNITED STATES

Chlorine is a greenish-yellow gas with a pungent, irritating odor. It is stored and transported as a liquid under pressure. Chlorine is transported as either a liquid or a gas through pipelines within chemical plants or over distances of several kilometers. When chlorine is released into the environment, it reacts with both organic and inorganic substances that it comes into contact with. When chlorine gas is released into water, such as during water chlorination, it quickly dissolves in the water and then disproportionates within seconds to form chloride and hypochlorous acid. Chlorine may be released into the environment from facilities where it is produced or used, or during accidents, such as a chlorine tank rupture or a liquid chlorine spill. Most of the chlorine released during these types of incidents is expected to volatilize into the air forming a greenish-yellow chlorine gas cloud. Because chlorine is approximately 2.5 times heavier than air, the chlorine cloud remains near the ground. This cloud can be carried away from the site of release by the wind. Chlorine undergoes direct photolysis in the air and its half-life in the troposphere is on the order of several minutes.

Because it is so reactive, chlorine gas is normally not detected in the environment except at low levels in seawater aerosols. Therefore, background exposure of the general population to chlorine is not expected to represent a health concern. Human biomonitoring data are not available for chlorine. Analyzing human biological tissue and fluids for chlorine is not relevant because >95% of the chlorine that is inhaled (over a 1–5 ppm range) is removed in the upper respiratory tract and eventually joins the chloride pool in the body. The amount of chlorine that would need to be inhaled to induce a significant increase in extracellular chloride in the body is probably a lethal amount.

There are two primary means by which the general population may be exposed to chlorine. Individuals located near an accidental release of chlorine, either from a manufacturing facility or the transportation of liquefied chlorine may be exposed to high levels of this gas through inhalation and dermal contact if the cloud travels in their direction. In addition, people who mix acidic solutions with hypochlorite solutions, such as bleach or certain types of swimming pool chemicals, may accidentally be exposed to chlorine gas. Children may be exposed to chlorine through the same routes that affect adults, except for occupational exposures. Occupational exposure to low levels of chlorine gas in air may occur for individuals who work at facilities that produce or use chlorine. These individuals may also be exposed to high chlorine concentrations if an accidental release occurs inside the facility.

Exposure to chlorine through drinking water is expected to be very low. Free chlorine in drinking water is defined as the sum of dissolved chlorine gas, hypochlorous acid, and hypochlorite anion. As discussed in Chapters 4 and 6, the level of dissolved chlorine in water is extremely low, except under acidic conditions; therefore, the term free chlorine in public water systems typically refers to the concentration of hypochlorous acid and hypochlorite anion. The term total chlorine as it pertains to water sanitation practices usually refers to the amount of free chlorine plus chloroamines (sometimes called combined chlorines) produced during the sanitation process. It is important to recognize that these compounds are different from molecular chlorine even though the terminology is often used interchangeably.

2.2 SUMMARY OF HEALTH EFFECTS

Chlorine Gas. The principal targets of exposure to chlorine gas are the respiratory airways and the eyes. Exposure can occur only by direct contact of inhaled chlorine gas with the respiratory epithelium or via direct contact of the eyes with the gas. The skin seems to be a less sensitive target to direct contact with chlorine gas possibly because it lacks the moisture of mucous membranes. The effects of acute-duration exposures to high concentrations of chlorine have been known for almost a century, starting with its use as a chemical weapon at Ypres, Belgium, during World War I. Additional information regarding the effects of brief high-level exposures to chlorine has been collected from accidental exposures following leaks during transport of tanks containing liquid chlorine, leaks from storage tanks, domestic accidents involving bleach solutions, mishandling of chemicals used at swimming pools, and even accidents in high school science experiments. These and many additional studies, including studies in volunteers exposed to controlled concentrations of chlorine, indicate that exposures to 1-3 ppm produce mild irritation of the nose that can be tolerated for about 1 hour; 5 ppm may produce eye irritation; headache and throat irritation may occur at concentrations of 5-15 ppm; 30 ppm produces immediate chest pain, nausea and vomiting, dyspnea, and cough: 40–60 ppm produces toxic pneumonitis and pulmonary edema; 430 ppm usually causes death in 30 minutes, and 1,000 ppm is fatal within a few minutes. In most cases, death is the result of pulmonary edema. Accidental releases of chlorine have affected adults and children, and a few reports suggested that children might be more susceptible than adults to the effects of chlorine. This may be due to the smaller diameter of the airways of children compared to adults.

The effects of exposure to chlorine seem to depend, at least above a certain minimal exposure concentration, on the duration of exposure and exposure concentration, and the moisture content of the surface contacted by the gas (i.e., the respiratory epithelium). Exposures to relatively low concentrations

of chlorine (<5 ppm) are not expected to affect deep lung structures since most of the inhaled chlorine (>95%) is scrubbed in the upper portion of the respiratory tract, whether breathing is through the nose or through the mouth. With the exception of cough, substernal pain, and respiratory distress, the symptoms occurring after exposure to moderate concentrations of chlorine generally subside within 24 hours. Edema, observed following high exposures, is caused by marked alveolar capillary congestion followed immediately by focal and confluent area of fluid with a high content of fibrinogen. Pulmonary edema peaks in 12–24 hours and the resulting hypoxia further increases capillary permeability, which creates a vicious cycle. Initially, the pulmonary fluid is interstitial, but if it overwhelms the capacity of the lymphatic system to drain it, the alveoli become filled. A further complication is the formation of hyaline membranes from the alveolar fluid with high-fibrinogen content, which along with developing areas of atelectasis (collapse), and right to left shunting of blood, explains the poor oxygen diffusion with resultant hypoxemia and later hypercapnea. Subjects surviving the acute phase of exposure to high concentrations of chlorine may still be in danger of delayed death due to bronchial pneumonia or pneumonia. The complications of chlorine inhalation fit the histological condition known as diffuse alveolar damage that is associated with the clinical condition known as the adult respiratory distress syndrome.

Not all of the signs and symptoms exhibited by subjects exposed to moderate to high concentrations of chlorine gas are caused directly by chlorine. In general, it is believed that effects such as nausea and vomiting are reflex in origin, and headache and loss of consciousness are probably due to the hypoxia caused by pulmonary edema. Leukocytosis is almost always found in subjects admitted to emergency departments following exposure to high chlorine gas and is most likely a general response to inflammation. Anxiety and changes in blood pressure and heart rate also are commonly mentioned in case reports. While cardiovascular alterations can be due in part to a ventilation perfusion mismatch, they may also represent a general response to the stress and anxiety of having been involved in a chemical accident and being admitted to a health facility.

Prolonged exposures to relatively low concentrations of chlorine in occupational settings have not given indications of respiratory or other health problems among the workers, but additional better-controlled studies are necessary to add confidence to these early findings. Workers occasionally experience brief episodes of high exposure ("gassing" incidents), in some cases to concentrations high enough to warrant a visit to the emergency room. In some of these cases and also in some cases of high exposure of the general population, long-term follow-up has shown persistent respiratory alterations that included airway obstruction and reactive airway dysfunction syndrome (RADS). RADS is defined as an asthma-like illness after a single acute exposure to a respiratory irritant in otherwise healthy individuals, characterized

by increased responsiveness to methacholine challenge. There are many factors that can play a role in whether residual effects will be present, including exposure level and duration of exposure, medical treatment following exposure, length of the follow-up, underlying respiratory disease, and smoking status.

A series of reports by Kilburn suggested that acute exposure to high concentrations of chlorine produced long-term neurobehavioral effects (i.e., memory loss, slow reaction time, impaired balance, hearing loss, visual alterations). No other study of chlorine-exposed subjects has included neurobehavioral testing, but this could potentially be examined in animal models. It is not known whether exposure to chlorine gas can affect reproduction or development in humans. Only one early study reported that pregnancy outcome was not affected among female workers at a chlorine plant. There is also no relevant information regarding effects of chlorine exposure on the immune system. A few studies of workers in the chemical industry did not find any evidence that chlorine gas is carcinogenic. The EPA, the International Agency for Research on Cancer (IARC), and the Department of Health and Human Services (DHHS) have not classified chlorine gas as to its carcinogenicity.

The respiratory system is also the target of chlorine toxicity in animals. Animals exposed briefly to high concentrations of chlorine gas have shown respiratory effects similar to those observed in humans, with the added observations of severe gross and microscopic changes in the respiratory airways. Chlorine, in relatively low concentrations (1–3 ppm), also induced histological alterations in the respiratory tract, particularly the upper portion, in intermediate- and chronic-duration inhalation studies in animals. In these studies, there was no indication that chlorine exposure affects reproductive parameters. No studies are available that evaluated whether chlorine affects immunocompetence or the development of young organisms. Chlorine gas was not carcinogenic in rats and mice exposed to up to 2.5 ppm for 2 years.

Hypochlorite Solutions. At very low pH (<2), it is theoretically possible that chlorine gas can be formed; therefore, exposures to hypochlorite solutions are briefly discussed even though, under normal pH, the predominant species are expected to be hypochlorous acid and hypochlorite (for a detailed discussion and definitions of terms related to the chemistry of chlorine in water, please see Chapter 4). The principal targets of exposure to hypochlorite solutions are the upper gastrointestinal tract and the skin. Exposure to hypochlorite can occur via accidental or intentional ingestion of chlorine bleach or via direct contact of the skin with hypochlorite solutions. In most cases, ingestion of small amounts (less than a cup) of sodium hypochlorite bleach (approximately 5.25% sodium hypochlorite, 52,250 ppm, or 52,250 mg/L) (ppm in water = mg/L) can produce esophageal irritation without permanent consequences. However, fatalities due to ingestion of sodium hypochlorite have been reported. In a reported case, an autopsy of a

woman who drank an unknown amount of bleach revealed esophageal and gastric mucosal erosions, perforations at the gastroesophageal junction, and extensive necrosis of adjacent soft tissue. Aspiration of hypochlorite bleach into the lungs following ingestion of bleach also has been reported as a cause of death. The lethal dose of sodium hypochlorite in adults has been reported to be approximately 200 mL of a solution containing 3-6% chlorine. No significant additional toxicities have been reported in humans following oral exposure to hypochlorite. Two intermediate-duration studies in which volunteers were exposed to known amounts of chlorine in water provided no evidence of adverse effects. In one of them, consumption of water containing 5 mg/L chlorine (approximately 0.036 mg Cl/kg/day) had no significant effect on hematology, serum chemistry, urinalysis, or additional physiological parameters. Another study of limited scope showed that consumption of water containing 20 ppm chlorine (approximately 0.4 mg Cl/kg/day) had no significant effect on serum lipids or serum levels of thyroid hormones. It is not known whether oral exposure to chlorine can affect the immune and nervous systems, or reproduction or development in humans. There are no studies of cancer in humans exposed to chlorine itself. Based on inadequate evidence for carcinogenicity of hypochlorite salts in animals and no data from studies in humans, a study determined that hypochlorite salts are not classifiable as to their carcinogenicity in humans.

Direct contact of the skin with household chlorine bleach can cause skin irritation in humans. Although sodium hypochlorite generally is not considered a contact sensitizer, several cases of allergic contact dermatitis have been reported. Commercial household bleaches are prepared with sodium hydroxide and are typically very alkaline; it is this property that may result in the irritant contact dermatitis. The limited information regarding ocular effects of direct contact of the eye with hypochlorite solutions suggest that splashes in the eye with house solutions of sodium hypochlorite rarely result in serious consequences.

For the most part, the results of oral and dermal studies of chlorine in animals support the observations in humans. Studies in which hypochlorite bleach was placed in the esophagus of animals reproduced the observations following high exposure in humans. Additional intermediate- and chronic-duration studies that examined hematology and clinical chemistry parameters and conducted gross and microscopic examination of tissues from rats and mice following exposure to chlorinated water provided little evidence of chlorine-related toxicity. In the intermediate-duration studies, Sprague-Dawley rats, F344 rats, and B6C3F₁ mice were dosed for 90 days with up to 24.9, 85, and 39.2 mg Cl/kg/day, respectively. In the chronic-duration studies, rats were exposed for 2 years to up to 14.4 mg Cl/kg/day and mice to up 24.2 mg Cl/kg/day. Studies in animals have provided no evidence that exposure to hypochlorite ions adversely affects the immune or nervous system, although an 8-week study in rats

reported alterations in some immune parameters of unknown toxicological significance (reduced delayed-type hypersensitivity reaction, increased prostaglandin E2 synthesis by macrophages, and reduced oxidative metabolism by macrophages following stimulation with phorbol myristate acetate). Exposure of male and female rats to hypochlorite before and during breeding and of the females during gestation and lactation did not cause reproductive effects in either sex or adverse developmental effects in the offspring. Cancer bioassays in rats and mice have been negative except for equivocal evidence of increased incidence of leukemia in female F344 rats in one study's bioassay. It should be mentioned, however, that one study considered the overall evidence only weakly supportive of an association between the occurrence of mononuclear cell leukemia and the consumption of chlorinated water based on the following: (1) the increase in leukemia was slight and not clearly dose-related, (2) there was no decrease in tumor latency, and (3) the incidence in concurrent controls was less than in historical controls. A limited number of studies of *in vivo* genotoxicity of hypochlorite ion provided negative results.

2.3 MINIMAL RISK LEVELS (MRLs)

Estimates of exposure levels posing minimal risk to humans (MRLs) have been made for chlorine. An MRL is defined as an estimate of daily human exposure to a substance that is likely to be without an appreciable risk of adverse effects (noncarcinogenic) over a specified duration of exposure. MRLs are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. MRLs are based on noncancerous health effects only and do not consider carcinogenic effects. MRLs can be derived for acute, intermediate, and chronic duration exposures for inhalation and oral routes. Appropriate methodology does not exist to develop MRLs for dermal exposure.

Although methods have been established to derive these levels (Barnes and Dourson 1988; EPA 1990a), uncertainties are associated with these techniques. Furthermore, ATSDR acknowledges additional uncertainties inherent in the application of the procedures to derive less than lifetime MRLs. As an example, acute inhalation MRLs may not be protective for health effects that are delayed in development or are acquired following repeated acute insults, such as hypersensitivity reactions, asthma, or chronic bronchitis. As these kinds of health effects data become available and methods to assess levels of significant human exposure improve, these MRLs will be revised.

Inhalation MRLs

 An MRL of 0.06 ppm has been derived for acute-duration inhalation exposure (14 days or less) to chlorine gas.

The effects of acute-exposure of humans and animals to chlorine have been well characterized (see Sections 2.2 and 3.2.1.2). Chlorine is a sensory irritant (substance capable of eliciting sensory irritation) and the most sensitive target for chlorine toxicity in humans and in animals is the respiratory system. Information that could be used for quantitative risk assessment regarding effects from acute exposure of humans to chlorine is available from studies of volunteers exposed to chlorine gas for 15 minutes to 8 hours (Anglen 1981; D'Alessandro et al. 1996; Rotman et al. 1983; Schins et al. 2000; Shusterman et al. 1998, 2003b). Some of these studies, as detailed below, also included sensitive individuals. Collectively, the results of these studies suggest that brief exposures to concentrations of chlorine ≤ 0.5 ppm do not cause sensory irritation or significant alterations in pulmonary function tests, but exposure to ≥ 1 ppm chlorine can induce transient respiratory and eye irritation and slight alterations in pulmonary function tests. Evaluations of soldiers gassed during World War I provide information on the effects of acute exposure to very high concentrations of chlorine and also on potentially persistent effects of acute exposure (Berghoff 1919; DOA 1933; Meakins and Priestley 1919). Similar information is available in many reports of accidental exposures to chlorine gas of workers and members of the general population (i.e., Agabiti et al. 2001; Agency for Toxic Substances and Disease Registry 1998; Bonetto et al. 2006; CDC 1991, 2005; Chasis et al. 1947; Chester et al. 1977; Hasan et al. 1983; Schönhofer et al. 1996; Sexton and Pronchik 1998; Weill et al. 1969). In both the war cases and the accidental exposures to chlorine gas, the concentrations of chlorine were generally not known. These high exposure cases have provided data on respiratory effects and on additional signs and symptoms of intoxication with chlorine that are not due to a direct action of chlorine, but that probably represent reflex responses and/or general responses to inflammation and stress that were caused by products of chlorine's reaction with bodily fluids. Some of these responses include nausea, vomiting, headache, anxiety, alterations in blood pressure, and leukocytosis.

The acute-duration database in animals is extensive, and includes a great number of studies conducted after the use of chlorine as a chemical weapon during World War I (for review, see DOA [1933] and Withers and Lees [1985b]). Most of the early studies provide information regarding lethal concentrations of chlorine as well as descriptions of the pathology of the respiratory tract caused by exposure to relatively high concentrations of chlorine. Although qualitatively informative, the early data do not meet current guidelines for use in quantitative risk assessment. More recent studies in animals, mainly rodents,

have confirmed the earlier findings regarding sensory irritation and pathological changes in the respiratory tract (Barrow and Smith 1975; Buckley et al. 1984; Demnati et al. 1995; Jiang et al. 1983; Leustik et al. 2008; Tian et al. 2008; Yildirim et al. 2004). In general, morphological alterations in the nasal mucosa of rats and mice occurred with chlorine concentrations >5 ppm. Specific lowest-observed-adverse-effect levels (LOAELs) for sensory irritation in rodents are not available. However, in response to exposure to irritant substances, a reflex mechanism allows rodents to decrease the respiratory rate as a protective response (Alarie 1973). The concentration of the irritant that induces a 50% decrease in respiratory rate has been termed RD₅₀ and is commonly used to compare the irritant potencies of chemicals. This reflex reaction has also been demonstrated in humans, dogs, and cats (Alarie 1973). Acute-duration inhalation studies provided very little information regarding end points other than those involving the respiratory system. Body weight loss, which is due to reduced food consumption, was reported in some studies (Dodd et al. 1980; Jiang et al. 1983).

Evaluation of the acute-duration inhalation database summarized above indicates that sensory irritation and pulmonary function in humans are the most sensitive end points for exposure to chlorine and will serve as the basis for derivation of an acute-duration inhalation MRL for chlorine. These findings were reported in a group of studies that can serve as co-principal studies (Anglen 1981; D'Alessandro et al. 1996; Rotman et al. 1983; Schins et al. 2000; Shusterman et al. 1998, 2003b). A detailed description of these studies is provided in Appendix A.

Collectively, this group of studies provides evidence of sensory irritation and transient pulmonary changes occurring in humans exposed to 1 ppm chlorine for up to 8 hours/day. The pulmonary changes indicated increased airway resistance and reduced air flow. No such changes were reported in volunteers exposed to 0.5 ppm chlorine (0.4 ppm in the D'Alessandro et al. [1996]) study. The longest exposure duration was 8 hours (Anglen 1981; Rotman et al. 1983). These studies also included sensitive individuals: an atopic subject in the study by Rotman et al. (1983), subjects showing methacholine hyperresponsiveness in the study by D'Alessandro et al. (1996), and subjects exhibiting seasonal allergic rhinitis (Shusterman et al. 1998). Also of significance is the fact that Rotman et al. (1983) reported that exposure to 1 ppm for 8 hours induced greater changes in pulmonary function tests than exposure to the same concentration for 4 hours, suggesting that the response was related to duration in addition to concentration. Given this information, an acute-duration inhalation MRL for chlorine can be derived by duration adjustment of the no-observed-adverse-effect level (NOAEL) of 0.5 ppm for continuous exposure (0.5 ppm x 8 hours/24 hours = 0.167 ppm) (8 hours was the longest period of exposure for which there is information). Although sensitive individuals were tested in some of these studies, the

number of individuals tested at the region of the NOAEL (0.4–0.5 ppm) was small. Therefore, an uncertainty factor of 3 is used to account for sensitive populations. The resulting acute-duration inhalation MRL for chlorine is 0.06 ppm (0.167 ppm/3).

• An MRL of 0.002 ppm has been derived for intermediate-duration inhalation exposure (15–364 days to chlorine gas.

No human studies were available that could serve as the basis for derivation of an intermediate-duration inhalation MRL. The animal database for intermediate-duration exposure to chlorine is limited to two studies. In one study, male and female F344 rats were exposed to 0, 1, 3, or 9 ppm chlorine 6 hours/day, 5 days/week for 6 weeks (Barrow et al. 1979). In the other study, male and female F344 rats were exposed to 0, 0.5, 1.5, or 5 ppm chlorine 6 hours/day, 5 days/week for 62 days (Kutzman 1983). Aside from a reduction in final body weight of approximately 11% relative to controls in female rats exposed to 0.5 ppm chlorine (most likely due to reduced food consumption) in the Kutzman (1983) study, the most sensitive target for chlorine exposure was the respiratory tract. Barrow et al. (1979) described inflammation of the nasal turbinates in rats exposed to ≥1 ppm chlorine, whereas loss of cilia and epithelium in the trachea was seen in rats exposed to ≥0.5 ppm in the Kutzman (1983) study. No NOAELs for respiratory effects were established in either study. Since incidences of animals with respiratory lesions were presented in the Kutzman (1983) study, but not in the Barrow et al. (1979) study, the Kutzman (1983) study was selected as the principal study for derivation of an intermediate-duration inhalation MRL for chlorine (more complete descriptions of the end points evaluated and the reported results in these studies can be found in Section 3.2 and Appendix A).

There were no significant exposure-related increases in the incidences of animals with histological lesions in any of the examined tissues with the exception of a loss of cilia in the trachea (Kutzman 1983). The incidences of slight to moderate loss of tracheal cilia were 1/23, 12/23, 4/23, and 13/23 in the 0, 0.5, 1.5, and 5 ppm exposure groups, respectively. Although the incidence for this lesion in the mid-exposure group was not significantly different from the control incidence, a statistically significant (p=0.0055) Cochran-Armitage trend test for these data can be demonstrated. However, when attempts were made to apply dose-response models to the data, no adequate fits of EPA Benchmark Dose Software (BMDS) models to the data were obtained (p-values for chi-square goodness of fit statistics were <0.1). Thus, the LOAEL of 0.5 ppm was used as the point of departure for deriving an intermediate-duration inhalation MRL for chlorine, after it was converted to a human equivalent concentration (HEC) using the EPA cross-species dosimetric methodology (EPA 1994a) for a category 1 gas, as follows:

$$LOAEL_{[HEC]} = LOAEL_{[ADJ]} \times RGDR_{TB}$$

where:

 $LOAEL_{[ADJ]} = 0.5 \text{ ppm x } 6/24 \text{ hours x } 5/7 \text{ days} = 0.09 \text{ ppm and}$ $RGDR_{TB} = \text{ratio of the regional gas dose in rats to that of humans for the tracheobronchial region}$

$$RGDR_{TB} = (VE/SA_{TB})_A / (VE/SA_{TB})_H$$

where:

VE = minute volume (0.137 L/minute for rats, 13.8 L/minute for humans [EPA 1994a]) and SA_{TB} = surface area of the tracheobronchial region (22.5 cm² for rats and 3,200 cm² for humans [EPA 1994a])

 $LOAEL_{[HEC]} = 0.09 \text{ ppm x } (0.137 \text{ L/minute/22.5 cm}^2) / (13.8 \text{ L/minute/3,200 cm}^2) = 0.14 \text{ ppm}$

Applying an uncertainty factor of 90 (3 for extrapolation from animals to humans with dosimetric adjustment, 3 for the use of a minimal LOAEL, and 10 for human variability) to the LOAEL_[HEC] yields an intermediate-duration inhalation MRL of 0.002 ppm for chlorine.

• An MRL of 0.00005 ppm has been derived for chronic-duration inhalation exposure (365 days or more) to chlorine gas.

There is no information regarding chronic-duration exposure of the general population to chlorine because this type of exposure occurs only in occupational settings. There are few studies of chronicallyexposed workers in which there is some documentation regarding exposure levels and in which there is no evidence, at least explicitly mentioned in the studies, of the workers having being subjected to acute episodes of high exposure or "gassing" incidents. One of these studies involved 600 workers from 25 plants producing chlorine subjected to an evaluation of medical and occupational histories, blood and urine tests, pulmonary function tests, and electrocardiogram (Patil et al. 1970). Exposure data were available for 332 workers and showed a time-weighted average (TWA) 8-hour mean of 0.15±0.29 ppm (range, 0.006–1.42 ppm). Evaluation of the 332 workers who had exposure data showed that none of the end points examined (those subjected to recall or measured) showed a dose-response relationship. The mean concentration of 0.15 ppm may be considered a NOAEL for the study, but limitations such as unclear analytical methodology, no clear definition of the case/control populations, and insufficient detail regarding the method of analysis render the NOAEL questionable. A respiratory health assessment of 392 male pulp mill workers exposed predominantly to a mean 8-hour TWA of 0.18 ppm chlorine (other possible exposures included, sulfur dioxide, hydrogen sulfide, and methylmercaptan, in addition to various particulates) found that, relative to a control group, the pulp mill workers complained more frequently of usual phlegm, wheeze without cold, and chest illness (Enarson et al. 1984). However, the most significant finding was that a subgroup of nonsmokers (n=4) had a significantly lower fixed

expiratory flow rate at 25–75% vital capacity (FEF_{25–75%}) and forced expiratory volume in 1 second/forced vital capacity (FEV₁/FVC) ratio than the control workers. Given the small number of workers involved and the possibility of exposure to multiple chemicals, the validity of the 0.18 ppm as an effect level is questionable. An additional issue to consider is that neither one of these studies seemed adequate to detect possible mild alterations in the nasal cavity, a sensitive target of chlorine exposure in humans and animals, as described in Sections 2.2 and 3.2.1.2. Due to the limitations mentioned above, these long-term studies are insufficient for quantitative risk assessment.

There are only two chronic-duration inhalation studies of chlorine in animals. One is a 1-year study in monkeys (Klonne et al. 1987) and the other is a 2-year bioassay in rats and mice (Wolf et al. 1995). Both studies tested similar concentrations of chlorine (up to 2.3 ppm in monkeys and 2.5 ppm in rats and mice) and evaluated multiple end points including respiratory tract histopathology, hematology, and clinical chemistry. In both studies, the upper respiratory tract was the target for chlorine toxicity. In general, lesions were less severe in the monkeys than in rats and mice, but extended more distally in the respiratory tract. In rats and mice, an increased incidence of minimal to moderate alterations occurred with the lowest exposure concentration tested, 0.4 ppm chlorine. In general, the nasal lesions were sitespecific, but the severity and/or incidence were not always concentration-dependent. Lesions observed included respiratory and olfactory epithelial degeneration, septal fenestration, mucosal inflammation, respiratory epithelial hyperplasia, squamous metaplasia, and goblet cell hypertrophy and hyperplasia, and secretory metaplasia of the transitional epithelium of the lateral meatus. For the most part, monkeys exhibited only mild concentration-related respiratory epithelial hyperplasia with focal loss of cilia over the range of concentrations tested (0, 0.1, 0.5, and 2.3 ppm) and showed no evidence of the major nasal lesions seen in rats and mice. These differences are probably related to species-specific respiratory-tract airflow characteristics (Ibanes et al. 1996), which in turn, are determined by anatomical differences. Moreover, rats and mice are obligatory nose breathers with a greater surface-area-to-volume ratio of the upper respiratory tract than primates. Therefore, exposure of rodents and primates to equal concentrations for equal amounts of time will likely result in greater pathological changes in the nasal area of the rodent (Barrow et al. 1979). It appears, therefore, that primates are a better model to evaluate potential respiratory effects in humans than rodents. For these reasons, the study in monkeys (Klonne et al. 1987) was selected for deriving a chronic-duration inhalation MRL for chlorine.

In the principal study, male and female Rhesus monkeys (4/sex/exposure level) were exposed to 0, 0.1, 0.5, or 2.3 ppm chlorine 6 hours/day, 5 days/week for 1 year (Klonne et al. 1987). The only treatment-related histopathological effects consisted of focal epithelial hyperplasia characterized by increased cell

numbers and loss of cilia and goblet cells in the respiratory epithelium of the nose and trachea. The affected areas of the nasal passages showed hypercellularity with loss of goblet cells and cilia. In some of these areas, the nuclei showed altered polarity. Lesions were more frequent on the angular margins of the turbinates and less frequent on the lateral wall or septum adjacent to these margins. In some cases, the respiratory epithelial hyperplasia was associated with mild suppurative inflammatory response. Lesions in the trachea resembled those in the nose, but were less severe and involved only a small circumferential section of the ventral and ventrolateral trachea. The combined incidences of hyperplasia in the nasal epithelium with loss of goblet cells and cilia, characterized as trace and mild in males and females, were 1/8, 3/8, 6/8, and 8/8 in the control, 0.1, 0.5, and 2.3 ppm exposure groups, respectively. The exposure concentration of 0.1 ppm is considered a minimal LOAEL for nasal lesions in monkeys.

Incidence data for nasal lesions in male and female monkeys exposed to chlorine gas (Klonne et al. 1987) were analyzed using the benchmark dose (BMD) approach for MRL derivation (further details of the modeling are presented in Appendix A) (EPA 2008a). Models in the EPA BMDS (version 1.4.1) (i.e., Gamma, Logistic, Log-logistic, Multi-stage, Probit, Log-probit, Quantal linear, and Weibull) were fit to the nasal lesion data to determine potential points of departure for the MRL. A Quantal linear model provided the best fit to the data. From this model, the predicted exposure concentration associated with a 10% extra risk (BMC₁₀) for nasal lesions in monkeys was 0.04 ppm; the lower 95% confidence limit on this concentration (BMCL₁₀) was 0.02 ppm. The monkey BMCL₁₀ served as the point of departure for the chronic-duration MRL, after it was converted to a HEC (BMCL_{10[HEC]}) using the EPA cross-species dosimetric methodology (EPA 1994a) for a category 1 gas, as follows:

$$BMCL_{10fHEC1} = BMCL_{10fADII} \times RGDR_{ET}$$

where:

 $BMCL_{10[ADJ]} = 0.02 \text{ ppm x } 6/24 \text{ hours x } 5/7 \text{ days} = 0.004 \text{ ppm and}$ $RGDR_{ET} = \text{ratio of the regional gas dose in rats to that of humans for the extrathoracic region}$

$$RGDR_{ET} = (VE/SA_{ET})_A / (VE/SA_{ET})_H$$

where:

VE = minute volume 2.1 m³/day for monkeys, calculated using the allometric equation for monkeys in EPA (1988) assuming a body weight of 7 kg for Rhesus monkeys with nasal cavity surface area of 62 cm² (Gross and Morgan 1991); 20 m³/day for humans (EPA 1994a) and

SA_{ET} = 62 cm² surface area of the nasal cavity in Rhesus monkeys weighing 7 kg (Gross and Morgan 1991); 200 cm² for humans (EPA 1994a)

$$RGDR_{ET} = (2.1 \text{ m}^3/\text{day} / 62 \text{ cm}^2) / (20 \text{ m}^3/\text{day} / 200 \text{ cm}^2) = 0.34$$

$$BMCL_{10[HEC]} = 0.004 \text{ ppm x } 0.34 = 0.00136 \text{ ppm}$$

Applying an uncertainty factor of 30 (3 for extrapolation from animals to humans with dosimetric adjustment and 10 for human variability) to the $BMCL_{10[HEC]}$ yields a chronic-duration inhalation MRL of 0.00005 ppm for chlorine.

For the purpose of comparison, using the NOAEL/LOAEL approach would yield a chronic-duration inhalation MRL of 0.00007 ppm for chlorine. This results from duration-adjusting the LOAEL of 0.1 ppm (0.1 ppm x 6/24 x 5/7 = 0.02 ppm) and then multiplying the LOAEL_[ADJ] by the RGDR_{ET} of 0.34 calculated above (LOAEL_[HEC] = 0.02 ppm x 0.34 = 0.007 ppm). Applying an uncertainty factor of 100 (10 for extrapolation from a LOAEL to NOAEL, 3 for animal to human extrapolation using dosimetric adjustments, and 3 for human variability) to the LOAEL_[HEC] of 0.007 ppm would result in a chronic-duration inhalation MRL of 0.00007 ppm for chlorine, which is very close to the MRL calculated by benchmark analysis. If the LOAEL is considered a minimal LOAEL, then the composite uncertainty factor would be 30 and the resulting MRL would be 0.0002 ppm, which is 4 times higher than the MRL calculated by benchmark analysis.

Oral MRLs

Oral MRLs were not derived for hypochlorite solutions for the following reasons. MRLs are derived when reliable and sufficient data exist to identify a target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. Scientifically, as part of having sufficient and reliable data, it is important to be able to see the full, or at least a significant range, of the dose-response curve. In the case of the oral database for hypochlorite solutions, no reliable LOAEL could be identified at levels of hypochlorite that could reasonably be encountered in the environment. It is a matter of policy of ATSDR not to derive free-standing MRLs. A summary of the oral database is presented below.

Earlier acute-duration studies in animals tried to reproduce the lesions to the esophagus and/or stomach due to ingestion of bleach. In most studies, commercial hypochlorite bleach was administered through a tube directly into the esophagus and, in some cases, the distal end of the esophagus was artificially occluded to prolong and control the contact time between the solution and the mucosa (Hook and Lowry 1974; Landau and Saunders 1964; Strange et al. 1951; Yarington 1970). Only three acute modern studies in animals were available. Cunningham (1980) administered 0, 8, 40, or 200 ppm Cl/kg/day (as sodium

hypochlorite) to Wistar rats (10 females/dose group) by gavage in milk for 14 days and reported that this treatment had no significant effect on growth or on the weight of the brain, liver, kidney, or heart. No other end points were evaluated. In a limited scope study, Meier et al. (1985) exposed male B6C3F₁ mice (10/dose group) to 0, 1.6, 4, or 8 mg Cl/kg/day (from sodium hypochlorite or hypochlorous acid) for 5 days and reported that in mice treated with sodium hypochlorite and sacrificed 3 weeks after exposure, there were significant increases in sperm abnormalities at 1.6, 4, and 8 mg Cl/kg/day (not clearly dose-related), but no such increases were seen in mice sacrificed 1 or 5 weeks after exposure. In addition, no increases in sperm abnormalities were seen in mice treated with hypochlorous acid, which the investigators considered "somewhat surprising since hypochlorite should be converted to hypochlorous acid in the acid pH of the stomach." Furukawa et al. (1980) administered sodium hypochlorite in the water to male and female F344 rats for 14 days and reported weight loss at approximately ≥36 mg Cl/kg/day accompanied by marked reductions in water consumption. No histological evaluations were conducted in this study.

Two human studies were located in the available intermediate-duration oral database. A study of limited scope evaluated serum lipid profile and serum levels of thyroid hormones in volunteers who drank 1.5 L/day of distilled water containing 0 or 20 ppm chlorine (0 or 0.4 mg Cl/kg/day based on a mean body weight of 71 kg from the study) for 4 weeks (Wones et al. 1993). No significant deviations from normality were found. In the other study, consumption of water containing 5 mg/L chlorine (approximately 0.036 mg Cl/kg/day) for 12 weeks by 10 volunteers had no significant effect on hematology, serum chemistry, urinalysis, and additional physiological parameters (Lubbers et al. 1982). Since the study did not control for non-experimental ingestion of chlorine by the volunteers, the actual dose of chlorine cannot be estimated, but is likely to have been higher than 0.036 mg Cl/kg/day.

Few intermediate-duration studies in animals were located that examined a wide range of end points following exposure to hypochlorite solutions chlorine. These studies showed that the main effect of exposure to solutions of hypochlorous acid or sodium hypochlorite, particularly at the higher concentrations levels, is a reduction of water intake that is due to taste aversion. The available intermediate-duration oral studies evaluated systemic toxicity (body weight, tissue and organ histopathology, hematology, clinical chemistry) (Abdel-Rahman et al. 1984; Cunningham 1980; Daniel et al. 1990, 1991; Furukawa et al. 1980) and also provided information, albeit limited, on immunological/lymphoreticular (organ weight and histopathology and limited immunocompetence) (Daniel et al. 1990, 1991; Exon et al. 1987), neurological (weight and histopathology of the brain and sciatic nerve) (Daniel et al. 1990, 1991), reproductive (male and female reproductive organ weight and histopathology, fertility,

and sperm parameters) (Carlton et al. 1986; Daniel et al. 1990, 1991), and developmental effects (fetal viability and fetal weight in a gestation exposure study) (Carlton et al. 1986). None of the available studies reported effects that could be attributed directly to chlorine or only reported effects that were considered of unknown toxicological significance. LOAELs were not identified in the intermediate-duration studies, but in one of these studies, rats exposed to 4.1 mg Cl/kg/day (the highest dose tested) for 8 weeks showed a statistically significant reduction in delayed-type hypersensitivity reaction (DTH) to bovine serum albumin, increased prostaglandin E2 synthesis by macrophages, and reduced oxidative metabolism (Exon et al. 1987). The toxicological significance of these findings is unknown.

The highest NOAEL identified in intermediate-duration oral studies is 76 mg Cl/kg/day for reduction in body weight gain in rats dosed with sodium hypochlorite in the drinking water for 13 weeks (Hasegawa et al. 1986). Doses of 152 and 305 mg Cl/kg/day were associated with reductions in final body weight of 19 and 47%, respectively, relative to controls. This study, however, has serious limitations including no reporting of food or water consumption and lack of presentation of data on other end points. Three other 13-week studies are available. Furakawa et al. (1980) administered up to approximately 85 mg Cl/kg/day (from sodium hypochlorite) to F344 rats for 92 days and reported significant reductions in final body weight males at 85 and 50 mg Cl/kg/day (19 and 46%, respectively) and in females at 84 mg Cl/kg/day (30%) relative to controls. This was accompanied by significant reductions of up to 66% in water consumption. For the most part, clinical chemistry hematology tests were unremarkable. Gross necropsy showed bladder abnormalities (no further description) among all groups, while microscopic examination showed endocardial hyperplasia and fibrosis of the myocardium in males and females dosed with 84 mg Cl/kg/day. Daniel et al. (1990) exposed male and female Sprague-Dawley rats to chlorine in the drinking water and evaluated a number of end points including organ histopathology, hematology, and clinical chemistry. Daniel et al. (1991) conducted a similar study in male and female B6C3F₁ mice. In neither study were there significant toxic effects that could be attributed to exposure to chlorine. However, in both studies, water consumption was significantly decreased, particularly in the high-dose groups. In high-dose male (16.7 mg/ Cl/kg/day) and female (24.9 mg Cl/kg/day) rats, water consumption was reduced 36 and 38%, respectively; in high-dose male (34.4 mg Cl/kg/day) and female (39.2 mg Cl/kg/day) mice, water intake was reduced 30 and 20%, respectively. The decrease in water intake, which resulted in dehydration and possibly altered electrolyte balance, could explain the reductions in weight gain and sporadic changes in hematological parameters, clinical chemistry, and organ weights observed in both rats and mice. In the absence of effects that could be clearly attributed to chlorine toxicity, the highest doses tested represent the NOAELs for the studies; no LOAELs were defined.

No chronic-duration human study of exposure to hypochlorous acid or sodium hypochlorite was located; thus, a target for long-term exposure to chlorine in humans has not been identified. Three chronicduration studies were available, two in rats (Hasegawa et al. 1986; NTP 1992) and one in mice (NTP 1992). In the NTP (1992) study, the only treatment-related effect was a reduction in water consumption, particularly at the higher dose levels of chlorine, which, as generally agreed, is due to taste aversion. All three studies evaluated a comprehensive number of end points including hematology and clinical chemistry and tissue and organ histopathology. The highest NOAEL was 133 mg Cl/kg/day for female rats in the Hasegawa et al. (1986) study. However, final body weight in low- and high-dose females at termination (104 weeks) was 11 and 20% lower, respectively, than in controls. Hasegawa et al. (1986) stated that in females, the daily water intake was "somewhat lower" (no data provided) than in the other groups during the first year, but that this trend was not observed during the second year. In addition, the investigators indicated that in males, water intake was comparable among groups, except during the last 20 weeks of the study, during which time water intake was consistently 10–20% higher in the experimental groups than in the controls. The latter seems to be inconsistent with findings in other studies that reported marked reductions in water consumption at lower chlorine concentrations (i.e., Daniel et al. 1990, 1991; NTP 1992). In the NTP (1992) studies, F344 rats received doses of up to 14.4 mg Cl/kg/day and B6C3F₁ mice up to 24.2 mg Cl/kg/day for up to 2 years. No significant alterations attributable to chlorine exposure were noticed in either species for a wide range of end points assessed.

CHLORINE 25

3. HEALTH EFFECTS

3.1 INTRODUCTION

The primary purpose of this chapter is to provide public health officials, physicians, toxicologists, and other interested individuals and groups with an overall perspective on the toxicology of chlorine. It contains descriptions and evaluations of toxicological studies and epidemiological investigations and provides conclusions, where possible, on the relevance of toxicity and toxicokinetic data to public health.

A glossary and list of acronyms, abbreviations, and symbols can be found at the end of this profile.

3.2 DISCUSSION OF HEALTH EFFECTS BY ROUTE OF EXPOSURE

To help public health professionals and others address the needs of persons living or working near hazardous waste sites, the information in this section is organized first by route of exposure (inhalation, oral, and dermal) and then by health effect (death, systemic, immunological, neurological, reproductive, developmental, genotoxic, and carcinogenic effects). These data are discussed in terms of three exposure periods: acute (14 days or less), intermediate (15–364 days), and chronic (365 days or more).

Levels of significant exposure for each route and duration are presented in tables and illustrated in figures. The points in the figures showing no-observed-adverse-effect levels (NOAELs) or lowest-observed-adverse-effect levels (LOAELs) reflect the actual doses (levels of exposure) used in the studies. LOAELs have been classified into "less serious" or "serious" effects. "Serious" effects are those that evoke failure in a biological system and can lead to morbidity or mortality (e.g., acute respiratory distress or death). "Less serious" effects are those that are not expected to cause significant dysfunction or death, or those whose significance to the organism is not entirely clear. ATSDR acknowledges that a considerable amount of judgment may be required in establishing whether an end point should be classified as a NOAEL, "less serious" LOAEL, or "serious" LOAEL, and that in some cases, there will be insufficient data to decide whether the effect is indicative of significant dysfunction. However, the Agency has established guidelines and policies that are used to classify these end points. ATSDR believes that there is sufficient merit in this approach to warrant an attempt at distinguishing between "less serious" and "serious" effects. The distinction between "less serious" effects and "serious" effects is considered to be important because it helps the users of the profiles to identify levels of exposure at which major health effects start to appear. LOAELs or NOAELs should also help in determining whether or not

the effects vary with dose and/or duration, and place into perspective the possible significance of these effects to human health.

The significance of the exposure levels shown in the Levels of Significant Exposure (LSE) tables and figures may differ depending on the user's perspective. Public health officials and others concerned with appropriate actions to take at hazardous waste sites may want information on levels of exposure associated with more subtle effects in humans or animals (LOAELs) or exposure levels below which no adverse effects (NOAELs) have been observed. Estimates of levels posing minimal risk to humans (Minimal Risk Levels or MRLs) may be of interest to health professionals and citizens alike.

A User's Guide has been provided at the end of this profile (see Appendix B). This guide should aid in the interpretation of the tables and figures for Levels of Significant Exposure and the MRLs.

This toxicological profile discusses health effects that result from exposure to chlorine gas. However, ATSDR feels there is a need to inform the public about the role of chlorine in the process of disinfection of water, namely, the production of hypochlorous acid and hypochlorite ion when chlorine is introduced in water. As a way to inform the public, ATSDR has decided to include some information regarding the health effects of hypochlorite solutions in the Profile for chlorine. For chlorine gas, the most important route of exposure is inhalation. For hypochlorous acid and sodium hypochlorite, the most important routes of exposure are oral and dermal.

3.2.1 Inhalation Exposure

3.2.1.1 Death

There is extensive information regarding the lethal effects of exposure to high concentrations of chlorine. Much of the information available is derived from the use of chlorine gas as a chemical weapon at the battle of Ypres, Belgium, during World War I. Approximately 150 tons of chlorine released from 6,000 cylinders killed, by some accounts, 800 soldiers and incapacitated 2,500–3,000 (Joy 1997). In a review of the effects of warfare gases, the U.S. Department of Army (DOA 1933) stated that of the total 70,752 casualties from gas poisoning in the American Expeditionary Forces, 1,843 were gassed with chlorine. A study of 838 of these subjects revealed that 4 deaths were due to later-developing effects of chlorine gassing. The causes of death were broncho-pneumonia, lobar pneumonia, purulent pleurisy, and tuberculous meningitis. In an evaluation of the after effects of chlorine gassing in 700 members of the First Canadian Division, Meakins and Priestley (1919) reported that five deaths had occurred, three

apparently due to an acute pneumonic condition. One death occurred 5 months after exposure to chlorine and, although the exact cause of death was unknown, the symptoms recorded were pain in the left chest, dyspnea, orthopnea, and pronounced cyanosis. The concentration of chlorine to which the soldiers were exposed during the gas attacks is unknown.

DOA (1933) summarized the pathology of chlorine exposure leading to death in 24 hours as follows: an acute inflammation of the trachea and bronchi is followed by congestion and edema of the entire respiratory tract. The edema and consolidation of the lungs lead to acute dilation followed by passive congestion of the abdominal viscera. Acute death is the result of the pulmonary edema and respiratory and cardiovascular failure. Subacute death is generally due to pulmonary infection with resulting bronchitis and pneumonia (DOA 1933). Postmortem findings include: acute conjunctivitis, congestion of abdominal organs (especially the liver), increased lung volume, fluid in the pleural cavity, mottled appearance on lung surface with scattered areas of emphysema, pleural hemorrhage, perivascular edema, and dilation of blood vessels, frothy fluid filling the trachea and bronchi, red mucous membranes, and heart enlargement and dilated heart chambers (especially the right side) (DOA 1933).

There are also reports of deaths due to spills of chlorine gas to the environment following railroad accidents. CDC (2005) reported that in January 2005, a train in South Carolina carrying chlorine tanker cars collided with another train and an estimated 11,500 gallons of chlorine gas were immediately released to the air causing the death of nine persons. The primary cause of death of those who died at the scene was asphyxia (Van Sickle et al. 2009). Reported findings included congested lungs, pulmonary edema, and tracheal and bronchial erythema. CDC (2005) also reported that a similar accident on June 2004 in Texas released approximately 90,000 pounds of chlorine gas resulting in two fatalities among residents near the site. Eight people died as a direct result of lung injury caused by exposure to chlorine following a freight train accident in Florida in February 1978 (Jones et al. 1986). A train accident in San Luis Potosí, Mexico, in August 1981 caused the release of an unspecified amount of chlorine gas, which caused the death of 14 people (Costero and Falcón Escobedo 1983). An 11-monthold infant died after liquid chlorine spilled as a result of a derailment in La Barre, Louisiana, in January 1961 (Joyner and Durel 1962). The child died hours after exposure and the cause of death presumably was massive pulmonary edema. Joyner and Durel (1962) indicate that 7 hours after the accident, levels of 400 ppm chlorine were measured in areas 75 yards from the wreck. Citing an unpublished report, Baxter et al. (1989) stated that the rupture of a chlorine storage vessel in Romania in 1939 caused the death of 68 people.

Dixon and Drew (1968) reported the case of a worker at a factory producing chlorine who was exposed to chlorine gas that leaked from a faulty valve for about 30 minutes and died 3–3.5 hours later; postmortem examination showed that pulmonary edema was the prime cause of death. Adelson and Kaufman (1971) reported the death of two healthy adults 25 and 76 hours after being exposed to chlorine gas that leaked into their home from a nearby water filtration plant while they were sleeping. An autopsy of the earlier death showed congested and edematous lungs whose cut surfaces released substantial amounts of water and frothy liquid on pressure, and injected tracheobronchial mucosa. Postmortem examination of the later death showed a similar picture of the lungs with the additional findings of swollen brain with flattening of convolutions and subarachnoid hemorrhage. Suzuki et al. (2001) reported the case of a worker who inhaled concentrated chlorine gas and died from pulmonary thrombosis 6 days after exposure.

Using information on lethal effects of chlorine in animals and humans from the literature, Withers and Lees (1985a) developed probit equations to derive a revised estimate of the lethal toxicity of chlorine that considers physical activity, particularly inhalation rate, effectiveness of medical treatment, and the form of the lethal toxic load function. The LC₅₀ values for a 10-minute exposure with standard level of activity were estimated at 433, 173, and 364 ppm for the regular, vulnerable, and average population, respectively. For a 30-minute exposure, the corresponding LC₅₀ values were estimated at 250, 100, and 210 ppm, respectively. Prater (1990) indicates that concentrations of 400 ppm can be lethal to humans in 30 minutes and that immediate death follows inhalation of a concentration of 1,000 ppm. Concentrations between 1,000 and 1,200 ppm for 30 minutes are lethal to humans according to a review of earlier data from various American authors by Withers and Lees (1985b).

Even more abundant information exists regarding lethal effects of chlorine in animals. For data regarding work beginning with World War I, the reader is referred to a review by Withers and Lees (1985b). Work conducted at the U.S. Army's Medical Research Laboratory of the Chemical Warfare Service cited by DOA (1933) indicates that acute exposures to concentrations >870 ppm were usually lethal to dogs, whereas concentrations below 656 ppm were rarely fatal. Additional studies also in dogs, also summarized by DOA (1933), indicate that dogs that died within 24 hours of gassing showed severe injury to the mucous membranes of the upper respiratory tract, congestion, and edema of the entire respiratory tract including the peribronchial tissues and the sheaths of the large blood vessels. There was also acute inflammatory reaction of the lungs that developed into pneumonia. In dogs dying 2–5 days after gassing, the most important feature was the inflammatory process, whereas in dogs dying 5–14 days after gassing death was due to pulmonary infection. Since the original sources were not available, these studies in dogs are not listed in Table 3-1.

Weedon et al. (1940) conducted lethality studies in unspecified strains of rats and mice. In a group of eight rats exposed to 1,000 ppm chlorine, the first death occurred in 20 minutes and all were dead in 1.7 hours. The exposure level of 1,000 ppm was the LC₅₀ in 53 minutes, and 250 ppm was the LC₅₀ in 440 minutes. Rats that died immediately after exposure showed slight brain congestion; lungs distended and hemorrhagic with cut surfaces wet and foamy; distended heart; liver congestion; distended and hemorrhagic stomach; distended intestines; and congested kidneys. In mice, the first death occurred in 21 minutes, and all eight were dead in 50 minutes. The 1,000 ppm exposure level was a 28-minute LC₅₀, whereas 250 ppm was a 440-minute LC₅₀. Mice that died immediately after exposure had slight brain congestion; lungs partly collapsed and hemorrhagic; moderately distended heart; liver congestion; distended and hemorrhagic stomach; slightly distended intestines, and congested kidneys.

In lethality studies conducted by Zwart and Woutersen (1988), 5,484 ppm was a 5-minute LC_{50} in Wistar rats, whereas 447 ppm was a 60-minute LC_{50} ; in Swiss-Webster mice, 1,032 ppm was a 10-minute LC_{50} and 516 ppm was a 30-minute LC_{50} .

In a different study in mice, a 10-minute LC_{50} of 302 ppm was calculated in intact mice and 131 ppm in mice that were exposed via a cannula placed in the trachea, suggesting that the nose is an effective scrubber of chlorine (Alarie 1981). In a study in mechanically-ventilated pigs, exposure to 140 ppm chlorine for 10 minutes caused the death of four out of five pigs within 6 hours of exposure (Gunnarsson et al. 1998). Death was caused by cardiovascular failure triggered by a severe mismatching of ventilation and perfusion. Additional information on lethal concentrations of chlorine in various animal species can be found in World Health Organization (WHO 1982).

Lethality was also reported in an intermediate-duration study in rats (10/sex/exposure level) exposed to 1, 3, or 9 ppm 6 hours/day, 5 days/week for 6 weeks (Barrow et al. 1979). Three rats from the 9 ppm exposure level group died before day 30 and had lesions consistent with chlorine exposure (widespread inflammation of the respiratory tract with hyperplasia and hypertrophy of epithelial cells of the respiratory bronchioles, alveolar ducts, and alveoli). The specific times of death were not reported.

The LOAEL values for death in each species and duration category are recorded in Table 3-1 and plotted in Figure 3-1.

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

		Exposure/ Duration/							
Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (ppm)	Less Serious (ppm)		rious (ppm)	Reference Chemical Form	Comments
	TE EXPOS	SURE			_				
Death 1	Rat (Sprague- Dawley)	1 hr				293	(1-hour LC50)	Vernot et al. 1977	
2	Rat (NS)	1 min - 16 hr				1000	(50% killed in 53 minutes)	Weedon et al. 1940	
						250	(50% killed in 440 minutes)		
3	Rat (Wistar)	5-60 min				688	(30-minute LC50)	Zwart and Woutersen 1988	
						1926	(10-minute LC50)		
						5486	(5-minute LC50)		
						447	(60-minute LC50)		
4	Mouse (Swiss- Webster)	10 min				302 N	If (10-minute LC50 followed by 3-hour observation period)	Alarie 1981	
5	Mouse (NS)	16 hr				1000	(50% killed in 28 minutes)	Weedon et al. 1940	
						250	(50% killed in 440 minutes)		

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

(continued)

		Exposure/ Duration/				LOAEL			
a Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (ppm)	Less Serious (ppm)	Seri (J	ious ppm)	Reference Chemical Form	Comments
=	Mouse (Swiss- Webster)	10-30 min				1032 516	(10-minute LC50) (30-minute LC50)	Zwart and Woutersen 1988	
System	ic								
7	Human	4-8 hr	Resp	0.5	(itching and burning nose and throat; all pulmonary function	tered		Anglen 1981	
			Ocular	0.5	1 (eye irritation)				
3	Human	1 hr	Resp	0.4	(transient alteration pulmonary function			D'Alessandro et al. 1996	
В	Human	8 hr	Resp	0.5 M	1 M (changes in pulmor function tests; runn nose; throat burnin	У		Rotman et al. 1983	
			Ocular	0.5 M	1 M (eye irritation)				
10	Human	3 d 6 hr/d	Resp	0.5 M				Schins et al. 2000	NOAEL is for pulmonary function.
11	Human	15 min	Resp	0.5				Shusterman et al. 1998	NOAEL is for nasal airway resistance ar pulmonary peak flov

(continued)

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

		Exposure/				LOAEL		
Key to Figure	Species (Strain)	Duration/ Frequency (Route)	System	NOAEL (ppm)	Less Serious (ppm)	Serious (ppm)	Reference Chemical Form	Comments
12	Human	15 min	Resp		(increased nasal airwa resistance)	у	Shusterman et al. 2003b	
13	Rat (Fischer- 3	10 min 44)	Resp		25 M (50% decrease in respiratory rate, RD50))	Barrow and Steinhagen 1982	
14	Rat (Sprague- Dawley)	2-10 min	Resp	100 M	200 M (slight perivascular edema in the lung)		Demnati et al. 1995	NOAEL and LOAEL are for 2-minute exposure and evaluation 72 hours later.
15	Rat (Fischer- 3	1-10 d 44) 5 d/wk 6 hr/d	Resp		12 M (swelling of the nose a wheezing)	nd	Dodd et al. 1980	
			Ocular		12 M (swelling around the eyes)			
			Bd Wt			12 M (approximately 20% weight loss)		

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

		Exposure/			L	OAEL		
a Key to Figure	Species (Strain)	Duration/ Frequency (Route)	System	NOAEL (ppm)	Less Serious (ppm)	Serious (ppm)	Reference Chemical Form	Comments
16	Rat (Fischer- 34	1-5 d ₄₄₎ 6 hr/d	Resp			9.1 M (erosion and ulceration of olfactory epithelium)	Jiang et al. 1983	
			Bd Wt			9.1 M (13% weight loss on day 5)		
	Rat (Sprague- Dawley)	30 min	Resp		184 M (histological and biochemical evidence of lung injury)		Leustik et al. 2008	
	Rat (Fischer- 34	1-12 hr 14)	Resp	2.5 M	5 M (reduced total sulfydryl content in nasal respiratory epithelium after 6-hour exposure)		McNulty et al. 1983	
	Rat (Sprague- Dawley)	15 min	Resp			1330 M (pulmonary edema and hemorrhage)	Yildirim et al. 2004	
	Mouse (Swiss- Webster)	10 min	Resp		9.3 M (50% reduction in respiratory rate, RD50)		Barrow et al. 1977	

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

(continued)

		Exposure/ Duration/				LO			
a Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (ppm)		Serious ppm)	Serious (ppm)	Reference Chemical Form	Comments
	Mouse (Swiss- Webster)	5 d 6 hr/d	Resp				9.3 M (exfoliation, erosion, ulceration, necrosis of nasal respiratory epithelium)	Buckley et al. 1984	
_	Mouse (Hybrid)	60 min	Resp		3.5 M	(50% reduction in respiratory rate, RD50)		Gagnaire et al. 1994	
	Mouse (Swiss- Webster)	1-5 d 6 hr/d	Resp				9.1 M (erosion and ulceration o the olfactory epithelium)	f Jiang et al. 1983	
			Bd VVt				9.1 M (21% weight loss on day 5)		
	Mouse (Hybrid)	5 min	Resp		100 M	(flattening of pulmonary epithelium)		Martin et al. 2003	
_	Mouse (C57BL/6N)	15 min	Resp		2.3 F	(50% decrease in breathing frequency, RD50)		Morris et al. 2005	
•	Mouse (C57BL/6N)	60 min	Resp		221	(biochemical and histological evidence of lung injury)		Tian et al. 2008	

(continued)

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

		Exposure/				LOAEL		
a Key to Figure	Species (Strain)	Duration/ Frequency (Route)	System	NOAEL (ppm)	Less Serious (ppm)	Serious (ppm)	Reference Chemical Form	Comments
27	Rabbit (NS)	30 min	Resp	50		100 (pulmonary edema followed by emphysema	Barrow and Smith 1975)	
Death 28	Rat	E EXPOSUR	E			9 (death in 3/10 before da	v. Barrow et al. 1979	
	(Fischer- 34	44) 5 d/wk 6 hr/d				30 of exposure)	y Bullow et al. 1070	
System 29	nic Rat (Fischer- 34	6 wk 44) 5 d/wk 6 hr/d	Resp		1 F (inflammation of turbinates)	nasal 9 (erosion of nasal mucosal epithelium)	Barrow et al. 1979	
			Gastro	3		9 (focal erosion of gastric mucosa)		
			Hemato	5	9 (increased hema females and segr neutrophils in ma	mented		
			Hepatic	1	3 (cytoplasmic vac- of hepatocytes)	uolation		
			Renal	3	9 (slight to modera kidney congestio			
			Ocular	1	3 (ocular irritation)			
			Bd Wt	1	3 F (15% decreased weight)	final 9 M (43% decreased final weight)		

(continued)

		Exposure/ Duration/			LC	AEL		
a Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (ppm)	Less Serious (ppm)	Serious (ppm)	Reference Chemical Form	Comments
30	Rat (Fischer- 3	62 d 344 ₎ 5 d/wk 6 hr/d	Resp		0.5 M (loss of cilia and epithelium in the trachea)	5 (severe upper respirator irritation)	ry Kutzman 1983	NOAELs are for organ histopathology.
			Cardio	5				
			Hepatic	5 M				
			Renal	5 M				
			Ocular	0.5	1.5 (occasional signs of eye irritation)	5 (severe eye irritation)		
			Bd Wt		0.5 F (final weight 11% lower than controls)	5 F (weight loss; final weigh 32% lower than controls	t ;)	
lmmun	o/ Lympho	ret						
31	Rat (Fischer- 3	62 d 344) 5 d/wk 6 hr/d		5			Kutzman 1983	NOAEL is for histopathology of spleen and peribronchial lymph nodes.
Neurolo								
32	Rat (Fischer- 3	62 d 344 ₎ 5 d/wk 6 hr/d		5			Kutzman 1983	NOAEL is for histopathology of the brain.
Reprod	uctive Rat	62 d						NOAEL C. C. C.
JJ	(Fischer- 3	62 d 344 ₎ 5 d/wk 6 hr/d		5			Kutzman 1983	NOAEL is for fertility in males and females and sperm morphology.

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

			Table 3-1 Lev	els of Signific	cant Exposure to Chlorine -	Inhalation	(continued)	
		Exposure/				LOAEL		
Key to Figure	Species (Strain)	Duration/ Frequency (Route)	System	NOAEL (ppm)	Less Serious (ppm)	Serious (ppm)	Reference Chemical Form	Comments
CHR(System	ONIC EXF	POSURE						
34	Monkey (Rhesus)	1 yr 5 d/wk 6 hr/d	Resp		0.1 F (minimal nasal epith hyperplasia)	nelial	Klonne et al. 1987	NOAELs are for organ histopathology.
			Cardio	2.3				
			Gastro	2.3				
			Hemato	2.3				
			Musc/skel	2.3				
			Hepatic	2.3				
			Renal	2.3				
			Endocr	2.3				
			Dermal	2.3				
			Ocular	0.5	2.3 (conjunctival irritation	n)		
			Bd VVt	2.3				

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

		Ta	able 3-1 Lev	els of Signific	cant Exposure to Chlorine - Inhalation				(continued)		
		Exposure/					LOAEL				
Key to Figure	Species (Strain)	Duration/ Frequency (Route)	System	NOAEL (ppm)	Les	s Serious (ppm)	Serious (ppm)		erence mical Form	Comments	
35	Rat (Fischer- 3	M: 2 yr, 5 d/wk, 44) 6 hr/d F: 2 yr, 3 d/wk, 6 hr/d	Resp		0.4	(alterations of minims severity in nasal epithelium)	al	Wol	lf et al. 1995	NOAELs are for gross and microscopic pathology of organs and tissues.	
			Cardio	2.5							
			Gastro	2.5							
			Hemato	2.5							
			Musc/skel	2.5							
			Hepatic	2.5							
			Renal	2.5							
			Endocr	2.5							
			Dermal	2.5							
			Ocular	2.5							
			Bd Wt	2.5							

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

		Т	able 3-1 Lev	els of Signific	ant Exp	oosure to Chlorine - Inha	ation	(continued)		
		Exposure/ Duration/				L	OAEL			
Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (ppm)		s Serious (ppm)	Serious (ppm)	Reference Chemical Form		Comments
36	Mouse (B6C3F1)	2 yr 5 d/wk 6 hr/d	Resp		0.4	(minimal to moderate alterations in the nasal epithelium)		Wolf et al. 1995		NOAELs are for gross and microscopic pathology of organs and tissues.
			Cardio Gastro Hemato Musc/skel Hepatic Renal Endocr Dermal Ocular	2.5 2.5 2.5 2.5 2.5 2.5 2.5 2.5 2.5 2.5						
			Bd Wt	2.5						
lmmun 37	no/ Lymphor Monkey (Rhesus)	et 1 yr 5 d/wk 6 hr/d		2.3				Klonne et al. 1987		NOAEL is for histopathology of lymph nodes and spleen.
38	Rat (Fischer- 34	M: 2 yr, 5 d/wk, 44) 6 hr/d F: 2 yr, 3 d/wk, 6 hr/d		2.5				Wolf et al. 1995		NOAEL is for gross and microscopic pathology of lymphoreticular tissues.

3. HEALTH EFFECTS

		Exposure/				LOAEL		
Key to Figure	Species (Strain)	Duration/ Frequency (Route)	System	NOAEL (ppm)	Less Serious (ppm)	Serious (ppm)	Reference Chemical Form	Comments
39	Mouse (B6C3F1)	2 yr 5 d/wk 6 hr/d		2.5			Wolf et al. 1995	NOAELs are for gross and microscopic pathology of immunoreticular organs and tissues.
Neurol 40	ogical Monkey (Rhesus)	1 yr 5 d/wk 6 hr/d		2.3			Klonne et al. 1987	NOAEL is for histopathology of central and peripheral components of the nervous system.
41	Rat (Fischer- 3-	M: 2 yr, 5 d/wk, 44) 6 hr/d F: 2 yr, 3 d/wk, 6 hr/d		2.5			Wolf et al. 1995	NOAEL is for gross and microscopic pathology of central and peripheral components of the nervous system.
42	Mouse (B6C3F1)	2 yr 5 d/wk 6 hr/d		2.5			Wolf et al. 1995	NOAELs are for gross and microscopic pathology of the brain, spinal, and sciatic nerve.
Reprod 43	ductive Monkey (Rhesus)	1 yr 5 d/wk 6 hr/d		2.3			Klonne et al. 1987	NOAEL is for histopathology of reproductive organs and tissues.

	Species (Strain)	Exposure/ Duration/ Frequency (Route)	System	NOAEL (ppm)		LOAEL		
					Less Serious (ppm)	Serious (ppm)	Reference Chemical Form	
44	Rat (Fischer- 3	M: 2 yr, 5 d/wk, 44) 6 hr/d F: 2 yr, 3 d/wk, 6 hr/d		2.5			Wolf et al. 1995	NOAEL is for gross and microscopic pathology of reproductive organs.

(continued)

Table 3-1 Levels of Significant Exposure to Chlorine - Inhalation

b Used to derive an acute-duration inhalation minimal risk level (MRL) of 0.06 ppm; the MRL was derived by adjusting the NOAEL of 0.5 ppm for continuous exposure (0.5 ppm x 8/24) and dividing by an uncertainty factor of 3 to account for human variability.

c Used to derive an intermediate-duration MRL of 0.002 ppm; the MRL was derived by dividing the LOAEL[HEC] of 0.14 ppm by an uncertainty factor of 90 (3 for extrapolation from animals to humans with dosimetric adjustment. 3 for use of a minimal LOAEL, and 10 for human variability).

d Used to derive a chronic-duration inhalation MRL of 0.00005 ppm; the MRL was derived by dividing the BMCL10[HEC] of 0.00136 ppm by an uncertainty factor of 30 (3 for extrapolation from animals to humans with dosimetric adjustment and 10 for human variability).

Bd Wt = body weight; Cardio = cardiovascular; d = day(s); Endocr = endocrine; F = Female; Gastro = gastrointestinal; Hemato = hematological; hr = hour(s); Immuno/Lymphoret = immunological/lymphoreticular; LC50 = lethal concentration, 50% kill, LOAEL = lowest-observed-adverse-effect level; M = male; min = minute(s); Musc/skel = musculoskeletal; NOAEL = no-observed-adverse-effect level; NS = not specified; occup = occupational; ppm = parts per million; RD50 = 50% decrease in respiration rate; Resp = respiratory; wk = week(s); yr = year(s)

a The number corresponds to entries in Figure 3-1.

Figure 3-1 Levels of Significant Exposure to Chlorine - Inhalation

Acute (≤14 days)

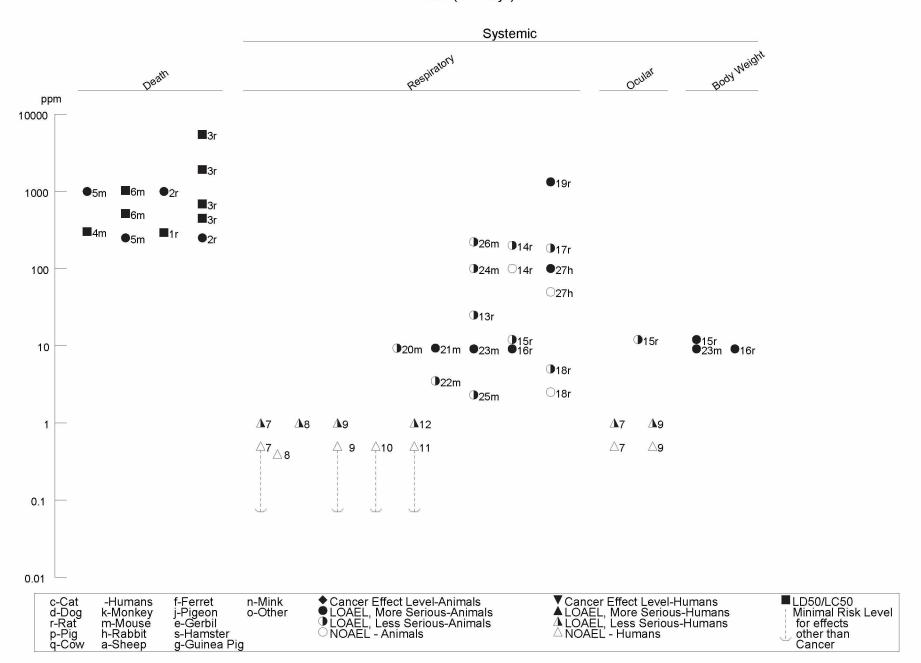


Figure 3-1 Levels of Significant Exposure to Chlorine - Inhalation (Continued)
Intermediate (15-364 days)

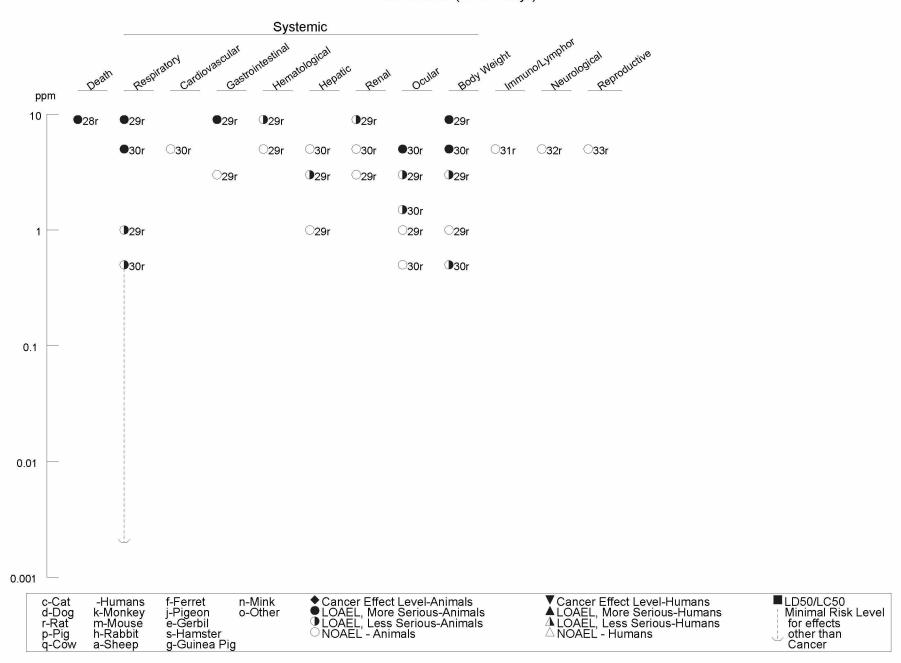


Figure 3-1 Levels of Significant Exposure to Chlorine - Inhalation (Continued)

Chronic (≥365 days)

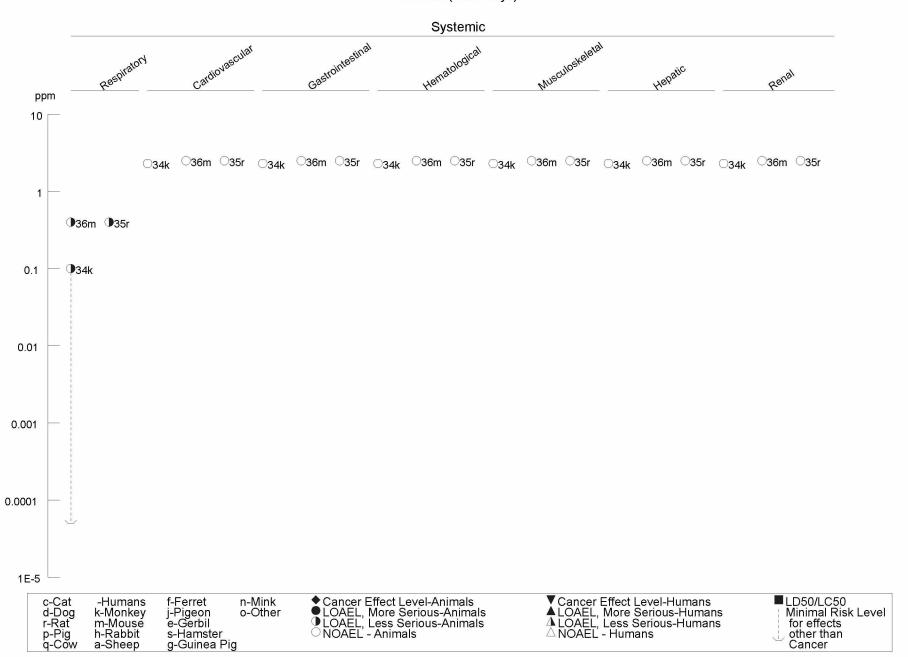
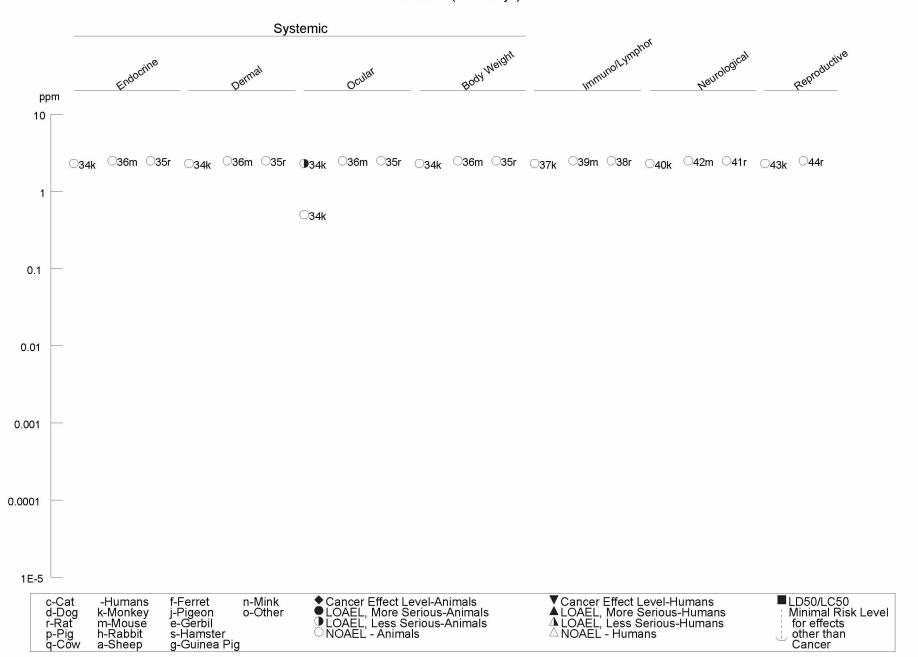


Figure 3-1 Levels of Significant Exposure to Chlorine - Inhalation (Continued)

Chronic (≥365 days)



3.2.1.2 Systemic Effects

The highest NOAEL and all reliable LOAEL values from each study for systemic effects in each species and duration category are recorded in Table 3-1 and plotted in Figure 3-1.

Respiratory Effects. The respiratory system, particularly the upper portion of the respiratory system, is the target for exposure to chlorine gas, and there is a considerable amount of information to support this observation. Chlorine is a respiratory irritant and its effects depend on its concentration, as well as the duration of exposure and the water content of the tissue involved.

Table 3-2 lists clinical effects associated with exposure concentrations compiled from epidemiological and toxicological studies by Ellenhorn and Barceloux (1988). Whether acute exposure to high levels of chlorine induces long-term respiratory effects is still a matter of debate. As mentioned below, some studies have reported a clinical picture of persistent airflow obstruction as well as increased bronchial reactivity, while many others have not found significant persistent alterations. There are many factors that can determine whether residual effects are detected, including exposure level and duration of exposure, medical treatment following exposure, length of the follow-up, underlying respiratory disease, and smoking status. The preponderance of the evidence suggests that no serious long-term respiratory alterations result from acute exposure to low to moderate (up to approximately 20 ppm) concentrations of chlorine gas.

Effects caused by several types of exposures are summarized below including exposure to high concentrations as it occurred during gassing attacks in World War I, accidental exposures of the general population, occupational exposures, and acute, low-level, controlled exposure in volunteers. Information on odor perception is also summarized. Due to the high volume of studies in some of these categories, representative examples are presented.

Odor Perception. A study in which a trained odor panel of four members was exposed to chlorine under controlled laboratory conditions reported that the first concentration at which all members cold detect the odor was 0.314 ppm (Leonardos et al. 1968). Ryazanov (1962) reported that the odor threshold of a group of volunteers was 0.3–0.4 ppm. In a study by Rupp and Henschler (1967), chlorine was slowly introduced to a test room so that the concentration increased from 0 to 1.3 ppm in a 50-minute period.

Table 3-2. Acute Effects of Chlorine Exposure on the Respiratory Tract of Humans

Concentration (ppm)	Effects			
0.2–3.5	Odor detection			
1–3	Mild mucous membrane irritation tolerated up to 1 hour			
5–15	Moderate irritation			
30	Immediate chest pain, dyspnea, cough			
40–60	Toxic pneumonitis and pulmonary edema			
430	Lethal over 30 minutes			
1,000	Lethal within minutes			

Source: Ellenhorn and Barceloux 1988

The odor of chlorine was first detected at 0.06 ppm and all the subjects (number not specified) could smell 0.2 ppm chlorine. In a summary of information on odor perception, NIOSH (1976) states that Stayzhkin reported a threshold of 0.2 ppm for chlorine in a group of 12 subjects who were asked to discern clean air from chlorine by inhaling from two tanks and that Beck found that subjects who were exposed to increasing concentrations of chlorine the length of time of perception was positively related to the exposure concentrations (i.e., the perception of higher concentrations lasted longer than that of low concentrations suggesting that tolerance to low chlorine concentrations may develop). In general, there seems to be consensus that the mean odor threshold lies between 0.2 and 0.4 ppm, which is between 1/25 and 1/50 of the concentration considered to represent immediate danger to life and health of 10 ppm (NIOSH 2005). This means that individuals with a normal sense of smell should be able to take appropriate avoidance measures before serious health effects due to chlorine exposure can occur.

War Exposures. A review of the effects of chlorine as a result of its use during World War I indicates that subjects exposed to very high concentrations (not specified, but presumably >100 ppm) experienced burning of the throat, cough, and feeling of suffocation, dyspnea, and usually death within 24 hours of acute pulmonary edema (DOA 1933). Those surviving 48 hours usually recovered, but bronchitis persisted for weeks and, in some cases, there was evidence of pulmonary emphysema. Data summarized by DOA (1933) on studies that evaluated residual effects indicate that bronchitis and asthma were common months to years after gassing. Dyspnea and chest pain after exercise were also frequent complaints. Citing a study of 838 U.S. soldiers who had been subjected to chlorine gassing, DOA (1933) states that 9 soldiers were discharged due to conditions attributable to gassing; these conditions included pulmonary tuberculosis, bronchitis, pleurisy, and dyspnea. An additional 39 cases were found to have been disabled at the time of discharge, and bronchitis and pleurisy were among the disabling conditions. The extent to which these residual effects were truly due to chlorine gassing is difficult to ascertain due to lack of information on pre-existing conditions, smoking status, and even factors such as the climatic conditions that existed at the time.

Exposures of the General Population. There is considerable information on the effects of exposures of the general population to chlorine derived from various types of situations, including storage tank leaks, railroad accidents, mishandling of bleach cleaning solutions, accidents involving swimming pool chemicals, and accidents in high school chemistry laboratories. Representative examples are summarized below.

Chasis et al. (1947) provided a comprehensive description of immediate and delayed respiratory effects in a group of subjects exposed to chlorine that leaked from a defective cylinder that was carried in a truck in Brooklyn, New York. The accident involved several hundred people. Although the exposure concentration was not known, it was high enough to cause a visible cloud. Chasis et al. (1947) report on 33 persons admitted to a local hospital. Immediate symptoms included rhinorrhea, cough, choking sensation, and substernal burning, pain, and constriction. Substernal pain, burning, and constriction, and choking sensation were present within 2 hours of exposure and, in most patients, subsided completely within 72 hours. Respiratory distress subsided within 3 days in 27 patients and within 6 days in 5 patients. During the first week, almost all 33 patients complained of substernal soreness, which was interpreted as indicating the presence of tracheobronchitis. Physical examination on admission revealed acutely ill patients in moderate to marked respiratory distress, increased respiratory rate of costal abdominal type, and both dry and moist rales. Laboratory data showed sputum with large numbers of epithelial cells showing pronounced degenerative changes. Most cultures showed microorganisms representative of the normal pharyngeal flora. Chest x-rays showed mottling of the lungs and patches of irregular density and differences in the degree of aeration between the two pulmonary fields. Spirometry was conducted on 8 patients 48 hours after exposure and showed markedly reduced vital capacity (VC) and maximal breathing capacity (1 minute); these changes showed improvement in subsequent days. The diagnosis of pulmonary changes was: pulmonary edema, tracheobronchitis, and pneumonia. In 29 patients who were followed for up to 16 months after exposure, there was no evidence of permanent pulmonary disease.

Hasan et al. (1983) reported that exposure of 28 subjects to chlorine that leaked from a storage tank caused cough, dyspnea, and nasopharyngeal irritation. Pulmonary tests conducted 18 hours after exposure showed diminished forced expiratory volume in 1 second (FEV₁), and low forced expiratory flow rate at 50 and 25% vital capacity (FEF₅₀ and FEF₂₅) and FEF_{25-75%}. These abnormalities were still present 14 days after exposure in subjects whose chief initial complaint was dyspnea. Evaluation of nine subjects 5 months after chlorine exposure showed pulmonary parameters within normal limits.

In contrast to the findings of the above two studies, some studies have reported long-term effects of acute high chlorine exposure. For example, Chester et al. (1977) reported the case of a woman who was exposed following a leak in a liquid storage tank and suffered severe cough and chest pain within minutes after exposure. Chest x-rays at the time showed bilateral infiltrates in the midpulmonary zones, but 1 year after the accident x-rays were normal. However, dyspnea and chronic cough with occasional production of white to yellow sputum persisted over the next 4 years. Schönhofer et al. (1996) studied three cases

that experienced nose and throat irritation, cough, shortness of breath, wheezing, chest tightness, and a feeling of suffocation minutes after exposure to chlorine gas that leaked from a tank. Chest x-rays showed no evidence of pulmonary edema. Four months after the accident, bronchoalveolar lavage showed inflammatory changes, but no such changes were seen 16 months later. However, moderate to severe bronchial hyperresponsiveness was observed up to 30 months after the accident. Schönhofer et al. (1996) noted that the condition showed the typical feature of the reactive airways dysfunction syndrome (RADS), defined as an asthma-like occupational illness after an acute exposure to concentrated respiratory irritants characterized by increased responsiveness to methacholine (Brooks et al. 1985).

Weill et al. (1969) studied 12 subjects who were exposed to chlorine after the derailment of a tank car containing 30 tons of liquid chlorine near the town of Morganza, Louisiana. Chlorine concentrations of 400 ppm were measured 75 yards from the wreck 7 hours after the accident. Immediate effects included shortness of breath, cough, and x-rays revealed pulmonary edema. Three and/or 7 years after the accident, all patients were free of respiratory symptoms and pulmonary tests, including total lung capacity (TLC), VC, residual volume (RV), FEV₁, and single-breath diffusing capacity for carbon monoxide (DL_{CO}) were within 2 standard deviations of the predicted value. Weill et al. (1969) concluded that acute exposure to chlorine does not result in significant permanent lung damage. A previous account of this accident was given by Joyner and Durel (1962).

The Agency for Toxic Substances and Disease Registry (1998) conducted a health assessment of a population in Alberton, Montana after tanker cars derailed and released chlorine gas, solid sodium chlorate, and potassium cresylate. Readings at the site of the accident occasionally reached 1,000 ppm, but it was estimated that members of the community had been exposed to up 20 ppm chlorine. Analysis of soil and wipe samples determined that chemicals other than chlorine had not migrated offsite. A total of 682 persons were interviewed within 2 weeks of the accident. The most frequent conditions were cough, nose and throat irritation, and difficulty breathing. The report also indicates that children aged 0–5 years had the highest prevalence of respiratory infections, that persons who had previous respiratory problems (i.e., asthma, chronic bronchitis, hay fever) reported a higher prevalence of respiratory health problems, and that the reported frequencies of respiratory symptoms were higher among current and former smokers than in nonsmokers.

Accidental domestic exposure to chlorine gas can occur when bleach (sodium hypochlorite) is mixed with other cleaning substances that contain an acid, for example, phosphoric acid cleaners (used to remove hard water deposits); the chemical reaction generates chlorine gas. Many such incidents have been

reported. For example, CDC (1991) reported five episodes in which subjects experienced nose irritation, sore throat, chest tightness, cough, and difficulty breathing; in most cases, respiratory symptoms subsided within a day. Gapany-Gapanavičius et al. (1982) reported two similar cases in which subjects inhaled chlorine after mixing bleach and hydrochloric acid and immediately experienced a burning sensation in the throat, cough, and shortness of breath. Chest x-rays taken hours after the accident showed air in the mediastinal space (space in the middle of the chest separating the two pleurae), probably the result of severe coughing, which resolved 7–10 days after the accident. None of these cases were followed to evaluate possible long-term effects. Additional information from similar exposure scenarios can be found in a review by Mrvos et al. (1993).

Chlorine gas can be released around swimming pools when chlorinating agents are handled improperly or due to malfunction of the chlorination equipment, which underscores a strong need for training pool owners and operators in pool maintenance. Pertinent information can be found in CDC (2009).

Sexton and Pronchik (1998) described the effects of such an exposure on 13 children who presented to the emergency department. On admission to the emergency department, most patients complained of throat irritation, chest pain, shortness of breath, wheezing, and chest tightness. Five patients who were admitted to the hospital had normal chest x-rays. At follow-up interviews 2 weeks later, the patients did not complain of residual respiratory symptoms. Ploysongsang et al. (1982) studied four patients who inhaled, for 2-5 minutes, an undetermined amount of chlorine gas that leaked from a container at a public swimming pool and experienced cough, a feeling of irritation of the upper respiratory tract, and tightness in the chest. Pulmonary function studies conducted 12–14 hours after the accident showed values within normal ranges. However, tests done 1 month later showed a significant increase in measurements of volumes, suggesting that there had been an acute reduction of lung volumes after the exposure. Ploysongsang et al. (1982) concluded that exposure to chlorine had produced an insignificant and inconsistent obstruction in large airways. Agabiti et al. (2001) reported the effects of accidental inhalation of chlorine gas among a total of 282 subjects attending a pool. Cough and shortness of breath were common acute symptoms after the accident and 27% reported some respiratory symptoms 15– 30 days after exposure. Lung function measurements at that time revealed a tendency to lower levels among those with the highest perceived exposure, but only a decrease in FEV₁ was significant. The study also found that among children (approximately half the sample), the incidences of all symptoms tended to be higher among those who had a history of chronic respiratory disease, among those who were engaged in physical exercise when the accident occurred, among those who were slow to evacuate the pool, and among those who reported higher exposure (as judged by eye irritation). Also, incidences were higher

among smokers and former smokers than among never smokers. A recent study of 18 children exposed to chlorine in a swimming pool accident found that a biomarker of pulmonary inflammation, leukotriene B₄, was still significantly increased in exhaled breath condensate 2 months after exposure, long after pulmonary function parameters had returned to normal values (Bonetto et al. 2006). Immediately after exposure, the children had experienced dyspnea and burning of the throat and spirometry tests done within the first 24 hours showed reduced forced vital capacity (FVC) and FEV₁. The authors also found that hours after exposure a Clara cell-specific protein, CC16, was significantly elevated in serum compared to non-exposed children, and suggested that this may be interpreted as a sign of injury to the lung epithelial permeability barrier. Transient increases in serum CC16 have been reported following exposure to certain pulmonary irritants (see Lakind et al. 2007 for review). Additional information regarding swimming pool accidents can be found in Almagro et al. (2008), Babu et al. (2008), Grasemann et al. (2007), and Thomas and Murray (2008).

Controlled Low-level Exposure of Volunteers. Anglen (1981) exposed up to 29 male and female volunteers to 0, 0.5, 1, or 2 ppm chlorine for either 4 or 8 hours. Sensations were recorded before and during exposure and pulmonary function was monitored by measuring FVC and FEV₁ before and at various times during exposure. Itching and burning of the throat were the highest responses and were most prevalent by the end of an 8-hour exposure to 1 ppm chlorine. Responses for sensations of itching or burning of the nose and eyes were also prevalent at 1 ppm chlorine. In general, males provided stronger irritation responses than females. Exposure to 1 or 2 ppm chlorine for 8 hours produced significant changes in pulmonary function but similar exposures to 0.5 ppm did not. Exposure to 2 ppm for up to 30 minutes produced no increase in subjective irritation and exposure to 2 ppm for 2 hours did not alter pulmonary function. In another study from the same group of investigators (it is unclear whether a follow-up to Anglen [1981] or a separate study), eight healthy male volunteers exposed to target concentrations of 0, 0.5, or 1 ppm chlorine (Rotman et al. 1983). Pulmonary tests were conducted before exposure, after a 4- and 8-hour exposure period and again 2 and 24 hours after exposure ceased. During exposure, the subjects exercised on a treadmill for 15 minutes of each hour to simulate light-to-moderate work that raised the heart rate to 100 beats per minute. Specific respiratory parameters measured included FVC, FEV₁, forced expired volume in 1 second as percent FVC (FEV₁%), peak expiratory flow rate (PEFR), FEF₅₀ and FEF₂₅, TLC, expiratory reserve volume (ERV), functional residual capacity (FRC), residual volume, airway resistance (Raw), single-breath DL_{co}, closing volume, and difference in nitrogen concentrations between 750 and 1,250 mL of inhaled vital capacity (ΔN_2). Exposure to 1 ppm chlorine caused runny nose and mild burning in the throat, but no such effects were reported at 0.5 ppm. Significant changes in pulmonary function tests were mostly restricted to the 1 ppm exposure level and

were evident after 4 hours of exposure. Changes were observed in FEV_1 , PEFR, FEF_{50} , FEF_{25} , TLC, Raw, and ΔN_2 . Greater changes in some of these parameters were seen after 8 hours of exposure. Few changes were still evident 24 hours after exposure, but most parameters had returned to pre-exposure values by that time. It should be noted that one volunteer who was atopic experienced severe distress during exposure to 1 ppm and was forced to exit the chamber before the full 8-hour period due to shortness of breath and wheezing.

D'Alessandro et al. (1996) evaluated pulmonary function in subjects with (n=10) and without (n=5) airway hyperresponsiveness (HR, defined by baseline methacholine hyperresponsiveness). The HR subjects were exposed to 0.4 or 1.0 ppm chlorine, whereas the healthy subjects were exposed to 1.0 ppm chlorine. All exposures lasted 60 minutes. Airflow and airway resistance were measured immediately before and immediately after exposure. Also, lung volumes, airflow, diffusing capacity, airway resistance, and responsiveness to methacholine were measured 24 hours before and 24 hours after exposure. Exposure of the HR group to 0.4 ppm chlorine resulted in no significant change in airflow or resistance either immediately or 24 hours after exposure. Exposure to 1.0 ppm chlorine resulted in an immediate decrease in FEV₁ and FEF_{25-75%} and increase in airway resistance among normal and HR subjects, but the magnitude of the effects among HR subjects was significantly greater than in healthy subjects. Twenty-four hours after exposure, there were no significant changes for healthy or HR subjects in airflow, lung volumes, diffusing capacity, resistance, or methacholine responsiveness. Comparing relative changes from baseline immediately after exposure between normal and HR subjects showed that HR subjects had much greater changes in pulmonary function tests.

A similar study was conducted in eight volunteers exposed to chlorine 6 hours/day on 3 consecutive days to each of the four exposure conditions, 0, 0.1, 0.3, and 0.5 ppm chlorine (Schins et al. 2000). Pulmonary function including effort-dependent parameters and effort-independent parameters were evaluated before and after exposures. In addition, nasal lavage measurements were performed before and after each exposure and 1 and 4 days after each exposure. The nasal lavage fluid was examined for total cells, epithelial cells, neutrophils, lymphocytes, eosinophils, monocytes, albumin (an indicator of epithelial permeability), and interleukin-8 (indicator of inflammatory response). Subjective complaints by the subjects were judged to be not treatment-related, but objective physiological measures of nasal irritant response were not included. Examination of the nasal lavages gave no indication of an inflammatory response or irritant effects on the nasal epithelium. The results of the pulmonary function tests showed that the only significant effect related to chlorine exposure was a difference in maximal mid expiratory

flow (MMEF) between 0 and 0.5 ppm exposure; however, this was attributed to an unexplained shift in baseline values during control exposure (0 ppm).

Shusterman et al. (2003b) measured nasal airway resistance in 52 healthy adults (24 males and 28 females) before and after exposure to 0 or 1 ppm chlorine for 15 minutes. Subjects were stratified on age (18–34, 35–51, 52–69 years), gender, and allergic rhinitis status (27 were positive). Nasal airway resistance was measured by active posterior rhinomanometry. Exposures to air and chlorine were a week apart. Subjects with allergic rhinitis showed a significantly greater increase in nasal airway resistance (49% increase from baseline) than healthy subjects (10% increase from baseline) 15 minutes after exposure. The increase in nasal airway resistance was most pronounced in older subjects and least pronounced in the youngest group. No significant differences were seen between males and females. In an earlier study, the same group of investigators had reported that subjects with SAR exposed to 0.5 ppm chlorine for 15 minutes experienced a much greater increase in nasal airway resistance than subjects without SAR, as measured by active posterior rhinomanometry (Shusterman et al. 1998). However, when subjective responses to odor, nasal irritation, and nasal congestion were analyzed separately by rhinitis status, no significant exposure-related changes were observed for rhinorrhea, postnasal drip, or headache either on a pool or stratified basis. In addition, within either the SAR or non-SAR group, there was no relationship between subjective and objective congestion after chlorine exposure. Pulmonary peak flow tests showed that none of the subjects exhibited clinically significant changes in peak flow, nor did they complain of cough, wheezing, or chest tightness on chlorine exposure days.

As a whole, these studies indicate that acute-duration exposures to 1 ppm chlorine can induce upper respiratory tract irritation and transient alterations in parameters of respiratory function and exposure concentrations of 0.5 ppm are generally devoid of such effects and, therefore, 0.5 ppm can be considered an acute (1–8 hours) NOAEL for sensory irritation and pulmonary function. The 0.5 ppm level caused no significant effects in an atopic subject (Rotman et al. 1983) or in subjects with airway hyperresponsiveness (D'Alessandro et al. 1996), and, while increased nasal airway resistance measured instrumentally in subjects with SAR, the latter did not clearly perceive the effect as an adverse effect (Shusterman et al. 1998). These studies also show that individuals with compromised respiratory function constitute a susceptible group for exposure to chlorine. The NOAEL of 0.5 ppm and LOAEL of 1 ppm from these studies as a group, serve as the basis for derivation of an acute-duration inhalation MRL for chlorine.

Long-term, Low-level Occupational Exposures. Relatively few studies have examined the effects of long-term exposure to low levels of chlorine in humans, and the ones that have done so have not provided conclusive answers largely because of study limitations.

Patil et al. (1970) studied the health effects of chlorine in 600 workers from 25 plants producing chlorine in North America. A group of 382 workers not considered to be routinely exposed to chlorine served as controls. The average duration of exposure was 11.9 years. Each worker received one physical examination that included evaluation of medical and occupational histories, blood and urine tests, pulmonary function tests and electrocardiogram (EKG). Tobacco and alcohol use were also monitored. The concentration of chlorine was monitored in each plant every 2 months over a period of 1 year in several representative areas, but otherwise unspecified. Exposure data were available for 332 workers and showed a time-weighted average (TWA) 8-hour mean of 0.15±0.29 ppm (range, 0.006−1.42 ppm). It also showed that almost all workers were exposed to <1 ppm chlorine, 94% were exposed to ≤0.5 ppm, and 70% were exposed to ≤0.2 ppm. Evaluation of the 332 workers who had exposure data showed that none of the end points examined (those subjected to recall or measured) showed a dose-response relationship. The mean concentration of 0.15 ppm may be considered a NOAEL for the study, but there are limitations such as unclear analytical methodology, no clear definition of the case/control populations, and insufficient detail regarding the method of analysis that render the NOAEL questionable; thus, it is not included in Table 3-1.

Ferris et al. (1967) examined the prevalence of chronic respiratory disease among 147 workers in a pulp mill and 124 controls who worked in a paper mill and found no significant differences in respiratory symptoms or in tests for FVC and FEV₁ (tests were conducted without a nose clip) between the two groups. Duration of exposure was not provided. Chlorine levels were measured on three different occasions in 3 years; in one occasion, the mean was 7.4 ppm and only traces were reported in the other two occasions. The limit of detection of the method was 1 ppm. Examination of the same cohort 10 years later did not reveal any increased mortality or increased specific cause of death (Ferris et al. 1979). Evaluation of 200 men seen at both times did not reveal any differences in respiratory symptoms or chronic nonspecific respiratory disease.

Enarson et al. (1984) evaluated respiratory effects and pulmonary function in a group of 392 male pulp mill workers exposed to chlorine, sulfur dioxide, hydrogen sulfide, and methylmercaptan, in addition to various particulates (i.e., wood dust, ash, lime dust), for a mean duration of 101.5±86.6 months. A control, unexposed group, consisted of 310 male rail yard workers who lived in the same community and

who performed similar manual labor. End points examined included prevalence of respiratory symptoms (usual cough, usual phlegm, wheezing without a cold, dyspnea when hurrying, chest tightness, and chest illness). Pulmonary function tests conducted included FEC, FEV₁, FEF_{25-75%}, and FEV₁/FVC ratio. Chlorine was the main contaminant in two areas of the pulp mill, the bleach plant and the machine room (mean 8-hour TWA 0.18 and 0.02 ppm, respectively). Overall, pulp mill workers complained more frequently of usual phlegm, wheeze without cold, and chest illness than rail workers. However, the most significant finding was that among bleach workers (n=15) and machine room workers (n=22), nonsmokers (n=4) had a significantly lower FEF_{25-75%} and FEV₁/FVC ratio than rail yard workers. Given the small number of workers involved, the possibility of exposure to multiple chemicals, and the lack of information on chlorine peak exposure levels, the validity of the 0.18 ppm as an effect level is questionable.

A study at a chlorine plant in Sweden compared the changes in vital capacity (VC) and FEV₁ that occurred between measurements separated by 10 years among 44 workers exposed to chlorine and 33 white-collar workers matched for age and smoking status (Hyback 1999). The author stated that the concentration of chlorine was measured continuously over the years and was always below 0.5 ppm, as a set standard. The results of the tests showed that in fact, over the years, VC and FEV₁ declined more in white-collar workers (significantly for FEV₁) than in the workers exposed to chlorine. Hyback (1999) speculated that perhaps the low concentrations of chlorine gas may protect workers from contracting respiratory infections that over time contribute to a decline in respiratory function.

The limited information available does not suggest that long-term exposure to low levels of chlorine gas affects respiratory function, but additional, better-conducted studies are necessary to confirm this view.

High-level Occupational Exposure. Schwartz et al. (1990) studied a group of 20 workers who were briefly (minutes) exposed in a pulp mill to chlorine gas when liquid chlorine leaked from a tank and evaporated. Acute symptoms included burning of the nose and throat, and dry cough with chest tightness. Pulmonary tests were conducted within 24 hours of exposure and several times over the next 12 years. The most significant findings were a high prevalence of airflow obstruction (FEV₁/VC ratio <65%) that persisted over the observation period and a prevalence of low residual volume (RV) that increased during the follow-up period. Schwartz et al. (1990) also found that 5 of 13 subjects tested at year 12 had increased airway reactivity to inhaled methacholine. While the findings were suggestive of long-term pulmonary complication, the investigators acknowledged that without pre-exposure pulmonary function

tests and individual measures of exposure, it is difficult to determine whether the changes were due to chlorine exposure.

Moulick et al. (1992) evaluated 82 patients exposed to approximately 66 ppm chlorine that leaked from a storage tank at a chemical factory in Bombay, India. Acute symptoms of exposure included dyspnea, cough, and irritation of the throat. Pulmonary tests performed in 62 cases within 48 hours of exposure indicated obstruction in 17 cases, restriction in 2 cases, and a mixed pattern in 33 cases. Also, bronchoscopy showed tracheobronchial mucosal congestion and hemorrhagic spots. Four out of 16 patients who were followed for 1 year showed persistent cough 4–6 weeks after exposure, but after 1 year, there were no residual symptoms and x-rays and pulmonary function tests were normal. Evaluation of five nonsmoking patients 3 years after exposure did not reveal any residual symptoms and pulmonary function tests were normal (tests were not specified).

Lemière et al. (1997) reported the case of a nonsmoking worker at a water-filtration plant man who was exposed to chlorine levels high enough to induce immediate burning of the nose and throat and retrosternal burning and wheezing. Five years earlier, he experienced similar symptoms after chlorine inhalation, but the symptoms had been transient. Two days after exposure, FEV₁ was significantly reduced (66% of predicted) and the response to methacholine provocation was slightly abnormal. A bronchial biopsy showed almost complete replacement of the epithelium by a fibrinohemorrhagic exudate. Subsequent biopsies taken over a 5-month period showed considerable epithelial desquamation 15 days after exposure followed by signs of regeneration 5 weeks after exposure and considerable improvement 5 months after exposure, although an inflammatory infiltrate was still present. The airway responsiveness to methacholine paralleled the inflammatory changes, but could be significantly improved by inhaled steroids.

Kowitz et al. (1967) described the effects of chlorine exposure that occurred when a cylinder containing chlorine that was being unloaded from a freighter leaked. Neither exposure concentration nor exposure duration was available. At least 150 men were involved and almost all experienced acute symptoms. Eleven of 17 subjects who were admitted to a hospital were evaluated over a 3-year period. All showed respiratory distress on admission; other common signs included rales, wheeze, or rhonchi, or both, and pulmonary edema. Pulmonary function testing conducted over the 3-year evaluation period showed a persistent decrease in lung volume and diffusing capacity and increased airway resistance. According to Kowitz et al. (1967), these alterations were compatible with an alveolar-capillary injury.

There are also a number of studies that describe the effects of occasional occupational exposures to elevated concentrations of chlorine over a background of low level exposure ("gassing" incidents). Some examples are summarized below.

In a study of 321 workers at a British Columbia pulp mill, 189 reported one or more gassing incidents, although no data were available to determine the severity of the exposure incidents (Kennedy et al. 1991). The average chlorine levels in the pulp mill were <1 ppm, but estimates of concentrations during the gassing incidents are not available. Pulp mill workers who reported having being gassed were significantly more likely to report wheezing on occasion than individuals who had not been gassed. While there were no significant differences in lung function tests between the overall pulp mill group and a control group, nonsmoking and formerly smoking pulp mill workers who reported being gassed had significantly lower MMEF and FEV₁/FVC ratio than those who had not been gassed. Kennedy et al. (1991) hypothesized that the first accidental high exposure incident may cause an inflammatory reaction in the small airways that does not completely resolve because of continuous or repeated presence of the stimulus. A longitudinal analysis over a 7-year period (1981–1988) of 67 pulp mill workers that underwent pulmonary function tests both in 1981 and 1988 reported that there was a significantly greater decline in FEV₁/FVC ratio and MMEF in the gassed group than in the unexposed group matched for age and smoking status (Salisbury et al. 1991).

In a survey of 281 construction workers involved in renovation work at a pulp mill, 257 workers reported an average of 24 gassing episodes over a 3–6-month period (Courteau et al. 1994). Exposure data during the gassings were not available, but 36% of 483 air samples taken in the bleach plant after the episodes showed chlorine concentrations of <0.5 ppm, 58% were between 0.5 and 8 ppm, and 6% were >8 ppm. The most commonly reported symptoms were irritation of the throat (78%), cough (67%), and shortness of breath (54%); the latter was not associated with age, smoking state, or history of asthma or chronic bronchitis. Over 60% of the workers described a flu-like syndrome that lasted for an average of 11 days. Visits to the first aid department were associated with greater reporting of most symptoms including dyspnea and cough. Seventy-one workers identified as having moderate to high risk based on exposure data and onset of respiratory symptoms were evaluated 18–24 months later (Bhérer et al. 1994). At this time, a questionnaire completed by 64 workers (90%) suggested that 58 (91%) still had respiratory symptoms and 51 underwent spirometry and methacholine challenge tests. Bronchial obstruction (FEV₁ <80% predicted) was observed in 16 workers, whereas 29 showed significantly increased airway responsiveness. Based on the fact that workers who had been to an emergency room were more likely to be left with airway hyper-responsiveness, Bhérer et al. (1994) speculated that severity of one or more

gassing episodes may be a more significant determinant of the likelihood of developing permanent alterations than the number of episodes.

In a study of 300 pulp and paper workers in New Hampshire, 105 reported ever having experienced an episode of high exposure to chlorine gases (Henneberger et al. 1996). Spirometry (FEV₁, FVC, FEV₁/FVC, MMEF) showed that the prevalence of obstructive pattern was >3 times greater for the gassed subjects compared with the others. For workers who had 26-pack years of cigarette smoking, an obstructive pattern (abnormally low FEV₁ and FEV₁/FVC) was observed only among those with a history of gassing. In addition, the combination of high cigarette smoking and gassing had a greater-than-additive effect on obstruction. Monitoring data were not available in this report.

Gautrin et al. (1999) evaluated 239 workers in a metal production plant who, 3 years earlier, had taken part in a cross-sectional study (Gautrin et al. 1995). In the first evaluation, the authors reported that FVC was higher in workers who had no symptoms after a gassing episode compared with those who had mild symptoms. Also, FEV₁ and FVC were significantly lower in workers who experience >10 gassing incidents with mild symptoms than in workers who experienced no symptoms. In both cases, the differences were significant only in workers who had ever smoked. Increased airway responsiveness was also found in subjects who experienced >10 gassing incidents with mild symptoms. In 98% of the accidental exposures, chlorine was reported as the gas involved. Among 211 workers seen at follow-up (Gautrin et al. 1999), heavy smokers showed a decrease in FEV₁/FVC% that was predicted by the number of gassing episodes causing mild symptoms between the two evaluations. Nineteen workers showed increased airway responsiveness which was associated with accidents reported to the first aid unit during the previous 2 years. The same group of investigators also reported that chronic rhinitis reported in the second assessment was significantly associated with acute exposure to chlorine and that chronic lower airway symptoms were more frequent in the second assessment among workers reporting chronic rhinitis on both assessments than in others (Leroyer et al. 1999).

Effects in Animals. The studies in animals support the findings in humans, particularly the nature of the acute effects of exposure to high concentrations of chlorine gas. A great deal of information regarding the toxicity of chlorine in animals was generated during World War I and the years that followed triggered by the use of chlorine as a chemical weapon during the war. A summary of the earlier studies conducted in various European countries as well as in the United States and Russia can be found in Withers and Lees (1985b). Additional information on the early literature, particularly from extensive studies in dogs, can be found in DOA (1933).

More recently, studies in rodents have confirmed the earlier observations regarding high exposures and have provided valuable information regarding the irritant properties of chlorine.

Acute exposure to low-to-moderate concentrations of chlorine induces a reduction in the respiratory rate, a protective reflex response mediated by stimulation of trigeminal nerve endings in the nasal mucosa. The concentration of the chemical that induces a 50% decrease in respiratory rate is termed RD_{50} . For example, RD_{50} values of 9.3 and 3.5 ppm were determined in mice exposed for 10 and 60 minutes, respectively (Barrow et al. 1977; Gagnaire et al. 1994). An RD_{50} of 25 ppm was determined in male F344 rats exposed to chlorine for 10 minutes (Barrow and Steinhagen 1982). This study also demonstrated the development of tolerance to chlorine since in rats pre-exposed to chlorine at 1, 5, or 10 ppm intermittently for 32 weeks, the RD_{50} values were 90, 71, and 454 ppm, respectively, measured 16–24 hours after the last day of pre-exposure. Barrow and Steinhagen (1982) speculated that the mechanism of tolerance may involve reactions of chlorine with sulfhydryl groups in the receptors or that chlorine exposure may damage the free nerve endings in the respiratory nasal mucosa. Rats pre-exposed to chlorine also developed cross-tolerance to formaldehyde (Chang and Barrow 1984). Interestingly, rats pre-exposed to 15 ppm formaldehyde did not develop tolerance to formaldehyde, but did develop cross-tolerance to chlorine, which suggested the existence of different reactive sites for the two gases (Chang and Barrow 1984).

A study by the same group of investigators examined the effects of chlorine on lung -SH content and on the enzymes that maintain non-protein -SH levels, glucose-6-phosphate dehydrogenase (G6PD) and glutathione reductase (GSSG-RED) in rats exposed to 0 or 12 ppm chlorine for up to 2 weeks and sacrificed at various times after cessation of exposure (Dodd et al. 1980). The results showed no significant alterations in lung protein -SH, non-protein -SH, G6PD, or GSSG-RED in rats sacrificed immediately after 1, 5, or 10 days of exposure. Rats sacrificed 3 or 6 days after exposure showed an increase in lung -SH, G6PD, and GSSG-RED. These parameters returned to control values after 10 days of recovery. The investigators concluded that the increase in lung -SH and enzymatic activities observed during the recovery periods may reflect reparative processes subsequent to damage induced by chlorine. A different study by the same group showed that exposures to up to 10 ppm chlorine for 12 hours did not alter the total sulfhydryl content (TSH) of the olfactory mucosa but lower concentrations did reduce TSH in the respiratory mucosa, suggesting that inhaled chlorine can oxidize tissue sulfhydryl groups at the point of entry, but not at deeper regions of the respiratory tract (McNulty et al. 1983). McNulty et al. (1983) also found that exposure to 5 ppm for 6 hours or 10 ppm for 3 hours (concentration times

exposure=30) produced similar reductions in TSH, but exposure to 2.5 ppm for up to 12 hours did not significantly affect TSH content. The investigators speculated that a threshold concentration may be needed to overwhelm the protective mechanism in the respiratory mucosa (perhaps the mucociliary flow) allowing chlorine to penetrate deeper into the underlying tissue.

Acute studies also have examined respiratory function in animals.

Barrow and Smith (1975) evaluated inspiratory-expiratory flow rate ratios (Vi/Ve) and volume-pressure relationships (lung compliance) in rabbits exposed to 0, 50, 100, and 200 ppm chlorine for 10 minutes. The tests were conducted 0.5 hours after exposure and after 3, 14, and 60 days without exposure. After the last test, the rabbits were killed and the lungs were removed for gross and microscopic examination. Rabbits exposed to 50 ppm showed mild pneumonitis, which was also observed in control animals; this exposure level did not induce significant changes in air flow ratios, but transiently decreased lung compliance. Exposure to 100 or 200 ppm induced transient concentration related increases in Vi/Ve and a decrease, followed by an increase, in pulmonary compliance; these changes are related to gross signs of pulmonary edema and microscopic changes characterized by chronic pneumonitis and anatomic emphysema. Rabbits allowed to recover for 14 or 60 days showed no "specific airway pathology."

In another study, mice exposed for 15 minutes to 0.8, 2, 3.1, or 3.8 ppm chlorine showed concentration-related decreases in respiratory frequency and increases in specific airway resistance (Morris et al. 2005). Pretreatment with atropine did not alter the increase in airway resistance, suggesting that this response does not involve parasympathetic cholinergic endings. However, pretreatment with capsaicin, a sensory nerve toxin, dramatically reduced respiratory irritation and the obstructive response, suggesting the involvement of sensory nerves. Mice exposed to much higher concentrations of chlorine (100–800 ppm) for 15 minutes showed increased airways resistance and increased responsiveness to methacholine and microscopic examination of the lungs showed flattening of the epithelium and epithelial cell loss and changes associated with oxidative stress (Martin et al. 2003). Since the increased responsiveness to methacholine could be prevented by inhibition of nitric oxide synthase, it appeared that nitric oxide (NO) production may have contributed to the airway response to inhaled chlorine.

Jiang et al. (1983) studied the time course of the histopathological alterations in the respiratory tract of rats and mice exposed to the RD_{50} of 9.1 ppm chlorine 6 hours/day for 1–5 days. The animals were killed immediately after the last exposure and the nose, larynx, trachea, and lungs were processed for microscopic examination. In both species, lesions were seen in the nasal passages with less severe

changes in the nasopharynx, larynx, trachea, and lungs. The lesions in the nasal passages involved both the olfactory and respiratory epithelia; the nasal squamous epithelium showed minimal change. Lesions in the respiratory epithelium included acute epithelial degeneration with epithelial cell exfoliation, erosion, and ulceration after 1 and 3 days of exposure, infiltration by neutrophils after 3 and 5 days, and squamous metaplasia after 5 days. After 3 and 5 days of exposure, the epithelial lesions and inflammatory response found in the respiratory and olfactory mucosa became progressively more severe and extended more posteriorly. The most severe changes occurred in the olfactory mucosa of the anterior portion of the dorsal meatus, which showed extensive epithelial erosion and ulceration. Less severely affected areas showed necrosis and variable loss of sensory cells. Scanning electron microscopy showed loss of olfactory cilia and cellular exfoliation. The larynx and trachea showed acute degeneration of respiratory epithelial cells, whereas the lungs showed moderate to severe peribronchiolitis. Exposure of mice to the RD_{50} of 9.3 ppm for 5 days induced severe exfoliation, erosion, ulceration, and necrosis of the nasal respiratory epithelium (Buckley et al. 1984). It also induced minimal inflammation and squamous metaplasia. In the olfactory epithelium, chlorine induced severe ulcerations and necrosis and degeneration of sensory nerve endings. In the trachea, the lesions ranged from mild to moderate epithelial exfoliation, hyperplasia, and squamous metaplasia. In the lungs, chlorine induced a moderately severe terminal bronchiolitis with occlusion of affected bronchioles by serocellular exudate. Mice killed 72 hours after the last exposure had reduced inflammation and exudate, but there was little difference in the extent of ulceration and degenerative lesions, suggesting that the 3-day period was insufficient for complete repair of the lesions.

Exposure of rats to much higher concentrations of chlorine (50–1,500 ppm for 1–10 minutes) induced increasing lung damage that partially resolved within 24 hours of exposure (Demnati et al. 1995). In rats exposed to 1,330 ppm chlorine for 15 minutes, pulmonary changes observed 45 days after exposure included interstitial fibrosis and thickening of the alveolar septa due to thickening of the basement membrane (Yildirim et al. 2004). Exposure of rats to ≥184 ppm chlorine for 30 minutes induced hypoxemia, respiratory acidosis, and hypercapnia, and increased the surfactant surface tension and protein content of the bronchoalveolar lavage fluid, suggesting alveolar epithelial injury (Leustik et al. 2008). Less evidence of lung injury was seen in rats that were injected before exposure to chlorine with a mixture of ascorbate, deferoxamine, and N-acetyl-L-cysteine, suggesting that damage could be ameliorated by low-molecular antioxidants (Leustik et al. 2008). Exposure of mice to 221–289 ppm chlorine for 60 minutes caused severe lung inflammation as evidenced by widespread neutrophil influx into the lung parenchyma 6 hours after exposure followed by a clustering of neutrophils around damaged

airways 24 hours after exposure (Tian et al. 2008). Histologically, chlorine caused massive sloughing of the airway epithelium that was evident 6 hours after exposure.

Two intermediate-duration studies in animals were located.

Barrow et al. (1979) evaluated the respiratory response in F344 rats exposed to 0, 1, 3, or 9 ppm chlorine 6 hours/day, 5 days/week for 6 weeks. Nasal discharge was seen occasionally in rats exposed to 1 ppm, but was common in rats exposed to 3 and 9 ppm. Respiratory difficulty was also apparent in some rats exposed to 9 ppm. At termination, gross necropsy revealed accumulation of inflammatory reactions in the upper nasal passages in rats exposed to 3 and 9 ppm chlorine. Microscopic evaluations showed indications of inflammatory reactions in the upper and lower respiratory tract of high-dose males and females. The nasal turbinates showed mucopurulent inflammation with secretory material and erosions of the mucosal epithelium. Changes in the trachea and bronchi consisted mostly of hyperplasia of the epithelial lining and inflammatory reactions. The alveolar sacs contained macrophages and secretory material and epithelial cells showed necrosis, hypertrophy and hyperplasia. Alterations in rats exposed to 1 and 3 ppm were less extensive and were limited to focal mucopurulent inflammation of the nasal turbinates in females. Males exposed to 1 or 3 ppm showed deeper pulmonary changes consisting of slight to moderate inflammatory reaction around the respiratory bronchioles and alveolar ducts, increased alveolar macrophages, and isolated areas of atelectasis (incomplete expansion). A LOAEL of 1 ppm for respiratory effects can be defined in this study based on the presence of inflammatory changes in the nasal turbinates of females and in the lungs of males; no NOAEL was established.

A similar study examined clinical signs, lung function, and histopathology of the nasal turbinates and lungs from F344 rats exposed to 0, 0.5, 1.5, or 5 ppm chlorine 6 hours/day, 5 days/week for 62 days (Kutzman 1983). Pulmonary function tests (plethysmograph-based assessment of multiple end points, including lung and tidal volumes, breathing frequency, transpulmonary pressure, lung compliance, N₂ washout, diffusing capacity for CO, maximum expiratory flow volume, peak expiratory flow, and airway resistance) were conducted in 21–24 anesthetized males 6 hours after the last exposure. Respiratory tissues from these rats were prepared for histopathology. The lungs from some of these rats were also examined for collagen, elastin, total protein, and DNA. Exposure to 5 ppm cause severe upper respiratory irritation; exposure to 1.5 ppm showed occasionally less severe signs of irritation, whereas exposure to 0.5 ppm caused no obvious signs of irritation or discomfort. The tests of pulmonary function did not reveal marked abnormalities. The most significant effect was a reduction in airflow at 75% of exhaled vital capacity in all exposed groups, indicating some degree of small airway involvement. There were no

histopathological alterations in the lungs and nasal turbinates, but there was a tendency in the trachea for loss of cilia and epithelium at 0.5 and 5 ppm chlorine. The lung biochemistry only showed an increased collagen concentration at 1.5 and 5 ppm. Based on upper respiratory irritation and loss of cilia and epithelium in the trachea, the exposure level of 0.5 ppm can be defined as a LOAEL for respiratory effects; no NOAEL was defined in this study. This study was used as the basis for derivation of an intermediate-duration inhalation MRL for chlorine.

Two studies have examined the effects of chronic exposure to chlorine on respiratory parameters in animals.

Wolf et al. (1995) exposed groups of F344 rats and B6C3F₁ mice (approximately 70/sex/exposure level) to 0, 0.4, 1, or 2.5 ppm chlorine gas for 2 years. Males from both species and female mice were exposed 6 hours/day, 5 days/week, whereas female rats were exposed 6 hours/day, 3 days/week. The reduced exposure of female rats was based on unpublished data from the investigators that showed female rats to have a greater sensitivity to repeated long-term exposure to chlorine. End points evaluated included gross and microscopic examination of the respiratory tract; the nasal passages were examined microscopically at five different levels. Both in rats and mice, there were no gross lesions attributable to exposure to chlorine, and microscopic evaluation of the respiratory tract showed that chlorine-related effects were restricted to the nasal passages. In the study, the incidences were presented as percentages of all animals for which the nasal passages were adequate for microscopic examination, but the numbers of animals examined were not provided. No lesions were seen in the larvnx, trachea, bronchi, or bronchioles. In general, rats and mice exhibited similar types of lesions. For the most part, the nasal lesions were sitespecific, but the severity and/or incidence were not always concentration-dependent. The majority of the nasal responses exhibited a rostral-to-caudal severity gradient. The lesions rarely extended to the nasopharyngeal meatus. Lesions observed included respiratory and olfactory epithelial degeneration, septal fenestration, mucosal inflammation, respiratory epithelial hyperplasia, squamous metaplasia, and goblet cell (rats only) hypertrophy and hyperplasia, and secretory metaplasia of the transitional epithelium of the lateral meatus. Also observed was intracellular accumulation of eosinophilic proteinaceous material involving the respiratory, transitional, and olfactory epithelia. Lesions were also observed in controls, but the incidences were significantly lower than in the treated groups. One of the lesions with the lowest incidence in controls was goblet cell hyperplasia in female rats (4%); the respective incidences in the 0.4, 1, and 2.5 ppm group were 71, 90, and 91%. In mice, olfactory epithelium atrophy exhibited one of the lowest incidences in controls (3%); the respective incidences in the 0.4, 1, and 2.5 ppm group were 20, 21, and 39%. In both cases, severity also was concentration-related. Based on the increased

incidence of various types of lesions in the nasal passages, the exposure level of 0.4 ppm constitutes a LOAEL for respiratory effects in rats and mice; a NOAEL was not defined.

Klonne et al. (1987) exposed Rhesus monkeys to 0, 0.1, 0.5, or 2.3 ppm chlorine 6 hours/day, 5 days/week for 1 year. Pulmonary diffusing capacity for CO and distribution of ventilation were evaluated monthly during the study. At termination, the nasal tissues (at the first palatine ridge and just posterior to the third, fifth, and seventh palatine ridges), trachea, and lungs were examined. There was no evidence of treatment-related effects on pulmonary function at any interval during the study. The only treatment-related histopathological effects consisted of focal epithelial hyperplasia characterized by increased cell numbers and loss of cilia and goblet cells in the respiratory epithelium of the nose and trachea. The affected areas of the nasal passages showed hypercellularity with loss of goblet cells and cilia. In some of these areas, the nuclei showed altered polarity. Lesions were more frequent on the angular margins of the turbinates and less frequent on the lateral wall or septum adjacent to these margins. In some cases, the respiratory epithelial hyperplasia was associated with mild suppurative inflammatory response. Lesions in the trachea resembled those in the nose, but were less severe and involved only a small circumferential section of the ventral and ventrolateral trachea. The combined incidences of lesions in the nasal mucosa, characterized as trace and mild in males and females, were 1/8, 3/8, 6/8, and 8/8 in the control, 0, 0.1, 0.5, and 2.3 ppm exposure groups, respectively. The lowest exposure concentration of 0.1 ppm chlorine is a LOAEL for nasal lesions in monkeys. This study was used as the basis for derivation of a chronic-duration inhalation MRL for chlorine.

Ibanes et al. (1996) re-examined the respiratory tissues from the chronic studies in monkeys, rats, and mice summarized above to better characterize the lesions and to improve human risk assessments based on these data sets. In general, the re-evaluation found a good correlation between the subjective scores of tissue responses in the original studies and quantitative analyses in the re-evaluation. The investigators also noted that the lesions in monkeys and rodents exhibited both differences and similarities. One notable difference was that monkeys showed less severe lesions, but extended more distally in the respiratory tract. The investigators concluded that respiratory tract airflow characteristics play a major role in lesion distributions in monkeys and rodents. Yet, with appropriate exposure and response adjustments, both rodents and Rhesus monkeys appear to be valid models for human risk assessment.

Cardiovascular Effects. It is not uncommon to find reports of tachycardia and elevated blood pressure in individuals admitted to emergency rooms following accidental exposure to high concentrations of chlorine, but how much of this can be attributed to chlorine exposure or to a stressful

situation is not clear. Some cases in which EKGs were performed shortly after exposure found no significant alterations (Chasis et al. 1947; Güloğlu et al. 2002; Ramachandran et al. 1990). Tachypnea and hypertension were frequently observed in persons who were hospitalized following a train derailment that released chlorine gas in South Carolina in 2005 (Van Sickle et al. 2009). In most persons in whom the conditions were observed, tachypnea and tachycardia persisted across examination. No specific mention of cardiovascular effects was found in studies that evaluated possible long-lasting effects of chlorine exposure.

Rats and mice that were exposed to lethal concentrations of chlorine had distended hearts (Weedon et al. 1940), but little additional information is available regarding cardiovascular effects in animals following exposure to chlorine. It is reasonable to assume that some cardiovascular parameters would be affected after acute exposure to high concentrations of chlorine as a result of the severe ventilation-perfusion mismatching and hypoxia caused by pulmonary edema; however, no evaluations were made in the studies available.

In an intermediate-duration study, rats exposed to 5 ppm chlorine for 62 days had no significant gross or microscopic alterations in the heart (Kutzman 1983). Intermittent exposure of monkeys to 2.3 ppm chlorine for 1 year (Klonne et al. 1987) or of rats and mice to 2.5 ppm chlorine for 2 years (Wolf et al. 1995) did not cause any significant gross or microscopic alterations in the heart and aorta.

Gastrointestinal Effects. Nausea and vomiting are common acute symptoms following exposure to high concentrations of chlorine and are believed to be reflex reactions and not a specific effect of chlorine.

Barrow et al. (1979) reported that rats exposed to 9 ppm chlorine 6 hours/day, 5 days/week for 6 weeks had focal erosion of the gastric mucosa, which was usually accompanied by focal inflammation of the adjacent submucosal areas. No such effects were seen in rats exposed to 3 ppm chlorine. The investigators suggested that general stress or perhaps ingestion of hydrochloric acid and/or hypochlorous acid as a result of grooming or deposition in the oral cavity may have caused the condition. The results of Barrow et al. (1979) could not be confirmed or refuted in a study of very similar design by Kutzman (1983) because the latter did not examine the gastrointestinal tract.

Intermittent exposure of monkeys to up to 2.3 ppm chlorine for 1 year or of rats and mice to up to 2.5 ppm for 2 years did not produce gross or microscopic alterations in the gastrointestinal tract (Klonne et al. 1987; Wolf et al. 1995).

Hematological Effects. Leukocytosis is a common finding in individuals acutely exposed to high concentrations of chlorine, and is most likely a clinical manifestation of acute inflammation and tissue destruction rather than a specific effect of chlorine. Hypoxemia is also characteristic of exposures high enough to cause pulmonary edema and consequently, interference in gas exchange.

Experiments in dogs conducted by Underhill (1920) following World War I showed that following gassing with high concentrations of chlorine, there was a relative increase in red blood cells and hemoglobin that reached a peak approximately 5 hours after exposure and was due to a decrease in intravascular fluid content. He also observed that in dogs exposed to relatively low concentrations of chlorine, there was slight leukocytosis, which was caused solely by an increase in polymorphonuclear cells. An intermediate-duration study in rats reported that exposure to 9 ppm chlorine for 6 weeks induced a significant increase in hematocrit (females) and segmented neutrophils (males and females) (Barrow et al. 1979). No significant changes in hematological parameters were observed in monkeys, rats, or mice exposed to 2.3–2.5 ppm chlorine in chronic duration studies (Klonne et al. 1987; Wolf et al. 1995).

Musculoskeletal Effects. The only information regarding musculoskeletal effects and exposure to chlorine is a report in which a 25-year-old man was diagnosed with myasthenia gravis manifested as laryngeal stridor after accidental exposure to chlorine gas (Foulks 1981). Myasthenia gravis is an autoimmune disorder that causes a reduction in the number and/or sensitivity of acetylcholine receptors at the neuromuscular junction resulting in muscle weakness. Foulks (1981) speculated that the massive exposure of the laryngeal muscles to chlorine may have altered the neuromuscular junction in a way that caused an autoimmune stimulus that resulted in myasthenia gravis.

Exposure of monkeys to up to 2.3 ppm chlorine for 1 year or of rats and mice to up to 2.5 ppm for 2 years did not produce gross or microscopic alterations in skeletal muscle (Klonne et al. 1987; Wolf et al. 1995).

Hepatic Effects. Very limited information was located regarding evaluation of liver end points in patients after acute high exposure to chlorine. Leube and Kreiter (1971) reported that 1 day after exposure to chlorine, 26 out of 65 patients (40%) had elevated serum ALT and 2 out of 13 (15%) had elevated

serum aspartate aminotransferase (AST) values. Without any information on other factors that impact the levels of these enzymes in serum, such as alcohol consumption, it is difficult to ascertain the toxicological significance of this finding. Moulick et al. (1992) mention that measurements of serum AST activity in patients admitted to the hospital following exposure to chlorine did not show any significant abnormality. The exposure concentration was not known, but 2 hours after the massive leak, the level of chlorine was found to be 66 ppm. Hasan et al. (1983) reported that subjects exposed to chlorine gas following a leak from a liquid storage tank had normal liver function tests; the tests were not specified.

Liver congestion has been observed in rodents exposed to acute lethal concentrations of chlorine (i.e., Weedon et al. 1940), but generally, other low-concentration, acute-duration studies did not examine the liver, or if they did, the findings were not reported. In an intermediate-duration study, rats exposed to ≥3 ppm chlorine, but not 1 ppm, 6 hours/day, 5 days/week for 6 weeks had cytoplasmic vacuolation in the hepatocytes (Barrow et al. 1979). This was accompanied by a significant increase in serum alkaline phosphatase at 3 and 9 ppm in males and females and an increase in serum alanine aminotransferase (ALT) in females exposed to 9 ppm. In contrast, in a study of very similar design, rats exposed to up to 5 ppm chlorine 6 hours/day, 5 days/week for 62 days showed no significant histological alterations in the liver (Kutzman 1983). The reason for this apparent discrepancy is not apparent.

Intermittent exposure of monkeys to up to 2.3 ppm chlorine for 1 year or of rats and mice to up to 2.5 ppm for 2 years did not produce gross or microscopic alterations in the liver (Klonne et al. 1987; Wolf et al. 1995).

Renal Effects. No reports were located of renal effects in humans following inhalation exposure to chlorine. However, it would have not been unexpected to find some abnormal indices of renal function in individuals acutely exposed to high levels of chlorine as a consequence of severe cardiopulmonary alterations.

Slight to moderate congestion of the kidneys was reported in male and female rats exposed intermittently to 9 ppm chlorine for 6 weeks, but no significant alterations were seen in rats exposed to 3 ppm (Barrow et al. 1979). Kutzman (1983) examined the kidneys of rats exposed to up to 5 ppm chlorine for 62 days and reported no significant gross or microscopic alterations. Intermittent exposure of monkeys to up to 2.3 ppm chlorine for 1 year or of rats and mice to up to 2.5 ppm for 2 years did not produce gross or microscopic alterations in the kidney (Klonne et al. 1987; Wolf et al. 1995).

Endocrine Effects. No information was located regarding endocrine effects in humans following inhalation exposure to chlorine. No gross or microscopic alterations were reported in the adrenals, pancreas, parathyroid, pituitary, or thyroid gland from monkeys exposed to up to 2.3 ppm chlorine for 1 year (Klonne et al. 1987). Similarly, no gross or histological alterations were found in the pancreas, parathyroid, pituitary, or thyroid from rats and mice exposed to up to 2.5 ppm chlorine for 2 years (Wolf et al. 1995). No further information on endocrine effects in animals exposed to chlorine was located in the literature.

Dermal Effects. Considering that chlorine is an irritating gas, very few reports have described skin effects in subjects exposed to high concentrations of the gas. A health assessment of a population that may have been exposed to up to 200 ppm chlorine after tanker cars derailed reported that skin rashes and skin burns affected 16–25% of a total of 682 persons interviewed 2 weeks after the accident (Agency for Toxic Substances and Disease Registry 1998). In another train derailment, some exposed subjects had minor first-degree skin burns resulting from the vapor exposure (Joyner and Durel 1962). A health evaluation report of firefighters that responded to a chlorine gas leak in Henderson, Nevada, mentions that many of the firefighters complained of skin irritation (NIOSH 1995). During the incident, chlorine concentrations in the air ranged from <0.2 to 17 ppm.

Studies in animals have not described skin alterations following exposure to chlorine gas, even in high exposure acute-duration studies in which animals experienced whole-body exposures. In monkeys exposed to up 2.3 ppm chlorine for 1 year, histological examination of samples of skin did not reveal any significant exposure-related alterations (Klonne et al. 1987). The same negative observations were made in rats and mice exposed to up to 2.5 ppm chlorine for 2 years (Wolf et al. 1995).

Ocular Effects. Almost all reports of acute exposure to high concentrations of chlorine mention eye irritation (lacrimation, conjunctivitis, burning sensation) as one of the most prevalent complains among the patients. These effects usually resolve within a few days after exposure. In a controlled exposure study with volunteers, complaints of itching or burning of the eyes were significantly greater during exposures to 1 ppm chlorine than during control exposures (Anglen 1981).

Swelling around the eyes was reported in rats exposed to 12 ppm chlorine 6 hours/day, 5 days/week for 10 days, but not for 5 days (Dodd et al. 1980). Ocular irritation was also reported in rats exposed intermittently to 3 ppm chlorine for 6 weeks, but no irritation was apparent at 1 ppm (Barrow et al. 1979). This is more or less consistent with the results of another intermediate-duration study that reported

occasional signs of eye irritation in rats exposed to 1.5 ppm for 62 days and severe irritation in rats exposed to 5 ppm chlorine (Kutzman 1983). Monkeys exposed intermittently to 2.3 ppm for 1 year showed conjunctival irritation with some exudation at the end of the study, but there was no gross or microscopic evidence of chronic changes in the conjunctiva or in the cornea (Klonne et al. 1987). No ocular effects were observed in rats or mice exposed to up to 2.5 ppm chlorine for 2 years (Wolf et al. 1995).

Body Weight Effects. No relevant information was located regarding body weight effects in humans following inhalation exposure to chlorine. Reduced body weight gain or weight loss has been commonly reported in acute high-exposure studies in animals of enough duration to allow evaluation of weight changes. Although rarely reported, the reduced growth or weight loss is largely due to reduced food consumption. For example, rats exposed to 12 ppm chlorine 5 days/week for 2 weeks lost approximately 20% of their body weight during the second week relative to their starting weight (Dodd et al. 1980). Pair-fed rats also lost weight, confirming that the effect in the chlorine-exposed rats was due to reduced food intake. In another acute-duration study, rats exposed to 9.1 ppm chlorine for 3 or 5 days lost 11 and 13%, respectively, of the initial body weight (Jiang et al. 1983). Under the same exposure conditions, mice lost 17 and 21% of the initial body weight (Jiang et al. 1983).

Similar results have been obtained in intermediate-duration studies. Final body weight of rats exposed to 9 ppm chlorine for 6 weeks (Barrow et al. 1979) or to 5 ppm for 62 days (Kutzman 1983) were >30% lower than controls. Even a relatively low concentration of chlorine of 0.5 ppm for 62 days lowered body weight in female rats by approximately 11% relative to controls (Kutzman 1983). Neither study provided information on food or water consumption. Exposure of monkeys to 2.3 ppm chlorine for 1 year (Klonne et al. 1987) or of rats and mice to 2.5 ppm for 2 years (Wolf et al. 1995) reduced body weight gain during the studies, but the final weights, judging by figures in the papers, appeared to be <10% lower than the respective controls.

3.2.1.3 Immunological and Lymphoreticular Effects

No studies were located that evaluated immunocompetence or effects on lymphoreticular organs in humans following exposure to chlorine gas.

Minimal information is available from studies in animals. Barrow et al. (1979) reported that the spleen and thymus from rats exposed to 9 ppm chlorine for 6 weeks had a decreased content of lymphoid

elements, but no such changes were seen in rats exposed to 1 or 3 ppm. According to the investigators, this may have been a function of the poor physical condition and decreased nutritional state of the rats in that dosing group. In a very similar study, no histological alterations were reported in the spleen and peribronchial lymph nodes from rats exposed to up to 5 ppm chlorine for 62 days (Kutzman 1983). Studies in monkeys, rats, and mice also have found no gross or microscopic lesions in lymphoreticular organs and tissues following intermittent chronic exposure to 2.3–2.5 ppm chlorine (Klonne et al. 1987; Wolf et al. 1995).

The highest NOAEL values and all LOAEL values from each reliable study for lymphoreticular effects in each species and duration category are recorded in Table 3-1 and plotted in Figure 3-1.

3.2.1.4 Neurological Effects

Symptoms effects such as headache, dizziness, anxiety, and syncope are commonly reported following acute high exposures to chlorine and are thought to be due, at least in part, to asphyxia induced by chlorine.

In a case of high exposure to chlorine that resulted in the death of the patient, postmortem examination showed a swollen brain with flattening of convolutions and subarachnoid hemorrhage (Adelson and Kaufman 1971). The investigators speculated that the lesions could have been caused by hypoxia that resulted from the severe pulmonary effects. In another case report, a 60-year-old man who accidentally inhaled chlorine gas in a swimming pool accident had a magnetic resonance scan of the head conducted 2 years after the accident that showed multiple areas of decreased signal in the periventricular white matter (Levy et al. 1986). Other neurological tests showed no evidence of cranial nerve abnormalities or sensory deficits. This brief communication does not mention what might have prompted the subject to undergo the scan.

Kilburn (1995, 2000, 2003b) published a series of reports describing long-lasting neurological effects in subjects accidentally exposed to high concentrations of chlorine gas under various scenarios. The earliest study (Kilburn 1995) reported that six subjects exposed to an undetermined concentration of chlorine for 3 minutes to 5 hours had difficulty concentrating and sleeping, dizziness, loss of balance, excessive fatigue, loss of strength, depression, and irritability during a period of 1–3 years after the accident. Neurobehavioral tests were conducted 15–50 months after exposure and the results were compared to a control group matched for sex, age, and education. It should be noted that the testers were aware of the

exposure status of the subjects. The results showed impaired balance with the eyes closed and hearing loss in all of the exposed subjects. Five had decreased vibration sensitivity, color discrimination, and verbal recall; four had prolonged blink reflex latency; three had prolonged simple and choice reaction times, and three had nerve defects or constricted visual fields. In a subsequent study, 22 patients exposed briefly (the reports mentions seconds to a few minutes in one section and minutes to a few hours in another section) to chlorine gas were evaluated with a battery of tests 7–48 months after exposure. A total of 296 unexposed subjects served as controls. The results showed significant impairment among the exposed group in a number of areas including balance, reaction time, color identification, visual field performance, blink latency, cognition, verbal recall, and making trails. A similar study was conducted with subjects exposed to chlorine as a result of a train derailment (see Agency for Toxic Substances and Disease Registry [1998] under Respiratory Effects) (Kilburn 2003b). Ninety-seven subjects were tested 7 weeks after exposure and 57 were tested 3 years later; 26 were tested on both occasions. Seven weeks after exposure, the exposed subjects showed impairment in five neurobehavioral functions compared with unexposed subjects recruited for the 3-year testing. At 3 years, the patients showed impairment in seven additional tests compared with controls. Because of lack of exposure data, these studies are not listed in Table 3-1.

There are no studies in animals that could confirm or refute Kilburn's findings mentioned above. The only information regarding neurological effects of chlorine in animals is limited to reports of no gross or microscopic alterations in the brain of rats exposed intermittently to up to 5 ppm chlorine for 62 days (Kutzman 1983), or in the brain, spinal cord, and sciatic nerve of rats and mice exposed to up to 2.5 ppm chlorine for 2 years (Wolf et al. 1995). Also, in monkeys exposed intermittently for 1 year to up to 2.3 ppm chlorine, there were no gross or histological alterations in central or peripheral nervous system tissues (Klonne et al. 1987). The investigators also mentioned that clinical observational neurological examinations conducted in the monkeys prior to sacrifice were unremarkable, but the scope of these tests was not specified.

The highest NOAEL values from each reliable study for neurological effects in each species and duration category are recorded in Table 3-1 and plotted in Figure 3-1.

3.2.1.5 Reproductive Effects

The only information available regarding effect in humans is that evaluation of the outcome of 15 pregnancies among female workers at a chlorine plant in 1932–1933 did not provide any evidence of reproductive toxicity (Skljanskaya et al. 1935).

In an intermediate-duration inhalation study, male and female rats were exposed intermittently to up to 5 ppm chlorine for 62 days (Kutzman 1983). At the end of the exposure period, 8 exposed males were mated with unexposed females and 10 exposed females were mated with unexposed males and all females were sacrificed on gestation day 19 for evaluation of reproductive end points. The results showed no significant effects of chlorine exposure on fertility, number of corpora lutea, viable embryos, early or late deaths, or pre-implantation loses. In addition, in males exposed for 62 days, there were no histological alterations in the testes, and sperm morphology was unremarkable.

In chronic-duration studies, exposure of male and females monkeys for 1 year or male and female rats and male mice for 2 years to up to 2.5 ppm chlorine did not result in gross or microscopic alterations of the reproductive organs (Klonne et al. 1987; Wolf et al. 1995). Female mice developed a concentration-related increase in ovarian abscesses and of suppurative inflammation of the uterus. Without a further discussion, the investigators stated that it was unlikely that chlorine induced the inflammatory response, but no alternative explanation was provided.

The highest NOAEL values from each reliable study for reproductive effects in each species and duration category are recorded in Table 3-1 and plotted in Figure 3-1.

3.2.1.6 Developmental Effects

In a summary of the limited information available, WHO (1982) reports that early studies from the Russian literature found no evidence of teratogenic in offspring from effects female workers at a chlorine plant in 1932–1933 and that rabbits exposed to low chlorine concentrations (0.6–1.6 ppm) during pregnancy gave birth to healthy offspring. No additional information was located regarding developmental effects in animals following inhalation exposure to chlorine.

3.2.1.7 Cancer

There are several studies of cancer in humans that involve exposure to chlorine, among other chemicals, in occupational settings.

A study of 28 employees at a Texas chemical plant who had died of primary intracranial neoplasms found no evidence that exposure to chlorine may have played a role (Bond et al. 1983). Chlorine was one of the chemicals to which cases and controls had the most frequent potential for exposure. Bond et al. (1985) conducted a case-control study of 26 renal cancer deaths among employees of a multiple process chemical production facility. Although an increased odds ratio (OR) for renal cancer was found in a chlorine production area, it was not attributed to chlorine exposure, but to asbestos and caustic materials. In addition, in the magnesium processing area, where large amounts of chlorine were used, there was a decreased risk of renal cancer. A nested case-control study of 306 lung cancer deaths among 19,608 employees of a chemical plant provided no evidence that chlorine had a role in the deaths (Bond et al. 1986). The OR for chlorine exposure was 1.08 (95% confidence interval [CI], 0.81–1.44).

A study of 51 lung cancer cases at a dye and resin manufacturing plant reported an increased OR for lung cancer for employees who were seen at the plant infirmary for acute exposure to chlorine after adjusting for smoking (OR=27; 95% CI, 3.5–205) (Barbone et al. 1992). However, the number of cases was small (6 cases versus 3 out of 102 controls), and four of the six cases worked in the anthraquinone dye and epichlorohydrin production area. Barbone et al. (1992) stated that based on small number of cases, the small association between lung cancer and acute exposure to chlorine may be due to chance or to confounding by other unidentified exposures.

In a larger study of 2,391 male workers producing magnesium metal, Heldaas et al. (1989) found 4 cases of lung cancer versus 1.3 expected in a subset of workers who experienced chlorine intoxication and had at least 20 years since first employment (95% CI, 0.8–7.8). However, the rate ratios for lung cancer were higher in those workers who were not registered in the chlorine exposure list. The authors speculated that the use of respiratory protective gear (mouthpieces) may have been a reason for the difference.

In a study of 1,190 workers at chlor alkali plants, there was a marginally significant excess of lung cancers (10 observed versus 4.9 expected; 95% CI, 1.0–3.8) which, according to the authors, was possibly due to previous use of asbestos (Barregård et al. 1990). Jäppinen et al. (1987) conducted a retrospective cohort study of 3,545 workers in the Finnish pulp and paper industry and found 78 cases of lung cancer

versus 62.6 expected (95% CI, 98–155). The excess was most prominent in board mill workers (40 observed versus 18.1 expected; 95% CI, 158–302), particularly after 20 years of latency (25 observed versus 7.8 expected; 95% CI, 209–476). However, there is no mention in the study of the chemicals to which the various subcohorts (based on work histories) may have been more intensely exposed.

Only the study by Wolf et al. (1995) provided information on carcinogenicity of inhaled chlorine in animals. In that study, male and female rats and mice were exposed intermittently to up to 2.5 ppm chlorine for 2 years. Gross and histological examination of all major tissues and organs, including the nasal cavity at five levels did not show any biologically or statistically significant increase in neoplasms.

The EPA, the International Agency for Research on Cancer (IARC), and the Department of Health and Human Services (DHHS) have not classified chlorine gas as to its carcinogenicity.

3.2.2 Oral Exposure

Studies that used tap water as the medium for delivery of chlorine to experimental animals are not included in this section to eliminate potential confounding effects of chlorination byproducts.

Although people are not likely to be exposed to chlorine itself through oral routes, there are reports of accidental or intentional exposure to bleach (typically a 3–6% solution of sodium hypochlorite). Information from some of those reports is also included in this section.

3.2.2.1 Death

Human deaths have been reported following ingestion of sodium hypochlorite. In a review of the literature, Racioppi et al. (1994) state that the lethal dose of sodium hypochlorite in adults has been reported to be approximately 200 mL of a solution containing 3–6% chlorine. Racioppi et al. (1994) also indicate that the aspiration of bleach in the lungs following ingestion as been reported as the cause of fatalities. Ross and Spiller (1999) described a fatal case of a 66-year-old woman who ingested an unknown quantity of bleach and died of cardiac arrest 4.5 hours after the bleach ingestion. Autopsy revealed esophageal and gastric mucosal erosions, perforations at the gastroesophageal junction, and extensive necrosis of adjacent soft tissue. Jakobsson et al. (1991) reported the case of a fatal poisoning in a 1-year-old girl after ingestion of a household cleanser containing 4.5% sodium hypochlorite in an alkaline solution. Examination of the upper gastrointestinal tract showed severe gross and microscopic damage. According to unpublished information reviewed by Racioppi et al. (1994), the oral LD₅₀ for a

5.25% solution of sodium hypochlorite household bleach in rats is 13,000 mg/kg; for a 13% solution, LD_{50} values of 5,000 and 8,200 mg/kg are listed. A review of the literature by BG-Chemie (1991) indicates that LD_{50} values of 6,800 and 5,800 mg/kg were determined for male and female ddY rats, respectively.

3.2.2.2 Systemic Effects

The highest NOAEL values and all LOAEL values from each reliable study for systemic effects in each species and duration category are recorded in Table 3-3 and plotted in Figure 3-2.

Respiratory Effects. Respiratory rate was not altered in a group of 10 volunteers who drank 0.5 L/day of drinking water containing 5 mg/L chlorine for 12 weeks (Lubbers et al. 1982). This corresponds approximately to doses of 0.036 mg Cl/kg/day, assuming a body weight of 70 kg. Evaluations of the respiratory rate were done weekly during the dosing period and continued for an additional 8 weeks post-dosing. Since the study did not control for non-experimental ingestion of chlorine by the volunteers, the total daily dose of chlorine consumed is likely to have been higher than 0.036 mg Cl/kg/day. In a review of the literature, Racioppi et al. (1994) mention the case of a woman who developed aspiration pneumonitis after ingesting an unknown amount of hypochlorite bleach. Bracco et al. (2005) described a similar case. In this case, chest x-rays performed 2 hours after intoxication showed bilateral bibasal infiltrate suggestive of aspiration pneumonia. Over the next 24 hours, the patient's respiratory condition declined and she required ventilatory support. The patient eventually recovered after 26 days of mechanical ventilation. In the fatal case of bleach ingestion described by Jakobsson et al. (1991), microscopic examination of the lungs showed aspiration of epithelium derived from the upper respiratory tract, and signs of acute bronchitis.

Limited data on respiratory effects are available in animals. Ninety-day drinking water studies in Sprague-Dawley rats (Daniel et al. 1990) and B6C3F₁ mice (Daniel et al. 1991) dosed with up to 24.9 and 39.2 mg Cl/kg/day, respectively, reported no gross or microscopic alterations in the lungs, trachea, and nasal turbinates. Furukawa et al. (1980) also reported no significant histological alterations in the lungs and bronchial tube of F344 rats dosed with up to 85 mg Cl/kg/day for 92 days. Similar results were reported in chronic-duration studies in F344 rats (Hasegawa et al. 1986; NTP 1992) and B6C3F₁ mice (NTP 1992). Rats received doses of up to 133 mg Cl/kg/day and mice received doses up to 24.2 mg Cl/kg/day for 2 years.

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution _ Oral

		Exposure/			LC	DAEL		
	Species (Strain)	Duration/ Frequency (Route)	System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments
	E EXPOS	SURE						
System								
1	Rat (Sprague- Dawley)	once (G)	Hepatic		20 M (increased triacylglycerols in whole liver homogenate)		Chang et al. 1981	No other end points were evaluated.
•	Rat	14 d						
2	(Wistar)	(GW)	Cardio	200 F			Cunningham 1980	Only organ weight was assessed.
			Hepatic	200 F				
			Renal	200 F				
			Bd Wt	200 F				
Neurolo	ogical							
3	Rat (Wistar)	14 d (GW)		200 F			Cunningham 1980	Only brain weight was assessed.
INTER System		E EXPOSURE						
4	Human	4 wk (W)	Hepatic	0.4			Wones et al. 1993	NOAELs are for serum thyroid hormones and serum lipid profile.
			Endocr	0.4				
5	Rat (Sprague- Dawley)	12 mo (W)	Hemato	12 M			Abdel-Rahman et al. 1984	

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

Hepatic

Renal Endocr

Dermal Bd Wt

24.9 F 24.9 F

24.9 F 24.9 F

24.9 F

			Table 3-3 Level	s of Significant	Exposure to Hypochlorit	e Solution - Oral	(continued)		
		Exposure/ Duration/				LOAEL			
Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments	
6	Rat (Long- Eva	66-76 d ns) (GW)	Hemato	3.4			Cariton et al. 1986	NOAELs are for blood counts and serum thyroid hormone levels.	
			Endocr	3.4					
			Bd Wt	3.4					
7	Rat (Wistar)	6 wk (W)	Bd Wt	15.7 M			Cunningham 1980		ķ
8	Rat (Sprague- Dawley)	90 d (W)	Resp	24.9 F			Daniel et al. 1990	NOAELs are for gross and microscopic evaluation of tissues and organs, and hematology and clinical chemistry.	HEALTH EFFECTS
								onomically.	Ø
			Cardio	24.9 F					
			Gastro	24.9 F					
			Hemato	24.9 F					
			Musc/skel	24.9 F					

Exposure/ Duration/ Frequency (Route)			LO	AEL		
	System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments
8 wk (VV)	Bd Wt	4.1 M			Exon et al. 1987	
92 d (44) (W)	Resp	85 M			Furukawa et al. 1980 hypochlorous acid and sodio hypochlorite	NOAELs are for histopathology of tissues and organs clinical chemistry at hematology.
	Cardio	52 F	84 F (endocardial hyperplasia; myocardial fibrosis)			

85 M (46% reduction in final

Hasegawa et al. 1986

body weight)

305 M (final body weight

reduced 47%)

Key to Species Figure (Strain)

Rat (Sprague-Dawley)

Rat

(Fischer- 344) (W)

9

10

11

Rat

(Fischer- 344) (W)

13 wk

Gastro

Hemato

Hepatic

Renal

Endocr

Dermal

Bd Wt

Bd Wt

85 M

85 M

85 M

85 M

85 M

85 M

26 M

76

50 M (19% reduction in final

body weight)

152 M (final body weight reduced 19%)

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

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		Exposure/ Duration/				LOAEL		
a Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments
	Mouse (B6C3F1)	90 d (W)	Resp	39.2 F			Daniel et al. 1991	NOAELs are for gross and histological evaluation of tissues and organs, adn hematology and clinical chemistry.
			Cardio	39.2 F				
			Gastro	39.2 F				
			Hemato	39.2 F				
			Musc/skel	39.2 F				
			Hepatic	39.2 F				
			Renal	39.2 F				
			Endocr	39.2 F				
			Dermal	39.2 F				
			Bd Wt	39.2 F				
	Gn Pig (albino)	5 wk (W)	Bd Wt	26 M			Cunningham 1980	
lmmun	o/ Lymphoi							
14	Rat (Sprague- Dawley)	90 d (W)		24.9 F			Daniel et al. 1990	NOAEL is for gross and microscopic evaluation of lymphoreticular organs.

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

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		Exposure/ Duration/				LOAEL		
Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments
15	Rat (Sprague- Dawley)	8 wk (W)		4.1 M			Exon et al. 1987	Some reported changes in immune parameters of unknown toxicological significance were not considered adverse.
16	Rat (Fischer- 34	92 d (4) (W)		85 F			Furukawa et al. 1980 hypochlorous acid and sodium hypochlorite	NOAEL is for gross and microscopic evaluation of lymphoreticular organs.
17	Mouse (B6C3F1)	90 d (W)		39.2 F			Daniel et al. 1991	NOAEL is for gross and histological evaluation of lymphoreticular organs.
Neurol	ogical Rat (Sprague- Dawley)	90 d (W)		24.9 F			Daniel et al. 1990	NOAEL is for gross and microscopic evaluation of the brain and sciatic nerve.
19	Rat (Fischer- 34	92 d 4) (W)		85 F			Furukawa et al. 1980 hypochlorous acid and sodium hypochlorite	NOAEL is for gross and microscopic examination of the brain.

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

on		

		Exposure/				LOAEL		
a Key to Figure	Species (Strain)	Duration/ Frequency (Route)	System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments
	Mouse (B6C3F1)	90 d (W)		39.2 F			Daniel et al. 1991	NOAEL is for gross and histological evaluation of the brain and sciatic nerve.
Reprod	luctive Rat (Long- Evar	66-76 d ns) (GW)		3.4			Carlton et al. 1986	NOAEL is for fertility and histopathology of the reproductive tract.
	Rat (Sprague- Dawley)	90 d (W)		24.9 F			Daniel et al. 1990	NOAEL is for gross and microscopic evaluation of reproductive organs.
23	Rat (Fischer- 34	92 d 14) (W)		85 F			Furukawa et al. 1980 hypochlorous acid and sodium hypochlorite	NOAEL is for gross and microscopic evaluation of reproductive organs.
	Mouse (B6C3F1)	90 d (W)		39.2 F			Daniel et al. 1991	NOAEL is for gross and histological evaluation of reproductive organs.
25	pmental Rat (Long- Evar	42 d ns) (GW)		3.4			Carlton et al. 1986	NOAEL is for litter viability and size and pup weight.

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

		Table 3-3 Leve	ls of Significan	t Exposure to Hypochlorite	Solution - Oral	(continued)	
	Exposure/ Duration/				LOAEL		
a Key to Species Figure (Strain)	Frequency	System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments
CHRONIC EXI	POSURE						
26 Rat (Fischer- 3	2 yr 344) (W)	Resp	133 F			Hasegawa et al. 1986	NOAELs are for gross and microscopic alterations in organs and tissues.
		Cardio	133 F				
		Gastro	133 F				
		Hemato	133 F				
		Hepatic	133 F				
		Renal	133 F				
		Endocr	133 F				
		Dermal	133 F				
		Ocular	133 F				
		Bd VVt		67 F (11% reduction in fir body weight)	nal		

(continued)

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

		Exposure/ Duration/				LOAEL		
a Key to Figure	Species (Strain)	Frequency (Route)	System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments
	Rat (Fischer- 3	2 yr 44) (W)	Resp	14.4 F			NTP 1992	NOAELs are for histopathology of tissues and organs and hematology parameters.
			Cardio	14.4 F				
			Gastro	14.4 F				
			Hemato	14.4 F				
			Hepatic	14.4 F				
			Renal	14.4 F				
			Endocr	14.4 F				
			Dermal	14.4 F				
			Bd Wt	14.4 F				
			Other	14.4 F				

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

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Key to Figure		Exposure/ Duration/ Frequency (Route)				LOAEL		
	Species (Strain)		System	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	Comments
28	Mouse (B6C3F1)	2 yr (W)	Resp	24.2 F			NTP 1992	NOAELs are for histopathology of tissues and organs and hematology parameters.
			Cardio	24.2 F				
			Gastro	24.2 F				
			Hemato	24.2 F				
			Hepatic	24.2 F				
			Renal	24.2 F				
			Endocr	24.2 F				
			Dermal	24.2 F				
			Bd Wt	24.2 F				
			Other	24.2 F				
lmmun 29	o/ Lympho Rat (Fischer- 3	2 yr		133 F			Hasegawa et al. 1986	NOAEL is for gross and microscopic appearence of the spleen.
30	Rat (Fischer- 3	2 yr 44) (W)		14.4 F			NTP 1992	NOAEL is for histopathology of lymphoreticular organs.

(continued)

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

	Exposure/ Duration/ Frequency (Route)				LOAEL		Comments
		NOAEL System (mg/kg/d	NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	
 Mouse (B6C3F1)	2 yr (W)		24.2 F			NTP 1992	NOAEL is for histopathology of lymphoreticular organs.
o gical Rat (Fischer- 3	2 yr 44) (W)		133 F			Hasegawa et al. 1986	NOAEL is for gross and microscopic appearence of the brain.
Rat (Fischer- 3-	2 yr 44) (W)		14.4 F			NTP 1992	NOAEL is for histopathology of various brain areas.
Mouse (B6C3F1)	2 yr (W)		24.2 F			NTP 1992	NOAELs are for histopathology of various brain areas.
uctive Rat (Fischer- 34	2 yr 44) (W)		133 F			Hasegawa et al. 1986	NOAEL is for gross and microscopic appearence of the reproductive organs.
Rat (Fischer- 34	2 yr 44) (W)		14.4 F			NTP 1992	NOAEL is for histopathology of reproductive organs.

	Species (Strain)	Exposure/ Duration/ Frequency (Route)		LOAEL			
Key to Figure			NOAEL (mg/kg/day)	Less Serious (mg/kg/day)	Serious (mg/kg/day)	Reference Chemical Form	orm Comments
37	Mouse (B6C3F1)	2 yr (W)	24.2 F			NTP 1992	NOAEL is for histopathology of reproductive organs.

(continued)

Table 3-3 Levels of Significant Exposure to Hypochlorite Solution - Oral

a The number corresponds to entries in Figure 3-2.

Bd Wt = body weight; Cardio = cardiovascular; d = day(s); Endocr = endocrine; F = Female; Gastro = gastrointestinal; Gn pig = guinea pig; (GW) = gavage in water; Hemato = hematological; Immuno/Lymphoret = immunological/lymphoreticular; LOAEL = lowest-observed-adverse-effect level; M = male; mo = month(s); Musc/skel = musculoskeletal; NOAEL = no-observed-adverse-effect level; Resp = respiratory; (W) = drinking water; wk = week(s); yr = year(s)

Figure 3-2 Levels of Significant Exposure to Hypochlorite Solution - Oral Acute (≤14 days)

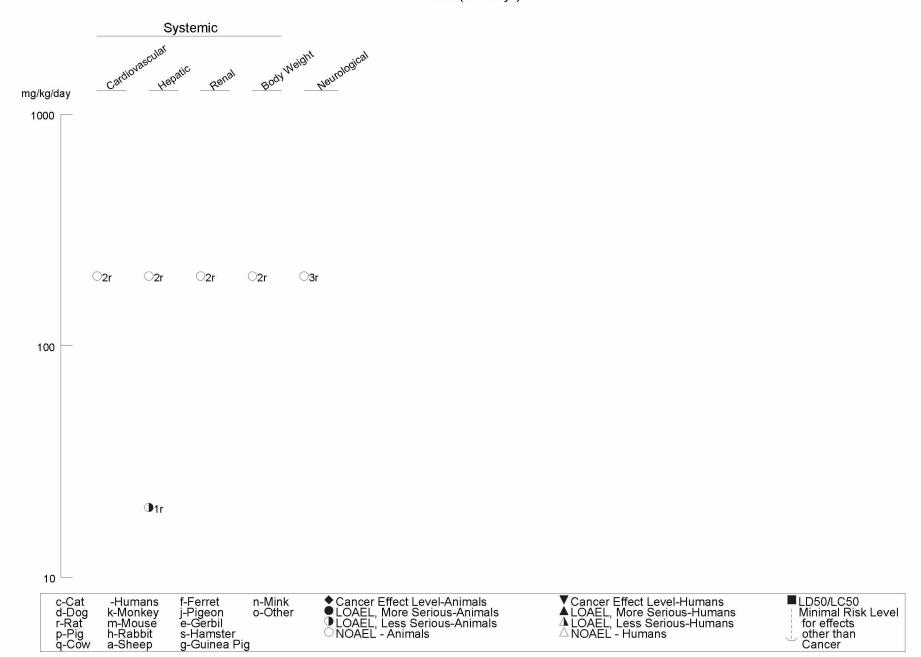


Figure 3-2 Levels of Significant Exposure to Hypochlorite Solution - Oral *(Continued)*Intermediate (15-364 days)

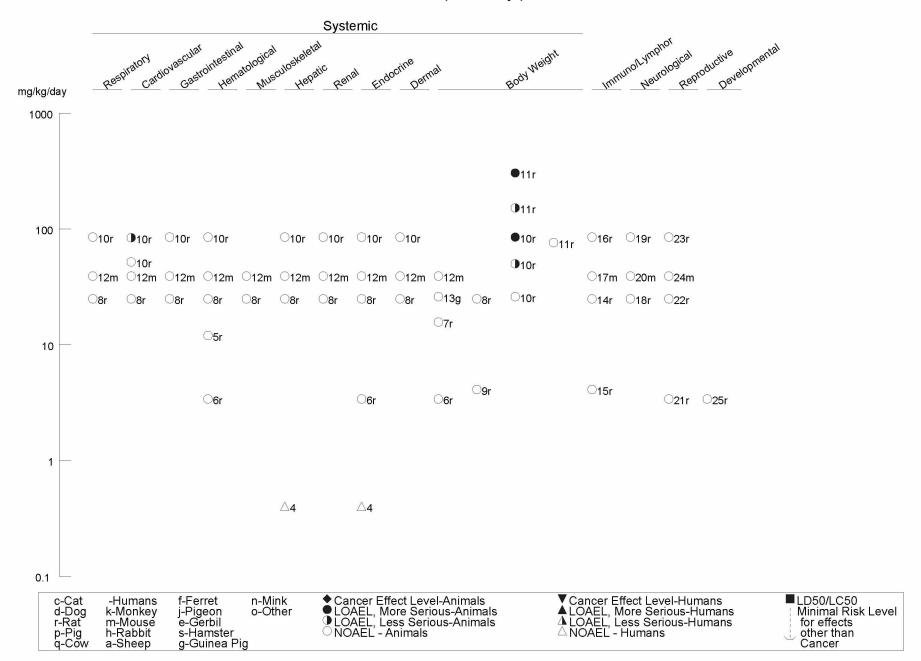
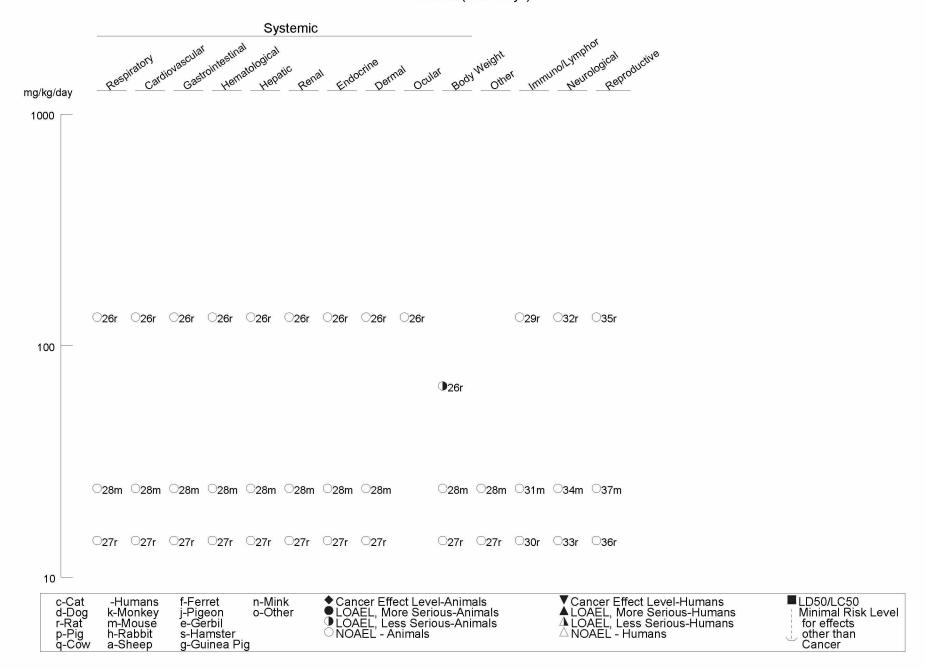


Figure 3-2 Levels of Significant Exposure to Hypochlorite Solution - Oral (Continued)

Chronic (≥365 days)



Cardiovascular Effects. In the study by Lubbers et al. (1982) in volunteers mentioned above, drinking water that provided 0.036 mg Cl/kg/day for 12 weeks did not alter systolic or diastolic blood pressure or pulse rate, and electrocardiograms were unremarkable. No additional relevant information was located regarding cardiovascular effects in humans following oral exposure to hypochlorite solutions.

A 14-day study in which Wistar rats were dosed by gavage with sodium hypochlorite in milk (up to 200 mg Cl/kg/day) reported no significant alterations in heart weight, but no additional cardiovascular parameters were examined (Cunningham 1980). Two intermediate-duration studies reported no significant gross or microscopic alterations in the heart and aorta from rats and mice exposed to up to 24.9 and 39.2 mg Cl/kg/day, respectively, for 90 days (Daniel et al. 1990, 1991). Furukawa et al. (1980) reported that endocardial hyperplasia and fibrosis of the myocardium were observed in male and female F344 rats dosed with approximately 84 mg Cl/kg/day in the drinking water for 92 days. No such effects were seen in rats dosed with approximately 50 mg Cl/kg/day. Two-year studies also did not find histological alterations in the heart from rats and mice that received doses of up to 133 and 24.2 mg Cl/kg/day, respectively (Hasegawa et al. 1986; NTP 1992).

Gastrointestinal Effects. In general, ingestion of small amounts (less than a cup) of sodium hypochlorite bleach (approximately 5.3% sodium hypochlorite) does not cause serious or permanent damage to the upper gastrointestinal tract. Pike et al. (1963) reviewed 129 cases of children who ingested Clorox® and reported that no complications of consequences were found. Sixty-five cases were examined by esophagoscopy within 96 hours of the ingestion and only 2 showed evidence of esophageal injury. The children were between 12 month and 7 years old and the amounts of bleach ingested ranged from "½ ounce to 1 cup." Landau and Saunders (1964) state that among 393 children who ingested bleach and were seen at a hospital, there were no esophageal strictures or perforations, and about 50% of the patients received no treatment. Hook and Lowry (1974) reported that among 23 definite cases of children who ingested Clorox®, severe irritation of the esophageal mucosa was observed in only 1 case. Minor transient irritation was observed in some of the patients. A report from the German literature of 23 children who accidentally ingested 3–5% sodium hypochlorite indicates that there was only 1 case with signs of superficial burns in the esophagus, which had disappeared 2 weeks later when controlled by esophagoscopy (Mühlendahl et al. 1978). Liquid bleach is a strong emetic, which helps reduce the time of residence in the stomach, but on the other hand, it increases the potential for aspiration.

Examination of fatal cases following ingestion of unknown quantities has revealed esophageal and gastric mucosal erosions, perforations at the gastroesophageal junction, and extensive necrosis of adjacent soft tissue (Ross and Spiller 1999). In a fatal case of a child who drank 4.5% sodium hypochlorite in an alkaline solution (pH 12), severe gross lesions were seen in the mouth, tongue, glottis, epiglottis, esophagus, and stomach (Jakobsson et al. 1991). Glottic and subglottic edema was described by Babl et al. (1998) in a child who drank household bleach from a cup.

In some earlier studies in animals, commercial bleach was administered through a tube directly into the esophagus and, in some cases, the distal end of the esophagus was artificially occluded to prolong and control the contact time between the solution and the mucosa (Hook and Lowry 1974; Landau and Saunders 1964; Strange et al. 1951; Yarington 1970). For example, commercial bleach placed in the esophagus of 151 dogs for several minutes caused the immediate death of 8 dogs from perforations into their pleural cavities (Landau and Saunders 1964). Necropsy performed 3 months later on the seven dogs that survived revealed no abnormalities. Yarington (1970) reported that, in dogs, the minimum amount of bleach that caused a burn in the esophagus was 10 cm³ applied over a 5-minute period. A volume of 30 cm³ applied for 2 minutes caused minimal edema of the esophagus.

Few more recent studies are available. Gross and microscopic examination of multiple levels of the gastrointestinal tract of Sprague-Dawley rats that drank water that provided up to 24.9 mg Cl/kg/day for 90 days did not reveal any significant gross or microscopic alterations (Daniel et al. 1990). The same negative results were reported in F344 rats and B6C3F₁ mice that drank water with up to 85 mg Cl/kg/day or 39.2 mg Cl/kg/day, respectively, for 90 days (Daniel et al. 1991; Furukawa et al. 1980). Two-year studies also did not find histological alterations in the gastrointestinal tract from F344 rats and B6C3F₁ mice that received doses of up to 133 and 24.2 mg Cl/kg/day, respectively (Hasegawa et al. 1986; NTP 1992).

Hematological Effects. No significant treatment-related alterations in a comprehensive number of hematological parameters were reported in volunteers who drank 0.5 L/day of drinking water containing 5 mg/L chlorine (approximately 0.036 mg Cl/kg/day) for 12 weeks (Lubbers et al. 1982).

A 12-month study in Sprague-Dawley rats exposed to approximately 12 mg Cl/kg/day reported alterations in red blood cell fragility at various times during the study; however, the alterations were sporadic and not dose- or duration-related, and varied in direction suggesting that they may have not been caused by exposure to chlorine (Abdel-Rahman et al. 1984). Red blood cell count and hematocrit were reduced

significantly after 3 months of treatment with 12 mg Cl/kg/day, but not at lower doses of chlorine (0.12 and 1.2 mg Cl/kg/day). Evaluations conducted after 6 months of treatment showed no significant difference in hematology parameters between treated and control groups (Abdel-Rahman et al. 1984). In another intermediate-duration study, exposure of male and female Long-Evans rats to up to 3.4 mg Cl/kg/day did not induce significant alterations in blood cell counts (Carlton et al. 1986). In yet additional studies, standard hematology parameters were not significantly altered in rats or mice dosed with up to 85 and 39.2 mg Cl/kg/day, respectively, for 90 days (Daniel et al. 1990, 1991; Furukawa et al. 1980). Hematology parameters were also evaluated in F344 rats and B6C3F1 mice after 14–15 and 66 months of treatment with chlorine in the 2-year drinking water bioassay (NTP 1992) and at termination in the Hasegawa et al. (1986) study in F344 rats. The results showed no significant treatment-related alterations in rats dosed with \leq 133 mg Cl/kg/day or in mice dosed with \leq 24.2 mg Cl/kg/day.

Musculoskeletal Effects. No information was located regarding musculoskeletal effect in humans following oral exposure to hypochlorite and limited information is available in animals. No significant gross or microscopic alterations were seen in skeletal muscle and sternebrae from Sprague-Dawley rats following exposure to up to 24.9 mg Cl/kg/day in the drinking water for 90 days (Daniel et al. 1990). Similar results were reported in B6C3F₁ mice exposed to up to 39.2 mg Cl/kg/day for 90 days (Daniel et al. 1991). The 2-year NTP (1992) study reported no significant alterations in the femur from F344 rats or B6C3F₁ mice that received up to 14.4 or 24.2 mg Cl/kg/day, respectively, in the drinking water.

Hepatic Effects. Serum chemistry tests used to evaluate liver function did not reveal any significant alteration in volunteers during 12 weeks of exposure to approximately 0.036 mg Cl/kg/day in the drinking water and during a period of 8 weeks after exposure ceased (Lubbers et al. 1982). In another study of controlled exposure in volunteers, exposure to approximately 0.4 mg Cl/kg/day in the drinking water for 4 weeks had no significant effect on the lipid profile in serum (Wones et al. 1993).

Very limited information is available regarding liver effects in acute-duration oral studies in animals. Liver weight was not significantly altered in Wistar rats after 14 days of daily gavage doses of sodium hypochlorite in milk (up to 200 mg Cl/kg/day) (Cunningham 1980); no other liver parameter was monitored in this study. In another study of limited scope, triacylglycerols were significantly increased in whole liver homogenates from rats 2 days after a single gavage dose of approximately 20 mg Cl/kg (Chang et al. 1981). Ten days after dosing, the levels of triacylglycerols had returned to pre-dosing levels.

No significant gross or microscopic alterations were seen in the liver of Sprague-Dawley rats, F344 rats, or B6C3F₁ mice exposed to up to 24.9, 85, and 39.2 mg Cl/kg/day, respectively, in the drinking water for 90 days (Daniel et al. 1990, 1991; Furukawa et al. 1980). Levels of serum liver enzymes also were not significantly affected by treatment. Similarly, no significant hepatic effects were reported in F344 rats or B6C3F₁ mice that received up to 133 or 24.2 mg Cl/kg/day, respectively, in the drinking water for 2 years (Hasegawa et al. 1986; NTP 1992). Clinical chemistry tests conducted at termination in rats in the Hasegawa et al. (1986) study did not provide any indication of liver toxicity.

Renal Effects. The only relevant information regarding renal effects in humans following oral exposure to hypochlorite is that urinalysis performed weekly on volunteers during a 12-week controlled intake of approximately 0.036 mg Cl/kg/day and during a subsequent period of 8 weeks after treatment was unremarkable (Lubbers et al. 1982).

The only information available in acute-duration oral studies in animals is that kidney weight was not altered in rats in a 14-day study in which received daily doses of up to 200 Cl/kg/day by gavage in milk (Cunningham 1980). No other renal parameter was evaluated. No significant gross or microscopic alterations were seen in the kidney of Sprague-Dawley rats, F344 rats, or B6C3F₁ mice exposed to up to 24.9, 85, and 39.2 mg Cl/kg/day, respectively, in the drinking water for 90 days (Daniel et al. 1990, 1991; Furukawa et al. 1980). However, Furukawa reported increased incidence of gross abnormalities (no further details provided) in the bladder of rats in all treated groups (8–85 mg Cl/kg/day) in the 90-day study. No significant kidney effects were reported in F344 rats or B6C3F₁ mice that received up to 133 or 24.2 mg Cl/kg/day, respectively, in the drinking water for 2 years (Hasegawa et al. 1986; NTP 1992).

Endocrine Effects. A study in male and female volunteers who consumed daily for 4 weeks 1.5 L of distilled water that provided a dose of approximately 0.4 mg Cl/kg/day reported that there was a slight reduction in serum levels of thyroxine (T4) and triiodothyronine (T3) in men, which was not accompanied by any meaningful change in thyroid-stimulating hormone (TSH) levels (Wones et al. 1993). In another study in volunteers, ingestion of approximately 0.036 mg Cl/kg/day in the drinking water for 12 weeks did not significantly alter serum levels T3 or T4 (Lubbers et al. 1982).

No information was located regarding endocrine effects in acute-duration studies in animals. In an intermediate-duration reproductive study in male and female Long-Evans rats, dosing with up to 3.4 mg Cl/kg/day by gavage (hypochlorous acid) had no significant effect on serum thyroid hormone (unspecified) levels in males or females (Carlton et al. 1986). In the 90-day drinking water studies in

Sprague-Dawley and F344 rats and B6C3F₁ mice administered up to 24.9, 85, and 39.2 mg Cl/kg/day, respectively, there were no gross or microscopic alterations in the pancreas, adrenals, pituitary, thyroid, and parathyroid glands (Daniel et al. 1990, 1991; Furukawa et al. 1980). Similar results were obtained in the 2-year drinking water bioassay in F344 rats and B6C3F₁ mice (NTP 1992). Rats were exposed to up to 14.4 mg Cl/kg/day and mice were exposed to up to 24.2 mg Cl/kg/day. Hasegawa et al. (1986) also reported no alterations in the adrenal glands from F344 rats exposed to up 133 mg Cl/kg/day in the drinking water for 2 years.

Dermal Effects. No information was located regarding dermal effects in humans following oral exposure to hypochlorite solutions. The skin of rats and mice was examined in the 90-day (Daniel et al. 1990, 1991; Furukawa et al. 1980) and 2-year drinking water studies (Hasegawa et al. 1986; NTP 1992) and no significant gross or microscopic alterations related to treatment with chlorine were reported. In the 90-day studies, high doses in rats and mice were approximately 85 and 39.2 mg Cl/kg/day, respectively. In the 2-year studies, high doses in rats and mice were 133 and 24.2 mg Cl/kg/day.

Ocular Effects. No studies were located regarding ocular effects in humans following oral exposure to hypochlorite solutions. The only information from studies in animals is that exposure of F344 rats to up to 133 mg Cl/kg/day for 2 years did not increase the incidence of cataracts (Hasegawa et al. 1986). The eyes were not examined in rats or mice in the 90-day studies (Daniel et al. 1990, 1991) or in the NTP (1992) 2-year study.

Body Weight Effects. No studies were located regarding body weight effects in humans following oral exposure to hypochlorite solutions. Reduced growth was not observed in Wistar rats that received 200 mg Cl/kg/day (from sodium hypochlorite) by gavage in milk for 14 days (Cunningham 1980). Body weight was also not affected in Sprague-Dawley rats and B6C3F₁ mice exposed to up to 24.9 and 39.2 mg Cl/kg/day, respectively, in the drinking water for 90 days (Daniel et al. 1990, 1991) or in F344 rats and B6C3F₁ mice exposed to up to 14.4 and 24.2 mg Cl/kg/day, respectively, in the drinking water for 2 years (NTP 1992). Hasegawa et al. (1986) reported that male Fisher-24 rats exposed to approximately 152 or 305 mg Cl/kg/day (from sodium hypochlorite) in the drinking water for 90 days had reductions of 19 and 47% in final body weight compared with control rats; however, neither food nor water consumption data were provided in the study. In another 90-day study, male F344 dosed with approximately 50 mg Cl/kg/day (as sodium hypochlorite) in water had a final body weight 19% lower than controls; this group of rats also consumed 42% less water daily than controls (Furukawa et al. 1980). Rats dosed with 84 mg Cl/kg/day consumed 66% less water than controls and their final body weight was 46% lower than

controls (Furukawa et al. 1980). In the 2-year study conducted by the same investigators, final body weights were reduced 11 and 20% in female rats dosed with 67 and 133 mg Cl/kg/day, respectively. The investigators stated that in high-dose females water intake was somewhat lower (no quantitative data provided) during the first year, but not in the second year.

Metabolic Effects. Hypernatremia and hyperchloremic acidosis were reported in a woman who intentionally drank about 500 mL of a strong bleach solution (10% sodium hypochlorite) (Ward and Routledge 1988). Treatment with 5% dextrose gradually improved her electrolytes and 5 days later, she had recovered. Hyperchloremic metabolic acidosis also was reported in a woman who intentionally ingested an unknown amount of bleach (5.25% sodium hypochlorite, pH 11.4) and eventually died of cardiac arrest 4.5 hours after ingestion of the bleach solution (Ross and Spiller 1999). No other relevant information was located regarding metabolic effects in humans or animals following oral exposure to hypochlorite solutions.

3.2.2.3 Immunological and Lymphoreticular Effects

No studies were located regarding immunological effects in humans following oral exposure to hypochlorite solutions. Limited information is available in animals. In an 8-week study in Sprague-Dawley rats, exposure to 4.1 mg Cl/kg/day (the highest dose tested) in the drinking water (as sodium hypochlorite) resulted in alterations in several immunological parameters including reduced delayed-type hypersensitivity (DTH) reaction, increased prostaglandin E2 synthesis by macrophages, and reduced oxidative metabolism by macrophages following stimulation with phorbol myristate acetate (Exon et al. 1987). In the absence of systemic toxicity in rats dosed with much higher dose of chlorine in longer-term studies, the toxicological significance of these changes is unknown. In 90-day (Daniel et al. 1990, 1991; Furukawa et al. 1980) and 2-year (Hasegawa et al. 1986; NTP 1992) studies in rats and mice dosed with chlorine in the drinking water, there were no significant gross or microscopic alterations in the spleen, thymus, and lymph nodes. In the 90-day studies Sprague-Dawley and F344 rats and B6C3F₁ mice were exposed to up to 24.9, 85, and 39.2 mg Cl/kg/day, respectively. In the 2-year studies, high-doses in F344 and rats and B6C3F₁ mice were 133 and 24.2 mg Cl/kg/day, respectively. In none of these long-term studies was immunocompetence evaluated.

The highest NOAEL values from each reliable study for immunological and lymphoreticular effects in each species and duration category are recorded in Table 3-3 and plotted in Figure 3-2.

3.2.2.4 Neurological Effects

No studies were located regarding neurological effects in humans following exposure to hypochlorite solutions. One acute-duration study reported that exposure of Wistar rats to up to 200 mg Cl/kg/day (as sodium hypochlorite) by gavage in milk had no significant effect on brain weight, but no other neurological end point was evaluated in this study (Cunningham 1980). No gross or microscopic alterations were reported in the brain and sciatic nerve of Sprague-Dawley or F344 rats and B6C3F₁ mice in 90-days drinking water studies (Daniel et al. 1990, 1991; Furukawa et al. 1980). In these studies, rats and mice received doses of up to 85 and 39.2 mg Cl/kg/day, respectively. In the NTP (1992) 2-year drinking water study in F344 rats and B6C3F₁ mice, gross and microscopic examination of several brain areas did not show any significant alteration that could be attributed to treatment with chlorine. Hasegawa et al. (1986) also reported no histological alterations in the brain of F344 rats dosed with up to 133 mg Cl/kg/day (as sodium hypochlorite) in the drinking water for 2 years. None of these studies reported any adverse neurological sign in the animals throughout the studies, but no neurological tests were performed.

The highest NOAEL values from each reliable study for neurological effects in each species and duration category are recorded in Table 3-3 and plotted in Figure 3-2.

3.2.2.5 Reproductive Effects

No information was located regarding reproductive effects in humans following oral exposure to hypochlorite solutions. In an acute-duration drinking water study, exposure of male B6C3F₁ mice to ≥1.6 mg Cl/kg/day (as sodium hypochlorite) for 5 days resulted in significant increases in sperm abnormalities in mice sacrificed 3 weeks after dosing ceased, but not in mice sacrificed 1 or 5 weeks after dosing (Meier et al. 1985). In addition, no sperm abnormalities were seen in mice that were treated in the same manner with hypochlorous acid. According to the investigators, the results were somewhat surprising since sodium hypochlorite should be converted in to hypochlorous acid in acid pH of the stomach. In the absence of corroborating information from other studies and lack of internal consistency of the results, the toxicological significance of these results is difficult to ascertain; therefore, this study is not listed in Table 3-3.

In an intermediate-duration study, exposure of male and female Long-Evans rats to up to 3.4 mg Cl/kg/day (as hypochlorous acid) by gavage before and during breeding and of the females throughout gestation and lactation did not affect fertility, the gross and microscopic appearance of the reproductive

organs of males or females, and did not induce sperm abnormalities (Carlton et al. 1986). Neither the 90-day (Daniel et al. 1990, 1991; Furukawa et al. 1980) nor the 2-year (Hasegawa et al. 1986; NTP 1992) drinking water studies reported any gross or histological alterations in the reproductive organs of male and female rats and mice. These studies, however, did not assess fertility.

The highest NOAEL values from each reliable study for reproductive effects in each species and duration category are recorded in Table 3-3 and plotted in Figure 3-2.

3.2.2.6 Developmental Effects

No information was located regarding developmental effects in humans following oral exposure to hypochlorite solutions. Abdel-Rahman et al. (1982) exposed female Sprague-Dawley rats to 0, 0.1, 1, or 10 mg Cl/kg/day (as hypochlorous acid) in the drinking water for 2.5 months before mating with untreated males and continuing throughout pregnancy until sacrifice on gestation day 20. Exposure to chlorine had no significant effects on fetal viability or on mean fetal body weight. Skeletal anomalies were increased at 1 and 10 mg/kg/day and total soft tissue defects at 10 mg/kg/day relative to controls. However, since neither maternal body weight nor water consumption data were provided in the study, it also appeared that the incidence of fetal anomalies in the control group were higher than in the low-dose group, interpretation of these results is problematic; therefore, this study is not listed in Table 3-3. In the study by Carlton et al. (1986) mentioned above under *Reproductive Effects*, exposure of rats during gestation to maternal doses of 3.4 mg Cl/kg/day had no significant effect on neonate viability, weight gain, or on the incidence of gross external abnormalities. Pups sacrificed at 21 days of age had normal blood counts and serum levels of thyroid hormones. Developmental landmarks such as mean day of eye opening and average day of observed vaginal patency were unaltered in pups evaluated at age 28 and 40 days. The developmental NOAEL of 3.4 mg Cl/kg/day is listed in Table 3-3 and plotted in Figure 3-2.

3.2.2.7 Cancer

Studies of the carcinogenicity of trihalomethanes or other organic chemicals that form in water as a result of the chlorination of drinking water are not discussed in this section since these studies were not intended to assess whether chlorine itself is responsible for cancer. For reviews on this issue, the reader is referred to IARC (1991), Koivusalo and Vartiainen (1997), and EPA (1994b).

Cancer bioassays of chlorine in drinking water have been conducted in rats and mice. In the NTP (1992) bioassay, F344 rats (70/sex/dose group) were exposed to 0, 70, 140, or 275 ppm sodium hypochlorite in

the drinking water for 103–104 weeks. This provided doses of 0, 4.2, 7.3, or 13.6 mg Cl/kg/day to males and 0, 4.2, 7.8, or 14.4 mg Cl/kg/day to females. The water used in the study was deionized charcoalfiltered water. Interim sacrifices (10 rats/sex/dose) were conducted at 14 and 66 weeks. The only significant finding was an increased incidence of leukemia in female rats. The incidences were: 8/50, 7/50/, 19/51, and 16/50 in the control, low-, mid-, and high-dose females, respectively. Pair-wise comparison showed a statistically significant difference between controls and the mid-dose (p=0.014) and a trend test was also significant (p=0.037). In males, the respective incidences were 25/51, 25/51, 27/50, and 29/51. These results led NTP (1992) to conclude that there was equivocal evidence of carcinogenicity in female rats based on the fact that there was no clear dose-related response or reduced latency, there was no decrease in tumor latency, and the incidence in concurrent controls (16%) was significantly lower than in historical controls (25%). In a similar study, Hasegawa et al. (1986) administered sodium hypochlorite in distilled water to groups of F344 rats (50/sex/dose) in concentrations of 0, 500, or 1,000 ppm to males and 0, 1,000, or 2,000 ppm to females for 104 weeks; this was followed by a period of 8 weeks of drinking untreated water. This corresponds to doses of approximately 0, 33, or 67 mg Cl/kg/day for males and 0, 67, or 133 mg Cl/kg/day for females. The results showed no significant treatment-related increased incidence of neoplasms or alterations in latency of neoplasms.

In the NTP (1992) study, B6C3F₁ mice (70/sex/dose group) drank water with 0, 70, 140, or 275 ppm sodium hypochlorite for 103–104 weeks. This corresponds to doses of approximately 0, 7.4, 14, or 24 mg Cl/kg/day for males and 0, 7.6, 14.2, or 24.2 mg Cl/kg/day for females. The water used in the study was deionized charcoal-filtered water. Interim sacrifices (10 mice/sex/dose) were conducted at 15 and 66 weeks. The results showed that treatment with chlorine did not induce significant treatment-related increases in neoplasms. In a study by Kurokawa et al. (1986), groups of B6C3F₁ mice (50/sex/dose) were exposed to sodium hypochlorite in drinking water in concentrations of 0, 0.5, or 1% for 103 weeks. These concentrations provided doses of approximately 0, 33, or 55 mg Cl/kg/day to males and 0, 27, or 52 mg Cl/kg/day to females. Controls consisted of 73 males and 72 females. At termination, examination of tissues and organs did not show any statistically significant differences in tumor incidences between controls and treated mice.

Neither the EPA nor the DHHS has classified chlorine (elemental) or hypochlorite salts as to their carcinogenicity. Based on inadequate evidence for carcinogenicity of hypochlorite salts in animals and no data from studies in humans, IARC determined that hypochlorite salts are not classifiable as to their carcinogenicity in humans (Group 3) (IARC 1991).

3.2.3 Dermal Exposure

3.2.3.1 Death

No studies were located regarding death in humans or in animals following dermal exposure to hypochlorite solutions.

3.2.3.2 Systemic Effects

No studies were located regarding respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, endocrine, and body weight effects in humans or in animals following dermal exposure to hypochlorite solutions.

The highest NOAEL values and all LOAEL values from each reliable study for systemic effects in each species and duration category are recorded in Table 3-4.

Dermal Effects. Nixon et al. (1975) reported that patch application of bleach containing 5.25% sodium hypochlorite, pH 10.7, to intact human skin for 4 hours was severely irritating. Habets et al. (1986) reported that a solution of 2% sodium hypochlorite in water caused weak to moderate skin irritation in 15 out of 69 individuals tested; the volume applied was not disclosed. Twenty individuals who were tested with 0.1 and 0.5% solutions showed no irritation. Hostynek et al. (1989) tested 10 subjects with 100 μL of a 6% strength sodium hypochlorite solution (pH 11.2) and 4 of them developed a non-immunologic form of contact urticaria within 20 minutes of the application to skin of the forehead. The same group of investigators studied the skin irritation of 20 or 100 μL of hypochlorite bleach containing 1% sodium hypochlorite and various amounts of sodium hydroxide following 24 hours exposure under occluded patch conditions in 50 volunteers (Hostynek et al. 1990). The results showed that 20 μL 1% sodium hypochlorite and 1% sodium hydroxide produced no irritation, whereas 100 μL produced significant irritation. Goffin et al. (1997) reported that patch test exposure of 15 women to 150 μL of a commercial bleaching agent containing 4% sodium hypochlorite and 0.2% sodium hydroxide for up to 90 minutes produced no clinical signs of irritation. However, instrumental tests (reflectance colorimetry, transepidermal water loss, and skin conductance) revealed subclinical damage to the stratum corneum. The investigators concluded that a 4% solution of sodium hypochlorite can alter the superficial part of the stratum corneum without modifying the barrier function of the skin.

Table 3-4 Levels of Significant Exposure to Hypochlorous Acid and/or Sodium Hypochlorite _ Dermal

Species (Strain)	Exposure/ Duration/ Frequency (Route)			LOAE			
		System	NOAEL	Less Serious	Serious	Reference Chemical Form	Comments
	XPOSURE						
Systemic Human	90 min (C)	Dermal	4 F Percent (%)			Goffin et al. 1997	Instrumental tests revealed subclinical damage to the stratum corneum.
Human	48 hr (C)	Dermal	0.5 Percent (%)	2 (weak to moderate skin Percent (%) irritation)		Habets et al. 1986	
Human	20 min (C)	Dermal		6 (non-immunologic Percent (%) contact urticaria)		Hostynek et al. 1989	Non-immunologic contact urticaria developed within 20 minutes of application.
Human	24 hr (C)	Dermal	1 Percent (%)			Hostynek et al. 1990	Application of 100 microliters induced irritation; 20 microliters did not.
Human	4 hr (C)	Dermal		5.25 (severe skin irritation in Percent (%) 4/7 subjects)		Nixon et al. 1975	
Mouse (CD-1)	2 d 80%/d	Dermal		0.53 F (histologic changes indicating moderate irritation response)		Hess et al. 1991	The solution was applied as a spray.

Table 3-4 Levels of Significant Exposure to Hypochlorous Acid and/or Sodium Hypochlorite _ Dermal (continued)

Species (Strain)	Exposure/ Duration/ Frequency (Route)	LOAEL					
		System	NOAEL	Less Serious	Serious	Reference Chemical Form	Comments
Gn Pig (Hartley)	4 hr (C)	Dermal	5.25 Percent (%)			Nixon et al. 1975	
Rabbit (New Zealand)	once (C)	Ocular		5 B (moderate eye irritation) Percent (%)		Griffith et al. 1980	Median day to clea varied from 7 to ov 21 depending on volume applied.
Rabbit NS)	4 hr (C)	Dermal	5.25 Percent (%)			Nixon et al. 1975	

B = both male and female; (C) = capsule; d = day; F = female; Gn pig = guinea pig; hr = hour; LOAEL = lowest-observed-adverse-effect level; min = minute; NOAEL = no-observed-adverse-effect level

Nixon et al. (1975) conducted 4-hour patch tests with a 5.25% hypochlorite bleach solutions in rabbits and guinea pigs and reported that in both species, hypochlorite caused only slight irritation to both intact and abraded skin and concluded that neither species provide an accurate model for human skin. Strange et al. (1951) studied the effect of Clorox® mixed with various biological media on the skin of rabbits and rats. Clorox® was mixed with water, saliva, gastric juice, or plasma in ratios of 2:1, 1:1, and 1:2 and applied to the shaved abdominal skin of anesthetized rabbits in inverted tubes for 15 and 30 minutes. The mixtures with saliva and water were the most damaging, whereas the mixture with plasma was the least damaging, regardless of the dilution. Strange et al. (1951) speculated that the proteins in solutions containing plasma had a buffer effect on the harmful action of Clorox® on tissue. Strange et al. (1951) also reported that submerging the feet of anesthetized rats for 15 minutes into Clorox® mixed with proteins (plasma, egg white, milk) protected the tissue from the action of Clorox®. Mixtures of Clorox® with saliva or Clorox® with water induced edema, bleeding, and actual ulceration and destruction of tissue.

Hess et al. (1991) sprayed 0.8 mL of a 1:10 dilution of a commercial bleach solution onto a shaved abdominal area of female CD-1 mice 8 times/day for 2 consecutive days. Eighteen to 24 hours after the last application, the mice were sacrificed and the skin was processed for microscopic examination of the epidermis, dermis, and hypodermis. Exposure to bleach caused a "moderate" response. Grossly, the skin appeared dry with scattered brown crusty patches. Acanthosis, intraepithelial edema, hyperkeratosis, and atypical epithelial cells were seen in the epidermis. The dermis showed some evidence of edema, whereas the hypodermis showed a mild infiltration of neutrophils, macrophages, and lymphocytes.

Ocular Effects. Very limited information was located regarding ocular effects of direct contact of the eye with hypochlorite solutions. In their text *Toxicology of the Eye*, Grant and Schuman (1993) state that "because most accidental splashes in the eye have been with the relatively weak 5% household solutions of sodium hypochlorite, very few human eye injuries have been reported, and recovery has been rapid and complete."

Experiments conducted in male and female New Zealand albino rabbits showed that instillation of 0.1 mL of household bleach directly to the central corneal surface and followed over a 21-day period produced moderate irritation (Griffith et al. 1980). The median day to clear was 7 days. In a review of the literature, Racioppi et al. (1994) mention unpublished data indicating that in rabbits, 0.1 mL of an 8% solution of sodium hypochlorite (without rinsing) caused moderate irritation and that the recovery time

was 7 days; under similar conditions, 0.01 mL of the same solution had low irritation potential and the recovery time was 3 days.

3.2.3.3 Immunological and Lymphoreticular Effects

Although sodium hypochlorite generally is not considered a contact sensitizer, several cases of allergic contact dermatitis have been reported. Osmundsen (1978) reported that case of a woman had a strong reaction to patch testing with 0.5% sodium hypochlorite in water years after having had dermal contact with chloramine. Further tests showed positive reactions to sodium hypochlorite in 3 out of 225 patients. Habets et al. (1986) reported two cases of hand dermatitis related to sodium hypochlorite allergy, as diagnosed by patch tests. Both patients showed a positive reaction to sodium hypochlorite up to a concentration of 0.1%. Van Joost et al. (1987) reported one additional case among 40 housewives who apparently had used bleaching agents for long periods. Eun et al. (1984) also reported a case of allergic contact dermatitis in a veterinarian who occasionally washed his hands with a commercial solution containing 4–6% sodium hypochlorite.

No information was located regarding immunological and lymphoreticular effects in animals following dermal exposure to hypochlorite solutions.

No studies were located regarding the following effects in humans or animals after dermal exposure to hypochlorite solutions.

- 3.2.3.4 Neurological Effects
- 3.2.3.5 Reproductive Effects
- 3.2.3.6 Developmental Effects

3.2.3.7 Cancer

No information was located regarding cancer in humans due to dermal exposure to hypochlorite solutions. No bioassays have been conducted in animals by the dermal route of exposure, but the co-carcinogenic properties of hypochlorite have been examined. In a two-stage study in female ddN mice using 4-nitroquinoline 1-oxide in benzene as initiator, application of a commercial sodium hypochlorite solution (45 times) to a shaved area in the back of the mice after 20 applications of the nitroquinoline solution resulted in an incidence of 9/32 skin tumors compared with 0/29 in the nitroquinoline alone group and 0/27 in the hypochlorite group alone (Hayatsu et al. 1971). Although there is suggestive evidence of

promotion activity for sodium hypochlorite, the study is limited by insufficient information on dose, survival, and age at termination. In another initiation-promotion study, skin application of single initiating doses of 7,12-dimethylbenz[a]anthracene in acetone to female Sencar mice followed by twice weekly applications of sodium hypochlorite resulted in squamous cell carcinomas in one mouse initiated with 7,12-dimethylbenz[a]anthracene followed by sodium hypochlorite applications (Kurokawa et al. 1984). No skin carcinomas were seen in uninitiated mice promoted with sodium hypochlorite or in control groups. In yet another study, application of sodium hypochlorite to the shaved back of female NMRI mice twice per week for 10 weeks before the mice were applied benzo[a]pyrene twice per week for 10 weeks resulted in a decrease (approximately 40%) in the number of skin carcinomas induced by benzo[a]pyrene (Pfeiffer 1978). Mice that were treated with sodium hypochlorite during or after benzo[a]pyrene had tumor incidences comparable to benzo[a]pyrene alone.

3.3 GENOTOXICITY

No studies were located regarding genotoxic effects of chlorine gas in humans. The only information available in animals exposed to chlorine gas is that from an intermediate-duration study by Kutzman (1983). In that study, rats were exposed to 0, 0.5, 1.5, or 5 ppm chlorine 6 hours/day, 5 days/week for 62 days. At termination, samples of blood and bone marrow were collected and evaluated for cytogenetic effects. The results showed no evidence on increased incidence of sister chromatid exchanges or of cellular proliferation in the bone marrow from exposed rats. There was also no evidence of increased sister chromatid exchanges or of chromosomal aberrations in peripheral lymphocytes.

There is limited information regarding the *in vivo* genotoxicity of hypochlorite ions. A study in which male B6C3F₁ mice were administered chlorine in the drinking water as sodium hypochlorite or hypochlorous acid for 5 days and that provided doses of up to 8 mg Cl/kg/day found no evidence of increased incidences of chromosomal aberrations or micronuclei in bone marrow (Meier et al. 1985). In another study, administration of a single intraperitoneal dose of up to 2,500 mg/kg sodium hypochlorite (1,175 mg Cl/kg/day) to male ddY mice did not increase the incidence of micronuclei in bone marrow evaluated 24 hours after dosing (Hayashi et al. 1988). Exposure of newt larvae to sodium hypochlorite in the surrounding water (0.12 or 0.25 μg/mL) for 12 days increased the frequency of micronuclei in blood erythrocytes (Le Curieux et al. 1993). However, the study did not specify in what type of water the larvae were kept. If the larvae were kept in tap water, it is possible that chlorination byproducts rather than chlorine or the hypochlorite anion were the clastogenic agents. Table 3-5 summarizes the genotoxicity of sodium hypochlorite *in vitro* are summarized

Table 3-5. Genotoxicity of Sodium Hypochlorite In Vivo

Species (test system)	End point	Results	Reference
Mice bone marrow cells	Micronuclei	_	Meier et al. 1985
Mice bone marrow cells	Chromosomal aberrations	-	Meier et al. 1985
Mice bone marrow erythrocytes	Micronuclei	-	Hayashi et al. 1988
Newt (<i>Pleurodeles waltl</i>) larvae erythrocytes	Micronuclei	+	Le Curieux et al. 1993

^{- =} negative result; + = positive result

in Table 3-6. As the table shows, the results have been mixed and no general statements can be made. The variability of the results may be due to differences in the experimental protocols used.

3.4 TOXICOKINETICS

3.4.1 Absorption

3.4.1.1 Inhalation Exposure

Nodelman and Ultman (1999a) measured the fraction of an inspired chlorine bolus cleared during a single breath as a function of the bolus penetration into the respiratory system of five nonsmoker males and females during both nasal and oral breathing at a respiratory flow of 250 mL/second using a noninvasive procedure. Measurements of the chlorine concentrations were made by means of a fast-responding thermionic chlorine analyzer. Peak concentrations of 0.5 and 3 ppm chlorine were used in nasal breathing experiments and 3 ppm in oral breathing experiments. The results indicated that almost all of the chlorine inhaled was absorbed in the upper airways (above the cords) whether the subjects inhaled through the nose or through the mouth. By comparing mass transfer parameters, the investigators also determined that total absorption rates for the mouth and nose were similar. When the peak concentration in the nasal breathing experiments was increased from 0.5 to 3 ppm, the mass transfer parameters remained unchanged, indicating that the dissolution, diffusion, and chemical reactions governing the absorption of the gas by the nasal mucosa are all linear processes. In other words, over the 0.5–3 ppm concentration range, absorption appeared to be non-saturable. In a separate experimental series, the investigators determined the longitudinal distribution of a bolus of 3 ppm chlorine as a function of the flow rate (Nodelman and Ultman 1999b). Using flow rates of 150, 250, or 1,000 mL/second, the authors determined that irrespective of the mode of breathing, nasal or oral, and respiratory flow rate, >95% of the inspired chlorine was absorbed in the upper airways and <5% was delivered to the lower airways.

Some studies in animals also have provided indirect evidence that the upper respiratory tract is an efficient scrubber of chlorine. For example, Alarie (1981) reported that the 10-minute LC₅₀ in intact mice was 302 ppm, but in mice that breathed through a cannula in the trachea, the LC₅₀ was only 131 ppm. In another study, the surgically isolated upper respiratory tract of anesthetized mice was shown to scrub chlorine with a mean efficiency of approximately 98% (Morris et al. 2005). Preliminary studies in F344 rats similar to those conducted by Nodelman and Ultman (1999a, 1999b) in humans also showed that >90% of a chlorine dose (0.5, 1.0, 2.5 ppm) is cleared in the upper respiratory tract (Roberts et al. 2007).

Table 3-6. Genotoxicity of Sodium Hypochlorite *In Vitro*

		Results		
Species (test system)	End point	With activation	Without activation	Reference
Salmonella typhimurium (TA100, TA102, TA98)	Gene mutation	Not tested	-	Le Curieux et al. 1993
S. typhimurium (TA100)	Gene mutation	Not tested	+	Ishidate et al. 1984
S. typhimurium (TA1530)	Gene mutation	Not tested	+	Wlodkowski and Rosenkranz 1975
S. typhimurium (TA1538)	Gene mutation	Not tested	-	Wlodkowski and Rosenkranz 1975
Escherichia coli (PQ37)	DNA repair	Not tested	-	Le Curieux et al. 1993
<i>E. coli</i> (DNA polymerase deficient)	DNA damage	Not tested	+	Rosenkranz 1973
Human fibroblasts cell line HE2144	Sister chromatid exchange	Not tested	+	Sasaki et al. 1980
Humans fibroblasts cell line HE1244	Chromosome breakage	Not tested	_	Sasaki et al. 1980
Syrian hamster embryo cells	Sister chromatid exchange	Not tested	+	Miyachi and Tsutsui 2005
Chinese hamster fibroblast cell line	Chromosomal aberrations	Equivocal	+	Ishidate et al. 1984
Chinese hamster lung cell line	Chromosomal aberrations	Toxic	+	Matsuoka et al. 1979
Syrian hamster embryo cells	Chromosomal aberratons	+	-	Hagiwara et al. 2006

^{— =} negative result; + = positive result; DNA = deoxyribonucleic acid

3.4.1.2 Oral Exposure

No relevant data were located regarding oral absorption of hypochlorite ions in humans and limited information exists in studies in animals. Administration of a single dose of approximately 3.3 mg/kg of radiolabeled hypochlorous acid (³⁶Cl) to fasted Sprague-Dawley resulted in a peak ³⁶Cl in plasma of 7.9 μg/mL 2 hours after dosing (Abdel-Rahman et al. 1983). In non-fasted rats, administration of the same dose resulted in a peak concentration of 10.7 μg/mL ³⁶Cl in plasma 4 hours after dosing, suggesting that food residues slow down absorption perhaps due to the reaction of chlorine with food components (Fukayama et al. 1986). For both fasted and non-fasted rats, the absorption half-life was 2.2 hours. The elimination half-life from plasma was 44.1 and 88.5 hours in fasted and non-fasted rats, respectively.

3.4.1.3 Dermal Exposure

No information was located regarding absorption of hypochlorite ions in humans or in animals after dermal exposure to hypochlorite solutions. Studies of dermal exposure of humans or animals to aqueous chlorine have focused almost exclusively of effects on the skin; therefore, no systemic effects have been described that could provide indirect evidence of dermal absorption of hypochlorite ions.

3.4.2 Distribution

3.4.2.1 Inhalation Exposure

No information was located regarding distribution of chlorine following inhalation exposure to chlorine gas.

3.4.2.2 Oral Exposure

In rats, 96 hours after the administration of a single gavage dose of approximately 3 mg/kg of hypochlorous acid labeled with 36 Cl, the plasma had the highest amount of radioactivity (1.92 µg/g) followed by whole blood, bone marrow, testes, skin, kidneys, lungs, packed cells, duodenum, stomach, spleen, thyroid, thymus, liver, carcass, and fat (0.09 µg/g) (Abdel-Rahman et al. 1983). Examination of the subcellular distribution of radioactivity in the liver 24 hours after dosing with hypochlorous acid showed that 75% of the total radioactivity of the whole homogenate was associated with the cytosol, 2.5% with the microsomal fraction, 1.5% with the nuclear fraction, and <0.1% with the mitochondrial fraction.

3.4.2.3 Dermal Exposure

No information was located regarding distribution of chlorine following dermal exposure to chlorine gas or hypochlorite ions.

3.4.3 Metabolism

Limited information is available regarding the metabolism of chlorine. In a study in which rats received a single oral gavage dose of radiolabeled (³⁶Cl) hypochlorous acid showed that, 96 hours after dosing, 81% of radioactivity detected in plasma was chloride ion (Abdel-Rahman et al. 1983).

Hypochlorous acid is a very reactive chemical and has been shown to react with biomolecules found in food (Fuyakama et al. 1986). Hypochlorous acid reacts with proteins, amino acids, and unsaturated lipids to form chlorinated compounds, whereas the reaction with carbohydrates yields oxidation products. Scully et al. (1986) reported that chlorination of the stomach from rats resulted in the production of N-chloramines, tentatively identified as N-chloroalanine, N-chloroglycine, and N-chlorophenylalanine. Chemicals such as chloroform, dichloroacetonitrile and di- and trichloroacetic acids were shown to form *in vivo* in the stomach of rats following oral administration of sodium hypochlorite (Mink et al. 1983).

3.4.4 Elimination and Excretion

3.4.4.1 Inhalation Exposure

No information was located regarding elimination and excretion of chlorine following inhalation exposure to the gas.

3.4.4.2 Oral Exposure

The excretion of ³⁶Cl was studied in rats following administration of a single gavage dose of approximately 2.6 mg/kg of radiolabeled hypochlorous acid (Abdel-Rahman et al. 1983). Urine, feces, and expired air were collected over a 4-day period after dosing. During the first 24 hours, 7 and 7.5% of the administered radioactivity was excreted in the urine and feces, respectively. At the end of the 4-day period, 36 and 15% of the administered radioactivity had been recovered in the urine and feces, respectively, for a combined total of about 51% of the administered dose. No radioactivity was recovered in expired air during the study period. Since all or some of the amount recovered in the feces could have

been un-absorbed radioactivity, the 36% recovered in the urine represents the minimum that was absorbed.

3.4.4.3 Dermal Exposure

No information was located regarding elimination and excretion of chlorine gas or hypochlorite ions in humans or in animals following dermal exposure to these chemicals.

3.4.5 Physiologically Based Pharmacokinetic (PBPK)/Pharmacodynamic (PD) Models

Physiologically based pharmacokinetic (PBPK) models use mathematical descriptions of the uptake and disposition of chemical substances to quantitatively describe the relationships among critical biological processes (Krishnan et al. 1994). PBPK models are also called biologically based tissue dosimetry models. PBPK models are increasingly used in risk assessments, primarily to predict the concentration of potentially toxic moieties of a chemical that will be delivered to any given target tissue following various combinations of route, dose level, and test species (Clewell and Andersen 1985). Physiologically based pharmacodynamic (PBPD) models use mathematical descriptions of the dose-response function to quantitatively describe the relationship between target tissue dose and toxic end points.

PBPK/PD models refine our understanding of complex quantitative dose behaviors by helping to delineate and characterize the relationships between: (1) the external/exposure concentration and target tissue dose of the toxic moiety, and (2) the target tissue dose and observed responses (Andersen and Krishnan 1994; Andersen et al. 1987). These models are biologically and mechanistically based and can be used to extrapolate the pharmacokinetic behavior of chemical substances from high to low dose, from route to route, between species, and between subpopulations within a species. The biological basis of PBPK models results in more meaningful extrapolations than those generated with the more conventional use of uncertainty factors.

The PBPK model for a chemical substance is developed in four interconnected steps: (1) model representation, (2) model parameterization, (3) model simulation, and (4) model validation (Krishnan and Andersen 1994). In the early 1990s, validated PBPK models were developed for a number of toxicologically important chemical substances, both volatile and nonvolatile (Krishnan and Andersen 1994; Leung 1993). PBPK models for a particular substance require estimates of the chemical substance-specific physicochemical parameters, and species-specific physiological and biological parameters. The numerical estimates of these model parameters are incorporated within a set of differential and algebraic

equations that describe the pharmacokinetic processes. Solving these differential and algebraic equations provides the predictions of tissue dose. Computers then provide process simulations based on these solutions.

The structure and mathematical expressions used in PBPK models significantly simplify the true complexities of biological systems. If the uptake and disposition of the chemical substance(s) are adequately described, however, this simplification is desirable because data are often unavailable for many biological processes. A simplified scheme reduces the magnitude of cumulative uncertainty. The adequacy of the model is, therefore, of great importance, and model validation is essential to the use of PBPK models in risk assessment.

PBPK models improve the pharmacokinetic extrapolations used in risk assessments that identify the maximal (i.e., the safe) levels for human exposure to chemical substances (Andersen and Krishnan 1994). PBPK models provide a scientifically sound means to predict the target tissue dose of chemicals in humans who are exposed to environmental levels (for example, levels that might occur at hazardous waste sites) based on the results of studies where doses were higher or were administered in different species. Figure 3-3 shows a conceptualized representation of a PBPK model.

If PBPK models for chlorine exist, the overall results and individual models are discussed in this section in terms of their use in risk assessment, tissue dosimetry, and dose, route, and species extrapolations.

A computational fluid dynamics-physiologically based pharmacokinetic model is being developed for chlorine (Jarabek et al. 2007). The model is intended to address experimental dosimetry data on chlorine in rats, including uptake of chlorine delivered unidirectionally at various flow rates and concentrations measured in the isolated upper respiratory tract of F344 rats, and measurement of chlorotyrosines (as biomarkers) in samples taken from four regions from the respiratory and olfactory tissues.

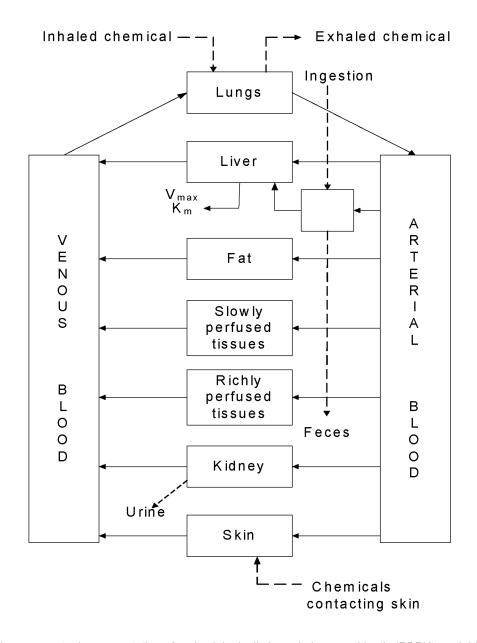
3.5 MECHANISMS OF ACTION

3.5.1 Pharmacokinetic Mechanisms

Chlorine is a strong oxidizer that hydrolyzes in water forming hydrochloric and hypochlorous acids. Because of the high water content of the epithelial lining fluid and the local concentration of chloride and pH, the hydrolysis reaction of chlorine has a large equilibrium constant, such that the concentration of chlorine in the form of hypochlorite is 120,000 times that of chlorine gas (Nodelman and Ultman 1999a,

Figure 3-3. Conceptual Representation of a Physiologically Based Pharmacokinetic (PBPK) Model for a Hypothetical

Chemical Substance



Note: This is a conceptual representation of a physiologically based pharmacokinetic (PBPK) model for a hypothetical chemical substance. The chemical substance is shown to be absorbed via the skin, by inhalation, or by ingestion, metabolized in the liver, and excreted in the urine or by exhalation.

Source: Adapted from Krishnan and Andersen 1994

1999b). This means that the effective solubility of chlorine between the inhaled gas and the mucus phase is 5 orders of magnitude larger than the physical solubility, which explains why >95% of the inhaled chlorine was absorbed in the upper airways in the studies with volunteers mentioned above (Nodelman and Ultman 1999a).

In a review of the toxicological significance of the chemical reactions of hypochlorite ions, Scully et al. (1989) point out that because hypochlorite is a potent oxidant, pharmacokinetic studies of radiolabeled hypochlorous acid (³⁶Cl) in animals do not reveal what happens to the parent compound, but rather to the product of the reactions of these compounds *in vivo*. However, as discussed by Scully et al. (1986), since hypochlorous acid undergoes rapid isotope exchange with unlabeled chloride, it is unclear whether the radioactive chloride detected in plasma of rats in the study by Abdel-Rahman et al. (1983) is due to complete reduction of the hypochlorous acid or to isotope exchange followed by elimination of chloride.

3.5.2 Mechanisms of Toxicity

Chlorine Gas. The toxicity of chlorine is strongly related to its oxidizing capacity. Chlorine reacts with water in the epithelial lining of the upper respiratory airways according to the following equation:

$$Cl_2 + H_2O \leftrightarrow HCl + HOCl$$

Chlorine gas has been shown to be 33 times more potent as a sensory irritant in mice than hydrochloric acid (HCl) (Barrow et al. 1977), which led the investigators to suggest that, in terms of sensory irritation, the response observed must be due to hypochlorous acid (HOCl) rather than to hydrochloric acid. More recently, Morris et al. (2005) reported that in mice, (a) an aerosol of sodium hypochlorite and (b) chlorine gas, at equivalent air concentrations, induced similar decreases in respiratory rate and increases in specific airway resistance, which suggested to the investigators that the oxidant properties of chlorine alone are sufficient to account for the observed responses. The precise mechanism by which this might occur is not known, but the assumption is that products of the reaction of chlorine with water are able to interact with functional groups in components from cells in the respiratory epithelium. At low concentrations, only sensory receptors may be affected, triggering only changes in respiratory dynamics, but higher concentrations produce frank tissue damage due to disruption of cellular components.

Hypochlorous Acid/Hypochlorite. The mechanism of toxicity of hypochlorous acid/hypochlorite is basically the same as that for chlorine gas. However, hypochlorous acid is a stronger oxidant than

chlorine gas as reflected by its higher redox potential. Damage to the upper gastrointestinal tract, as may occur following ingestion of sodium hypochlorite bleach, is likely the result of oxidation reactions of hypochlorous acid with a range of biological molecules. Exposure to dilute solutions of bleach usually result in only minor esophageal irritation, but ingestion of concentrated solutions of bleach can produce serious tissue damage. These properties of hypochlorous acid form the basis for the use of this chemical as a disinfectant (i.e., Lapenna and Cuccurullo 1996; Schraufstätter et al. 1990; Wang et al. 2007). Due to its high reactivity, ingested hypochlorous acid also reacts with organic compounds present in the stomach fluid such as proteins, polysaccharides, lipids, and vitamins, which may result in the formation of potentially harmful compounds (for review, see Fukayama et al. 1986; Mink et al. 1983; Scully et al. 1989).

3.5.3 Animal-to-Human Extrapolations

The respiratory system is the target for exposure to chlorine gas in humans and animals, and for the most part, humans and animals exhibit similar effects, particularly following acute-duration high exposures. Less is known regarding long-term effects of acute high exposures and of prolonged low level exposures, especially with regard to pulmonary function parameters. An evaluation of respiratory lesions in monkeys, rats, and mice following chronic exposure to comparable concentrations of chlorine noted that respiratory tract airflow characteristics play a major role in the distribution of the lesions and that the lesions in monkeys and rodents exhibited both differences and similarities (Ibanes et al. 1996). The conclusion was that with appropriate exposure and response adjustments, both rodents and Rhesus monkeys appear to be valid models for human risk assessment.

The gastrointestinal tract is the target for oral exposure to hypochlorite in humans and animals. Ingestion of concentrated solutions of hypochlorite bleach induces similar effects in humans and animals, and no single animal species has emerged as a preferred animal model for human gastric toxicity. A comparative study of dermal exposure of humans, rabbits, and guinea pigs to a 5.25% hypochlorite bleach solution reported that rabbits and guinea pigs were much more sensitive to the irritating properties of sodium hypochlorite, and therefore, neither species provide an accurate model for human skin (Nixon et al. 1975).

3.6 TOXICITIES MEDIATED THROUGH THE NEUROENDOCRINE AXIS

Recently, attention has focused on the potential hazardous effects of certain chemicals on the endocrine system because of the ability of these chemicals to mimic or block endogenous hormones. Chemicals with this type of activity are most commonly referred to as *endocrine disruptors*. However, appropriate

terminology to describe such effects remains controversial. The terminology endocrine disruptors, initially used by Thomas and Colborn (1992), was also used in 1996 when Congress mandated the EPA to develop a screening program for "...certain substances [which] may have an effect produced by a naturally occurring estrogen, or other such endocrine effect[s]...". To meet this mandate, EPA convened a panel called the Endocrine Disruptors Screening and Testing Advisory Committee (EDSTAC), and in 1998, the EDSTAC completed its deliberations and made recommendations to EPA concerning endocrine disruptors. In 1999, the National Academy of Sciences released a report that referred to these same types of chemicals as hormonally active agents. The terminology endocrine modulators has also been used to convey the fact that effects caused by such chemicals may not necessarily be adverse. Many scientists agree that chemicals with the ability to disrupt or modulate the endocrine system are a potential threat to the health of humans, aquatic animals, and wildlife. However, others think that endocrine-active chemicals do not pose a significant health risk, particularly in view of the fact that hormone mimics exist in the natural environment. Examples of natural hormone mimics are the isoflavinoid phytoestrogens (Adlercreutz 1995; Livingston 1978; Mayr et al. 1992). These chemicals are derived from plants and are similar in structure and action to endogenous estrogen. Although the public health significance and descriptive terminology of substances capable of affecting the endocrine system remains controversial, scientists agree that these chemicals may affect the synthesis, secretion, transport, binding, action, or elimination of natural hormones in the body responsible for maintaining homeostasis, reproduction, development, and/or behavior (EPA 1997). Stated differently, such compounds may cause toxicities that are mediated through the neuroendocrine axis. As a result, these chemicals may play a role in altering, for example, metabolic, sexual, immune, and neurobehavioral function. Such chemicals are also thought to be involved in inducing breast, testicular, and prostate cancers, as well as endometriosis (Berger 1994; Giwercman et al. 1993; Hoel et al. 1992).

There are no studies that have tested whether chlorine gas has properties of endocrine disruptor, but given its mechanism of toxicity, such effects are very unlikely.

Oral exposure to hypochlorite solutions has provided no evidence of endocrine disruption in humans or in animals, but the available studies have not been designed to carefully evaluate that possibility. A study in male and female volunteers who consumed for 4 weeks 1.5 L of distilled water that provided a dose of approximately 0.4 mg Cl/kg/day reported that there was a slight reduction in serum levels of T4 and T3 in men which was not accompanied by any meaningful change in thyroid-stimulating hormone levels (Wones et al. 1993). In another study in volunteers, ingestion of approximately 0.036 mg Cl/kg/day in the drinking water for 12 weeks did not significantly alter serum levels T3 or T4 (Lubbers et al. 1982).

Relevant information in animals is very limited. Exposure of rats during gestation to maternal doses of up to 3.4 mg Cl/kg/day (as hypochlorous acid) had no significant effect on pups' serum levels of thyroid hormones at 21 days of age (Carlton et al. 1986). In addition, developmental landmarks such as mean day of eye opening and average day of observed vaginal patency were unaltered in pups evaluated at age 28 and 40 days. Studies in rats and mice found no significant gross or microscopic alterations in endocrine glands following long-term exposure to chlorine the drinking water (Daniel et al. 1990, 1991; Furukawa et al. 1980; Hasegawa et al. 1986; NTP 1992).

3.7 CHILDREN'S SUSCEPTIBILITY

This section discusses potential health effects from exposures during the period from conception to maturity at 18 years of age in humans, when all biological systems will have fully developed. Potential effects on offspring resulting from exposures of parental germ cells are considered, as well as any indirect effects on the fetus and neonate resulting from maternal exposure during gestation and lactation. Relevant animal and *in vitro* models are also discussed.

Children are not small adults. They differ from adults in their exposures and may differ in their susceptibility to hazardous chemicals. Children's unique physiology and behavior can influence the extent of their exposure. Exposures of children are discussed in Section 6.6, Exposures of Children.

Children sometimes differ from adults in their susceptibility to hazardous chemicals, but whether there is a difference depends on the chemical (Guzelian et al. 1992; NRC 1993). Children may be more or less susceptible than adults to health effects, and the relationship may change with developmental age (Guzelian et al. 1992; NRC 1993). Vulnerability often depends on developmental stage. There are critical periods of structural and functional development during both prenatal and postnatal life, and a particular structure or function will be most sensitive to disruption during its critical period(s). Damage may not be evident until a later stage of development. There are often differences in pharmacokinetics and metabolism between children and adults. For example, absorption may be different in neonates because of the immaturity of their gastrointestinal tract and their larger skin surface area in proportion to body weight (Morselli et al. 1980; NRC 1993); the gastrointestinal absorption of lead is greatest in infants and young children (Ziegler et al. 1978). Distribution of xenobiotics may be different; for example, infants have a larger proportion of their bodies as extracellular water, and their brains and livers are proportionately larger (Altman and Dittmer 1974; Fomon 1966; Fomon et al. 1982; Owen and Brozek

1966; Widdowson and Dickerson 1964). The infant also has an immature blood-brain barrier (Adinolfi 1985; Johanson 1980) and probably an immature blood-testis barrier (Setchell and Waites 1975). Many xenobiotic metabolizing enzymes have distinctive developmental patterns. At various stages of growth and development, levels of particular enzymes may be higher or lower than those of adults, and sometimes unique enzymes may exist at particular developmental stages (Komori et al. 1990; Leeder and Kearns 1997; NRC 1993; Vieira et al. 1996). Whether differences in xenobiotic metabolism make the child more or less susceptible also depends on whether the relevant enzymes are involved in activation of the parent compound to its toxic form or in detoxification. There may also be differences in excretion, particularly in newborns who all have a low glomerular filtration rate and have not developed efficient tubular secretion and resorption capacities (Altman and Dittmer 1974; NRC 1993; West et al. 1948). Children and adults may differ in their capacity to repair damage from chemical insults. Children also have a longer remaining lifetime in which to express damage from chemicals; this potential is particularly relevant to cancer.

Certain characteristics of the developing human may increase exposure or susceptibility, whereas others may decrease susceptibility to the same chemical. For example, although infants breathe more air per kilogram of body weight than adults breathe, this difference might be somewhat counterbalanced by their alveoli being less developed, which results in a disproportionately smaller surface area for alveolar absorption (NRC 1993).

The respiratory tract is also the target for chlorine gas toxicity in children, and children exposed to chlorine exhibit the same signs and symptoms observed in adults detailed in Section 3.2.1.2. Whether children are more susceptible than adults to the effects of chlorine exposure is not known with certainty, but there are some reports that suggest that they might be. There are many reports of intoxication of children following a variety of exposure scenarios: tank explosions, train derailments, accidents at swimming pools, and accidents during school science class. Some representative examples are summarized below.

Following a train derailment in which members of a community, including over 100 children, may have been exposed to up 20 ppm chlorine, the most frequent conditions were coughing, headache, throat irritation, and a burning sensation in the eyes (Agency for Toxic Substances and Disease Registry 1998). Children aged 0–5 years had the highest prevalence of respiratory infections, rash, and vomiting. Fleta et al. (1986) reported the case of a leak of chlorine from a tank that exposed 76 children. The most prevalent symptoms were irritative cough (91%), nasal-pharyngeal pruritus (66%), chest pain (25%),

tachypnea (20%), and dyspnea (14%). Other conditions included headache, vomiting, and nausea. The report indicates that the symptoms of most children subsided naturally upon leaving the center of contamination. Seventy of the 76 children were released within 2 hours of the accident. In a similar case, 106 individuals were affected and 60 of them were children and adolescents <18 years (Güloğlu et al. 2002). Of those hospitalized due to their severe condition, patients 0–1 and 2–7 years had the longest duration of hospitalization, suggesting an increased susceptibility among children than adults.

Sexton and Pronchik (1998) described the case of 13 children, ages 6-18, who were overcome by chlorine gas at two swimming pools. On admission to the Emergency Department, most children complained of eve and throat irritation, chest pain, anxiety, shortness of breath, wheezing, and chest tightness. Five patients were admitted to the hospital with hypoxia, but all were released after 2 days. Two weeks after the exposures, no patients complained of residual symptoms. In another swimming pool accident, 134 children inhaled chlorine vapors and acute respiratory symptoms occurred in 72% (Agabiti et al. 2001). The incidence of all symptoms tended to be higher among those who had a history of chronic respiratory disease and among those who were engaged in physical exercise during the accident. Lung function tests conducted in 82 children 15–30 days after exposure showed lower FVC, FEV₁, and FEF_{25-75%}. A study of 10 children exposed to chlorine gas at a swimming pool reported that all children had respiratory distress and reduced lung function on admission to the hospital (Bonetto et al. 2006). Although lung function returned to normal values 15 days after the accident, biochemical markers of pulmonary inflammation were still elevated in exhaled breath condensate several months after the accident. Persistent respiratory alterations were also observed in a child who developed dyspnea, hypoxemia, and pneumonitis approximately 12 hours after exposure to chlorine gas (Vohra and Clark 2006). Pulmonary testing 4 months after the episode revealed the presence of mild obstructive reactivity of the airways. After being exposed to excess chlorine in an indoor pool, 18 children were followed for up to 28 days (Grasemann et al. 2007). Fractional exhaled NO (a measure of airway inflammation) was significantly increased on day 1 compared to days 8 and 28 in 13 of the children who had lower airway symptoms with cough or shortness of breath on day 1, but not in the total study group or in asymptomatic children. In the total group of children, FVC was decreased on day 1 compared to days 8 and 28, whereas a marker of pulmonary hyperinflation was elevated on day 1 compared to day 8 or 28.

There are no studies in animals that addressed possible differential susceptibilities to exposure to chlorine gas between young and older animals.

In children, as in adults, the gastrointestinal tract is the target for oral exposure to hypochlorite bleach. Children usually constitute a significant percentage of the reported cases of accidental ingestion of chlorine bleach. In general, ingestion of small amounts (less than a cup) of household bleach (5– 5.5% sodium hypochlorite) does not result in serious effects, but fatalities have been reported. For example, a review of 129 cases of children who ingested Clorox® reported that no complications of consequences were found (Pike et al. 1963). Sixty-five cases were examined by esophagoscopy within 96 hours of the ingestion and only 2 showed evidence of esophageal injury. Another review of 393 children seen at a hospital who ingested bleach reported that there were no esophageal strictures or perforations and that about 50% of the patients received no treatment (Landau and Saunders 1964). Hook and Lowry (1974) reported that among 23 definite cases of children who ingested Clorox[®], severe irritation of the esophageal mucosa was observed in only one case. Minor transient irritation was observed in some of the patients. A report from the German literature of 23 children who accidentally ingested 3–5% sodium hypochlorite indicates that there was only one case with signs of superficial burns in the esophagus, which had disappeared 2 weeks later when controlled by esophagoscopy (Mühlendahl et al. 1978). Severe gross lesions were seen in the mouth, tongue, glottis, epiglottis, esophagus, and stomach in a child who died after drinking an unknown amount of a 4.5% sodium hypochlorite solution in alkali (pH 12.0) (Jakobsson et al. 1991). The information available does not suggest that children are more or less sensitive to oral exposure to chlorine than adults.

There are no studies in animals that have examined whether young animals are more susceptible to the effects of ingestion of hypochlorite bleach than adult animals.

3.8 BIOMARKERS OF EXPOSURE AND EFFECT

Biomarkers are broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility (NAS/NRC 1989).

A biomarker of exposure is a xenobiotic substance or its metabolite(s) or the product of an interaction between a xenobiotic agent and some target molecule(s) or cell(s) that is measured within a compartment of an organism (NAS/NRC 1989). The preferred biomarkers of exposure are generally the substance itself, substance-specific metabolites in readily obtainable body fluid(s), or excreta. However, several factors can confound the use and interpretation of biomarkers of exposure. The body burden of a substance may be the result of exposures from more than one source. The substance being measured may

be a metabolite of another xenobiotic substance (e.g., high urinary levels of phenol can result from exposure to several different aromatic compounds). Depending on the properties of the substance (e.g., biologic half-life) and environmental conditions (e.g., duration and route of exposure), the substance and all of its metabolites may have left the body by the time samples can be taken. It may be difficult to identify individuals exposed to hazardous substances that are commonly found in body tissues and fluids (e.g., essential mineral nutrients such as copper, zinc, and selenium). Biomarkers of exposure to chlorine are discussed in Section 3.8.1.

Biomarkers of effect are defined as any measurable biochemical, physiologic, or other alteration within an organism that, depending on magnitude, can be recognized as an established or potential health impairment or disease (NAS/NRC 1989). This definition encompasses biochemical or cellular signals of tissue dysfunction (e.g., increased liver enzyme activity or pathologic changes in female genital epithelial cells), as well as physiologic signs of dysfunction such as increased blood pressure or decreased lung capacity. Note that these markers are not often substance specific. They also may not be directly adverse, but can indicate potential health impairment (e.g., DNA adducts). Biomarkers of effects caused by chlorine are discussed in Section 3.8.2.

A biomarker of susceptibility is an indicator of an inherent or acquired limitation of an organism's ability to respond to the challenge of exposure to a specific xenobiotic substance. It can be an intrinsic genetic or other characteristic or a preexisting disease that results in an increase in absorbed dose, a decrease in the biologically effective dose, or a target tissue response. If biomarkers of susceptibility exist, they are discussed in Section 3.10, Populations That Are Unusually Susceptible.

3.8.1 Biomarkers Used to Identify or Quantify Exposure to Chlorine

There are no biomarkers that can be used to quantify exposure to chlorine gas or sodium hypochlorite in humans. However, a recent study documented a statistically significant exposure concentration-dependent increase in 3-chlorotyrosine (CY) and 3,5-dichlorotyrosine (dCY) in nasal tissue from F344 rats exposed to chlorine gas concentrations ranging from 0.5 to 2.5 ppm for 90 minutes (Sochaski et al. 2008). Preferential formation of CY and dCY was found in the respiratory and transitional epithelium versus the olfactory epithelium of the nasal cavity. The investigators point out that the method presented in the study is only applicable to animals, as the collection of tissue samples would be too invasive for use in humans; nevertheless, it represents a good starting point for future method development. It is not

known whether exposure to other chlorinated compounds could also produce CY and dCY, thus compromising the specificity of these biomarkers.

3.8.2 Biomarkers Used to Characterize Effects Caused by Chlorine

There are no specific biomarkers that can be used to characterize the effects of chlorine. Chlorine gas is a sensory and pulmonary irritant, and similar effects can be observed after exposure to many other irritants. Ingestion of sodium hypochlorite can irritate the upper gastrointestinal tract, but this effect is not specific to chlorine.

3.9 INTERACTIONS WITH OTHER CHEMICALS

The only relevant information is that exposure to chlorine gas may result in the development of cross-tolerance to other chemicals. A study showed rats pre-exposed to chlorine developed cross-tolerance to formaldehyde and vice versa, and the development of tolerance was a function of the duration of pre-treatment (Chang and Barrow 1984). Slight loss of cross-tolerance was observed following a recovery period of a few days. Interestingly, exposure of rats to 15 ppm formaldehyde did not induce tolerance to formaldehyde, but resulted in cross-tolerance to chlorine and, according to Chang and Barrow (1984), suggested the existence of different reactive sites at the trigeminal nerve endings.

As previously mentioned, hypochlorous acid is a very reactive chemical and has been shown to react with biomolecules found in food (Fuyakama et al. 1986). Hypochlorous acid reacts with proteins, amino acids, and unsaturated lipids to form chlorinated compounds, whereas the reaction with carbohydrates yields oxidation products. Scully et al. (1986) reported that chlorination of the stomach from rats resulted in the production of N-chloramines, tentatively identified as N-chloroalanine, N-chloroglycine, and N-chlorophenylalanine. Chemicals such as chloroform, dichloroacetonitrile, and di- and trichloroacetic acids were shown to form *in vivo* in the stomach of rats following oral administration of sodium hypochlorite (Mink et al. 1983).

3.10 POPULATIONS THAT ARE UNUSUALLY SUSCEPTIBLE

A susceptible population will exhibit a different or enhanced response to chlorine than will most persons exposed to the same level of chlorine in the environment. Reasons may include genetic makeup, age, health and nutritional status, and exposure to other toxic substances (e.g., cigarette smoke). These parameters result in reduced detoxification or excretion of chlorine, or compromised function of organs

affected by chlorine. Populations who are at greater risk due to their unusually high exposure to chlorine are discussed in Section 6.7, Populations with Potentially High Exposures.

Populations unusually susceptible to chlorine gas exposure include individuals with respiratory conditions such as asthma, hay fever, and chronic bronchitis, heavy smokers, and children. Rotman et al. (1983) described the case of an atopic individual who experienced severe distress during exposure to 1 ppm chlorine, a concentration that was tolerated by healthy subjects. D'Alessandro et al. (1996) also reported that subjects with airway hyperresponsiveness to methacholine exhibited a much more pronounced decrease in FEV₁ and FEF_{25-75%} than healthy subjects during exposure to 1 ppm chlorine. Shusterman et al. (1998) reported that subjects with seasonal allergic rhinitis experienced a significantly greater increase in nasal airway resistance (congestion) than non-rhinitic subjects following exposure to 0.5 ppm chlorine for 15 minutes. Following an accidental leak of chlorine, individuals who had a more prevent history of smoking and asthma exhibited more hypoxemia and were more likely to have tachypnea, crackles, and wheezes during examination than subjects without asthma and/or who smoked less (Hasan et al. 1983). In the former, signs and symptoms of chlorine intoxication resolved more slowly reduced and flow rates and lung volumes were still evident 2 weeks after acute exposure to chlorine. Similar observations regarding smokers have been made in studies of workers who have experienced occasional high exposures or "gassing" episodes (Chester et al. 1969; Gautrin et al. 1999; Henneberger et al. 1996).

In a swimming pool accident involving 126 adult and 134 children, among both children and adults, the incidences of all symptoms (eye, nose, and throat irritation) and respiratory problems (shortness of breath, wheezing, cough) were higher among those who had a history of chronic respiratory disease than among healthy people (Agabiti et al. 2001). In addition, in adults, incidences were higher among smokers and former smokers than among never smokers.

Some reports in which adults and children were accidentally exposed to high concentrations of chlorine have suggested that children might be more susceptible to the effects of chlorine than adults. For example, in a case involving 106 individuals, 60 of whom were children and adolescents <18 years old, of those hospitalized due to their severe condition, patients 0–1 and 2–7 years old had the longest duration of hospitalization, suggesting an increased susceptibility among children than adults (Güloğlu et al. 2002). In another case in which over 100 children may have been exposed to up 20 ppm chlorine, children aged 0–5 years had the highest prevalence of respiratory infections, rash, and vomiting (Agency for Toxic Substances and Disease Registry 1998).

No information was located regarding populations unusually susceptible to exposure to hypochlorite bleach.

3.11 METHODS FOR REDUCING TOXIC EFFECTS

This section will describe clinical practice and research concerning methods for reducing toxic effects of exposure to chlorine. However, because some of the treatments discussed may be experimental and unproven, this section should not be used as a guide for treatment of exposures to chlorine. When specific exposures have occurred, poison control centers and medical toxicologists should be consulted for medical advice. The following texts provide specific information about treatment following exposures to chlorine:

Ellenhorn MJ, Barceloux DG. 1988. Medical Toxicology: Diagnosis and Treatment of Human Poisoning. New York, NY: Elsevier, 878-879.

Ellenhorn MJ. 1997. Medical Toxicology: Diagnosis and Treatment of Human Poisoning. 2th ed. Philadelphia, PA: Williams and Wilkins, 1521.

Goldfrank LR, Flomenbaum NE, Lewin NA, et al. 2002. Goldfrank's Toxicologic Emergencies. 7th ed. New York, NY: McGraw-Hill, 1458-1459.

Viccellio P, Bania T, Brent J, et al. 1998. Chlorine gas. In: Emergency toxicology. 2nd ed. Philadelphia, PA: Lippincott-Raven Press, 444-445.

Additional information can be found in ATSDR's Medical Management Guideline for chlorine (ATSDR 2007).

3.11.1 Reducing Peak Absorption Following Exposure

There are no specific methods for reducing absorption of chlorine gas other than removing the patient from the source of the chlorine gas to fresh air and monitor for respiratory distress. It should be noted that rescuers should wear self-contained breathing apparatus and have protective clothing, if needed.

Sodium hypochlorite or hypochlorous acid is not absorbed in the gastrointestinal tract as such. Either substance will react with the acid in the stomach to form chlorine gas and/or with organic compounds present in the stomach fluid to form chlorinated compounds (Mink et al. 1983; Scully et al. 1989). Gastrointestinal decontamination procedures such as emesis, gastric lavage, and activated charcoal should be avoided following ingestion of chlorine bleach. However, dilution with water or milk is recommended, but the dilution amount should be small to avoid inducing vomiting. In case of exposure

of the skin to hypochlorite solutions, flushing with copious amounts of plain tepid water is recommended. In case of exposure of the eyes, irrigation with saline or Ringer's lactate is recommended.

3.11.2 Reducing Body Burden

There are no standard methods for reducing chlorine body burden. Studies in humans have shown that under low exposure conditions (<5 ppm), >95% of the inspired chlorine is absorbed in the upper airways and <5% is delivered to the lower airways (Nodelman and Ultman 1999a, 1999b). Chlorine that reacts with the mucosa of the upper respiratory airways eventually joins the pool of chloride ions in the body. Studies in animals also have shown that most of the chlorine ingested as hypochlorous acid is transformed and eliminated as chloride (Abdel-Rahman et al. 1983).

3.11.3 Interfering with the Mechanism of Action for Toxic Effects

The toxic effects of chlorine gas are due to its oxidant properties and also to the added tissue damage caused by the hypochlorous and hydrochloric acids that result from the reaction of chlorine with water. There are no established methods to interfere with the oxidant properties of chlorine, but nebulized sodium bicarbonate has been used to neutralize the acid (Bosse 1994; Douidar 1997).

The treatment of exposure to chlorine gas is symptomatic, exposure to low concentrations may require only treatment for sensory irritation, but exposure to high concentrations may cause serious respiratory symptoms including pulmonary edema and respiratory failure and death. The information below has been extracted from the texts listed above and also from Baxter et al. (1989).

Before any treatment, the patient should be assessed for signs of corrosive injury to mucous membrane, eyes, and skin. The assessment should also include a check for lung sounds, peak flow, and vital signs. Patients heavily exposed who show breathing difficulties at rest should undergo baseline x-ray examination. The initial treatment consists of irrigation with water or saline and vasoconstrictive ophthalmic solutions for eye irritation, but eye damage may require referral to a health care facility. Nausea may be treated with Phenergan® and administration of clear liquids, whereas sore throat can be treated with throat lozenges or spray or a humidifier. Decongestants are recommended for rhinitis and antitussive agents for the treatment of cough. Skin burns should be treated as thermal burns. Patients exhibiting respiratory effects should receive 100% humidified oxygen, unless it is contraindicated by the medical history. As mentioned above, 5% nebulized bicarbonate has been used in patient with respiratory effects with favorable responses in at least some patients (Bosse 1994; Douidar 1997). Nebulized

bronchodilators may be used to treat bronchospasm. Therapy with corticosteroids has not been proved to produce improvement in chlorine gas poisoning (Baxter et al. 1989). Monitoring of respiratory function and arterial blood gases in important because pulmonary edema may occur up to 24 hours after exposure. If pulmonary edema occurs, emergent treatment and monitoring in an intensive care unit is often required. Caution should be exercised with the administration of intravenous fluids and because fluid overload is extremely dangerous in such patients. If fluid overload occurs, diuretics such as furosemide may be useful as indicated. Survivors of high chlorine exposure should be monitored periodically to determine possible persistent loss of pulmonary function.

3.12 ADEQUACY OF THE DATABASE

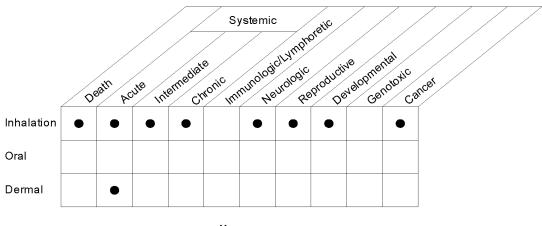
Section 104(I)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of chlorine is available. Where adequate information is not available, ATSDR, in conjunction with the National Toxicology Program (NTP), is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of chlorine.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

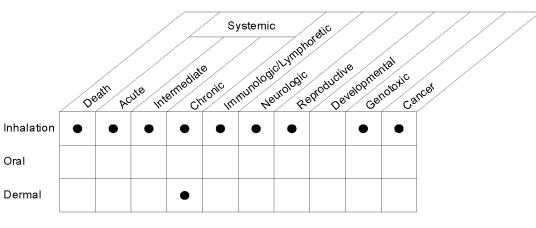
3.12.1 Existing Information on Health Effects of Chlorine

The existing data on health effects of inhalation, oral, and dermal exposure of humans and animals to chlorine gas and hypochlorite solutions chlorine are summarized in Figures 3-4 and 3-5, respectively. The purpose of these figures is to illustrate the existing information concerning the health effects of chlorine. Each dot in the figure indicates that one or more studies provide information associated with that particular effect. The dot does not necessarily imply anything about the quality of the study or studies, nor should missing information in this figure be interpreted as a "data need". A data need, as defined in ATSDR's *Decision Guide for Identifying Substance-Specific Data Needs Related to Toxicological Profiles* (Agency for Toxic Substances and Disease Registry 1989), is substance-specific

Figure 3-4. Existing Information on Health Effects of Chlorine Gas



Human

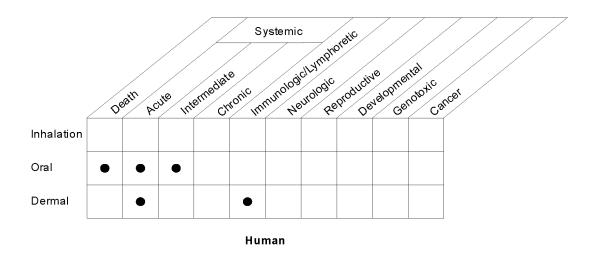


Animal

Existing Studies

3. HEALTH EFFECTS

Figure 3-5. Existing Information on Health Effects of Hypochlorite Solution



Existing Studies

information necessary to conduct comprehensive public health assessments. Generally, ATSDR defines a data gap more broadly as any substance-specific information missing from the scientific literature.

The existing information on the health effects of chlorine gas in humans comes from accounts of soldiers exposed during gas attacks in World War I, subjects exposed to chlorine at work, and members of the general population accidentally exposed due to leaks or explosions of storage tanks or due to the mishandling of bleach solutions or swimming pool chemicals. Regardless of the exposure scenario, the target for chlorine gas toxicity is the respiratory system. Effects of exposure to low concentrations may be limited to irritation of the eyes, upper respiratory tract, and skin, whereas exposure to high concentrations may cause serious pulmonary effects and death. There is limited information on neurological effects in humans exposed to chlorine gas. Oral exposure is not a relevant route of exposure to chlorine gas in humans or animals.

Acute-, intermediate-, and chronic inhalation studies of chlorine gas in animals are available. In these studies, the respiratory tract was also established as the target for chlorine toxicity. There are minimal data on neurological, lymphoreticular, reproductive, and genotoxic effects, as well as cancer, in animals.

The available information on health effects of hypochlorous acid or sodium hypochlorite in humans is derived almost exclusively from cases of accidental or intentional ingestion of chlorine bleach. These observations indicate that the principal target for oral exposure to hypochlorite is the upper gastrointestinal tract. Ingestion of small amounts of chlorine bleach may only cause esophageal irritation, but ingestion of strong solutions of bleach can cause severe damage to the upper gastrointestinal tract and even death. There are no data on long-term exposure of humans to hypochlorite bleach. Since no target for chlorine toxicity has been identified in oral studies of various durations with dose levels much higher than those that could be generally encountered by the general population, oral MRLs for hypochlorite were not derived. Additional oral studies are not considered necessary except for the few exceptions indicated in Section 3.12.2 below.

Information is available from acute-duration oral studies with hypochlorous acid or sodium hypochlorite in animals, which also indicates that the upper gastrointestinal tract is the main target of toxicity for concentrated bleach. Intermediate- and chronic-duration studies with hypochlorous acid or sodium hypochlorite in animals have examined systemic end points and also have provided limited information on immunological, neurological, reproductive, and developmental effects. There are cancer bioassays available with sodium hypochlorite in rats and mice.

There are data that indicate that dermal exposure to hypochlorite bleach can cause skin irritation in humans and in animals.

3.12.2 Identification of Data Needs

Acute-Duration Exposure. Information regarding health effects of acute exposure to chlorine gas is available from studies with volunteers (Anglen 1981; D'Alessandro et al. 1996; Rotman et al. 1983; Schins et al. 2000; Shusterman et al. 1998, 2003b), from exposures of soldiers during World War I (i.e., Berghoff 1919; DOA 1933; Joy 1997; Meakins and Priestley; Sandall 1922), and from accidental exposures of workers and the general public following chlorine leaks in a variety of scenarios (i.e., Agabiti et al. 2001; Agency for Toxic Substances and Disease Registry 1998; Bhérer et al. 1994; Bonetto et al. 2006; CDC 1991, 2005; Chasis et al. 1947; Chester et al. 1977; Edwards et al. 1983; Hasan et al. 1983; Jones et al. 1986; Kowitz et al. 1967; Moulick et al. 1992; Salisbury et al. 1991; Schönhofer et al. 1996; Sexton and Pronchik 1998; Weill et al. 1969). These and many additional studies showed that the respiratory tract is the target for chlorine toxicity and that the effects range from sensory irritation at low exposures (<5 ppm) to severe pulmonary effects (40–60 ppm) and possibly death (>100 ppm). Information is also available regarding long-term effects of acute high exposures to chlorine; some studies did not find persistent effects (i.e., Chasis et al. 1947; Jones et al. 1986; Moulick et al. 1992; Weill et al. 1969), whereas others did (i.e., Bhérer et al. 1994; Kowitz et al. 1967; Salisbury et al. 1991). Further research on this issue is needed. The studies in volunteers provided sufficient information for derivation of an acute-duration inhalation MRL for chlorine gas. The studies in animals support the findings in humans and provided additional information regarding histopathological changes in the respiratory tract and duration and reversibility of the effects. Standard additional acute-duration inhalation studies of chlorine gas in animals do not seem necessary at this time.

Information regarding health effects of hypochlorous acid and sodium hypochlorite in humans is derived exclusively from cases of accidental or intentional ingestion of hypochlorite bleach. These reports indicate that the upper gastrointestinal tract is the main target of toxicity for the oral route of exposure (Landau and Saunders 1964; Pike et al. 1963; Ross and Spiller 1999). The animal data support the findings in humans, but the available studies are inadequate for constructing dose-response relationships. Earlier studies in animals tried to reproduce the lesions to the esophagus and/or stomach due to ingestion of bleach. In most of these studies, commercial bleach was administered through a tube directly into the esophagus and, in some cases, the distal end of the esophagus was artificially occluded to prolong and

monitor the contact time between the solution and the mucosa (Hook and Lowry 1974; Landau and Saunders 1964; Strange et al. 1951; Yarington 1970). Two more recent studies were of very limited scope (Cunningham 1980) or reported ambiguous results (Meier et al. 1985).

Dermal effects have been reported in a few cases of direct acute contact of the skin with high concentrations of chlorine gas in humans (Agency for Toxic Substances and Disease Registry 1998; Joyner and Durel 1962; NIOSH 1995), and eye irritation was reported in volunteers exposed to 1 ppm chlorine for up to 8 hours (Anglen 1981; Rotman et al. 1983). Information on dose-response for sensory irritation was used along with data on pulmonary effects to derive the acute-duration inhalation MRL for chlorine. Additional studies of sensory irritation with chlorine gas do not appear necessary at this time. Chlorine gas is not absorbed through the skin, so systemic effects due to contact of the skin with chlorine are not expected to occur. Dermal effects of hypochlorite bleach have been reported in humans and in animals (Habets et al. 1986; Hostynek et al. 1989, 1990; Nixon et al. 1975; Strange et al. 1951); therefore, additional dermal studies do not seem necessary at this time.

Intermediate-Duration Exposure. No studies of humans exposed specifically for intermediate duration to chlorine gas were located. However, it is likely that in many of the occupational studies available, some workers were exposed for intermediate durations. Only two intermediate-duration studies in animals are available (Barrow et al. 1979; Kutzman 1983). Both studies utilized rats and in both studies, the most sensitive target for chlorine exposure was the respiratory tract. Barrow et al. (1979) described inflammation of the nasal turbinates in rats exposed to ≥ 1 ppm chlorine, whereas loss of cilia and epithelium in the trachea was seen in rats exposed to ≥ 0.5 ppm in the Kutzman (1983) study. The Kutzman (1983) study was selected as the principal study for derivation of an intermediate-duration inhalation MRL for chlorine. Additional intermediate-duration inhalation studies in animals do not seem necessary at this time.

Two intermediate-duration studies in which volunteers were exposed to known amounts of total chlorine from chlorinated water provided no evidence of adverse effects (Lubbers et al. 1982; Wones et al. 1993). Few intermediate-duration studies in animals were located that examined a wide range of end points following exposure to hypochlorite. These studies showed that the main effect of exposure to solutions of hypochlorous acid or sodium hypochlorite, particularly at the higher concentrations levels, is a reduction of water intake that is due to taste aversion. The available intermediate-duration oral studies evaluated systemic toxicity (Abdel-Rahman et al. 1984; Cunningham 1980; Daniel et al. 1990, 1991; Furukawa et al. 1980) and also provided information, albeit limited, on immunological/lymphoreticular (Daniel et al.

1990, 1991; Exon et al. 1987), neurological (Daniel et al. 1990, 1991), reproductive (Carlton et al. 1986; Daniel et al. 1990, 1991), and developmental effects (Carlton et al. 1986). None of the available studies reported effects that could be attributed directly to chlorine or only reported effects that were considered of unknown toxicological significance. Additional intermediate-duration oral studies with hypochlorite do not seem necessary. It is also unclear what needed information would be provided in additional intermediate-duration dermal studies.

Chronic-Duration Exposure and Cancer. There are relatively few long-term studies in workers exposed to chlorine gas (Enarson et al. 1984; Ferris et al. 1967, 1979; Hyback 1999; Patil et al. 1970). Health evaluations of the workers in these studies, including pulmonary function monitoring, did not provide evidence of significant health problems. In the Enarson et al. (1984) and Patil et al. (1970) studies, it was estimated that the workers were exposed to a TWA mean of 0.15-0.18 ppm chlorine. Yet, due to limitations, these long-term studies were insufficient for quantitative risk assessment. In none of the studies available was the nasal cavity, a sensitive target of chlorine exposure in humans and animals, examined. Therefore, evaluations of workers currently exposed to chlorine should include examination of the nasal cavity. In addition, future studies should include more reliable methods to estimate exposure. Two chronic-duration inhalation studies in animals are available. One of them studied the effects of chlorine inhalation in monkeys (Klonne et al. 1987) and the other in rats and mice (Wolf et al. 1995). In both, the upper respiratory tract was most sensitive end point and the study in monkeys was selected as basis for derivation of a chronic-duration inhalation MRL for chlorine gas. These studies also evaluated hematology and clinical chemistry parameters and, for the most part, no significant alterations were found. Additional chronic-duration inhalation studies of chlorine gas in animals do not seem necessary at this time.

No chronic-duration human studies of exposure to hypochlorous acid or sodium hypochlorite were located. Three chronic-duration studies were available in rats and mice (Hasegawa et al. 1986; NTP 1992). All three studies evaluated a comprehensive number of end points including hematology and clinical chemistry and tissue and organ histopathology and did not find any significant toxicity attributed to exposure to hypochlorite solutions. Additional chronic-duration oral studies with hypochlorite solutions are not necessary at this time.

There are several studies of cancer in humans exposed to chlorine gas, and probably simultaneously to other chemicals that did not find any evidence that chlorine gas is carcinogenic (Barbone et al. 1992; Barregård et al. 1990; Bond et al. 1983, 1985, 1986; Heldaas et al. 1989). The chronic-duration

inhalation study in rats and mice exposed to chlorine for 2 years found no evidence of carcinogenicity at termination (Wolf et al. 1995). It is unclear what useful information additional studies with chlorine gas would provide. There are no studies of cancer in humans exposed to hypochlorite ions, and it is unlikely that such a cohort can ever be found. Studies of cancer in humans exposed to chlorinated water have been conducted, but the focus of these studies has not been the hypochlorite ion but the chlorinated byproducts derived from the reaction of chlorine with organic matter in the water (see Kantor [1994] for review). Long-term drinking water bioassays have been conducted in rats (Hasegawa et al. 1986; NTP 1992) and mice (Kurokawa et al. 1986; NTP 1992) and, with one exception, the results were negative. Equivocal evidence of increased leukemia was reported in female rats in the NTP (1992) study. A need to conduct additional drinking water studies in the F344 strain may be unwarranted given that: (1) mononuclear cell leukemia occurs with high incidence in F344 rats; (2) the increase in leukemia was slight and not clearly dose-related; (3) there was no decrease in tumor latency; and (4) the incidence in concurrent controls was less than in historical controls. Although an option would be to conduct a study in a different strain of rat, the need would still have to be balanced with the fact that epidemiologic data do not suggest such an effect in humans.

Genotoxicity. No studies were located regarding genotoxic effects of chlorine gas in humans. The only information available in animals is that from a study in which blood and bone marrow from rats exposed to chlorine gas for 62 days showed no evidence on increased incidence of sister chromatid exchanges, chromosomal aberrations, or of cellular proliferation (Kutzman 1983). It does not appear that there is a need for additional genotoxicity studies for chlorine gas. Studies examining the *in vivo* genotoxicity of hypochlorite ions in mammals gave negative results (Hayashi et al. 1988; Meier et al. 1985) and studies *in vitro* in animal cells and bacteria gave mixed results (Hagiwara et al. 2006; Ishidate et al. 1984; Le Curieux et al. 1993; Miyachi and Tsutsui 2005; Sasaki et al. 1980). It is unlikely that additional studies with hypochlorite will settle the issue.

Reproductive Toxicity. The only information available regarding effect of chlorine gas in humans is that evaluation of the outcome of 15 pregnancies among female workers at a chlorine plant in 1932–1933 did not provide any evidence of reproductive toxicity (Skljanskaja et al. 1935). Pre-exposure of male or female rats to up to 5 ppm chlorine for 62 days followed by mating with untreated rats resulted in no significant effects on fertility, number of corpora lutea, viable embryos, early or late deaths, or pre-implantation loses. In addition, in males exposed for 62 days there were no histological alterations in the testes, and sperm morphology was unremarkable (Kutzman 1983). Chronic-duration inhalation studies with monkeys, rats, and mice exposed to up to 2.5 ppm chlorine did not observe gross or microscopic

lesions in the reproductive organs (Klonne et al. 1987; Wolf et al. 1995). It is plausible that accidental exposure of pregnant women to concentrations of chlorine high enough to produce severe hypoxia may affect pregnancy outcomes (i.e., stillbirth, abortions). Therefore, identification and evaluation of women who were pregnant during past chlorine accidents may provide valuable information. This can also be tested in animal models.

There is no information regarding reproductive effects in humans exposed to hypochlorite solutions and there are limited data in animals. An acute-duration study reported sperm abnormalities in mice treated with sodium hypochlorite, but not in mice treated with hypochlorous acid (Meier et al. 1985). This and other internal inconsistencies make this finding of unknown toxicological significance. A study in rats exposed to hypochlorous acid by gavage before and during breeding reported no significant effects on fertility or on histopathology of the reproductive organs of males or females (Carlton et al. 1986). Sperm was examined in this study and no significant alterations were reported. Long-term drinking water studies in rats and mice did not report any gross or microscopic alterations in the reproductive organs of males and females (Daniel et al. 1991, 1991; Furukawa et al. 1980; Hasegawa et al. 1986; NTP 1992). Although 2-generation reproductive studies with sodium hypochlorite (in water devoid of organic matter to eliminate the formation of chlorination byproducts) have not been conducted, the available information does not suggest that reproductive parameters are sensitive targets for hypochlorite. Thus, additional oral studies do not appear necessary at this time. There are no studies that evaluated reproductive parameters in humans or animals following dermal exposure to hypochlorite. However, dermal exposure to hypochlorite bleach is expected to affect only the site of exposure.

Developmental Toxicity. The only information available in humans exposed to chlorine gas is that evaluation of the outcome of 15 pregnancies among female workers at a chlorine plant in 1932–1933 did not provide any evidence of teratogenic effects (Skljanskaja et al. 1935). The same investigators reported that rabbits exposed to low chlorine concentrations (0.6–1.6 ppm) during pregnancy gave birth to healthy offspring (Skljanskaja and Rappoport 1935). There are no modern developmental studies of chlorine gas in animals. As mentioned in the preceding paragraph, exposure to high concentrations of chlorine gas during pregnancy could affect fetal or neonatal development, although so far, no study has examined that possibility. Therefore, identification and evaluation of women that were pregnant during past accidents in which chlorine gas was released may provide valuable information. Again, this can also be tested in animal models.

There is no information regarding developmental effects in humans exposed to hypochlorite. Only one reliable study in animals was available. In that study, exposure of pregnant rats to hypochlorous acid by gavage had no effect on neonate viability, weight gain, incidence of gross external abnormalities, or developmental landmarks (Carlton et al. 1986). Additional studies using a relevant means of administering chlorine, such as drinking water rather than gavage, may be necessary to confirm or refute the results of Carlton et al. (1986). There are no studies that evaluated developmental parameters in humans or animals following dermal exposure to hypochlorite bleach. However, as mentioned above, dermal exposure to bleach is expected to affect only the site of exposure.

Immunotoxicity. No studies were located that evaluated immunocompetence or effects on lymphoreticular organs in humans following exposure to chlorine gas. Studies in workers did not provide any evidence that exposure to chlorine gas may affect immunocompetence. A 6-week study reported that the spleen and thymus from rats exposed to 9 ppm chlorine showed decreased content of lymphoid elements, but according to the investigators, this may have been a function of the poor physical condition and decreased nutritional state of the rats in that dosing group (Barrow et al. 1979). Chronic-duration inhalation studies in moneys, rats, and mice found no gross or microscopic lesions in lymphoreticular organs and tissues (Klonne et al. 1987; Wolf et al. 1995). The immune system does not seem to be a sensitive target for chlorine gas toxicity; additional studies of the immune system in animals exposed by inhalation to chlorine gas are not necessary at this time.

There is no information regarding effects on the immune system in humans following oral exposure to chlorine. One study reported that exposure of rats to chlorine in the drinking water for 8 weeks resulted in alterations in some immune parameters (Exon et al. 1987). The toxicological significance of these findings is difficult to ascertain because there is no known mechanism by which oral administration of chlorine could induce immunological effects and no additional studies that could corroborate these findings. It is possible that oxidative reactions play a role in the effects reported by Exon et al. (1987). Other intermediate-duration studies and chronic-duration studies in rats and mice dosed with much higher doses of chlorine in the drinking water found no gross or microscopic alterations in lymphoreticular organs, but did not examine immunocompetence (Daniel et al. 1990, 1991; Furukawa et al. 1980; Hasegawa et al. 1986; NTP 1992). It would be useful to try to replicate the findings from Exon et al. (1987) adding to the protocol challenges with microorganisms to determine whether the reported alterations translate into decreased immunity.

Sodium hypochlorite is generally not considered a skin sensitizer, but several cases of allergic contact dermatitis have been reported (Eun et al. 1984; Habets et al. 1986; Osmundsen 1978; Van Joost et al. 1987). Additional dermal studies are not necessary

Neurotoxicity. A series of studies by Kilburn (1995, 2000, 2003b) suggested that brief exposures to high concentrations of chlorine gas can result in long-term neurological alterations in humans. No other high-exposure studies in humans have reported similar effects, but no neurobehavioral tests have been conducted in these studies. Therefore, it would be useful to conduct neurobehavioral evaluations in subjects known to have been exposed to high concentrations of chlorine to confirm or refute Kilburn's findings. The study should include comparison populations matched for the prior occurrence of a non-chemically related traumatic event. In addition, there are several validated animal models that have been used to test for neurobehavioral effects of chemicals (i.e., lead) that could be used to test for possible chlorine effects. The available studies in animals have provided no evidence of gross or microscopic alterations in tissues of the central or peripheral nervous system following exposure to chlorine gas.

There is no information regarding neurological effects in humans following exposure to hypochlorite bleach. The only information relevant information in animals is that from 90-day and 2-year studies that found no gross or microscopic alterations in the brain from rats and mice exposed to chlorine in the drinking water (Daniel et al. 1990, 1991; Furukawa et al. 1980; Hasegawa et al. 1986; NTP 1992). No further neurological end points were evaluated in these studies. Since the nervous system does not seem to be a sensitive target for oral chlorine, there is no compelling reason to conduct additional studies.

Epidemiological and Human Dosimetry Studies. There is a considerable number of studies of humans exposed to chlorine gas (see Section 3.2.1.2 for representative references). The effects of acute exposure to high concentrations of chlorine gas are known and concentration-response relationships have been established (Ellenhorn and Barceloux 1988). However, less is known about long-term effects of high acute exposures to chlorine gas and low-level, long-term exposure. As previously mentioned, some studies have described persistent pulmonary alterations following acute exposure to chlorine gas (i.e., Bhérer et al. 1994; Kowitz et al. 1967; Salisbury et al. 1991), whereas others have not (i.e., Chasis et al. 1947; Jones et al. 1986; Moulick et al. 1992; Weill et al. 1969). Better exposure data and baseline health information would seem necessary to establish reliable correlations between exposure and effects. Also, evaluation of the nasal cavity of low-level, long-term exposure to chlorine gas seems warranted in light of the findings of Klonne et al. (1987) in monkeys exposed to chlorine for 1 year.

Exposure to hypochlorous acid or sodium hypochlorite can occur by accidental or intentional ingestion of chlorine bleach. This type of exposure is generally of acute duration and, in most cases, the effects are restricted to esophageal irritation without long-term consequences (Hook and Lowry 1974; Landau and Saunders 1964; Pike et al. 1963). As discussed in Chapters 4 and 6, the level of dissolved chlorine in drinking water is extremely low and most of the free chlorine is as hypochlorous acid at the normal pH of drinking water. The highest level of chlorine allowed in drinking water is 4 ppm (EPA 2006a), which is considerably lower than the maximal concentration of chlorine used in long-term studies (275 ppm available chlorine) in rats and mice (NTP 1992), which caused no significant toxicity. Therefore, it seems unlikely that free chlorine in drinking water will represent a health concern for humans. It should be noted, however, that chlorinated water contains a variety of chlorinated byproducts whose biological effects continue to be studied.

Biomarkers of Exposure and Effect.

Exposure. There are no specific biomarkers of exposure for chlorine in humans. Chlorine gas that enters the airways or chlorine ingested as sodium hypochlorite eventually joins the chloride pool in the body.

A recent study showed that exposure of rats to chlorine gas (0.5–2.5 ppm) for 90 minutes resulted in dose-related formation of 3-chlorotyrosine and 3,5-dichlorotyrosine in nasal tissue (Sochaski et al. 2008). Since the method could not be used in humans due to the invasive nature of the tissue collection, additional research needs to be conducted to make the method less invasive. In addition, it would be valuable to determine whether exposure to other chlorinated compounds also result in the formation of these potential biomarkers.

Effect. There are no biomarkers of effect specific for chlorine. The sensory irritation and respiratory alterations caused by exposure to chlorine gas or the esophageal irritation caused by ingestion of hypochlorite bleach can also be caused by other chemicals.

Absorption, Distribution, Metabolism, and Excretion. The only information regarding pharmacokinetics of chlorine gas is that from experiments in volunteers conducted by Nodelman and Ultman (1999a, 1999b) that showed that almost all (>95%) of a bolus dose of chlorine gas inhaled through the mouth or the nose is cleared by the upper respiratory tract and none reaches the lungs. This was observed over a 0.5–3 ppm exposure range. The methodology used to generate the bolus and to monitor the concentrations of chlorine in the airways could probably be adapted to studies in animals,

particularly monkeys, to test a wider range of concentrations and to correlate internal concentrations of chlorine with lesions in the respiratory tract.

There is only one study of the pharmacokinetics of hypochlorite, the study by Abdel-Rahman et al. (1983) that evaluated absorption, metabolism, distribution, and excretion of chlorine in rats following gavage doses or radiolabeled (³⁶Cl) hypochlorous acid. Additional studies may be useful to confirm or refute the findings of Abdel-Rahman et al. (1983). On the other hand, as Scully et al. (1989) pointed out, because hypochlorite is a potent oxidant, pharmacokinetic studies of radiolabeled hypochlorous acid (³⁶Cl) in animals do not reveal what happens to the parent compound, but rather to the product of the reactions of these compounds *in vivo*. Therefore, the usefulness of additional studies is questionable. As previously mentioned, a computational fluid dynamics-physiologically based pharmacokinetic model is being developed for chlorine (Jarabek et al. 2007).

Comparative Toxicokinetics. The nature and distribution of lesions in the respiratory tract of monkeys were compared with those in rats and mice following chronic exposure to comparable concentrations (Ibanes et al. 1996). The investigators noted that monkeys and rodents exhibited both differences and similarities that were most likely related to the differences in airflow characteristics. Intuitively, it would seem that monkeys are a better model for human risk assessment because rodents are obligate nose breathers, whereas humans and monkeys are not. However, Ibanes et al. (1996) concluded that with appropriate exposure and response adjustments, both rodents and Rhesus monkeys appear to be valid models for human risk assessment.

An animal model for human exposure to hypochlorite has not been identified. Studies in rats and mice exposed to sodium hypochlorite by the oral route for 90 days or 2 years showed practically no toxicity of at the concentrations of chlorine tested (Daniel et al. 1990, 1991; NTP 1992). The gastrointestinal effects observed in humans after ingestion of high amounts of hypochlorite bleach (i.e., Ross and Spiller 1999) are similar to those described in earlier studies in dogs and rabbits exposed also to high amounts of hypochlorite bleach (Landau and Saunders 1964; Strange et al. 1951; Yarington 1970). Additional comparative studies do not seem necessary at this time.

Methods for Reducing Toxic Effects. The treatment of chlorine exposure is mostly supportive of respiratory and cardiovascular functions. The efficacy of some specific agents such as corticosteroids or nebulized sodium bicarbonate for the treatment of respiratory alterations due to exposure to chlorine gas has not been properly documented and further studies in animal models would be valuable.

Children's Susceptibility. Data needs relating to both prenatal and childhood exposures, and developmental effects expressed either prenatally or during childhood, are discussed in detail in the Developmental Toxicity subsection above.

Children exposed to high concentrations of chlorine gas or hypochlorite solutions have experienced effects similar to those observed in adults, although some reports have suggested that children may more susceptible to chlorine gas toxicity than adults (Agency for Toxic Substances and Disease Registry 1998; Güloğlu et al. 2002). Children may be at increased risk for exposure to chlorine gas because they have a greater lung surface area:body weight ratio and an increased minute volume:weight ratio. Children may also be more vulnerable than adults because of the smaller diameter of their airways. Prolonged low-level exposures to chlorine gas are not relevant to children since this type of exposure occurs only in occupational settings. There are no studies that have examined whether young animals are more or less susceptible than adults to chlorine gas or hypochlorite toxicity. Additional information on this issue would be useful.

Child health data needs relating to exposure are discussed in Section 6.8.1, Identification of Data Needs: Exposures of Children.

3.12.3 Ongoing Studies

The following ongoing studies pertaining to chlorine were identified in the Federal Research in Progress database (FEDRIP 2009):

Dr. M.D. Gunn and coworkers at Duke University, plan to conduct a series of murine studies to determine the extent to which specific pulmonary inflammatory cell types contribute to chlorine-induced lung injury and the consequent epithelial damage, bronchial reactivity, morbidity, and mortality. He will then examine the ability of the chemokine receptor antagonist to reduce these toxicities. The research is sponsored by the National Institute of Environmental Health Sciences (NIEHS).

Dr. S-E. Jordt and coworkers at the University of Alabama, Birmingham, intend to develop pharmacological approaches for the treatment of asthma patients after exposure to chlorine. Based on preliminary results indicating that the TRPA1, an ion channel of the transient receptor potential (TRP) gene family, is strongly activated by chlorine in primary and heterologous cells, the investigators

hypothesized that post-exposure administration of TRP channel antagonist, in combination with adrenergic agonists and antioxidants, will counteract life-threatening hypersensitivity responses in asthma patients to chlorine and other pulmonary chemical threats. Specifically the investigators intend to (1) compare responses of wild-type and TRP-channel deficient mice to chlorine in the background of the ovalbumin asthma model, (2) establish pharmacological measures to counteract chlorine hypersensitivity responses and chlorine-induced tissue damage in asthma, and (3) analyze the mechanism of TRPA1 activation and sensitization by chlorine, and its reversal. The research is sponsored by the NIEHS.

Dr. S. Matalon and associates at the University of Alabama, Birmingham, plan to test the hypothesis that the toxic effects of C12 will be heightened in animals infected with respiratory syncytial virus (RSV) or challenged with ova albumin. For that purpose, they will perform a number of biochemical, biophysical, physiological, and morphometric measurements in RSV infected and ova albumin challenged mice as well as normal rats prior to and following chlorine exposure to document the onset and progression of injury to lung epithelia and pulmonary and systemic vasculature. They will then treat the mice with antioxidants, TRP antagonists, and nitrite administered at various intervals post chlorine exposure either intratracheally or via aerosolization, or intraperitoneally (antioxidants and nitrite) and quantify recovery by specific functional measurements. The research is sponsored by the NIEHS.

Dr. S. Matalon and coworkers at the University of Alabama, Birmingham, also plan to determine whether systemic administration of reactive species scavengers, as well as agents that augment surfactant levels, ion transport and paracellular resistance, and a recently described peptide based on the lectin region of TNFa (tip peptide), shortly after exposure to chlorine will decrease lung injury, morbidity and mortality. This hypothesis will be tested by exposing either confluent monolayers of rat alveolar type II (ATII) epithelial cells or rats to chlorine (50–200 ppm for 30 minutes) and measuring a number physiological and biochemical indices of lung function 0.5, 6, 12, and 24 hours post exposure. The measurements will be repeated following intravenous injections of NAC, ascorbate, and deferoxamine as well as albuterol and the tip peptide. They will also assess the efficacy of intratracheally instilled ascorbate, NAC, deferoxamine, Infasurf (a surfactant replacement mixture), albuterol and the tip peptide, as well as aerosolized albuterol, in prolonging survival of rats with respiratory failure following chlorine exposure. The research is sponsored by the NIEHS.

Dr. P.R. Patel and coworkers at the University of Alabama, Birmingham, hypothesize that chlorine gas exposure induces both pulmonary and systemic vascular injury, which will be inhibited by nitrite-dependent NO formation. The investigators propose to test this via the following specific aims: (1) test

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the hypothesis that inactivation of eNOS leads to compromised pulmonary and systemic vascular function, (2) establish protective effects of nitrite administration to rats following chlorine gas exposure on cardiopulmonary function, and (3) determine the mechanism by which nitrite therapy protects against C12 gas induced cardiopulmonary toxicity.

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4. CHEMICAL AND PHYSICAL INFORMATION

4.1 CHEMICAL IDENTITY

Information regarding the chemical identity of chlorine is located in Table 4-1. This information includes synonyms, chemical formula and structure, and identification numbers. For the purpose of disambiguation, terms that are commonly used in reference to chlorinated water are defined in Table 4-2.

4.2 PHYSICAL AND CHEMICAL PROPERTIES

Information regarding the physical and chemical properties of chlorine is located in Table 4-3.

Chlorine (Cl₂) is a heavier-than-air, greenish-yellow gas with a pungent, irritating odor (HSDB 2009). The odor threshold for chlorine in air is generally between 0.2 and 0.4 ppm (Amoore and Hautala 1983; The Chlorine Institute 1998). Perceivable sensory irritation occurs at 1.0 ppm in air (EPA 1999). Chlorine is a nonflammable gas; however, it is a very strong oxidizing agent, reacting explosively or forming explosive compounds or mixtures with many common chemicals (O'Neil et al. 2001). Chlorine reacts directly with nearly all of the elements to form chlorides (Lide 2005; O'Neil et al. 2001). Chlorine is stored and transported as a liquid in pressurized containers (EPA 1999). Chlorine is transported as either a liquid or a gas through pipelines within chemical plants or over distances of several kilometers (Schmittinger et al. 2006).

Chlorine hydrolyzes rapidly and almost completely in water to form hydrochloric acid, hypochlorous acid, and hypochlorite as follows:

$$Cl_2 + H_2O$$
 \longrightarrow $HOCl + H^+ + Cl^ HOCl$ \longrightarrow $H^+ + OCl^-$

The equilibrium constants for these reactions are represented by:

$$K_{1} = \frac{[HOCl][H^{+}][Cl^{-}]}{[Cl_{2}]}$$
 (1)

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Table 4-1. Chemical Identity of Chlorine^a

Characteristic	Information		
Chemical name	Chlorine		
Synonyms/trade names	Chlorine gas ^b , Bertholite, molecular chlorine, chlorine mol, dichlorine		
Chemical formula	Cl_2		
Chemical structure	CI—CI		
Identification numbers:			
CAS registry	7782-50-5		
NIOSH RTECS	FO2100000°		
EPA hazardous waste	No data		
DOT/UN/NA/IMCO shipping	g UN1017; IMO 2.0		
HSDB	206		
EINECS	231-959-5°		
NCI	No data		

^aAll information obtained from HSDB 2009 except where noted.

CAS = Chemical Abstracts Service; DOT/UN/NA/IMO = Department of Transportation/United Nations/North America/International Maritime Dangerous Goods Code; EINECS = European Inventory of Existing Commercial Chemical Substances; EPA = Environmental Protection Agency; HSDB = Hazardous Substances Data Bank; NCI = National Cancer Institute; NIOSH = National Institute for Occupational Safety and Health; RTECS = Registry of Toxic Effects of Chemical Substances

^bEPA 1999

[°]IPCS 1996

Table 4-2. Commonly Used Terms Related to Chlorinated Water

Term	Meaning		
Chlorinated water	The solution that results when molecular chlorine or a hypochlorite salt is added to water for the purpose of water disinfection. Molecular chlorine reacts rapidly with water under environmental conditions to form hypochlorous acid, hypochlorite, and hydrochloric acid; therefore, chlorinated water does not contain molecular chlorine under normal conditions. Some other substances that are commonly formed in chlorinated water include oxidized inorganics, chloramines, and trihalomethanes.		
Free chlorine	The combination of the equilibrium species molecular chlorine, hypochlorous acid, and the hypochlorite ion in chlorinated water. Since molecular chlorine is usually not present in water samples, this term usually refers to the amount of hypochlorous acid and hypochlorite in water.		
Combined chlorine	The amount of chloramines (chlorine combined with nitrogen) present in chlorinated water.		
Total chlorine	The amount of free chlorine (hypochlorous acid and hypochlorite) plus combined chlorine (chloramines) present in chlorinated water.		
Available chlorine	A measure of the oxidizing strength of a solution. It is equal to the amount of molecular chlorine that when added to water would produce a solution with equivalent oxidizing power. It is commonly reported as weight percent.		
Residual chlorine	The amount of free chlorine remaining in a chlorinated water sample that has been collected at a point of use. This indicates whether the water has retained its disinfection properties.		
Aqueous chlorine	A term that usually has a meaning similar to that of free chlorine. In this sense, "aqueous chlorine" should not be misunderstood as the amount of molecular chlorine in water (aqueous molecular chlorine). The term "aqueous chlorine" is commonly used in reference to a prepared aqueous solution of hypochlorite and hypochlorous acid.		

Sources: APHA 1998a; Edstrom Industries 2003; Fukayama et al. 1986; IARC 1991; The Chlorine Institute 2006; Westerhoff et al. 2004; WHO 2007.

Table 4-3. Physical and Chemical Properties of Chlorine^a

Property	Information		
Molecular weight	70.905		
Color	Greenish-yellow		
Physical state	Gas		
Melting point	-101.00 °C		
Boiling point	-34.04 °C		
Density in air	2.482 (air=1) ^b		
Density, as liquid			
20 °C, 6.864 atm	1.4085 g/mL°		
-35 °C, 0.9949 atm	1.5648 g/mL°		
Odor	Pungent, irritating		
Odor threshold:			
Water	Not applicable ^d		
Air	0.2–0.4 ppm ^{c,e,f}		
Solubility:			
Water	14.6 g/L at 0 °C; 7.3 g/L at 20 °C ^{c,g}		
Other solvents	Glacial acetic acid, dimethylformamide, nitrobenzene, phosphoryl chloride, carbon tetrachloride, tetrachloroethane, pentachloroethane, hexachlorobutadiene, and chlorobenzene ^h		
Partition coefficients:			
Log K _{ow}	Not applicable		
Log K _{oc}	Not applicable		
Vapor pressure at 25 °C	5,830 mm Hg		
Henry's law constant	1.17x10 ⁻² atm-m ³ /mol ⁱ		
Autoignition temperature	Not applicable		
Flashpoint	Not applicable		
Reactivity	Strong oxidizer; reacts explosively with many materials		
Conversion factors	1 ppm=2.9 mg/m ³ ; 1 mg/m ³ =0.344 ppm ^c		

 $^{^{\}rm a}\text{All}$ information obtained from HSDB 2009, except where noted. $^{\rm b}\text{O'Neil}$ et al. 2001

^dAmoore and Hautala (1983) reported an odor threshold of 0.002 ppm for chlorine in water; however, these authors state that this solution lacks enough persistence for this value to be used for reference purposes.

^eAmoore and Hautala 1983

^fCI 1998

^gEPA 1994b

^hSchmittinger et al. 1996

Staudinger and Roberts 1996

4. CHEMICAL AND PHYSICAL INFORMATION

$$K_2 = \frac{[H^+][OCl^-]}{[HOCl]} \tag{2}$$

The relative percentage of Cl₂, HOCl, and OCl⁻ at some fixed concentration of Cl⁻ can be expressed as:

$$\%Cl_2 = \frac{[Cl_2]}{[Cl_2] + [HOCl] + [OCl^-]}$$
(3)

$$\%HOCl = \frac{[HOCl]}{[Cl_2] + [HOCl] + [OCl^-]}$$
(4)

$$\%OCl^{-} = \frac{[OCl^{-}]}{[Cl_{2}] + [HOCl] + [OCl^{-}]}$$
 (5)

Using the expressions for the equilibrium constants in Equations 1 and 2 and the relationship that pH is equivalent to the negative logarithm of the hydronium ion concentration, Equations 3–5 can be re-written as:

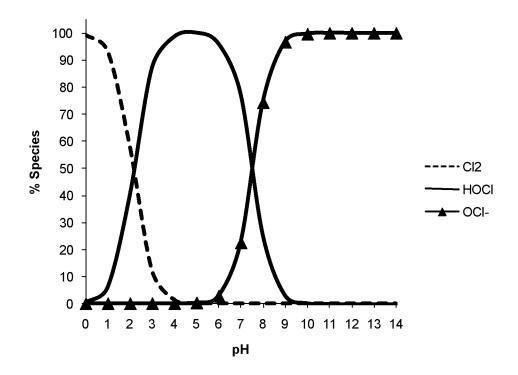
$$\%Cl_2 = \frac{1}{1 + \frac{K_1}{[CI^-]} 10^{pH} + \frac{K_1 K_2}{[CI^-]} 10^{2pH}}$$
(6)

$$\%HOCl = \frac{1}{1 + \frac{[Cl^{-}]}{K_{1}10^{pH}} + K_{2}10^{pH}}$$
 (7)

$$\%OCI^{-} = \frac{1}{1 + \frac{[CI^{-}]}{K_{1}K_{2}10^{2\,pH}} + \frac{1}{K_{2}10^{\,pH}}}$$
(8)

Figure 4-1 illustrates the speciation as a function of pH using values for $K_1 = 3.9 \times 10^{-4} \, \text{M}^2$ (Cotton et al. 1999; Farr et al. 2003) and $K_2 = 2.9 \times 10^{-8} \, \text{M}$ (Farr et al. 2003) at 25 °C. This figure shows the

Figure 4-1. Speciation of Cl₂, HOCl, and OCl⁻ as a Function of pH



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pH-dependant relationship between molecular chlorine, hypochlorous acid, and hypochlorite in aqueous solution. Sodium hypochlorite bleach solutions typically have a pH of 11–13 (The Chlorine Institute 2006). As illustrated in Figure 4-1, the addition of acid to a hypochlorite solution (e.g., mixing of sodium hypochlorite bleach with acid drain cleaner) can drive the pH low enough to result in the release of dangerous amounts of molecular chlorine gas.

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5. PRODUCTION, IMPORT/EXPORT, USE, AND DISPOSAL

5.1 PRODUCTION

The element chlorine was discovered in 1774, and the first patent for its use as a bleaching agent came as early as 1799 (Deutsch 1947). However, it wasn't until the late 1800s that adequate electrolytic equipment became available to produce chlorine on an industrial scale. Chlorine production increased steadily from 5,400 metric tons in 1900 to 63,500 metric tons in 1920 (Deutsch et al. 1963). U.S. chlorine production then underwent an extremely dramatic increase over the next 50 years. Production volumes in 1930, 1940, 1950, 1960, and 1970 were 181,000, 608,000, 1,814,000, 4,172,000, and 8,800,000 metric tons, respectively (Curlin et al. 1991; Deutsch et al. 1963; Robertson 1978). Reasons for this increase were the demand for use of chlorine as a bleaching agent, the demand for its use in the manufacture of other important industrial chemicals, and the further development of electrolytic cell technology, which improved plant production capacities by almost 200% (Bommaraju et al. 2004; Deutsch et al. 1963). Growth during this period was supported by the widespread construction of chlorine-producing plants by alkali producers who were interested in manufacturing chlorine and caustic soda (sodium hydroxide) as co-products, an effort that gave birth to the chlor-alkali industry (Bommaraju et al. 2004; Deutsch et al. 1963; Schmittinger et al. 2006). In 1915, there were only 15 chlorine-producing factories in the United States; by 1960, there were 240 (Deutsch et al. 1963).

During the 1970s and 1980s, U.S. chlorine production fluctuated between 11,200,000 metric tons in 1979 and 8,300,000 metric tons in 1982 (Curlin et al. 1991). Chlorine production rose steadily through the 1990s, reaching 12,100,000 metric tons in 1999 (The Chlorine Institute 2008). With the exception of a spike to 12,300,000 metric tons in 2004, production has steadily declined over the past decade (The Chlorine Institute 2008). The 2008 U.S. production volume for chlorine was reported to be 10,600,000 metric tons by the U.S. Census Bureau (2009). Environmental pressures have strained the chlorine market since the 1970s. Regulations have contributed to such changes as moving away from the use of mercury and asbestos in chlorine production, ending the use of chlorine in pulp and paper bleaching, and curtailing the production of certain chlorinated end products (Bommaraju et al. 2004; CMR 1977, 1980, 1989, 1992, 1995, 2000, 2003, 2006; Robertson 1978; Schmittinger et al. 2006). Negative effects on the market have been balanced by the development of alternative chlorine production methods and increases in demand for other chlorine end products, especially polyvinyl chloride. Although there are still operational facilities that use the older mercury and asbestos production methods, the newer chlorine production facilities are based on the more efficient membrane technology (Bommaraju et al. 2004; The Chlorine Institute 2008). The companies that produced chlorine in the

United States, their production sites, and their annual capacities for 2008 (the most recent year for which figures are available) are shown in Table 5-1 (SRI 2008).

Table 5-2 summarizes the number of facilities in each state that manufactured or processed chlorine (Cl₂) in 2006, the ranges of maximum amounts on site, if reported, and the activities and uses as reported in the Toxics Release Inventory (TRI) (TRI06 2008). The data listed in this table should be used with caution since only certain types of facilities are required to report. This is not an exhaustive list.

5.2 IMPORT/EXPORT

Annual U.S. chlorine import and export quantities reported for different years are listed in Table 5-3. The available data indicate that annual imports of chlorine into the United States have increased steadily over the past 20 years, rising from 251,000 metric tons in 1984 to 429,000 metric tons in 2007 (CMR 1989, 1992, 2000; HSDB 2009; ITA 2007; U.S. Census Bureau 2008). The decline in U.S. chlorine exports during the early 2000s (24,200 metric tons in 2000 to 10,400 metric tons in 2004) has been attributed to increasing energy costs, which have rendered the chlorine produced in the United States uncompetitive, especially in the Asian market (CMR 2006; ITA 2007). However, U.S. chlorine exports in 2006 were bolstered by a 9-fold increase in shipments to Mexico, accounting for approximately 82% (32,201 metric tons) of the 39,481 metric tons of chlorine exported during that year (ITA 2007). U.S. exports of chlorine fell to 25,740 in 2007 (US Census Bureau 2008).

5.3 USE

The major uses of chlorine during 2006 were the manufacturing of vinyl chloride to make polyvinyl chloride (PVC) plastics (36%), the manufacturing of other organic compounds (41%), the manufacturing of inorganic chemicals (15%), water treatment (4%), and pulp and paper bleaching (1%) (CMR 2006). Other miscellaneous uses accounted for 3% of total chlorine use during 2006. Chlorine is used in the production of a large number of commercial products (Bommaraju et al. 2004; Schmittinger et al. 2006). Some of the important end products for which chlorine plays a role in the production stream include refrigerants, aerosols, silicones, silicone rubber, plastics, solvents, polyethers, varnishes, foams, chlorinated rubber, polyurethane, detergents, dyes, insecticides, pesticides, disinfectants, bleaches, and white pigment enamel (Schmittinger et al. 2006). Chlorine has been used in the food industry as a bleaching agent for flour (Fukayama et al. 1986). Chlorine was used as a war gas during World War I (Compton 1987). Chlorine is also used to manufacture phosgene (O'Neil et al. 2001).

Table 5-1. Companies that Produce Chlorine in the United States and Annual Capacities for 2006

Company	Location	Capacity (thousands of short tons) ^a	Capacity (metric tons) ^a
ASHTA Chemicals, Inc.	Ashtabula, Ohio	44	40,000
Bayer MaterialScience LLC	Baytown, Texas	400	363,000
The Dow Chemical Company	Freeport, Texas	3,240	2,939,000
	Plaquemine, Louisiana	1,070	971,000
E.I. du Pont de Nemours and Company; DuPont Coatings and Color Technologies; DuPont Performance Coatings	Niagara Falls, New York	85	77,000
Equa-chlor LLC	Longview, Washington	87	79,000
ERCO Worldwide, Inc.	Port Edwards, Wisconsin	106	96,000
Formosa Plastics Corporation	Point Comfort, Texas	811	736,000
Georgia Gulf Corporation	Plaquemine, Louisiana	450	408,000
Georgia-Pacific Chemicals LLC	Green Bay, Wisconsin	9	8,000
	Muskogee, Oklahoma	6	5,000
	Rincon, Georgia	7	6,000
Kuehne Chemical Corporation	Delaware City, Delaware	16	15,000
Occidental Chemical Corporation;	Convent, Louisiana	389	353,000
Chloro-Vinyls Group	Corpus Christi, Texas	604	548,000
	Geismar, Louisiana	268	243,000
	Hahnville, Louisiana	750	680,000
	Mobile, Alabama	50	45,000
	Muscle Shoals, Alabama	150	136,000
	New Castle, Delaware	90	82,000
	Niagara Falls, New York	335	304,000
	Wichita, Kansas	263	239,000
Olin Corporation; Olin Chlor Alkali	Augusta, Georgia	120	109,000
Products Division	Charleston, Tennessee	245	222,000
	Henderson, Nevada	152	138,000
	McIntosh, Alabama	735	667,000
	Niagara Falls, New York	286	259,000
	St. Gabriel, Louisiana	180	163,000
OxyVinyls, L.P.	La Porte, Texas	580	526,000
PPG Industries, Inc.;	Lake Charles, Louisiana	1,375	1,247,000
Chemical Group	Natrium, West Virginia	510	463,000

Table 5-1. Companies that Produce Chlorine in the United States and Annual Capacities for 2006

Company	Location	Capacity (thousands of short tons) ^a	Capacity (metric tons) ^a
SABIC Innovative Plastics	Burkville, Alabama	90	82,000
	Mount Vernon, Indiana	96	87,000
Titanium Metals Corporation	Henderson, Nevada	5	5,000
Trans Carolina Products LLC	Hamlet, North Carolina	Not available	Not available
U.S. Magnesium, LLC	Rowley, Utah	47	43,000
Westlake Vinyls, Inc.	Calvert City, Kentucky	205	186,000
Total		13,856	12,570,000

^aMuch of the capacity is consumed captively.

Source: SRI 2008; The Chlorine Institute 2008

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Table 5-2. Facilities that Produce, Process, or Use Chlorine

State ^a facilities in pounds ^b Activities and uses ^c AK 13 0 9,999,999 1, 2, 3, 5, 6, 7, 10, 11, 12, AL 134 0 499,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, AR 67 0 49,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, AS 2 1,000 9,999 11, 12 AZ 44 100 9,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, CA 143 0 499,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	
AL 134 0 499,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, AR 67 0 49,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, AS 2 1,000 9,999 11, 12 AZ 44 100 9,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	
AR 67 0 49,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, AS 2 1,000 9,999 11, 12 AZ 44 100 9,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	
AS 2 1,000 9,999 11, 12 AZ 44 100 9,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	
AZ 44 100 9,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13
CA 143 0 499 999 999 1 2 3 4 5 6 7 8 9 10	11, 12, 14
1, 2, 0, 1, 0, 0, 10,	11, 12, 13
CO 19 0 49,999,999 1, 2, 3, 4, 5, 6, 9, 10, 11, 1	2
CT 28 0 999,999 1, 2, 3, 4, 5, 6, 7, 10, 11, 1	2
DC 1 100,000 999,999 12	
DE 31 100 499,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12
FL 109 0 499,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13
GA 105 0 9,999,999 1, 2, 3, 4, 5, 6, 7, 9, 10, 11	, 12, 13
GU 2 10,000 999,999 9, 12	
HI 18 0 9,999,999 1, 2, 3, 4, 6, 8, 10, 11, 12	
IA 46 0 9,999,999 1, 2, 3, 5, 6, 7, 8, 10, 11, 1	2
ID 36 0 9,999,999 1, 2, 3, 5, 6, 7, 10, 11, 12,	13, 14
IL 72 0 49,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13
IN 67 0 9,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13
KS 37 0 9,999,999 1, 2, 3, 4, 5, 6, 7, 9, 10, 11	, 12
KY 81 0 999,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13
LA 180 0 499,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13, 14
MA 25 0 999,999 1, 2, 3, 5, 6, 7, 9, 10, 11, 1	2
MD 41 100 49,999,999 1, 2, 3, 5, 6, 7, 8, 9, 10, 11	, 12, 13
ME 63 0 49,999,999 1, 2, 3, 4, 5, 6, 7, 10, 11, 1	2, 13
MI 104 0 499,999,999 1, 2, 3, 5, 6, 7, 8, 9, 10, 11	, 12, 13
MN 67 0 9,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13
MO 55 0 9,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13, 14
MS 76 0 49,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13, 14
MT 11 1,000 9,999,999 1, 4, 5, 7, 10, 11, 12, 13	
NC 118 0 499,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13
ND 9 100 999,999 1, 2, 3, 5, 6, 10, 11, 12	
NE 18 100 49,999,999 1, 2, 3, 4, 6, 9, 10, 11, 12	
NH 12 0 9,999,999 1, 2, 3, 5, 6, 9, 12, 13	
NJ 66 0 49,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13, 14
NM 19 0 9,999,999 1, 2, 3, 4, 5, 6, 9, 11, 12	
NV 31 0 49,999,999 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,	11, 12, 13

Table 5-2. Facilities that Produce, Process, or Use Chlorine

	Number of	Minimum amount on site	Maximum amount on site	
State	facilities	in pounds ^b	in pounds ^b	Activities and uses ^c
NY	106	0	49,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
OH	120	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
OK	48	0	49,999,999	1, 2, 3, 5, 6, 7, 8, 9, 10, 11, 12, 13
OR	55	0	9,999,999	1, 2, 3, 4, 5, 6, 8, 10, 11, 12, 13
PA	102	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
PR	27	0	49,999,999	2, 3, 4, 6, 7, 10, 11, 12
RI	15	100	9,999,999	1, 2, 3, 4, 6, 9, 10, 11, 12
SC	94	0	99,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
SD	9	100	999,999	7, 10, 11, 12
TN	99	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
TX	197	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
UT	40	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
VA	66	0	49,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
VT	2	1,000	9,999	11, 12
WA	102	0	499,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13
WI	107	0	9,999,999	1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14
WV	52	0	499,999,999	1, 2, 3, 4, 5, 6, 9, 10, 11, 12, 13
WY	13	1,000	999,999	1, 2, 3, 5, 10, 11, 12, 13

^aPost office state abbreviations used

- 1. Produce
- 2. Import
- 3. Onsite use/processing
- 4. Sale/Distribution
- 5. Byproduct

- 6. Impurity
- 7. Reactant
- 8. Formulation Component
- 9. Article Component
- 10. Repackaging

- 11. Chemical Processing Aid
- 12. Manufacturing Aid
- 13. Ancillary/Other Uses
- 14. Process Impurity

Source: TRI06 2008 (Data are from 2006)

^bAmounts on site reported by facilities in each state

^cActivities/Uses:

Table 5-3. U.S. Chlorine Imports and Exports by Year in Metric Tons

Year	Imports	Exports	Reference
1975	67,000	15,000	Robertson 1978
1984	251,000	39,500	HSDB 2007
1986	298,739	Not available	HSDB 2007
1987	Not available	3,787	HSDB 2007
1988	280,840	58,073	CMR 1989
1991	272,160	Not available	CMR 1992
1998	373,766	22,680	CMR 2000
1999	325,685	21,773	CMR 2000
2000	358,015	24,231	ITA 2007
2001	358,060	20,964	ITA 2007
2002	409,695	18,566	ITA 2007
2003	412,117	15,361	ITA 2007
2004	470,884	10,448	ITA 2007
2005	476,103	12,306	ITA 2007
2006	454,414	39,481	ITA 2007
2007	429,440	25,740	U.S. Census Bureau 2008

5.4 DISPOSAL

Chlorine is disposed of via a salt-forming reaction followed by neutralization (HSDB 2009). Chlorine gas is first introduced into a large volume solution of a reducing agent such as sodium thiosulfate, bisulfite, or ferrous salts or aqueous sodium hydroxide (HSDB 2009). The resulting salt solution is then neutralized and routed to a sewage treatment plant (HSDB 2009).

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6. POTENTIAL FOR HUMAN EXPOSURE

6.1 OVERVIEW

Although it has multiple uses and is released to the environment, chlorine is too reactive to be identified in any of the 1,704 hazardous waste sites that have been proposed for inclusion on the EPA National Priorities List (NPL) (HazDat 2007).

Chlorine may be released into the environment during accidents such as a chlorine gas leak from an industrial facility or a chlorine tank spill or rupture. Low concentrations of chlorine gas (<600 ppt) appear to be produced by the photolysis of seawater aerosol. Chlorine gas injected into the water during water chlorination quickly dissolves and forms chloride and hypochlorous acid within seconds. Liquid chlorine in a ruptured tank or spilled onto the ground or into water during an accident is expected to volatilize rapidly, forming a greenish-yellow cloud of chlorine gas. This gas cloud can be carried several miles away from the source of release while maintaining dangerous levels of chlorine.

Since chlorine gas is so reactive, it is not expected to remain in the environment very long after it is released. Chlorine immediately reacts with both organic and inorganic materials that it comes into contact with. As mentioned above, it is converted within seconds once it dissolves in water. Chlorine undergoes direct photolysis in the air and its half-life in the troposphere is on the order of several minutes. Chlorine levels in the ambient atmosphere, water, soil, or sediment are not available.

Exposure of the general population to chlorine gas is not expected except in the case of an accidental spill or industrial mishap. There have been several documented incidents in which large amounts of chlorine gas have been released, thereby exposing workers and the members of the general population following the derailment of trains carrying liquefied chlorine gas (Agency for Toxic Substances and Disease Registry 1998; NTSB 1998, 2005, 2006). Other accidental exposures may occur when individuals mix acidic household chemicals with bleach or pool sanitizing agents (see Section 6.3.2.2). Occupational exposure to low levels of chlorine gas may occur for individuals who work at facilities where it is produced or used. These individuals may be exposed to higher levels if an accidental release occurs within the facility. Children are expected to be affected by the same routes of exposure as adults, except for occupational exposure.

The general public is not exposed to molecular chlorine in drinking water as a result of water sanitation practices, even though chlorine gas may be used in these processes. Free chlorine in drinking water is

defined as the sum of dissolved chlorine gas, hypochlorous acid, and hypochlorite anion. As discussed in Chapter 4 and in Section 6.3.2.2, the level of dissolved chlorine in water is very low, except under acidic conditions. As a consequence, the term free chlorine in public water systems or swimming pools usually refers to the concentration of hypochlorous acid and hypochlorite anion (APHA 1998a, 1998b).

6.2 RELEASES TO THE ENVIRONMENT

The Toxics Release Inventory (TRI) data should be used with caution because only certain types of facilities are required to report (EPA 2005). This is not an exhaustive list. Manufacturing and processing facilities are required to report information to the TRI only if they employ 10 or more full-time employees; if their facility is included in Standard Industrial Classification (SIC) Codes 10 (except 1011, 1081, and 1094), 12 (except 1241), 20–39, 4911 (limited to facilities that combust coal and/or oil for the purpose of generating electricity for distribution in commerce), 4931 (limited to facilities that combust coal and/or oil for the purpose of generating electricity for distribution in commerce), 4939 (limited to facilities that combust coal and/or oil for the purpose of generating electricity for distribution in commerce), 4953 (limited to facilities regulated under RCRA Subtitle C, 42 U.S.C. section 6921 et seq.), 5169, 5171, and 7389 (limited to facilities primarily engaged in solvents recovery services on a contract or fee basis); and if their facility produces, imports, or processes ≥25,000 pounds of any TRI chemical or otherwise uses >10,000 pounds of a TRI chemical in a calendar year (EPA 2005).

6.2.1 Air

Estimated releases of 5,050,000 pounds (2,290 metric tons) of chlorine to the atmosphere from 915 domestic manufacturing and processing facilities in 2006, accounted for about 88% of the estimated total environmental releases from facilities required to report to the TRI (TRI06 2008). These releases are summarized in Table 6-1.

Chlorine may be released into the air in fugitive emissions from industrial facilities where it is produced or used. It may also be released into the air as a result of a spill or tank rupture (Henry et al. 2005; Horton et al. 2002). Between January 1993 and December 2000, 952 chlorine-related events (865 involved chlorine only) were reported to the ATSDR's Hazardous Substances Emergency Events Surveillance system (Horton et al. 2002). Of the 865 chlorine-only events, 592 (68.4%) involved air emissions. Of the 564 events for which release amount information was available in pounds, 511 events involved releases of ≤250 pounds. Examples of some recent large scale accidents involving the release of chlorine gas follow.

Table 6-1. Releases to the Environment from Facilities that Produce, Process, or Use Chlorine^a

				Reported	amounts	release	d in pounds pe	er year ^o	
							Total release		
State ^c	RF ^d	Air ^e	Water ^f	Ul ^g	Land ^h	Other ⁱ	On-site ^j	Off-site ^k	On- and off-site
AK	2	0	No data	0	0	0	0	0	0
AL	39	43,765	1,208	0	0	0	44,973	0	44,973
AR	23	54,016	637	0	0	0	54,653	0	54,653
ΑZ	10	668	250	0	517	0	1,435	0	1,435
CA	28	3,192	0	0	8,100	0	11,292	0	11,292
CO	9	45,635	0	0	0	0	45,635	0	45,635
CT	4	972	816	0	0	0	1,788	0	1,788
DC	2	1,008	0	0	0	0	1,008	0	1,008
DE	5	4,662	No data	0	0	0	4,662	0	4,662
FL	34	5,749	22	0	27,734	0	18,055	15,450	33,505
GA	34	5,332	2,702	0	0	0	8,034	0	8,034
HI	1	5	No data	0	0	0	5	0	5
IA	16	12,873	16,825	0	1,620	0	31,318	0	31,318
ID	6	8,965	250	0	5	0	9,215	5	9,220
IL	31	186,356	121,209	0	0	0	307,565	0	307,565
IN	34	77,356	1,501	0	278	0	78,862	273	79,135
KS	10	2,681	0	146,240	0	0	148,921	0	148,921
KY	21	78,635	490	0	0	0	79,125	0	79,125
LA	63	179,943	13,056	0	269	72	192,999	341	193,340
MA	5	385	No data	0	0	0	385	0	385
MD	9	781	0	0	0	0	781	0	781
ME	5	1,510	0	0	0	0	1,510	0	1,510
MI	26	30,950	679	0	0	0	31,628	0	31,628
MN	16	12,165	0	0	0	0	12,165	0	12,165
MO	20	15,545	1,157	0	0	0	16,702	0	16,702
MS	18	61,839	676	0	0	0	62,515	0	62,515
NC	23	121,870	12,308	0	8	0	134,186	0	134,186
ND	4	715	0	0	0	0	715	0	715
NE	9	2,126	220	0	0	0	2,346	0	2,346
NH	2	11	0	0	0	0	11	0	11
NJ	12	8,251	6,431	0	0	0	14,682	0	14,682
NM	6	3,133	No data	3,166	0	109	6,299	109	6,408
NV	9	5,246	No data	0	252,035	0	257,281	0	257,281
NY	23	61,884	1,209	0	0	694	63,093	694	63,787
ОН	42	51,278	1,206	0	0	7	52,484	7	52,491

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Table 6-1. Releases to the Environment from Facilities that Produce, Process, or Use Chlorine^a

				Reported	l amounts	released	d in pounds pe	er year ^b	
								Total rel	ease
$State^{\mathtt{c}}$	RF^{d}	Air ^e	Water ^f	UI ^g	Land ^h	Other ⁱ	On-site ^j	Off-site ^k	On- and off-site
OK	12	1,448	250	0	0	0	1,698	0	1,698
OR	10	10,744	301	0	0	0	11,045	0	11,045
PA	31	68,834	2,919	0	0	0	71,753	0	71,753
PR	7	3,644	No data	0	0	0	3,644	0	3,644
RI	3	1,796	No data	0	0	0	1,796	0	1,796
SC	18	12,836	23	0	0	0	12,859	0	12,859
SD	2	0	186	0	144	0	330	0	330
TN	26	138,054	0	0	0	0	138,054	0	138,054
TX	116	242,531	31,610	13,908	160	0	288,209	0	288,209
UT	6	3,330,619	0	0	0	0	3,330,619	0	3,330,619
VA	25	31,766	11,173	0	0	0	42,939	0	42,939
VT	1	0	No data	0	0	0	0	0	0
WA	18	2,268	17,006	0	5	0	19,279	0	19,279
WI	21	40,691	3,540	0	14	0	44,245	0	44,245
WV	14	17,997	4,000	0	0	0	21,997	0	21,997
WY	4	57,019	0	0	114	0	57,133	0	57,133
Total	915	5,049,751	253,859	163,314	291,003	882	5,741,931	16,879	5,758,810

^aThe TRI data should be used with caution since only certain types of facilities are required to report. This is not an exhaustive list. Data are rounded to nearest whole number.

RF = reporting facilities; UI = underground injection

Source: TRI06 2008 (Data are from 2006)

^bData in TRI are maximum amounts released by each facility.

^cPost office state abbreviations are used.

^dNumber of reporting facilities.

^eThe sum of fugitive and point source releases are included in releases to air by a given facility.

fSurface water discharges, waste water treatment-(metals only), and publicly owned treatment works (POTWs) (metal and metal compounds).

^gClass I wells, Class II-V wells, and underground injection.

^hResource Conservation and Recovery Act (RCRA) subtitle C landfills; other on-site landfills, land treatment, surface impoundments, other land disposal, other landfills.

Storage only, solidification/stabilization (metals only), other off-site management, transfers to waste broker for disposal, unknown

^jThe sum of all releases of the chemical to air, land, water, and underground injection wells.

^kTotal amount of chemical transferred off-site, including to POTWs.

A freight train derailment, which took place on April 11, 1996, 2 miles west of Alberton, Montana resulted in a spill releasing 64.8 tons of chlorine into the environment (Agency for Toxic Substances and Disease Registry 1998; NTSB 1998). On January 6, 2005, a train carrying sodium hydroxide, cresol, and liquefied chlorine collided with another train in Graniteville, South Carolina releasing approximately 120,000 pounds of chlorine gas to the surrounding atmosphere (NTSB 2005). On June 28, 2004, a westbound Union Pacific Railroad freight train collided with an eastbound freight train in Macdona, Texas (NTSB 2006). Consequently, a tank car carrying 180,000 pounds of liquefied chlorine was punctured, releasing a cloud of chlorine gas to the immediate area (NTSB 2006). An accident involving a broken cargo transfer pipe at the ATOFINA Chemicals plant in Riverview, Michigan, resulted in the release of flammable methyl mercaptan gas (NTSB 2002). A subsequent fire resulted in damage to an adjacent tank car loaded with chlorine. It was estimated that approximately 26,500 of the 178,560 pounds of chlorine in the tank car were released in this incident, which occurred in July 2001 (NTSB 2002).

Chlorine appears to be generated in very low concentrations by the photolysis of seawater aerosols above seawater (California Environmental Protection Agency 2002; Chang et al. 2004; Knipping and Dabdub 2003).

Based on its instability and reactivity, it is not expected to be identified in air at any of the 1,704 hazardous waste sites that have been proposed for inclusion on the EPA National Priorities List (NPL) (HazDat 2007).

6.2.2 Water

Estimated releases of 254,000 pounds (115 metric tons) of chlorine to surface water from 915 domestic manufacturing and processing facilities in 2006, accounted for about 4% of the estimated total environmental releases from facilities required to report to the TRI (TRI06 2008). These releases are summarized in Table 6-1.

Activities at industrial facilities where chlorine is produced or used may result in its unintentional release to surface water. Chlorine may also be released into water as a result of a spill or tank rupture. Chlorine gas is injected directly into water during certain disinfection processes; however, these processes take place under controlled conditions within treatment facilities to prevent the release of chlorine into the environment (Das 2002; Tchobanoglous and Schroeder 1985). Furthermore, the chlorine gas is

immediately converted into hypochlorous acid, chloride ion, and hypochlorite as soon as it enters the water. Chlorine may be formed in waters containing hypochlorite if the pH is lowered to levels below 4 (Farr et al. 2003).

Chlorine is too reactive to be identified in surface water or groundwater at any of the 1,704 hazardous waste sites that have been proposed for inclusion on the EPA National Priorities List (NPL) (HazDat 2007).

6.2.3 Soil

Estimated releases of 291,000 pounds (132 metric tons) of chlorine to soils from 915 domestic manufacturing and processing facilities in 2006, accounted for about 5% of the estimated total environmental releases from facilities required to report to the TRI (TRI06 2008). An additional 163,000 pounds (74 metric tons), constituting about 3% of the total environmental emissions, were released via underground injection (TRI06 2008). These releases are summarized in Table 6-1.

Activities at industrial facilities where chlorine is produced or used may result in its release to soil. Chlorine may also be released onto soil as a result of a spill. Of the 865 chlorine-only events reported to the ATSDR's Hazardous Substances Emergency Events Surveillance system between January 1993 and December 2000, 134 (15.5%) involved spills (Horton et al. 2002).

Chlorine is too reactive to be identified in soil or sediment at any of the 1,704 hazardous waste sites that have been proposed for inclusion on the EPA National Priorities List (NPL) (HazDat 2007).

6.3 ENVIRONMENTAL FATE

6.3.1 Transport and Partitioning

The boiling point of chlorine is -34.04 °C; therefore, chlorine is a gas under environmental conditions (HSDB 2009). Chlorine gas dissolves in water (7.3 g/L at 20 °C) and is immediately converted to hypochlorous acid and chloride ion at environmental pH. However, this conversion is hindered under very acidic conditions (pH<4). Molecular chlorine in water at this low pH is expected to volatilize rapidly based on a Henry's law constant of 1.17x10⁻² atm-m³/mol (Staudinger and Roberts 1996). Chlorine is not expected to bioaccumulate in plants or animals since it reacts with the moist tissues of living systems (Compton 1987; Schreuder and Brewer 2001; Schmittinger et al. 2006). Schreuder and Brewer (2001) observed severe damage to the foliage of two conifer species exposed to a chlorine gas

cloud released during an accidental spill. All buds on exposed trees located within 50 m of the release were killed.

If a large amount of liquid chlorine is released to a body of water, such as during a spill or an underwater release from a ruptured tank, some of the chlorine is expected to escape into the air before it can mix and react with the water. Similarly, when liquid chlorine is spilled onto the ground or when a tank containing liquid chlorine is ruptured, much of the chlorine will volatilize rapidly into the air creating a greenish-yellow cloud of chlorine gas (Agency for Toxic Substances and Disease Registry 1998; DOE 2005b). Since chlorine gas is heavier than air, a chlorine gas cloud will remain low to the ground. Movement and dissipation of the gas cloud is determined by such factors as the release volume, type of release, terrain, topography, temperature, humidity, atmospheric stability, and wind speed and direction (DOE 2005b; U.S. Chemical Safety and Hazard Investigation Board 2003).

Analysis of the chlorine gas cloud released during the 2005 Graniteville, South Carolina train derailment indicates that the dense cloud was initially driven southwest by gravity toward lower elevation, but was later dispersed by the wind toward the north and northeast as the boundary layer of the plume grew and as the gas began to mix with the surrounding air (DOE 2005a, 2005b). Wenck et al. (2007) reported that there were approximately 40 severe outcomes (≥3 nights of hospitalization and 9 deaths) and approximately 120 less severe outcomes (≤2 nights hospitalization) that occurred within a half-mile radius of the incident (Wenck et al. 2007). According to these authors, only a few less severe outcomes occurred between a half-mile and three-quarters of a mile from incident.

Movement of chlorine through soil is not expected to be relevant since chlorine will react and volatilize so quickly when spilled onto the ground (Agency for Toxic Substances and Disease Registry 1998; Schulte 1999).

6.3.2 Transformation and Degradation

6.3.2.1 Air

The primary removal mechanism for chlorine in air is direct photolysis (EPA 1993; Graedel 1978; Graedel et al. 1986). Sunlight at tropospheric wavelengths (<430 nm) breaks apart the chlorine molecule to form two chlorine radicals. These radicals then react with any available organic molecule to form hydrochloric acid. The mean atmospheric lifetime of chlorine has been reported as 440 seconds (the atmospheric half-life can be approximated by multiplying the lifetime by the natural logarithm of 2)

(Graedel 1978). A tropospheric lifetime of <15 minutes was calculated by Tanaka et al. (2003) under smog conditions in Houston, Texas. The rate of direct photolysis for any chemical species is dependent upon the intensity of sunlight, and therefore, factors such as time of year, geographic location, and time of day affect the photolysis rate. Hov (1985) discussed the diurnal variation of the photolysis rate of chlorine in an analysis of the effect that atmospheric chlorine had on the formation of various photochemical oxidants in southern Telemark, Norway. It was generally concluded that the photodissociation rate is rapid under both winter and summer sunlight conditions. Using absorption cross section data for chlorine at 330 nm, a maximum photodissociation rate constant of approximately 1.6×10^{-3} second⁻¹ was calculated for midsummer, midday sunlight conditions. This corresponds to a tropospheric half-life of approximately 7.2 minutes and a mean lifetime of slightly over 10 minutes. Using a photodissociation rate constant of approximately 0.2×10^{-3} second⁻¹ calculated for midday winter sunlight conditions, a half-life of 58 minutes is estimated.

Chlorine is also expected to react with cloud particulates and rain drops that it comes into contact with in the atmosphere, forming hydrochloric and hypochlorous acids (Vetrano 2001). These acids can then be washed out of the atmosphere by precipitation (Vetrano 2001).

6.3.2.2 Water

Water disinfection through chlorination has been in regular use since the early 1900s; consequently, the fate of chlorine in water has been well studied (Das 2002). Chlorine gas released into water first dissolves and then undergoes a disproportionation within seconds at environmental pH to form hydrochloric ($H^+ + Cl^-$) and hypochlorous acid (HOCl) (Cotton et al. 1999; Das 2002; EPA 1999; Farr et al. 2003; Morris 1946; Snoeyink and Jenkins 1980; Tchobanoglous and Schroeder 1985; Wang and Margerum 1994). The equilibrium that exists between hypochlorous acid and the hypochlorite anion is controlled by the pH of the water. Since the p K_a of hypochlorous acid is 7.49 (O'Neil et al. 2001), the formation of the conjugate base (hypochlorite anion) is favored under alkaline conditions and the protonated species (hypochlorous acid) is favored under neutral and mildly acidic conditions. Under strongly acidic conditions ($pH \le 2$), the formation of molecular chlorine is possible (see Chapter 4).

As illustrated in Figure 4-1, the equilibrium between chlorine, hypochlorous acid, and hypochlorite acid is dependant on the pH of the solution (Farr et al. 2003). As the pH is lowered to 4, the equilibrium begins to shift to the left, and small amounts of Cl₂ are present. At pH below 2, chlorine becomes the dominant species. Therefore, molecular chlorine will be formed in chlorinated water (containing hypochlorous

acid) that has been made very acidic. Under these conditions, chlorine is expected to react rapidly with both organic and inorganic mater that it comes into contact with in the water; it is also expected to volatilize rapidly into the air based on a Henry's law constant of $1.17x10^{-2}$ atm-m³/mol (Staudinger and Roberts 1996). Chlorine is toxic to microbial communities; therefore, biodegradation is not considered to be a relevant fate process (Vetrano 2001). The hypochlorous acid formed during the disproportionation of chlorine in natural waters reacts with organic and inorganic materials, ultimately forming chloride, oxidized inorganics, chloramines, trihalomethanes, oxygen, and nitrogen (IARC 1991; Vetrano 2001).

6.3.2.3 Sediment and Soil

If liquid chlorine is spilled onto soil, it will react with both organic and inorganic mater in the soil; however, much of the chlorine is expected to volatilize immediately (Agency for Toxic Substances and Disease Registry 1998; Schulte 1999). Chlorine is expected to dissolve and disproportionate in the water of moist soils to form chloride and hypochlorite (Cotton et al. 1999). Chlorine in a gas cloud is expected to react with soil surfaces that it comes into contact with. Chlorine is toxic to microbial communities; therefore, biodegradation is not considered to be a relevant fate process (Vetrano 2001).

When released into water, chlorine is expected to react with suspended solids and sediments that it comes into contact with.

6.3.2.4 Other Media

Specific information on the transformation of chlorine in other media is not available; however, chlorine is very unstable and is expected to react with most substances that it comes into contact with.

6.4 LEVELS MONITORED OR ESTIMATED IN THE ENVIRONMENT

Reliable evaluation of the potential for human exposure to chlorine depends in part on the reliability of supporting analytical data from environmental samples and biological specimens. Concentrations of chlorine in unpolluted atmospheres and in pristine surface waters are often so low as to be near the limits of current analytical methods. In reviewing data on chlorine levels monitored or estimated in the environment, it should also be noted that the amount of chemical identified analytically is not necessarily equivalent to the amount that is bioavailable. The analytical methods available for monitoring chlorine in a variety of environmental media are detailed in Chapter 7.

6.4.1 Air

Levels of chlorine monitored in the ambient atmosphere have not been located. Scientists have proposed that chlorine is generated during the photolysis of seawater aerosol (California Environmental Protection Agency 2002; Chang et al. 2004; Knipping and Dabdub 2003). According to California Environmental Protection Agency (2002), chlorine was detected in the air at a New York coastal site with a maximum concentration of 150 ppt. Chang et al. (2004) reported a maximum chlorine concentration of 580 ppt in air samples collected from coastal locations in Taiwan.

Little information is available regarding the levels of chlorine measured in air surrounding areas of accidental release. The concentrations of chlorine monitored in the air surrounding a chlorine release from a railroad tank car near Festus, Missouri were >1,000 ppm (U.S. Chemical Safety and Hazard Investigation Board 2003). Chlorine concentrations as high as 1,000 ppm were measured in the air at a train derailment chemical spill near Alberton, Montana that released chlorine gas in April 1996 (Agency for Toxic Substances and Disease Registry 1998).

6.4.2 Water

Levels of chlorine monitored in water are not available. Chlorine is not a predominant species in water at environmental pH since it disproportionates within seconds to form chloride anion and hypochlorous acid; therefore, chlorine is not expected to be detected in the aquatic environment (Das 2002; Morris 1946; Wang and Margerum 1994).

6.4.3 Sediment and Soil

Levels of chlorine monitored in sediment and soil are not available. Chlorine is not expected to be found in soil since it reacts and volatilizes so rapidly.

6.4.4 Other Environmental Media

Levels of chlorine monitored in food, animals, vegetation, and consumer products are not available. Chlorine is not expected to be found in these media because of its very high reactivity.

6.5 GENERAL POPULATION AND OCCUPATIONAL EXPOSURE

Chlorine is not normally detected in ambient air, soil, surface water, groundwater, or drinking water. Therefore, background exposure of the general population to chlorine is not expected. Greater than 95% of the chlorine that is inhaled (over a 1–5 ppm range) reacts in the upper respiratory tract (Nodelman and Ultman 1999a, 1999b; Winder 2001) and eventually joins the chloride pool in the body. Therefore, analysis of human biological materials such as blood, urine, and body tissue for chlorine is not considered relevant. The amount of chlorine that needs to be inhaled to induce a significant increase in extracellular chloride in the body is probably a lethal amount. Current procedures for gauging an individual's exposure to chlorine gas involve analysis of the lungs and respiratory airways for physical and functional damage (Lawson 1981; Winder 2001).

Members of the general population may be exposed to chlorine if they mix an acid with a solution containing sodium hypochlorite (CDC 1991). Examples include mixing toilet bowl cleaners containing hydrochloric, phosphoric, or oxalic acid with bleach (Becker and Forrester, 2008; CDC 1991; Gapany-Gapanavicius et al. 1982; Howard et al. 2007; Mrvos et al. 1993). If enough acid is added to lower the pH of the hypochlorite solution to below 4, chlorine gas will be released (Farr et al. 2003). Individuals may also be exposed to chlorine if swimming pool chemicals are accidentally mixed with acids or too much sodium hypochlorite is added to the water over a short period of time (Agabiti et al. 2001; Becker and Forrester 2008; Bonetto et al. 2006; Ngo et al. 2007; Sexton and Pronchik 1998). On October 22, 1998, hydrochloric acid accidentally spilled onto chlorinated swimming pool water at a recreational facility in Rome, Italy causing the liberation of chlorine gas to the entire center and exposing nearly 300 people to the gas (Agabiti et al. 2001). The exact amount of chlorine released in this accident was not reported. A similar accident occurred on February 17, 2004 in Parma, Italy in which 18 children were exposed to chlorine gas (Bonetto et al. 2006). In this case, the improper addition of an excessive amount of sodium hypochlorite was added to a chlorinated swimming pool to shock the system, resulting in the emission of chlorine gas.

Occupational exposure to low levels of chlorine gas in air may occur for individuals who work at facilities that produce, transport, or use chlorine (Gautrin et al. 1995). These individuals may also be exposed to high chlorine concentrations if an accidental release occurs at the facility (Beach et al. 1969; Chester et al. 1969; Gautrin et al. 1999; Kennedy et al. 1991; NTSB 2002, 2005, 2006; USB 2007).

Of the 865 chlorine-only events reported to the ATSDR's Hazardous Substances Emergency Events Surveillance system between January 1993 and December 2000, 275 events involved victims (1,071 individuals) (Horton et al. 2002). Of these, 759 were occupational exposures and 68 were first responders.

6.6 EXPOSURES OF CHILDREN

This section focuses on exposures from conception to maturity at 18 years in humans. Differences from adults in susceptibility to hazardous substances are discussed in Section 3.7, Children's Susceptibility.

Children are not small adults. A child's exposure may differ from an adult's exposure in many ways. Children drink more fluids, eat more food, breathe more air per kilogram of body weight, and have a larger skin surface in proportion to their body volume. A child's diet often differs from that of adults. The developing human's source of nutrition changes with age: from placental nourishment to breast milk or formula to the diet of older children who eat more of certain types of foods than adults. A child's behavior and lifestyle also influence exposure. Children crawl on the floor, put things in their mouths, sometimes eat inappropriate things (such as dirt or paint chips), and spend more time outdoors. Children also are closer to the ground, and they do not use the judgment of adults to avoid hazards (NRC 1993).

As with adults, biomonitoring is not considered relevant for assessing childhood exposure to chlorine. Therefore, information on chlorine levels in blood, urine, tissue, breast milk, neonatal blood, cord blood, and meconium fluid is not available. Children may be exposed to chlorine through the same routes that affect adults, except for occupational exposures. Children located near an accidental release of chlorine such as a leak from a factory or a chlorine tank spill or rupture may be exposed to high concentrations of chlorine through inhalation, skin contact, or eye contact. Children may be exposed to high levels of chlorine if they are in an area where swimming pool chemicals are being improperly used or where certain household chemicals are mixed together. Mixing an acid, such as toilet cleaner, with bleach can generate chlorine gas if the pH of the bleach is lowered to below 4.

6.7 POPULATIONS WITH POTENTIALLY HIGH EXPOSURES

Individuals located near an accidental release of chlorine may be exposed to high concentrations of this gas through inhalation, skin contact, and eye contact if the cloud travels in their direction (Horton et al. 2002; Wenck et al. 2007). Individuals who live near industrial facilities where chlorine gas is produced or used may be exposed to high concentrations if there is an accidental release of a large amount of

chlorine gas from the facility (USB 2007). Of the 865 chlorine-only events reported to ATSDR's Hazardous Substances Emergency Events Surveillance system between January 1993 and December 2000, 275 events involved victims (1,071 individuals) (Horton et al. 2002). Of these, 235 were members of the general public. A train derailment near Alberton, Montana on April 11, 1996 released approximately 130,000 pounds of chlorine gas to the atmosphere (NTSB 1998). According to the National Transportation Safety Board (NTSB), approximately 350 people were treated for chlorine inhalation. Nine people, including members of the train crew, were hospitalized. The NTSB reported that a transient riding the train died from acute chlorine toxicity (NTSB 1998). On January 6, 2005, a northbound Norfolk Southern Railway Company freight train traveling through Graniteville, South Carolina, encountered an improperly lined switch that diverted the train from the main track onto an industrial track, where it struck an unoccupied, parked train releasing approximately 120,000 pounds of chlorine gas from one of the cars of the northbound train (NTSB 2005). The train engineer and 8 other people died as a result of chlorine gas exposure and an additional 554 nearby residents were treated at local hospitals for respiratory illness as a result of this accident. On June 28, 2004 a westbound Union Pacific Railroad freight train collided with an eastbound freight train in Macdona, Texas (NTSB 2006). Consequently, a tank car loaded with liquefied chlorine was punctured, releasing a cloud of chlorine gas that surrounded the accident area. Three persons, including the conductor of the Union Pacific train and two local residents, died as a result of acute chlorine gas inhalation. The Union Pacific train engineer, 23 local residents, and 6 emergency responders were treated for respiratory illness or other injuries related to the collision and derailment.

6.8 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of chlorine is available. Where adequate information is not available, ATSDR, in conjunction with NTP, is required to assure the initiation of a program of research designed to determine the health effects (and techniques for developing methods to determine such health effects) of chlorine.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean

that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

6.8.1 Identification of Data Needs

Physical and Chemical Properties. The important chemical and physical properties for elemental chlorine are available (EPA 1999; HSDB 2009; Staudinger and Roberts 1996), as well as the important equilibrium constants describing the equilibrium reaction of chlorine in water (Cotton et al. 1999; Farr et al. 2003). No data need is identified at this time.

Production, Import/Export, Use, Release, and Disposal. According to the Emergency Planning and Community Right-to-Know Act of 1986, 42 U.S.C. Section 11023, industries are required to submit substance release and off-site transfer information to the EPA. The TRI, which contains this information for 2005, became available in May of 2007. This database is updated yearly and should provide a list of industrial production facilities and emissions.

Current production and U.S. import/export volumes are available for chlorine (CMR 2006; HSDB 2009; ITA 2007; The Chlorine Institute 2008; U.S. Census Bureau 2008), as well as adequate disposal methods (HSDB 2009). No data need is identified at this time.

Environmental Fate. The environmental fate of chlorine is understood. Chlorine is extremely reactive and will not remain in environmental media for long periods of time. The important equilibrium properties of chlorine in water are understood (Cotton et al. 1999; Farr et al. 2003). Scientists have proposed that minute quantities of chlorine are generated naturally during the photolysis of seawater aerosols (California Environmental Protection Agency 2002; Chang et al. 2004; Knipping and Dabdub 2003). Sunlight at tropospheric wavelengths (<430 nm) dissociates the chlorine molecule to form two chlorine radicals; a lifetime of <15 minutes (half-life of <11 minutes) was calculated for this reaction (Tanaka et al. 2003). No data need is identified at this time.

Bioavailability from Environmental Media. Chlorine is too reactive to be bioavailable from soil, water, or other environmental media. No data need is identified at this time.

Food Chain Bioaccumulation. Chlorine is too reactive to bioaccumulate in the food chain. No data need is identified at this time.

Exposure Levels in Environmental Media. Reliable monitoring data for the levels of chlorine in contaminated media at hazardous waste sites are needed so that the information obtained on levels of chlorine in the environment can be used in combination with the known body burden of chlorine to assess the potential risk of adverse health effects in populations living in the vicinity of hazardous waste sites.

Chlorine is too reactive to monitor for background levels in the environment. In the case of an accidental spill, levels >1,000 ppm have been observed in air around the accident site (U.S. Chemical Safety and Hazard Investigation Board 2003). Very low levels of chlorine (parts per trillion) are generated naturally during the photolysis of seawater aerosols (California Environmental Protection Agency 2002; Chang et al. 2004). No data need is identified at this time.

Exposure Levels in Humans. Humans can be exposed to chlorine following the accidental release at a manufacturing facility or an accident involving the transportation of liquefied chlorine gas (NTSB 1998, 2002, 2005, 2006); however, molecular chlorine levels in human tissues cannot be quantitatively assessed (see Chapter 7). In addition, people may be exposed to chlorine if they mix common household chemicals that are acidic with bleach or pool sanitizing chemicals (Agabiti et al. 2001; Bonetto et al. 2006). Continued monitoring of accidental chlorine releases and the health affects observed in humans is necessary.

This information is necessary for assessing the need to conduct health studies on these populations.

Exposures of Children. Similar to adults, exposure of children to chlorine gas primarily occurs from accidental industrial or transportation releases (NTSB 1998, 2002, 2005, 2006), or the inadvertent mixing of common household chemicals with bleach or pool sanitizing chemicals, resulting in the liberation of chlorine (Agabiti et al. 2001; Bonetto et al. 2006). Continued monitoring of accidental chlorine releases and the health affects observed in children is necessary.

Child health data needs relating to susceptibility are discussed in Section 3.12.2, Identification of Data Needs: Children's Susceptibility.

Exposure Registries. No exposure registries for chlorine were located. This substance is not currently one of the compounds for which a sub-registry has been established in the National Exposure Registry. The substance will be considered in the future when chemical selection is made for sub-

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registries to be established. The information that is amassed in the National Exposure Registry facilitates the epidemiological research needed to assess adverse health outcomes that may be related to exposure to this substance.

6.8.2 Ongoing Studies

Two ongoing studies have been located in the Federal Research in Progress Database (FEDRIP 2009) that will measure levels of chlorine in the marine atmosphere. These are listed in Table 6-2. No other ongoing studies regarding the potential for human exposure to chlorine were located.

Table 6-2. Ongoing Studies Regarding the Potential for Human Exposure to Chlorine

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Investigator	Affiliation	Research description	Sponsor
R. Talbot and A. Pszenny	University of New Hampshire	Study of reactive halogens in the marine boundary layer overlying the eastern tropical north Atlantic Ocean. This research will include measurement of inorganic chlorine gases.	National Science Foundation
E. Saltzman	University of California-Irvine	Study of the source, presence, and distribution of inorganic dihalogen gases in the marine atmosphere. This research will include measurement of chlorine gas in coastal and marine air.	National Science Foundation

Source: FEDRIP 2009

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7. ANALYTICAL METHODS

The purpose of this chapter is to describe the analytical methods that are available for detecting, measuring, and/or monitoring chlorine, its metabolites, and other biomarkers of exposure and effect to chlorine. The intent is not to provide an exhaustive list of analytical methods. Rather, the intention is to identify well-established methods that are used as the standard methods of analysis. Many of the analytical methods used for environmental samples are the methods approved by federal agencies and organizations such as EPA and the National Institute for Occupational Safety and Health (NIOSH). Other methods presented in this chapter are those that are approved by groups such as the Association of Official Analytical Chemists (AOAC) and the American Public Health Association (APHA). Additionally, analytical methods are included that modify previously used methods to obtain lower detection limits and/or to improve accuracy and precision.

7.1 BIOLOGICAL MATERIALS

Inhaled chlorine gas forms hypochlorous acid and hydrochloric acid upon contact with the moist mucous membranes of the upper respiratory tract (Vetrano 2001; Winder 2001). Since molecular chlorine reacts so quickly inside living systems, it is not found in biological materials. Therefore, analysis of these materials for chlorine is not relevant. Once they have been absorbed into the body, hypochlorous and hydrochloric acid are expected to react with proteins and nucleotides to produce a wide variety of chlorinated organic compounds (EPA 1999; Winder 2001). Based on a study that traced radiolabeled chlorine (as hypochlorite) through metabolism inside rats, it is expected that chlorine is ultimately converted to chloride in the blood and eliminated in the urine and feces of humans and animals primarily as the chloride ion (Abdel-Rahman et al. 1982, 1983; EPA 1999; Suh and Abdel-Rahman 1983). Chloroform has also been detected in the blood of rats exposed to hypochlorous acid (Abdel-Rahman et al. 1984). Since chloride is a natural component of blood, urine, and feces, monitoring chloride concentrations in these materials would not be helpful for assessing exposure to chlorine.

7.2 ENVIRONMENTAL SAMPLES

When chlorine is released into the environment, it reacts very quickly with both organic and inorganic matter forming chloride ion and chlorinated compounds. Therefore, aside from low levels in marine aerosols above the open ocean, and higher concentrations in the areas surrounding recent spills and leaks, molecular chlorine is not found in the environment.

Standardized methods have been established for analyzing chlorinated water for free chlorine (APHA 1998a, 1998b). Free chlorine refers to the combination of the equilibrium species aqueous molecular chlorine, hypochlorous acid, and the hypochlorite ion. These methods do not differentiate between the molecular chlorine and the hypochlorite species. Since molecular chlorine is usually not present in water samples, these tests typically measure the amounts of hypochlorous acid, hypochlorite, and chlorinated derivatives. The most popular of these tests is the DPD (N,N-diethyl-p-phenyldiamine) test (APHA 1998a, 1998b). A small amount of DPD is added to a water sample, which is immediately oxidized by free chlorine to produce a relatively stable free radical and results in a reddish-colored solution. The total chlorine is measured spectrophotometrically at 515 nm (APHA 1998a, 1998b). Some important residuals of water disinfection can also quantified by this method. Since chloramines are slow to react with DPD, they are quantified by the subsequent addition of potassium iodide. The iodide ion acts catalytically causing color production by monochloramine and dichloramine (APHA 1998a, 1998b). The free chlorine and the chloroamines are often referred to as the total chlorine content of the water.

Aside from the DPD test, free chlorine can be measured using the amperometric titration method or the starch-iodide titration method (APHA 1998a). The amperometric titration method involves the titration of the buffered sample with phenylarsine oxide (APHA 1998a). The decrease of free chlorine during the titration is detected by applying an electric potential across two electrodes and measuring the change in current through the solution. The starch-iodide titration method involves addition of potassium iodide and a starch indicator to the sample followed by titration with sodium thiosulfate (APHA 1998a). The end point is reached when the blue color of the solution disappears.

It should be noted that the free chlorine test methods described here work by detecting the presence of oxidizing species and are not actually specific and selective to free chlorine (hypochlorite and hypochlorous acid) (APHA 1998a). Therefore, care must be taken to avoid interference due to non-free chlorine oxidizing or reducing agents. The amperometric titration method is less affected by interference, temperature variations, turbidity, and color; however, this method requires greater operator skill to achieve reliable results (APHA 1998a).

Four standardized methods have been located that describe procedures for measuring molecular chlorine in air (EPA 2000b; NIOSH 1994; OSHA 2007a, 2007b). In EPA Method OAQPS-26, the air sample is passed through a particulate filter followed by a dilute sulfuric acid solution (EPA 2000b). Hydrogen chloride dissolves to form chloride in the acid solution, while chlorine, which is relatively insoluble in the acid, passes through to a dilute sodium hydroxide solution. Chlorine dissolves and disporportionates to

form both chloride and hypochlorous acid. Sodium thiosulfate is then added to the alkaline solution to ensure complete reaction with the hypochlorous acid, freeing the second chloride ion. Analysis is performed using ion chromatography.

OSHA Methods ID-101 and ID-126SGX are based on the reaction between chlorine and iodide to form iodine and chloride (OSHA 2007a, 2007b). In Method ID-101, chlorine is collected in a sulfamic acid solution, which is then reacted with potassium iodide and analyzed using a residual chlorine ion specific electrode. In Method ID-126SGX, chlorine is collected into a neutral solution of potassium iodide, which is then titrated with sodium thiosulfate. A second titration involving chlorine dioxide is performed next. The concentrations of chlorine and chlorine dioxide are determined using stoichiometric calculations. Disadvantages of this method are that it suffers from many interferences and that temperature and strong light affect solution solubility. Both of the OSHA methods recommend using a filter to eliminate particulates that may cause interference.

NIOSH Method 6011 describes a way to measure chlorine in air samples via collection onto a silver membrane filter, desorption into sodium thiosulfite, and subsequent analysis using ion chromatography (NIOSH 1994). This method is subject to positive interference from hydrogen chloride and negative interference from hydrogen sulfide. Also, silver chloride is photosensitive; therefore, the silver filter must be transferred to an amber bottle in the dark. Once the silver chloride has desorbed, it is no longer photosensitive. The detection limit for chlorine listed in this method is 0.007 ppm for a 90L air sample collected at a flow rate of 0.3–1 L per minute; however, Chang et al. (2004) was able to measure chlorine in air to a detection limit of 50 ppt (parts per trillion) using a Dionex DX-120 analyzer and longer sampling times.

Standardized methods for measuring chlorine in air and water including accuracy, detection limits, and additional details are listed in Table 7-1. Methods that analyze soil and sediment for chlorine are not available.

7.3 ADEQUACY OF THE DATABASE

Section 104(i)(5) of CERCLA, as amended, directs the Administrator of ATSDR (in consultation with the Administrator of EPA and agencies and programs of the Public Health Service) to assess whether adequate information on the health effects of chlorine is available. Where adequate information is not available, ATSDR, in conjunction with NTP, is required to assure the initiation of a program of research

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Table 7-1. Analytical Methods for Determining Chlorine in Environmental Samples

Sample	Description weatherd	Analytical	Sample	Percent	Deference
matrix	Preparation method	method	detection limit	recovery	Reference
Air	Sample is passed through a particulate filter into a dilute sulfuric acid solution followed by a dilute sodium hydroxide solution. Analysis is performed using IC.	EPA OAQPS- 26	0.1 ppm	NA	EPA 2000b
Air	Sample is passed through a teflon prefilter and dissolved in a sulfamic acid solution. Potassium iodide is added to the acid solution. Analysis is performed using RCE.	OSHA ID-101	0.14 ppm	NA	OSHA 2007b
Air	Sample is passed through a glass giber pre-filter and collected into a potassium iodide solution. The solution is then titrated with sodium thiosulfate in two steps to determine concentrations of chlorine and chlorine dioxide.	OSHA ID- 126SGX	0.3 ppm	NA	OSHA 2007a
Air	Sample is passed through a silver membrane filter. Silver chloride is desorbed from the membrane into a sodium thiosulfate solution, which is then analyzed using IC.	NIOSH 6011	0.007–0.5 ppm for a 90 L air sample (flow rate 0.3– 1.0 L/minute); 50 ppt	98.6%	NIOSH 1994; Chang et al. 2004
Water	Acetic acid and potassium iodide are added to the sample to create an acidic solution, which is then titrated using sodium thiosulfate.	APHA 4500-CI B	40 μg as free chlorine/L	NA	APHA 1998a, 1998b
Water	Acetic acid and potassium iodide are added to the sample to create an acidic solution, which is then titrated using sodium thiosulfate or phenylarsine oxide.	EPA 330.3	250–4,020 µg as free chlorine/L	NA	CAS 1978c

Table 7-1. Analytical Methods for Determining Chlorine in Environmental Samples

Sample matrix	Preparation method	Analytical method	Sample detection limit	Percent recovery	Reference
Water	Phenylarsine oxide, potassium iodide, and acetate buffer are combined with the sample. For starch iodide determination, a starch indicator is added followed by titration with iodine solution. For amperometric determination, titrate with an iodine solution using an amperometric titrator.	EPA 330.2	250–4,020 µg as free chlorine/L	NA	CAS 1978b
Water	Phosphate buffer is added to the sample, which is then titrated with phenylarsine oxide using a microammeter to observe current changes.	APHA 4500-CI D	200 μg as free chlorine/L	NA	APHA 1998a
Water	Potassium iodide and acetate buffer are added to the sample, which is then titrated with phenylarsine oxide or sodium thiosulfate using an amperometer to determine the end point.	EPA 330.1	380–3,500 µg as free chlorine/L	NA	CAS 1978a
Water	Add 10 mL of water to 0.5 mL o phosphate buffer solution and 0.5 mL of DPD reagent. Quantify by UV-VIS at 515 nm	f APHA 4500-CI G	10 μg as free chlorine/L	NA	APHA 1998a, 1998b

 $\label{eq:decomposition} \mbox{DPD} = \mbox{N,N-diethyl-p-phenyldiamine; IC = ion chromatography; NA = not available; RCE = residual chlorine ion specific electrode; UV-VIS = ultraviolet-visible$

designed to determine the health effects (and techniques for developing methods to determine such health effects) of chlorine.

The following categories of possible data needs have been identified by a joint team of scientists from ATSDR, NTP, and EPA. They are defined as substance-specific informational needs that if met would reduce the uncertainties of human health assessment. This definition should not be interpreted to mean that all data needs discussed in this section must be filled. In the future, the identified data needs will be evaluated and prioritized, and a substance-specific research agenda will be proposed.

7.3.1 Identification of Data Needs

Methods for Determining Biomarkers of Exposure and Effect.

Exposure. There are no biomarkers of exposure that are specific to chlorine. Chlorine reacts very quickly inside the body; therefore, analyzing for this substance in biological materials is not relevant. The chlorine reacts to form many chlorinated compounds that are ultimately broken down into chloride ion. Chloride is a natural component of blood, urine, and feces. Monitoring for chloride in biological materials would not be helpful in gauging exposure to chlorine.

Effect. There are no biomarkers of effect that are unique to chlorine exposure. The most obvious effect of exposure to high levels of chlorine is damage to the moist mucous membranes of the lungs and respiratory pathways. Other health effects that have been associated with exposure to chlorine include bronchitis, asthma, pulmonary edema, dermatitis, and conjunctivitis. The odor threshold for chlorine in air is 0.2–0.4 ppm and the lowest concentration in air at which there is perceivable sensory irritation is 1 ppm (EPA 1999; The Chlorine Institute 1998; WHO 1982). Current standardized methods for measuring chlorine in air have detection limits ranging from 0.007 to 0.3 ppm, which are below concentrations at which biological effects occur (EPA 2000b; NIOSH 1994; OSHA 2007a, 2007b).

Methods for Determining Parent Compounds and Degradation Products in Environmental

Media. Methods sensitive enough to measure background chlorine concentrations in ocean air are available. Chang et al. (2004) used a more sensitive analyzer and larger sampling volumes with the silver membrane filter method (NIOSH Method 6011) to lower the detection limit from 0.007 ppm to 50 ppt. Background chlorine concentrations measured were <600 ppt. Methods are also available for measuring chlorine in water; however, these methods do not distinguish between the equilibrium species molecular

chlorine, hypochlorous acid, or hypochlorite (APHA 1998a, 1998b). Since hypochlorous acid and hypochlorite are the dominant species at environmental pH, analyzing water for molecular chlorine is not relevant except under very acidic conditions (pH<4) (APHA 1998a, 1998b; Farr et al. 2003). Methods that analyze for chlorine in soil or sediment were not located. Chlorine is not expected to be found in these media since it is so reactive, rapidly oxidizing both organic and inorganic materials that it comes into contact with.

Among the analytical methods referred to in this chapter, only the detection limits reported for NIOSH Method 6011 are sensitive enough to detect chlorine in air at levels at or below the MRL values derived in Chapter 3. For reference, the acute, intermediate, and chronic MRLs derived for chlorine in air are 0.06 ppm, 0.002 ppm, and 50 ppt, respectively. Detection limits reported for NIOSH Method 6011 range from 0.007–0.5 ppm, while a detection limit of 50 ppt was achieved for a modification of this method (NIOSH 1994; Chang et al. 2004). The other detection methods for measuring chlorine levels in air report detection limits of >0.1 ppm (EPA 2000b). Based on the lack of sensitivity of the available methods relative to the derived MRL values, the development of standardized analytical methods that can detect chlorine in air at levels below the MRL values (below 50 ppt) is a data need.

7.3.2 Ongoing Studies

One ongoing study has been located in the Federal Research in Progress Database (FEDRIP 2009) related to the development of analytical methods for chlorine. This study will be led by investigator Eric Saltzman of the University of California-Irvine and is sponsored by the National Science Foundation (NSF). The study will explore the source, presence, and distribution of inorganic dihalogen gases in the marine atmosphere and will include the validation and further development of analytical methods for dihalogens, including chlorine, in air. No other ongoing studies regarding the development of methods for analyzing for chlorine in the environment were located in the available literature.

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8. REGULATIONS, ADVISORIES, AND GUIDELINES

MRLs are substance specific estimates, which are intended to serve as screening levels, are used by ATSDR health assessors and other responders to identify contaminants and potential health effects that may be of concern at hazardous waste sites.

ATSDR has derived an acute-duration inhalation MRL of 0.06 ppm for chlorine based on a NOAEL of 0.5 ppm for sensory irritation and pulmonary effects in volunteers exposed for up to 8 hours/day (Anglen 1981; D'Alessandro et al. 1996; Rotman et al. 1983; Schins et al. 2000; Shusterman et al. 1998, 2003b). The NOAEL was duration-adjusted for continuous exposure. An uncertainty factor of 3 was used to account for sensitive populations.

ATSDR has derived an intermediate-duration inhalation MRL of 0.002 ppm for chlorine based on a minimal LOAEL of 0.5 ppm for tracheal lesions in rats exposed 6 hours/day, 5 days/week for 62 days (Kutzman 1983). An uncertainty factor of 90 was used (3 for extrapolation from animals to humans with dosimetric adjustment, 3 for use of a minimal LOAEL, and 10 for human variability).

ATSDR has derived a chronic-duration inhalation MRL of 0.00005 ppm for chlorine based on an increased incidence of nasal lesions in monkeys exposed to chlorine 6 hours/day, 5 days/week for 1 year (Klonne et al. 1987). The MRL was derived using benchmark modeling of incidence data for nasal lesions in monkeys. The predicted exposure concentration associated with a 10% extra risk (BMC₁₀) for nasal lesions in monkeys was 0.04 ppm; the lower 95% confidence limit on this concentration (BMCL₁₀) was 0.02 ppm. An uncertainty factor of 30 was used (3 for extrapolation from animals to humans with dosimetric adjustment and 10 for human variability).

Oral MRLs were not derived for aqueous chlorine for the following reasons. MRLs are derived when reliable and sufficient data exist to identify a target organ(s) of effect or the most sensitive health effect(s) for a specific duration within a given route of exposure. Scientifically, as part of having sufficient and reliable data, it is important to be able to see the full, or at least a significant range, of the dose-response curve. In the case of the oral database for aqueous chlorine, no reliable LOAEL could be identified at levels of aqueous chlorine that could reasonably be encountered in the environment. It is a matter of policy of ATSDR not to derive free-standing MRLs.

EPA (IRIS 2007) has established an oral reference dose (RfD) for chlorine of 0.1 mg/kg/day based on a NOAEL of 14.4 mg/kg/day for systemic effects in F344/N rats exposed to chlorine in the drinking water for 2 years (NTP 1992). The uncertainty factor used in this assessment was 100 (10 for interspecies extrapolation and 10 for the protection of sensitive human subpopulations).

EPA has not derived an inhalation reference concentration (RfC) for chlorine gas.

The International Agency for Research on Cancer (IARC), the National Toxicology Program (NTP), and EPA has not classified chlorine, sodium hypochlorite, or hypochlorous acid for human carcinogenicity (IARC 2006; IRIS 2007; NTP 2005). The American Conference of Governmental Industrial Hygienists (ACGIH) has classified chlorine as an A4 carcinogen (not classifiable as a human carcinogen) (ACGIH 2006).

OSHA has required employers of workers who are occupationally exposed to chlorine to institute engineering controls and work practices to reduce and maintain employee exposure at or below permissible exposure limits (PELs) (OSHA 2006c). The employer must use engineering and work practice controls to reduce exposures to not exceed 1 ppm for chlorine at any time (ceiling) (OSHA 2006c).

EPA has designated chlorine as a hazardous air pollutant (HAP) under the Clean Air Act (CAA) (EPA 2007b). Chlorine and sodium hypochlorite are on the list of chemicals appearing in "Toxic Chemicals Subject to Section 313 of the Emergency Planning and Community Right-to-Know Act of 1986" and has been assigned a reportable quantity (RQ) limit of 10 and 1 pounds, respectively (EPA 2007e). Chlorine is also considered to be an extremely hazardous substance (EPA 2007f). The RQ represents the amount of a designated hazardous substance which, when released to the environment, must be reported to the appropriate authority.

Under the Federal Insecticide, Fungicide, and Rodenticide Act (FIFRA), chlorine gas is exempt from the requirement of a tolerance for pesticide chemicals in food when used as pre- or postharvest in solution on all raw agricultural commodities (EPA 2007h) and sodium hypochlorite is exempt from the requirement of a tolerance for residues in food (EPA 2007k).

The international and national regulations, advisories, and guidelines regarding chlorine in air, water, and other media are summarized in Table 8-1.

Table 8-1. Regulations, Advisories, and Guidelines Applicable to Chlorine and Chlorine Compounds

IARC Carcinogenicity classification No data IARC 2006 WHO Air quality guidelines No data WHO 2000 Drinking water quality guidelines WHO 2004 Chlorine 5 mg/L ^a NATIONAL Regulations and Guidelines:	Agency	Description	Information	Reference
IARC	INTERNATIONAL	=		
WHO Air quality guidelines Drinking water quality guidelines Chlorine No data WHO 2000 WHO 2004 NATIONAL Chlorine 5 mg/L³ Regulations and Guidelines: a. Air ACGIH TLV (8-hour TWA) (chlorine) 0.5 ppm ACGIH 2006 STEL (15-minute TWA) (chlorine) 1 ppm AIHA 2004 ERPG-1 ^b (chlorine) 3 ppm ERPG-2 ^b (chlorine) ERPG-3 ^b (chlorine) 20 ppm AIHA 2004 EPA AEGL-1 ^c (chlorine) 0.08 ppm EPA 2007a EPA AEGL-1 ^c (chlorine) EPA 2007a EPA 2007a EPA AEGL-1 ^c (chlorine) 0.5 ppm AEGL-2 ^c (chlorine) A hours 0.5 ppm AEGL-2 ^c (chlorine) AEGL-2 ^c (chlorine) 10 minutes 2.8 ppm 60 minutes 2.8 ppm 60 minutes 2.0 ppm AEGL-3 ^c (chlorine) AEGL-3 ^c (chlorine) 10 minutes 50 ppm AEGL-3 ^c (chlorine) AEGL-3 ^c (chlorine) 10 minutes 2.8 ppm AEGL-3 ^c (chlorine) AEGL-3 ^c (chlorine) 10 minutes 50 ppm AEGL-3 ^c (chlorine) <th< td=""><td>Guidelines:</td><td></td><td></td><td></td></th<>	Guidelines:			
Drinking water quality guidelines	IARC	Carcinogenicity classification	No data	IARC 2006
Chlorine S mg/La	WHO	Air quality guidelines	No data	WHO 2000
NATIONAL Regulations and Guidelines: a. Air		Drinking water quality guidelines		WHO 2004
Regulations and Guidelines: a. Air ACGIH TLV (8-hour TWA) (chlorine) STEL (15-minute TWA) (chlorine) 1 ppm AIHA ERPG-1 ^b (chlorine) ERPG-2 ^b (chlorine) ERPG-3 ^b (chlorine) Odor threshold (chlorine) 10 minutes 0.5 ppm 10 minutes 0.5 ppm 30 minutes 0.5 ppm 4 hours AEGL-2 ^c (chlorine) 10 minutes 2.5 ppm AEGL-2 ^c (chlorine) 10 minutes 0.5 ppm 4 hours 0.5 ppm 4 hours 0.5 ppm AEGL-2 ^c (chlorine) 10 minutes 0.5 ppm AEGL-2 ^c (chlorine) 10 minutes 2.8 ppm 60 minutes 2.9 ppm 4 hours 10 ppm AEGL-3 ^c (chlorine) 10 minutes 2.9 ppm 4 hours 30 minutes 50 ppm 4 hours 10 ppm AEGL-3 ^c (chlorine) 10 minutes 28 ppm 60 minutes 29 ppm 4 hours 10 minutes 20 ppm		Chlorine	5 mg/L ^a	
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4 hours 10 ppm		30 minutes	28 ppm	
		60 minutes	20 ppm	
8 hours 7.1 ppm		4 hours	10 ppm	
		8 hours	7.1 ppm	

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Table 8-1. Regulations, Advisories, and Guidelines Applicable to Chlorine and Chlorine Compounds

Agency	Description	Information	Reference
NATIONAL (cont.)			
	Hazardous air pollutant (chlorine)	Yes	EPA 2007b 42 USC 7412
	Regulated toxic substance and the threshold quantity for accidental release prevention (chlorine)	2,500 pounds ^d	EPA 2007d 40 CFR 68.130
NIOSH	REL (ceiling) (chlorine)	0.5 ppm	NIOSH 2005
	IDLH (chlorine)	10 ppm	
OSHA	PEL (ceiling) for general industry (chlorine)	1 ppm	OSHA 2006c 29 CFR 1910.1000
	PEL (ceiling) for shipyard industry (chlorine)	1 ppm	OSHA 2006a 29 CFR 1915.1000
	PEL (ceiling) for construction industry (chlorine)	1 ppm	OSHA 2006b 29 CFR 1926.55, Appendix A
	Toxic and reactive highly hazardous chemical which presents a potential for a catastrophic event at or above the threshold quantity (chlorine)	1,500 pounds	OSHA 2006d 29 CFR 1910.119
b. Water			
EPA	Designated as hazardous substances in accordance with Section 311(b)(2)(A) of the Clean Water Act (chlorine)		EPA 2007i 40 CFR 116.4
	Drinking water standards and health advisories (chlorine)		EPA 2006
	1-day health advisory for a 10-kg child	3 mg/L	
	10-day health advisory for a 10-kg child	3 mg/L	
	DWEL	5 mg/L	
	Lifetime	4 mg/L	
	10 ⁻⁴ Cancer risk	No data	
	National primary drinking water regulations (chlorine [as Cl ₂]) ^e		EPA 2003
	MRDL	4.0 mg/L	
	Public health goal (MRDLG)	4.0 mg/L	
	Reportable quantities of hazardous substances designated pursuant to Section 311 of the Clean Water Act		EPA 2007j 40 CFR 117.3
	Chlorine	10 pounds	
	Sodium hypochlorite	100 pounds	

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Table 8-1. Regulations, Advisories, and Guidelines Applicable to Chlorine and Chlorine Compounds

Αç	gency	Description	Information	Reference
N/	ATIONAL (cont.)			
c.	Food			
	EPA	Chlorine is exempt from the requirement of a tolerance for pesticide chemicals in food when used as preharvest or postharvest in solution on all raw agricultural commodities	Yes	EPA 2007h 40 CFR 180.1095
		Sodium hypochlorite is exempt from the requirement of a tolerance for residues in food	Yes	EPA 2007k 40 CFR 180.1235
	FDA	EAFUS (ingredient added directly to food that FDA has either approved as a food additive or listed or affirmed as GRAS) (chlorine and sodium hypochlorite)	Yes	FDA 2007a
		Food additives permitted for direct addition to food for human consumption (sodium hypochlorite)	Yes	FDA 2007b 21 CFR 172
		Indirect food additives: adhesives and components of coatings (sodium hypochlorite)	Yes	FDA 2006 21 CFR 175
d.	Other			
	ACGIH	Carcinogenicity classification (chlorine)	A4 ^f	ACGIH 2006
	EPA	Carcinogenicity classification (chlorine)	No data	IRIS 2007
		RfD (chlorine)	0.1 mg/kg/day	
		RfC (chlorine)	No data	
		Superfund, emergency planning, and community right-to-know		
		Designated CERCLA hazardous substance (chlorine and sodium hypochlorite)	Yes ^g	EPA 2007e 40 CFR 302.4
		Reportable quantity		
		Chlorine	10 pounds	
		Sodium hypochlorite	1 pound	
		Extremely hazardous substance and the threshold planning quantity (chlorine)	100 pounds	EPA 2007f 40 CFR 355, Appendix A
		Effective date of toxic chemical release reporting (chlorine)	01/01/87	EPA 2007g 40 CFR 372.65

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Table 8-1. Regulations, Advisories, and Guidelines Applicable to Chlorine and Chlorine Compounds

Agency	Description	Information	Reference
NATIONAL (
EPA	TSCA Master Testing List		EPA 2007c
	Chlorine	Yes ^h	
	Sodium hypochlorite	Yes ⁱ	
NTP	Carcinogenicity classification	No data	NTP 2005

^aFor effective disinfection, there should be a residual concentration of free chlorine of ≥0.5mg/litre after at least 30 minute contact time at pH <8.0 (WHO 2004).

ACGIH = American Conference of Governmental Industrial Hygienists; AEGL = Acute Exposure Guideline Levels; AIHA = American Industrial Hygiene Association; CERCLA = Comprehensive Environmental Response, Compensation, and Liability Act; CFR = Code of Federal Regulations; CTPU = Chemical Testing Program Underway; DWEL = drinking water equivalent level; EAFUS = Everything Added to Food in the United States; ECA = Enforceable Consent Agreement; EPA = Environmental Protection Agency; ERPG = Emergency Response Planning Guidelines; FDA = Food and Drug Administration; FRM = Final Rule-Making; GRAS = Generally Recognized As Safe; IARC = International Agency for Research on Cancer; IDLH = immediately dangerous to life or health; IRIS = Integrated Risk Information System; MCL = Maximum Contanimant Level; MRDL = Maximum Residual Disinfectant Level; MRDLG = Maximum Residual Disinfectant Level Goal; NIOSH = National Institute for Occupational Safety and Health; NTP = National Toxicology Program; OSHA = Occupational Safety and Health Administration; PEL = permissible exposure limit; REL = recommended exposure limit; RfC = inhalation reference concentration; RfD = oral reference dose; SIDS = Screening Information Data Set; STEL = short-term exposure limit; TLV = threshold limit values; TSCA = Toxic Substances Control Act; TWA = time-weighted average; USC = United States Code; VTA = Voluntary Testing Agreement; WHO = World Health Organization

^bERPG-1 is the maximum airborne concentration below which it is believed nearly all individuals could be exposed for up to 1 hour without experiencing or developing other than mild, transient adverse health effects or without perceiving a clearly defined objectionable odor; ERPG-2 is the maximum airborne concentration below which it is believed nearly all individuals could be exposed for up to 1 hour without experiencing or developing irreversible or other serious health effects or symptoms that could impair an individual's ability to take protective action; and ERPG-3 is the maximum airborne concentration below which it is believed nearly all individuals could be exposed for up to 1 hour without experiencing or developing life-threatening health effects (AIHA 2004).

^cAEGL-1 is the airborne concentration of a substance above which it is predicted that the general population, including susceptible individuals, could experience notable discomfort, irritation, or certain asymptomatic nonsensory effects, however, the effects are not disabling and are transient and reversible upon cessation of exposure; AEGL-2 is the airborne concentration of a substance above which it is predicted that the general population, including susceptible individuals, could experience irreversible or other serious, long-lasting adverse health effects or an impaired ability to escape; and AEGL-3 is the airborne concentration of a substance above which it is predicted that the general population, including susceptible individuals, could experience life-threatening health effects or death (EPA 2007a).

^dRegulated toxic and flammable substance under Section 112 of the Clean Air Act

^ePotential health effects from exposure above the MCL include anemia and nervous system effects in infants and young children; the common source of the contaminant in drinking water is the use of chlorine as a water additive used to control microbes (EPA 2003).

fA4: not classifiable as a human carcinogen

⁹Designated CERCLA hazardous substance pursuant to Section 112 of the Clean Air Act.

^hThe Office of Air and Radiation recommended chlorine for acute toxicity testing for health effects. Chlorine is a hazardous air pollutant and was added to the Master Testing List in 1995. EPA is initiating development of a testing action via TSCA Section 4 FRM, a TSCA Section 4 ECA, or a VTA (EPA 2007c).

ⁱThe Organization for Economic Cooperation and Development recommended sodium hypochlorite for a "base set" of screening level test data (SIDS), which include health effects and environmental effects and fate. Sodium hypochlorite was added to the Master Testing List in 1995 and EPA is initiating development of a testing action via CTPU-VTA (EPA 2007c).

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9. REFERENCES

Abdel-Rahman MS, Couri D, Bull RJ. 1982. Metabolism and pharmacokinetics of alternate drinking water disinfectants. Environ Health Perspect 46:19-23.

Abdel-Rahman MS, Sub DH, Bull RJ. 1984. Pharmacodynamics and toxicity of chlorine in drinking water in the rat. J Appl Toxicol 4(2):82-86.

Abdel-Rahman MS, Waldron DM, Bull RJ. 1983. A comparative kinetics study of monochloramine and hypochlorous acid in the rat. J Appl Toxicol 3:175-179.

ACGIH. 2006. Chlorine. Threshold limit values for chemical substances and physical agents and biological exposure indices. Cincinnati, OH: American Conference of Governmental Industrial Hygienists.

Adelson L, Kaufman J. 1971. Fatal chlorine poisoning: Report of two cases with clinicopathologic correlation. Am J Clin Pathol 56(4):430-442.

Adinolfi M. 1985. The development of the human blood-CSF-brain barrier. Dev Med Child Neurol 27(4):532-537.

Adlercreutz H. 1995. Phytoestrogens: Epidemiology and a possible role in cancer protection. Environ Health Perspect Suppl 103(7):103-112.

Agabiti N, Ancona C, Forastiere F, et al. 2001. Short term respiratory effects of acute exposure to chlorine due to a swimming pool accident. Occup Environ Med 58(6):399-404.

Agency for Toxic Substances and Disease Registry. 1989. Decision guide for identifying substance-specific data needs related to toxicological profiles; Notice. Agency for Toxic Substances and Disease Registry, Division of Toxicology. Fed Regist 54(174):37618-37634.

Agency for Toxic Substances and Disease Registry. 1990. Biomarkers of organ damage or dysfunction for the renal, hepatobiliary, and immune systems. Subcommittee on Biomarkers of Organ Damage and Dysfunction. Atlanta, GA: Agency for Toxic Substances and Disease Registry.

Agency for Toxic Substances and Disease Registry. 1998. Alberton chlorine spill; Alberton, Montana. Phase 1 study report. Atlanta, GA: Agency for Toxic Substances and Disease Registry. PB99115883.

Agency for Toxic Substances and Disease Registry. 2005. Notice of the revised priority list of hazardous substances that will be the subject of toxicological profiles. Fed Regist 70:72840-72842.

AIHA. 2004. Chlorine. Emergency Response Planning Guidelines (ERPG). Fairfax, VA: American Industrial Hygiene Association.

Alarie Y. 1973. Sensory irritation by airborne chemicals. CRC Crit Rev Toxicol 2:299-363.

^{*} Not cited in text

Alarie Y. 1981. Toxicological evaluation of airborne chemical irritants and allergens using respiratory reflex reactions. In: Leong BKJ, ed. Proceedings of the inhalation toxicology and technology symposium. Ann Arbor, MI: Ann Arbor Science Publishers, Inc., 207-231.

Almagro Nievas D, Acuria Castillo R, Hernandez Jerez A, et al. 2008. [Investigation of an outbreak of acute respiratory illness due to exposure to chlorine gas in a public swimming pool.] Gig Sanit 22(3):287-290. (Spanish)

Altman PL, Dittmer DS. 1974. Biological handbooks: Biology data book. Vol. III. 2nd ed. Bethesda, MD: Federation of American Societies for Experimental Biology, 1987-2008, 2041.

Amoore JE, Hautala E. 1983. Odor as an aid to chemical safety: Odor thresholds compared with threshold limit values and volatilities for 214 industrial chemicals in air and water dilution. J Appl Toxicol 3(6):272-290.

Andersen ME, Krishnan K. 1994. Relating *in vitro* to *in vivo* exposures with physiologically based tissue dosimetry and tissue response models. In: Salem H, ed. Animal test alternatives: Refinement, reduction, replacement. New York, NY: Marcel Dekker, Inc., 9-25.

Andersen ME, Clewell HJ, Gargas ML, et al. 1987. Physiologically based pharmacokinetics and the risk assessment process for methylene chloride. Toxicol Appl Pharmacol 87(2):185-205.

Anglen DA. 1981. Sensory response of human subjects to chlorine in air. Ann Arbor, MI: University of Michigan.

APHA. 1998a. Method 4500-Cl. Chlorine (residual). Standard methods for the examination of waste and wastewater. 20th ed. Washington, DC: American Public Health Association, 4-53 to 4-66.

APHA. 1998b. Method 4500 Cl-. Chloride. Standard methods for the examination of waste and wastewater. 20th ed. Washington, DC: American Public Health Association, 4-66 to 4-73.

Babl FE, Kharsch S, Woolf A. 1998. Airway edema following household bleach ingestion. Am J Emerg Med 16:514-516.

Babu RV, Cardenas V, Sharma G. 2008. Acute respiratory distress syndrome from chlorine inhalation during a swimming pool accident: A case report and review of the literature. J Intensive Care Med 23(4):275-280.

Barbone F, Delzell E, Austin H, et al. 1992. A case-control study of lung cancer at a dye and resin manufacturing plant. Am J Ind Med 22(6):835-849.

Barnes DG, Dourson M. 1988. Reference dose (RfD): Description and use in health risk assessments. Regul Toxicol Pharmacol 8(4):471-486.

Barregård L, Sallsten G, Jarvholm B. 1990. Mortality and cancer incidence in chloralkali workers exposed to inorganic mercury. Br J Ind Med 47(2):99-104.

Barrow CS, Steinhagen WH. 1982. Sensory irritation tolerance development to chlorine in F-344 rats following repeated inhalation. Toxicol Appl Pharmacol 65(3):383-389.

Barrow CS, Alarie Y, Warrick JC, et al. 1977. Comparison of the sensory irritation response in mice to chlorine and hydrogen chloride. Arch Environ Health 32(2):68-76.

Barrow CS, Kociba RJ, Rampy LW, et al. 1979. An inhalation toxicity study of chlorine in Fisher-344 rats following 30 days of exposure. Toxicol Appl Pharmacol 49:77-88.

Barrow RE, Smith RG. 1975. Chlorine-induced pulmonary function changes in rabbits. Am Ind Hyg Assoc J 36(5):398-403.

Baxter PJ, Davies PC, Murray V. 1989. Medical planning for toxic releases into the community: The example of chlorine gas. (Comment in: Br J Ind Med 46(10):752). Br J Ind Med 46(4):277-285.

Beach FX, Jones ES, Scarrow GD. 1969. Respiratory effects of chlorine gas. Br J Ind Med 26(3):231-236.

Becker M, Forrester M. 2008. Pattern of chlorine gas exposures reported to Texas poison control centers, 2000 through 2005. Tex Med 104(3):52-57.

Berger GS, ed. 1994. Epidemiology of endometriosis. In: Endometriosis: Advanced management and surgical techniques. New York, NY: Springer-Verlag, 3-7.

Berghoff RS. 1919. The more common gases; their effect on the respiratory tract. Arch Intern Med 24:678-684.

BG Chimie. 1991. Sodium hypochlorite. Toxicological evaluations. 4. Berlin: Springer. Berufgenossenschaft der Chemischen Industrie, 257-299.

Bhérer L, Cushman R, Courteau JP, et al. 1994. Survey of construction workers repeatedly exposed to chlorine over a three to six month period in a pulpmill: II. Follow-up of affected workers by questionnaire, spirometry, and assessment of bronchial responsiveness 18 to 24 months after exposure ended. Occup Environ Med 51(4):225-228.

Bommaraju TV, Luke B, O'Brien TF, et al. 2004. Chlorine. In: Kirk-Othmer encyclopedia of chemical technology. Volume 6. John Wiley & Sons, Inc. http://www.mrw.interscience.wiley.com/emrw/9780471238966/kirk/article/chlocurl.a01/current/pdf. March 29, 2007.

Bond GG, Cook RR, Wight PC, et al. 1983. A case-control study of brain tumor mortality at a Texas chemical plant. J Occup Med 25:377-386.

Bond GG, Flores GH, Shellenberger RJ, et al. 1986. Nested case-control study of lung cancer among chemical workers. Am J Epidemiol 124(1):53-66.

Bond GG, Shellenberger RJ, Flores GH, et al. 1985. A case-control study of renal cancer mortality at a Texas chemical plant. Am J Ind Med 7:123-139.

Bonetto G, Corradi M, Carraro S, et al. 2006. Longitudinal monitoring of lung injury in children after acute chlorine exposure in a swimming pool. Am J Respir Crit Care Med 174(5):545-549.

Bosse GM. 1994. Nebulized sodium bicarbonate in the treatment of chlorine gas inhalation. J Toxicol Clin Toxicol 32(3):233-241.

Bracco D, Dubois MJ, Bouali R. 2005. Intoxication by bleach ingestion. Can J Anaesth 52(1):118-119.

Brooks SM, Weiss MA, Bernstein IL. 1985. Reactive airways dysfunction syndrome. Case reports of persistent airways hyperactivity following high-level irritant exposures. J Occup Med 27(7):473-476.

Buckley LA, Jiang XZ, James RA, et al. 1984. Respiratory tract lesions induced by sensory irritants at the RD₅₀ concentration. Toxicol Appl Pharmacol 74:417-429.

California Environmental Protection Agency. 2002. Chlorine emissions from activated sea-salt aerosols and their potential impact on ozone. Sacramento, CA: California Environmental Protection Agency. PB2003100841. http://www.arb.ca.gov/research/apr/past/00324.pdf. May 17, 2007.

Carlton BD, Bartlett P, Basaran A, et al. 1986. Reproductive effects of alternative disinfectants. Environ Health Perspect 69:237-241.

CAS. 1978a. Method #: 330.1. Chlorine, total residual (titrimetric, amperometric) Columbia Analytical Services. http://www.caslab.com/EPA-Methods/PDF/EPA-Method-3301.pdf. May 27, 2010.

CAS. 1978b. Method #: 330.2. Chlorine, total residual (titrimetric, back, iodometric (starch or amperometric) Columbia Analytical Services. http://www.caslab.com/EPA-Methods/PDF/EPA-Method-3302.pdf. May 27, 2010.

CAS. 1978c. Method #: 330.3. Chlorine, total residual (titrimetric, iodometric) Columbia Analytical Services. http://www.caslab.com/EPA-Methods/PDF/EPA-Method-3303.pdf. May 27, 2010.

CDC. 1991. Chlorine gas toxicity from mixture of bleach with other cleaning products — California (Erratum in: MMWR Morb Mortal Wkly Rep 40(47):819). MMWR Morb Mortal Wkly Rep 40(36):619-629.

CDC. 2005. Public health consequences from hazardous substances acutely released during rail transit—South Carolina, 2005; selected states, 1999-2004. MMWR Morb Mortal Wkly Rep 54(3):64-67.

CDC. 2009. CDC Healthy swimming. http://cdc.gov/healthyswimming. February 27, 2009.

Chang CT, Liu TH, Jeng FT. 2004. Atmospheric concentrations of the Cl atom, ClO radical, and HO radical in the coastal marine boundary layer. Environ Res 94(1):67-74.

Chang JCF, Barrow CS. 1984. Sensory irritation tolerance and cross-tolerance in F-344 rats exposed to chlorine or formaldehyde gas. Toxicol Appl Pharmacol 76(2):319-327.

*Chang JH, Vogt CR, Sun AY. 1981. Effects of acute administration of chlorinated water on liver lipids. Lipids 16(5):336-340.

Chasis H, Zapp JA, Bannon JH, et al. 1947. Chlorine accident in Brooklyn. Occup Med (Lond) 4:152-170.

Chester EH, Gillespie DG, Krause FD. 1969. The prevalence of chronic obstructive pulmonary disease in chlorine gas workers. Am Rev Respir Dis 99:365-373.

CHLORINE 195 9. REFERENCES

Chester EH, Kaimal J, Payne CB, et al. 1977. Pulmonary injury following exposure to chlorine gas. Possible beneficial effects of steroid treatment. Chest 72(2):247-250.

Clewell HJ, Andersen ME. 1985. Risk assessment extrapolations and physiological modeling. Toxicol Ind Health 1(4):111-131.

CMR. 1977. Chemical profile: Chlorine. Chem Market Rep. June 27, 1977.

CMR. 1980. Chemical profile: Chlorine. Chem Market Rep. April 14, 1980.

CMR. 1989. Chemical profile: Chlorine. Chem Market Rep. June 12, 1989.

CMR. 1992. Chemical profile: Chlorine. Chem Market Rep. June 01, 1992.

CMR. 1995. Chemical profile: Chemical. Chem Market Rep. June 12, 1995.

CMR. 2000. Chemical profile: Chlorine. Chem Market Rep. September 04, 2000.

CMR. 2003. Chemical profile: Chlorine. Chem Market Rep. June 16, 2003.

CMR. 2006. Chemical profile: Chlorine. Chem Market Rep. May 21, 2006.

Compton JAF. 1987. Chlorine. In: Military chemical and biological agents. Chemical and toxicological properties. Caldwell, NJ: Telford Press, 113-118.

Costero C, Falcón Escobedo R. 1983. [Gas pneumopathy. Chlorine poisoning and the participation of oxygen in the pathological changes.] Salud Publica Mex 25(3):265-272. (Spanish)

Cotton FA, Wilkinson G, Murillo CA, et al., eds. 1999. Advanced inorganic chemistry. New York, NY: John Wiley & Sons, Inc., 550, 564, 565.

Courteau JP, Cushman R, Bouchard F, et al. 1994. Survey of construction workers repeatedly exposed to chlorine over a three to six month period in a pulpmill. I. Exposure and symptomatology. Occup Environ Med 51(4):219-224.

Cunningham HM. 1980. Effect of sodium hypochlorite on the growth of rats and guinea pigs. Am J Vet Res 41(2):295-297.

Curlin LC, Bommaraju TV, Hansson CB. 1991. Alkali and chlorine products. In: Kroschwitz JI, Howe-Grant M, eds. Kirk-Othmer encyclopedia of chemical technology. Vol. 1. New York, NY: John Wiley & Sons, Inc., 938-1025.

D'Alessandro A, Kuschner W, Wong H, et al. 1996. Exaggerated responses to chlorine inhalation among persons with nonspecific airway hyperreactivity. Chest 109(2):331-337.

Daniel FB, Condie LW, Robinson M, et al. 1990. Comparative subchronic toxicity studies of three disinfectants. J Am Water Works Assoc 82:61-69.

Daniel FB, Ringhand HP, Robinson M, et al. 1991. Comparative subchronic toxicity of chlorine and monochloramine in the B6C3F1 mouse. J Am Water Works Assoc 83(11):68-75.

Das TK. 2002. Disinfection. In: Kirk-Othmer encyclopedia of chemical toxicology. Vol. 8. New York, NY: John Wiley & Sons, Inc., 605-672.

http://www.mrw.interscience.wiley.com/emrw/9780471238966/kirk/article/disibloc.a01/current/pdf. March 28, 2007.

Decker WJ, Koch HF. 1978. Chlorine poisoning at the swimming pool. An overlooked hazard. Clin Toxicol 13(3):377-381.

Demnati R, Fraser R, Plaa G, et al. 1995. Histopathological effects of acute exposure to chlorine gas on Sprague-Dawley rat lungs. J Environ Pathol Toxicol Oncol 14(1):15-19.

Deutsch ZG. 1947. Alkali and chlorine industries. In: Kirk RE, Othmer DF, eds. Encyclopedia of chemical technology. Vol. 1. New York, NY: The Interscience Encyclopedia, Inc., 358-430.

Deutsch ZG. 1963. Alkali and chlorine industries. In: Standen A, ed. Kirk-Othmer encyclopedia of chemical technology. Vol. 1. New York, NY: Interscience Publishers, 668-758.

DOA. 1933. Chlorine. The residual effects of warfare gases. Washington, DC: U.S. Department of Army, 1-41.

Dodd DE, Bus JS, Barrow CS. 1980. Lung sulfhydryl changes in rats following chlorine inhalation. Toxicol Appl Pharmacol 52(2):199-208.

DOE. 2005a. Detailed numerical simulation of the Graniteville train collision. U.S. Department of Energy. WSRC-MS-2005-00635.

DOE. 2005b. The Savannah River National Laboratory's response to the Graniteville, SC train accident. Washington, DC: U.S. Department of Energy. WSRC-MS-2005-00612. DE2006881477.

Donnelly SC, FitzGerald MX. 1990. Reactive airways dysfunction syndrome (RADS) due to chlorine gas exposure. Isr J Med Sci 159(9-12):276-277.

Douidar SM. 1997. Nebulized sodium bicarbonate in acute chlorine inhalation. Pediatr Emerg Care 13(6):406-407.

Edstrom Industries. 2003. Forms of chlorine in water. Edstrom Industries, Inc. http://www.edstrom.com/DocLib/MI4148.pdf. May 27, 2010.

Eiserich JP, Cross CE, Jones AD, et al. 1996. Formation of nitrating and chlorinating species by reaction of nitrite with hypochlorous acid. A novel mechanism for nitric oxide-mediated protein modification. J Biol Chem 271(32):19199-19208.

Ellenhorn MJ. 1997. Chlorine. In: Ellenhorn's medical toxicology. Diagnosis and treatment of human poisoning. 2nd ed. Baltimore, MD: Williams and Williams, 1521.

Ellenhorn MJ, Barceloux DG. 1988. Chlorine. In: Medical toxicology. Diagnosis and treatment of human poisoning. New York, NY: Elsevier, 878-879.

Enarson DA, Maclean L, Dybuncio A, et al. 1984. Respiratory health at a pulpmill in British Columbia. Arch Environ Health 39(5):325-330.

EPA. 1988. Recommendations for and documentation of biological values for use in risk assessment. Cincinnati, OH: U.S. Environmental Protection Agency, Environmental Criteria and Assessment Office. EPA600687008.

EPA. 1990. Interim methods for development of inhalation reference concentrations. Washington, DC: U.S. Environmental Protection Agency, Office of Health and Environmental Assessment, Office of Research and Development, Environmental Criteria and Assessment Office. EPA600890066A.

EPA. 1993. A literature review of atmospheric transformation products of Clean Air Act Title III hazardous air pollutants. Research Triangle Park, NC: U.S. Environmental Protection Agency. EPA600R94088.

EPA. 1994a. Methods for Derivation of Inhalation Reference Concentrations and Application of Inhalation Dosimetry. Washington, DC: U.S. Environmental Protection Agency, Office of Research and Development. EPA600890066F.

EPA. 1994b. Final draft for the drinking water criteria document on chlorine dioxide, chlorite and chlorate. Cincinnati, OH: U.S. Environmental Protection Agency. EPA68C20139. PB94179884.

EPA. 1997. Special report on environmental endocrine disruption: An effects assessment and analysis. Washington, DC: U.S. Environmental Protection Agency, Risk Assessment Forum. EPA630R96012.

EPA. 1999. Reregistration eligibility decision (RED). Chlorine gas. U.S. Environmental Protection Agency. EPA738R99001. http://www.epa.gov/oppsrrd1/REDs/4022red.pdf. March 14, 2007.

EPA. 2000a. Benchmark dose technical guidance document. Washington, DC: U. S. Environmental Protection Agency, Risk Assessment Forum. EPA630R00001.

*EPA. 2000b. Method 26. Hydrogen chloride, halides, halogens. Emission Measurement Center. U.S. Environmental Protection Agency. http://www.epa.gov/ttn/emc/promgate/m-26.pdf. April 12, 2007.

EPA. 2003. National primary drinking water regulations. Washington, DC: U.S. Environmental Protection Agency, Office of Ground Water and Drinking Water. EPA816F03016. http://www.epa.gov/safewater/mcl.html. March 07, 2006.

EPA. 2005. Toxic chemical release inventory reporting forms and instructions: Revised 2004 version. Section 313 of the Emergency Planning and Community Right-to-Know Act (Title III of the Superfund Amendments and Reauthorization Act of 1986). U.S. Environmental Protection Agency. Office of Environmental Information. EPA260B05001.

EPA. 2006a. 2006 edition of drinking water standards and health advisories. Washington, DC: U.S. Environmental Protection Agency. EPA822R04005. http://epa.gov/waterscience/criteria/drinking/. April 11, 2007.

EPA. 2006b. Determining active oxidant species reacting with organophosphate pesticides in chlorinated drinking water. Washington, DC: U.S. Environmental Protection Agency, Office of Research and Development. EPA600R06103.

EPA. 2007a. Acute exposure guideline levels (AEGLs). Washington, DC: U.S. Environmental Protection Agency, Office of Pollution Prevention and Toxics. http://www.epa.gov/oppt/aegl/pubs/compiled.pdf. May 11, 2007.

EPA. 2007i. Designated as hazardous substances in accordance with Section 311(b)(2)(A) of the Clean Water Act. Code of Federal Regulations. 40 CFR 116.4. Washington, DC: U.S. Environmental Protection Agency. http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm. April 11, 2007.

EPA. 2007b. Hazardous air pollutants. Clean Air Act. U.S. Environmental Protection Agency. United States Code. 42 USC 7412. http://www.epa.gov/ttn/atw/orig189.html. April 11, 2007.

EPA. 2007c. Master testing list. Washington, DC: Office of Pollution Prevention and Toxics, U.S. Environmental Protection Agency. http://www.epa.gov/opptintr/chemtest/pubs/mtl.htm. May 10, 2007.

EPA. 2007d. Regulated toxic substances and threshold quantities for accidental release prevention. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 68.130. http://www.epa.gov/epacfr40/chapt-Linfo/chi-toc.htm. April 11, 2007.

EPA. 2007j. Reportable quantities of hazardous substances designated pursuant to Section 311 of the Clean Water Act. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 117.3. http://www.epa.gov/epacfr40/chapt-Linfo/chi-toc.htm. April 11, 2007.

EPA. 2007e. Superfund, emergency planning, and community right-to-know programs. Designation, reportable quantities, and notifications. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 302.4. http://www.epa.gov/epacfr40/chapt-Linfo/chi-toc.htm. April 11, 2007.

EPA. 2007f. Superfund, emergency planning, and community right-to-know programs. Extremely hazardous substances and their threshold planning quantities. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 355, Appendix A. http://www.epa.gov/epacfr40/chapt-I.info/chitoc.htm. April 11, 2007.

EPA. 2007g. Superfund, emergency planning, and community right-to-know programs. Toxic chemical release reporting. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 372.65. http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm. April 11, 2007.

EPA. 2007h. Tolerances and exemptions from tolerances for pesticide chemicals in food. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 180.1095. http://www.epa.gov/epacfr40/chapt-I.info/chi-toc.htm. May 11, 2007.

EPA. 2007k. Tolerances and exemptions from tolerances for pesticide chemicals in food. U.S. Environmental Protection Agency. Code of Federal Regulations. 40 CFR 180.1235. http://www.epa.gov/epacfr40/chapt-Linfo/chi-toc.htm. June 28, 2007.

Eun HC, Young Lee A, Lee YS. 1984. Sodium hypochlorite dermatitis. Contact Dermatitis 11:45.

Evans RB. 2004. Chlorine: State of the art. Lung 183(3):151-167.

Exon JH, Koller LD, O'Reillly CA, et al. 1987. Immunotoxicologic evaluation of chlorine-based drinking water disinfectants, sodium hypochlorite and monochloramine. Toxicology 44:257-269.

Farr JP, Smith WL, Steichen DS. 2003. Bleaching agents. In: Kirk-Othmer encyclopedia of chemical toxicology. Vol 4. John Wiley & Sons, Inc., 43-81. http://www.mrw.interscience.wiley.com/emrw/9780471238966/kirk/article/survfarr.a01/current/pdf. March 28, 2007. FDA. 2006. Indirect food additives: Adhesives and components of coatings. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 175. http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm. May 10, 2007.

FDA. 2007a. Everything added to food in the United States (EAFUS). Washington, DC: U.S. Food and Drug Administration. http://vm.cfsan.fda.gov/~dms/eafus.html. April 11, 2007.

FDA. 2007b. Food additives permitted for direct addition to food for human. U.S. Food and Drug Administration. Code of Federal Regulations. 21 CFR 172. http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm. May 15, 2007.

FEDRIP. 2009. Chlorine. Federal Research in Progress database. Springfield, VA: National Technical Information Service. January 21, 2009.

Ferris BG, Burgess WA, Worcester J. 1967. Prevalence of chronic respiratory disease in a pulp mill and a paper mill in the United States. Br J Ind Med 24:26-37.

Ferris BG, Puleo S, Chen HY. 1979. Mortality and morbidity in a pulp and a paper mill in the United States: A ten-year follow-up. Br J Ind Med 36:127-134.

Fleta J, Calvo C, Zuniga J, et al. 1986. Intoxication of 76 children by chlorine gas. Hum Toxicol 5(2):99-100.

Fomon SJ. 1966. Body composition of the infant: Part I: The male reference infant. In: Falkner F, ed. Human development. Philadelphia, PA: WB Saunders, 239-246.

Fomon SJ, Haschke F, Ziegler EE, et al. 1982. Body composition of reference children from birth to age 10 years. Am J Clin Nutr 35(Suppl 5):1169-1175.

Foulks CJ. 1981. Myasthenia gravis presenting as laryngeal stridor after exposure to chlorine gas. South Med J 74(11):1423-1424.

Fukayama MY, Tan H, Wheeler WB, et al. 1986. Reactions of aqueous chlorine and chlorine dioxide with model food compounds. Environ Health Perspect 69:267-274.

Furukawa F, Kurata Y, Kokubo T, et al. 1980. [Oral acute and subchronic toxicity studies for sodium hypochlorite in F-344 rats.] Eisei Shikensho Hokoku 98:62-69. (Japanese)

Gagnaire F, Azim S, Bonnet P, et al. 1994. Comparison of the sensory irritation response in mice to chlorine and nitrogen trichloride. J Appl Toxicol 14(6):405-409.

Gapany-Gapanavicius M, Yellin A, Almog S, et al. 1982. Pneumomediastinum: A complication of chlorine exposure from mixing household cleaning agents. J Am Med Assoc 248(3):349-350.

Gautrin D, Leroyer C, Infante-Rivard C, et al. 1999. Longitudinal assessment of airway caliber and responsiveness in workers exposed to chlorine. Am J Respir Crit Care Med 160:1232-1237.

Gautrin D, Leroyer C, L'Archeveque J, et al. 1995. Cross-sectional assessment of workers with repeated exposure to chlorine over a three year period. Eur Respir J 8(12):2046-2054.

Giwercman A, Carlsen E, Keiding N, et al. 1993. Evidence for increasing incidence of abnormalities of the human testis: A review. Environ Health Perspect Suppl 101(2):65-71.

*Goffin V, Pierard GE, Henry F, et al. 1997. Sodium hypochlorite, bleaching agents, and the stratum corneum. Ecotoxicol Environ Saf 37:199-202.

Goldfrank LR, Flomenbaum NE, Lewin NA, et al. 2002. Chlorine. In: Goldfrank's toxicologic emergencies. 7th ed. New York, NY: McGraw-Hill, 1458-1459.

Graedel TE. 1978. Chemical compounds in the atmosphere. Orlando, FL: Academic Press, Inc., 30, 31, 34, 35.

Graedel TE, Hawkins DT, Claxton LD. 1986. Atmospheric chemical compounds: Sources, occurrence, and bioassay. Orlando, FL: Academic Press, Inc., 70-71, 93.

Grasemann H, Tschiedel E, Groch M, et al. 2007. Exhaled nitric oxide in children after accidental exposure to chlorine gas. Inhal toxicol 19(10):895-898.

*Griffith JF, Nixon GA, Bruce RD, et al. 1980. Dose-response studies with chemical irritants in the albino rabbit eye as a basis for selecting optimum testing conditions for predicting hazard to the human eye. Toxicol Appl Pharmacol 55:501-513.

Gross EA, Morgan KT. 1991. Architecture of nasal passages and larynx. In: Comparative biology of the normal lung. Boca Raton, FL: CRC Press, 7-25.

Güloğlu C, Kaara IH, Erten PG. 2002. Acute accidental exposure to chlorine gas in the Southeast of Turkey. A study of 106 cases. Environ Res 88(2):89-93.

Gunnarsson M, Walther SM, Seidal T, et al. 1998. Exposure to chlorine gas: Effects on pulmonary function and morphology in anaesthetised and mechanically ventilated pigs. J Appl Toxicol 18(4):249-255.

Guzelian PS, Henry CJ, Olin SS, eds. 1992. Similarities and differences between children and adults: Implications for risk assessment. Washington, DC: International Life Sciences Institute Press.

*Habets JMW, Geursen-Reitsma AM, Stole E, et al. 1986. Sensitization to sodium hypochlorite causing hand dermatitis. Contact Dermatitis 15:140-142.

Hagiwara M, Watanabe E, Barrett JC, et al. 2006. Assessment of genotoxicity of 14 chemical agents used in dental practice. Ability to induce chromosome aberrations in Syrian hamster embryo cells. Mutat Res 603(2):111-120.

Hasan FM, Gehshan A, Fuleihan FJ. 1983. Resolution of pulmonary dysfunction following acute chlorine exposure. Arch Environ Health 38(2):76-80.

Hasegawa R, Takahashi M, Kokubo T, et al. 1986. Carcinogenicity study of sodium hypochlorite in F344 rats. Food Chem Toxicol 24(12):1295-1302.

Hayashi M, Kishi M, Sofuni T, et al. 1988. Micronucleus tests in mice on 39 additives and eight miscellaneous chemicals. Food Chem Toxicol 26(6):487-500.

Hayatsu H, Hoshino H, Kawazoe Y. 1971. Potential cocarcinogenicity of sodium hypochlorite. Nature 233:495.

HazDat. 2007. Chlorine. HazDat Database: ATSDR's Hazardous Substance Release and Health Effects Database. Atlanta, GA: Agency for Toxic Substances and Disease Registry. http://www.atsdr.cdc.gov/hazdat.html. May 17, 2007.

Heldaas SS, Langard S, Andersen A. 1998. Incidence of cancer in a cohort of magnesium production workers. Br J Ind Med 46:617-623.

Henneberger PK, Lax MB, Ferris BG Jr. 1996. Decrements in spirometry values associated with chlorine gassing events and pulpmill work. Am J Respir Crit Care Med 153(1):225-231.

*Hess JA, Molinari JA, Gleason MJ, et al. 1991. Epidermal toxicity of disinfectants. Am J Dent 41(1):51-56.

Hoel DG, Davis DL, Miller AB, et al. 1992. Trends in cancer mortality in 15 industrialized countries, 1969-1986. J Natl Cancer Inst 84(5):313-320.

Hook CT, Lowry LD. 1974. Effect of chlorine bleach on the esophagus. Ann Otol Rhinol Laryngol 83:709-713.

Horton DK, Berkowitz Z, Kaye WE. 2002. The public health consequences from acute chlorine releases, 1993-2000. J Occup Environ Med 44(10):906-913.

*Hostynek JJ, Patrick E, Younger B, et al. 1989. Hypochlorite sensitivity in man. Contact Dermatitis 20:32-37.

Hostynek JJ, Wilhelm KP, Cua AB, et al. 1990. Irritation factors of sodium hypochlorite solutions in human skin. Contact Dermatitis 23(5):316-324.

Hov O. 1985. The effect of chlorine on the formation of photochemical oxidants in southern Telemark, Norway. Atmos Environ 19(3):417-485.

Howard C, Ducre B, Burda AM, et al. 2007. Management of chlorine gas exposure. J Emerg Nurs 33(4):402-404.

HSDB. 2009. Chlorine. Hazardous Substances Data Bank. National Library of Medicine. http://toxnet.nlm.nih.gov. March 11, 2009.

Hyback B. 1999. A long-term study of pulmonary function at low exposures to chlorine. Int Arch Occup Environ Health 72:M24-M28.

IARC. 1991. IARC monographs on the evaluation of carcinogenic risks to humans. Volume 52. Lyon, France: World Health Organization. International Agency for Research on Cancer, 45-71.

IARC. 2006. Agents reviewed by the IARC monographs. Volumes 1-96. Lyon, France: International Agency for Research on Cancer. http://monographs.iarc.fr/ENG/Classification/index.php. April 11, 2007.

Ibanes JD, LeinigerJR, Jarabek AM, et al. 1996. Reexamination of respiratory tract response in rats, mice and rhesus monkeys chronically exposed to inhaled chlorine. Inhal Toxicol 8:859-876.

IPCS. 1996. Chlorine. Poisons information monograph 947. International Programme on Chemical Safety. PIM 947. http://www.inchem.org/documents/pims/chemical/pim947.htm. March 11, 2009.

IRIS. 2007. Chlorine. Integrated Risk Information System. Washington, DC: U.S. Environmental Protection Agency. http://www.epa.gov/iris/subst/index.html. May 11, 2007.

Ishidate M, Sofuni T, Yoshikawa K, et al. 1984. Primary mutagenicity screening of food additives currently used in Japan. Food Chem Toxicol 22(8):623-636.

ITA. 2007. 2801100000--Chlorine: U.S. Trade quick-reference tables. International Trade Administration, U.S. Department of Commerce. http://ita.doc.gov/td/industry/otea/trade-detail/index.html. April 2, 2007.

Jakobsson SW, Rajs J, Jonsson JA, et al. 1991. Poisoning with sodium hypochlorite solution. Report of a fatal case, supplemented with an experimental and clinic-epidemiological study. Am J Forensic Med Pathol 12(4):320-327.

Jappinen P, Hakulinen T, Pukkala E, et al. 1987. Cancer incidence of workers in the Finnish pulp and paper industry. Scand J Work Environ Health 13:197-202.

Jarabek AM, Schroeter JD, Andersen ME, et al. 2007. A hybrid CFD-PBPK model of chlorine gas uptake and tissue dosimetry in the upper respiratory tract (URT) of F344 rats. Toxicologist 96(1):83.

Jiang XZ, Buckley LA, Morgan KT. 1983. Pathology of toxic responses to the RD50 concentration of chlorine gas in the nasal passages of rats and mice. Toxicol Appl Pharmacol 71(2):225-236.

Johanson CE. 1980. Permeability and vascularity of the developing brain: Cerebellum vs. cerebral cortex. Brain Res 190(1):3-16.

Jones RN, Hughes JM, Glindmeyer H, et al. 1986. Lung function after acute chlorine exposure. Am Rev Respir Dis 134(6):1190-1195.

Joy RJT. 1997. Historical aspects of medical defense against chemical warfare. In: Office of the Surgeon General, Department of the Army, eds. Textbook of military medicine: Medical aspects of chemical and biological warfare. Washington, DC: U.S. Government Printing Office, 87-109.

Joyner RE, Durel EG. 1962. Accidental liquid chlorine spill in a rural community. J Occup Med 4:152-154.

Kaufman J, Burkons D. 1971. Clinical, roentgenologic, and physiologic effects of acute chlorine exposure. Arch Environ Health 23(1):29-34.

Kennedy SM, Enarson DA, Janssen RG, et al. 1991. Lung health consequences of reported accidental chlorine gas exposure among pulpmill workers. Am Rev Respir Dis 143(1):74-79.

Kilburn KH. 1995. Evidence that inhaled chlorine is neurotoxic and causes airways obstruction. Int J Occup Med Toxicol 4(2):267-276.

Kilburn KH. 2000. Chlorine-induced damage documented by neurophysiological, neuropsychological, and pulmonary testing. Arch Environ Health 55(1):31-37.

Kilburn KH. 2003b. Effects of chlorine and its cresylate byproducts on brain and lung performance. Arch Environ Health 58(12):746-755.

Klonne DR, Ulrich CE, Riley MG, et al. 1987. One-year inhalation toxicity study of chlorine in Rhesus monkeys (*Macaca mulatta*). Fundam Appl Toxicol 9(3):557-572.

Knipping EM, Dabdub D. 2003. Impact of chlorine emissions from sea-salt aerosol on coastal urban ozone. Environ Sci Technol 37(2):275-284.

Komori M, Nishio K, Kitada M, et al. 1990. Fetus-specific expression of a form of cytochrome P-450 in human livers. Biochemistry 29(18):4430-4433.

Kowitz TA, Reba RC, Parker RT, et al. 1967. Effects of chlorine gas upon respiratory function. Arch Environ Health 14(4):545-558.

Krishnan K, Andersen ME. 1994. Physiologically based pharmacokinetic modeling in toxicology. In: Hayes AW, ed. Principles and methods of toxicology. 3rd ed. New York, NY: Raven Press, Ltd., 149-188.

Krishnan K, Andersen ME, Clewell HJ, et al. 1994. Physiologically based pharmacokinetic modeling of chemical mixtures. In: Yang RSH, ed. Toxicology of chemical mixtures: Case studies, mechanisms, and novel approaches. San Diego, CA: Academic Press, 399-437.

Kurokawa Y, Takamura N, Matsushima Y, et al. 1984. Studies on the promoting and complete carcinogenic activities of some oxidizing chemicals in skin carcinogénesis. Cancer Lett 24:299-304.

*Kurokawa Y, Takayama S, Konishi Y, et al. 1986. Long-term *in vivo* carcinogenicity tests of potassium bromate, sodium hypochlorite, and sodium chlorite conducted in Japan. Environ Health Perspect 69:221-235.

Kutzman RS. 1983. A study of Fisher-344 rats subchronically exposed to 0, 0.5, 1.5, or 5.0 ppm chlorine. Upton, NY: Brookhaven National Laboratory. BNL 32710.

Landau GD, Saunders WH. 1964. The effect of chlorine bleach on the esophagus. Arch Otolaryngol 80:174-176.

Lakind JS, Holgate ST, Ownby DR, et al. 2007. A critical review of the use of Clara cell secretory protein (CC16) as a biomarker of acute or chronic pulmonary effects. Biomarkers 12(5):445-467.

Lapenna D, Cuccurullo F. 1996. Hypochlorous acid and its pharmacological antagonism: An update picture. Gen Pharmacol 27(7):1145-1147.

Lawson JJ. 1981. Chlorine exposure: A challenge to the physician. Am Fam Physician 23(1):135-138.

Le Curieux F, Marzin D, Erb F. 1993. Comparison of three short-term assays: Results on seven chemicals. Potential contribution to the control of water genotoxicity. Mutat Res 319(3):223-236.

Leeder JS, Kearns GL. 1997. Pharmacogenetics in pediatrics: Implications for practice. Pediatr Clin North Am 44(1):55-77.

Lemière C, Malo JL, Boutet M. 1997. Reactive airways dysfunction syndrome due to chlorine: Sequential bronchial biopsies and functional assessment. Eur Respir J 10(1):241-244.

Leonardos G, Kendall D, Barnard NJ. 1968. Odor threshold determination of 53 odorant chemicals In: 61st annual meeting of the Air Pollution Control Association, St. Paul, MN, June 23-27, 1968. Pittsburgh, PA: Air Pollution Control Association. Abstract 68-13.

Leroyer C, Malo JL, Girard D, et al. 1999. Chronic rhinitis in workers at risk of reactive airways dysfunction syndrome due to exposure to chlorine. Occup Environ Med 56(5):334-338.

Leroyer C, Malo JL, Infante-Rivard C, et al. 1998. Changes in airway function and bronchial responsiveness after acute occupational exposure to chlorine leading to treatment in a first aid unit. Occup Environ Med 55(5):356-359.

Leube G, Kreiter H. 1971. Akute chlorgasvergiftung. Med Klin [Klin] 66(10):354-357. (German)

Leung HW. 1993. Physiologically-based pharmacokinetic modelling. In: Ballantyne B, Marrs T, Turner P, eds. General and applied toxicology. Vol. 1. New York, NY: Stockton Press, 153-164.

Leustik M, Doran S, Bracher M, et al. 2008. Mitigation of chlorine-induced lung injury by low-molecular-weight antioxidants. Am J Physiol Lung Cell Mol Physiol 295(5):L733-L743.

Levy JM, Hessel SJ, Nykamp PW, et al. 1986. Detection of the cerebral lesions of chlorine intoxication by magnetic resonance imaging. Magn Reson Imaging 4(1):51-52.

Lewis RJ. 2000. Sax's dangerous properties of industrial materials. New York, NY: John Wiley & Sons, Inc., 776-777.

Lewis RJ. 2001. Hawley's condensed chemical dictionary. New York, NY: John Wiley & Sons, Inc., 246, 247.

Lide DR, ed. 2005. Chlorine. In: CRC handbook of chemistry and physics. Boca Raton: Taylor & Francis Group, 4-10, 4-57.

Livingston AL. 1978. Forage plant estrogens. J Toxicol Environ Health 4(2-3):301-324.

Lubbers JR, Chauan S, Bianchine JR. 1982. Controlled clinical evaluations of chlorine dioxide, chlorite and chlorate in man. Environ Health Perspect 46:57-62.

Martin JG, Campbell HR, Iijima H, et al. 2003. Chlorine-induced injury to the airways in mice. Am J Respir Crit Care Med 168(5):568-574.

Matsuoka A, Hayashi M, Ishidate N. 1979. Chromosomal aberration tests on 29 chemicals combined with S9 mix *in vitro*. Mutat Res 66:277-290.

Mayr U, Butsch A, Schneider S. 1992. Validation of two *in vitro* test systems for estrogenic activities with zearalenone, phytoestrogens and cereal extracts. Toxicology 74(2-3):135-149.

McNulty MJ, Chang JCF, Barrow CS. 1983. Sulfhydryl oxidation in rat nasal mucosal tissues after chlorine inhalation. Toxicol Lett 17:241-246.

Meakins JC, Priestley JG. 1919. The after-effects of chlorine gas poisoning. Can Med Assoc J 9:968-974.

Meier JR, Bull RJ, Stober JA, et al. 1985. Evaluation of chemicals used for drinking water disinfection for production of chromosomal damage and sperm-head abnormalities in mice. Environ Mutagen 7:201-211.

Mink FL, Coleman WE, Munch JW, et al. 1983. *In vivo* formation of halogenated reaction products following peroral sodium hypochlorite. Bull Environ Contam Toxicol 30:394-399.

Miyachi T, Tsutsui T. 2005. Ability of 13 chemical agents used in dental practice to induce sister-chromatid exchanges in Syrian hamster embryo cells. Odontology 93(1):24-29.

Morris JB, Wilkie WS, Shusterman DJ. 2005. Acute respiratory responses of the mouse to chlorine. Toxicol Sci 83:380-387.

Morris JC. 1946. The mechanism of the hydrolysis of chlorine. J Am Chem Soc 68(9):1692-1694.

Morselli PL, Franco-Morselli R, Bossi L. 1980. Clinical pharmacokinetics in newborns and infants: Age-related differences and therapeutic implications. Clin Pharmacokin 5(6):485-527.

Moulick ND, Banavali S, Abhyankar AD, et al. 1992. Acute accidental exposure to chlorine fumes. A study of 82 cases. Indian J Chest Dis Allied Sci 34(2):85-89.

Mrvos R, Dean BS, Krenzelok EP. 1993. Home exposures to chlorine/chloramine gas: Review of 216 cases. South Med J 86(6):654-657.

Mühlendahl KE, Oberdisse U, Krienke EG. 1978. Local injuries by accidental ingestion of corrosive substances by children. Arch Toxicol 39:299-314.

NAS/NRC. 1989. Report of the oversight committee. In: Biologic markers in reproductive toxicology. Washington, DC: National Academy of Sciences, National Research Council, National Academy Press, 15-35.

Nemery B, Hoet PHM, Nowak D. 2002. Indoor swimming pools, water chlorination and respiratory health. Eur Respir J 19:790-793.

Ngo A, Ponampalam R, Leong M, et al. 2007. Chlorine and its impact on an emergency department. Prehospital Disaster Med 22(2):136-139.

NIOSH. 1976. Criteria for a recommended standard. Occupational exposure to chlorine. Cincinnati, OH: National Institute for Occupational Safety and Health.

*NIOSH. 1991. Immune responsiveness in chlorine exposed rats. Cincinnati, OH: National Institute for Occupational Safety and Health. NIOSH-R03OH02425. PB92124478.

NIOSH. 1994. Method 6011. NIOSH manual of analytical methods. Atlanta, GA: National Institute for Occupational Safety and Health. http://www.cdc.gov/niosh/nman/methods-c/html. August 09, 2007.

NIOSH. 1995. Health hazard evaluation report. International association of fire fighters, Henderson, Nevada. National Institute for Occupational Safety and Health. HETA-91-0230-2543. http://www.cdc.gov/niosh/hhe/reports/pdfs/1991-0230-2543.pdf. March 19, 2007.

NIOSH. 2005. Chlorine. NIOSH pocket guide to chemical hazards. Atlanta, GA: National Institute for Occupational Safety and Health, Centers for Disease Control and Prevention. http://www.cdc.gov/niosh/npg/. May 11, 2007.

*Nixon GA, Tyson CA, Wertz WC. 1975. Interspecies comparisons of skin irritancy. Toxicol Appl Pharmacol 31:481-490.

Nodelman V, Ultman JS. 1999a. Longitudinal distribution of chlorine absorption in human airways: A comparison to ozone absorption. J Appl Physiol 87(6):2073-2080.

Nodelman V, Ultman JS. 1999b. Longitudinal distribution of chlorine absorption in human airways: Comparison of nasal and oral quiet breathing. J Appl Physiol 86(6):1984-1993.

NRC. 1993. Pesticides in the diets of infants and children. National Research Council. Washington, DC: National Academy Press.

NTP. 1992. Toxicology and carcinogenesis studies of chlorinated water (CAS Nos. 7782-50-5 and 7681-52-9) and chloraminated water (CAS No. 10599-90-3) (deionized and charcoal-filtered) in F344/N rats and B6C3F1 mice (drinking water studies). Research Triangle Park, NC: National Toxicology Program. NTP TR-392. NIH Publication No. 92-2847.

NTP. 2005. Introduction. Report on carcinogens. 11th ed. Research Triangle Park, NC: National Toxicology Program, U.S. Department of Health and Human Services, Public Health Service. http://ntp-server.niehs.nih.gov/ntp/roc/toc11.html. March 08, 2006.

NTSB. 1998. Railroad accident brief report. LAX 96 FR 010. Derailment and hazardous materials release with fatality. Montano Rail Lin, Alberton, Montana. April 11, 1996. Washington, DC: National Transportation Safety Board. RAB98-07.

NTSB. 2002. Hazardous materials accident report. Hazardous materials release from railroad tank car with subsequent fire at Riverview, Michigan, July 14, 2001. Washington, DC: National Transportation Safety Board. PB02917002.

NTSB. 2005. Collision of Norfolk Southern Freight Train 152 with standing Norfolk Southern Local Train P22 with subsequent hazardous materials release at Graniteville, South Carolina, January 06, 2005. Washington, DC: National Transportation Safety Board. PB05916304.

NTSB. 2006. National Transportation Safety Board railroad accident report: Collision of Union Pacific Railroad Train MHOTU-23 with BNSF Railway Company Train MEAP-TUL-126-D with subsequent derailment and hazardous materials release in Macdona, Texas, June 28, 2004. http://www.ntis.gov/search/product.asp?ABBR=PB2006916303&starDB=GRAHIST. July 05, 2007.

O'Neil MJ, Smith A, Heckelman PE, eds. 2001. Chlorine. In: The Merck index. Whitehouse Station, NJ: Merck & Co. Inc., 361-363.

OSHA. 2006a. Air contaminants. Occupational safety and health standards for shipyard employment. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1915.1000. http://www.osha.gov/comp-links.html. April 11, 2007.

OSHA. 2006b. Gases, vapors, fumes, dusts, and mists. Safety and health regulations for construction. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1926.55, Appendix A. http://www.osha.gov/comp-links.html. April 11, 2007.

OSHA. 2006c. Limits for air contaminants. Occupational safety and health standards. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1910.1000. http://www.osha.gov/comp-links.html. April 11, 2007.

OSHA. 2006d. List of highly hazardous chemicals, toxics, and reactives. Occupational safety and health standards. Occupational Safety and Health Administration. Code of Federal Regulations. 29 CFR 1910.119. http://www.osha.gov/comp-links.html. April 11, 2007.

OSHA. 2007a. Chlorine and chlorine dioxide in workplace atmospheres. Occupational Safety and Health Administration. http://www.osha.gov/dts/sltc/methods/inorganic/t-id126sgx-pv-01-0112-m/t-id126sgx-pv-0.html. July 05, 2007.

OSHA. 2007b. Chlorine in workplace atmospheres. Occupational Safety and Health Administration. http://www.osha.gov/dts/sltc/methods/inorganic/id101.html. July 05, 2007.

Osmundsen PE. 1978. Contact dermatitis due to sodium hypochlorite. Contact Dermatitis 4:177-178.

OTA. 1990. Neurotoxicity: Identifying and controlling poisons of the nervous system. Washington, DC: Office of Technology Assessment. OTABA438.

Owen GM, Brozek J. 1966. Influence of age, sex and nutrition on body composition during childhood and adolescence. In: Falkner F, ed. Human development. Philadelphia, PA: WB Saunders, 222-238.

Patil LR, Smith RG, Vorwald AJ, et al. 1970. The health of diaphragm cell workers exposed to chlorine. Am Ind Hyg Assoc J 31(6):678-686.

Pfeiffer EH. 1978. Health aspects of water chlorination with special consideration on the carcinogenicity of chlorine. Zentralb Bakteriol [B] 166:185-211.

Pike DG, Peabody JW, Davis EW, et al. 1963. A re-evaluation of the dangers of Clorox ingestion. J Pediatr 63(2):303-305.

Ploysongsang Y, Beach BC, DiLisio RE. 1982. Pulmonary function changes after acute inhalation of chlorine gas. South Med J 75(1):23-26.

Prater JF. 1990. Inhalation injury after exposure to chlorine gas leak. J Emerg Nurs 16(4):243-244.

Ramachandran KA, Chawla IS, Khokhar P. 1990. Acute chlorine poisoning: A study of 84 cases. J Assoc Physicians India 38(7):489-490.

Roberts KC, Moss OR, Sochaski MA, et al. 2007. Uptake and internal dosimetry of chlorine in the upper respiratory tract (URT) of F344 rats. Toxicologist 96(1):166.

Robertson AS. 1978. Alkali and chlorine products. In: Grayson M, Eckroth D, eds. Kirk-Othmer encyclopedia of chemical technology. Vol. 1. New York, NY: John Wiley & Sons, Inc., 799-883.

Rosenkranz HS. 1973. Sodium hypochlorite and sodium perborate: Preferential inhibitors of DNA polymerase-deficient bacteria. Mutat Res 21:171-174.

Ross MP, Spiller HA. 1999. Fatal ingestion of sodium hypochlorite bleach with associated hypernatremia and hyperchloremic metabolic acidosis. Vet Hum Toxicol 41(2):82-86.

Rotman HH, Fliegelman MJ, Moore T, et al. 1983. Effects of low concentrations of chlorine on pulmonary function in humans. J Appl Physiol 54(4):1120-1124.

RTECS. 2007. Chlorine. Registry of Toxic Effects on Chemical Substances. National Institute of Occupational Safety and Health. MDL Information Systems, Inc. May 15, 2007.

Rupp H, Henschler D. 1967. [Effect of low chlorine and bromine concentrations on man.] Int Arch Gewerbepathol Gewerbehyg 23(1):79-90. (German)

Ryazanov VA. 1962. Sensory physiology as basis for air quality standards. The approach used in the Soviet Union. Arch Environ Health 5:480-494.

Salisbury DA, Enarson DA, Chan-Yeung M, et al. 1991. First-aid reports of acute chlorine gassing among pulpmill workers as predictors of lung health consequences. Am J Ind Med 20:71-81.

Sandall TE. 1922. The later effects of gas poisoning. Lancet 2:857-859.

Sasaki M, Sugimura K, Mitsuaki AY, et al. 1980. Cytogenetic effects of 60 chemicals on cultured human and Chinese hamster cells. La Kromosom II 20:574-584.

Schins RP, Emmen H, Hoogendijk L, et al. 2000. Nasal inflammatory and respiratory parameters in human volunteers during and after repeated exposure to chlorine. (Comment in: Eur Respir J 19(2):381-382). Eur Respir J 16(4):626-632.

Schmittinger P, Florkiewicz T, Curlin LC, et al. 2006. Chlorine. In: Ullmann's encyclopedia of industrial chemistry. Wiley-VCH Verlag GmbH & Co.

http://www.mrw.interscience.wiley.com/emrw/9783527306732/ueic/article/a06_399/current/pdf. March 28, 2007.

Schraufstätter IU, Browne K, Harris A, et al. 1990. Mechanisms of hypochlorite injury of target cells. J Clin Invest 85:554-562.

Schreuder MDJ, Brewer CA. 2001. Effects of short-term exposure to chlorine gas on morphology and physiology of *Pinus ponderosa* and *Pseudotsuga menziesii*. Ann Bot 88:187-195.

Schulte EE. 1999. Soil land applied chlorine. http://learningstore.uvex.edu/pdf%5CA3556.pdf. May 16, 2007.

Schwartz DA, Smith DD, Lakshiminarayan S. 1990. The pulmonary sequelae associated with accidental inhalation of chlorine gas. Chest 97(4):820-825.

Setchell BP, Waites GMH. 1975. The blood-testis barrier. In: Creep RO, Astwood EB, Geiger SR, eds. Handbook of physiology: Endocrinology V. Washington, DC: American Physiological Society, 143-172.

Sexton JD, Pronchik DJ. 1998. Chlorine inhalation: The big picture. J Toxicol Clin Toxicol 36(1-2):87-93.

Shusterman D, Murphy MA, Balmes J. 1998. Subjects with seasonal allergic rhinitis and nonrhinitic subjects react differentially to nasal provocation with chlorine gas. J Allergy Clin Immunol 101(6 Pt 1):732-740.

Shusterman D, Murphy MA, Balmes J. 2003b. Influence of age, gender, and allergy status on nasal reactivity to inhaled chlorine. Inhal Toxicol 15(12):1179-1189.

Skljanskaja RM, Rappoport TL. 1935. [Experimental studies on chronic poisoning and the development of the offspring of chlorine-poisoned rabbits.] Naunyn Schmiedebergs Arch Exp Pathol Pharmakol 177:276-285. (German)

Snoeyink VL, Jenkins D. 1980. Water chemistry. New York, NY: John Wiley & Sons, Inc., 386-404.

Sochaski MA, Jarabek AM, Murphy J, et al. 2008. 3-Chlorotyrosine and 3,5-dichlorotyrosine as biomarkers of respiratory tract exposure to chlorine gas. J Anal Toxicol 32(1):99-105.

SRI. 2006. Chlorine. 2006 Directory of chemical producers. United States. Menlo Park, CA: SRI Consulting, 520-521.

SRI. 2008. Chlorine. 2008 Directory of chemical producers. United States. Menlo Park, CA: SRI Consulting, 486-487.

Staudinger J, Roberts PV. 1996. A critical review of Henry's law constants for environmental applications. Crit Rev Environ Sci 26:205-297.

Strange DC, Finneran JC, Shumacker HB, et al. 1951. Corrosive injury of the stomach. AMA Arch Surg 254(21):350-357.

Suh DH, Abdel-Rahman MS. 1983. Kinetics study of chlorine dioxide in rat. J Toxicol Environ Health 12:467-473.

Suzuki S, Sakamoto S, Maniwa K, et al. 2001. Fatal pulmonary arterial thrombosis associated with chlorine gas poisoning. Clin Appl Thromb Hemost 7(4):356-358.

Tanaka PL, Riemer DD, Chang S, et al. 2003. Direct evidence for chlorine-enhanced urban ozone formation in Houston, TX. Atmos Environ 37:1393-1400.

Tchobanoglous G, Schroeder ED. 1985. Water quality. Characteristics, modeling, modification. Reading, MA: Addison-Wesley Publishing Co., 559-570.

The Chlorine Institute. 1998. Molecular chlorine: Health and environmental effects. Pamphlet 90. Edition 2. The Chlorine Institute, Inc.

The Chlorine Institute. 2006. Pamphlet 96. Sodium hypochlorite manual. Edition 3. The Chlorine Institute, Inc.

The Chlorine Institute. 2008. North American chlor-alkali industry plants and production data report – 2007. Pamphlet 10. The Chlorine Institute, Inc.

Thomas HL, Murray V. 2008. Review of acute chemical incidents involving exposure to chlorine associated with swimming pools in England and Wales. J Public Health (Oxf) 30(4):391-397.

Thomas K, Colborn T. 1992. Organochlorine endocrine disruptors in human tissue. In: Colborn T, Clement C, eds. Chemically induced alterations in sexual and functional development: The wildlife/human connection. Princeton, NJ: Princeton Scientific Publishing, 365-394.

Tian X, Tao H, Brisolara J, et al. 2008. Acute lung injury induced by chlorine inhalation in C57BL/6 and FVB/N mice. Inhal Toxicol 20(9):783-793.

TRI06. 2008. TRI explorer: Providing access to EPA's toxics release inventory data. Washington, DC: Office of Information Analysis and Access. Office of Environmental Information. U.S. Environmental Protection Agency. Toxics Release Inventory. http://www.epa.gov/triexplorer/. February 04, 2009.

Underhill FP. 1920. Alterations in blood concentration. The lethal war gases. New Haven: Yale University Press, 40-83.

USB. 2007. Investigation report: Chlorine release. (16 Medically evaluated, community evacuated). U.S. Chemical Safety and Hazard Investigation Board. http://www.csb.gov/assets/document/DPC2-Final.pdf. May 27, 2010.

U.S. Census Bureau. 2008. Current industrial reports. Inorganic chemicals. http://www.census.gov/cir/www/325/mq325a.html. March 13, 2009.

U.S. Census Bureau. 2009. Current industrial reports (CIR) U.S. Census Bureau. http://www.census.gov/cir/www/325/mq325a.html. March 31, 2009.

U.S. Chemical Safety and Hazard Investigation Board. 2003. Investigation report, chlorine release. Washington, DC: U.S. Chemical Safety and Hazard Investigation Board. 2002-04-I-MO. PB2003103273.

Van Joost TH, Harris JMW, Stolz E, et al. 1987. Sodium hypochlorite sensitization. Contact Dermatitis 16:114-115.

Van Sickle D, Wenck MA, Belflower A, et al. 2009. Acute health effects after exposure to chlorine gas released after a train derailment. Am J Emerg Med 27(1):1-7.

Vernot EH, MacEwen JD, Haun CC, et al. 1977. Acute toxicity and skin corrosion data for some organic and inorganic compounds and aqueous solutions. Toxicol Appl Pharmacol 42:417-423.

Vetrano KM. 2001. Molecular chlorine: Health and environmental effects. Rev Environ Contam Toxicol 170:75-140.

Viccellio P, Bania T, Brent J, et al. 1998. Chlorine gas. In: Emergency toxicology. 2nd ed. Philadelphia, PA: Lippincott-Raven Press, 444-445.

Vieira I, Sonnier M, Cresteil T. 1996. Developmental expression of CYP2E1 in the human liver: Hypermethylation control of gene expression during the neonatal period. Eur J Biochem 238(2):476-483.

Vohra R, Clark RF. 2006. Chlorine-related inhalation injury from a swimming pool disinfectant in a 9-year-old girl. Pediatr Emerg Care 22(4):254-257.

Wang L, Najafi R, Najafi K, et al. 2007. Hypochloroous acid as a potential wound care agent. J Burns Wound 6:65-79.

Wang TZ, Margerum DW. 1994. Kinetics of reversible chlorine hydrolysis: Temperature dependence and general-acid/base-assisted mechanisms. Inorg Chem 33:1050-1055.

Ward MJ, Routledge PA. 1988. Hypernatraemia and hyperchloraemic acidosis after bleach ingestion. Hum Toxicol 7:37-38.

Weedon FR, Hartzell A, Setterstrom C. 1940. Toxicity of ammonia, chlorine, hydrogen cyanide, hydrogen sulphide, and sulphur dioxide gasses. V. Animals. Contrib Boyce Thompson Inst 11:365-385.

Wegman DH, Eisen EA. 1990. Acute irritants: More than a nuisance. Chest 97(4):773-775.

Weill H, George R, Schwarz M, et al. 1969. Late evaluation of pulmonary function after acute exposure to chlorine gas. Am Rev Respir Dis 99:374-379.

Wenck MA, van Sickle D, Drociuk D, et al. 2007. Rapid assessment of exposure to chlorine released from a train derailment and resulting health impact. Public Health Rep 122(6):784-792.

West JR, Smith HW, Chasis H. 1948. Glomerular filtration rate, effective renal blood flow, and maximal tubular excretory capacity in infancy. J Pediatr 32:10-18.

WHO. 1982. Chlorine and hydrogen chloride. Environmental Health Criteria 21. Geneva, Switzerland: World Health Organization.

WHO. 2000. Air quality guidelines. 2nd ed. Geneva, Switzerland: World Health Organization. http://www.euro.who.int/Document/AIQ/AirQualRepMtg.pdf. March 08, 2006.

WHO. 2004. Guidelines for drinking-water quality. Volume 1. Recommendations. 3rd ed. Geneva, Switzerland: World Health Organization. http://www.who.int/water_sanitation_health/dwq/gdwq3/en/. March 08, 2006.

WHO. 2007. How to measure chlorine residual in water. World Health Organization. Tech Note No. 11. http://www.who.int/entity/water sanitation health/hygiene/envsan/chlorineresid.pdf. May 27, 2010.

Widdowson EM, Dickerson JWT. 1964. Chemical composition of the body. In: Comar CL, Bronner F, eds. Mineral metabolism: An advanced treatise. Volume II: The elements Part A. New York: Academic Press, 1-247.

Winder C. 2001. The toxicology of chlorine. Environ Res 85(2):105-114.

Withers RMJ, Lees FP. 1985b. The assessment of major hazards: The lethal toxicity of chlorine. Part 2. Model of toxicity to man. J Hazard Mater 12:283-302.

Włodkowski TJ, Rosenkranz HS. 1975. Mutagenicity of sodium hypochlorite for *Salmonella typhimurium*. Mutat Res 31:39-42.

Wolf DC, Morgan KT, Gross EA, et al. 1995. Two-year inhalation exposure of female and male B6C3F1 mice and F344 rats to chlorine gas induces lesions confined to the nose. Fundam Appl Toxicol 24(1):111-131.

Wones RG, Deck CC, Stadler B, et al. 1993. Lack of effect of drinking water chlorine on lipid and thyroid metabolism in health humans. Environ Health Perspect 99:375-381.

Yarington CT. 1970. The experimental causticity of sodium hypochlorite in the esophagus. Ann Otol Rhinol Laryngol 79:895-899.

Yildirim C, Kocoglu H, Goksu S, et al. 2004. Long-term pulmonary histopathologic changes in rats following acute experimental exposure to chlorine gas. Inhal Toxicol 16(14):911-915.

Ziegler EE, Edwards BB, Jensen RL, et al. 1978. Absorption and retention of lead by infants. Pediatr Res 12(1):29-34.

Zwart A, Woutersen RA. 1988. Acute inhalation toxicity of chlorine in rats and mice: Time-concentration-mortality relationships and effects on respiration. J Hazard Mater 19(2):195-208.

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10. GLOSSARY

Absorption—The taking up of liquids by solids, or of gases by solids or liquids.

Acute Exposure—Exposure to a chemical for a duration of 14 days or less, as specified in the Toxicological Profiles.

Adsorption—The adhesion in an extremely thin layer of molecules (as of gases, solutes, or liquids) to the surfaces of solid bodies or liquids with which they are in contact.

Adsorption Coefficient (K_{oc})—The ratio of the amount of a chemical adsorbed per unit weight of organic carbon in the soil or sediment to the concentration of the chemical in solution at equilibrium.

Adsorption Ratio (Kd)—The amount of a chemical adsorbed by sediment or soil (i.e., the solid phase) divided by the amount of chemical in the solution phase, which is in equilibrium with the solid phase, at a fixed solid/solution ratio. It is generally expressed in micrograms of chemical sorbed per gram of soil or sediment

Benchmark Dose (BMD)—Usually defined as the lower confidence limit on the dose that produces a specified magnitude of changes in a specified adverse response. For example, a BMD₁₀ would be the dose at the 95% lower confidence limit on a 10% response, and the benchmark response (BMR) would be 10%. The BMD is determined by modeling the dose response curve in the region of the dose response relationship where biologically observable data are feasible.

Benchmark Dose Model—A statistical dose-response model applied to either experimental toxicological or epidemiological data to calculate a BMD.

Bioconcentration Factor (BCF)—The quotient of the concentration of a chemical in aquatic organisms at a specific time or during a discrete time period of exposure divided by the concentration in the surrounding water at the same time or during the same period.

Biomarkers—Broadly defined as indicators signaling events in biologic systems or samples. They have been classified as markers of exposure, markers of effect, and markers of susceptibility.

Cancer Effect Level (CEL)—The lowest dose of chemical in a study, or group of studies, that produces significant increases in the incidence of cancer (or tumors) between the exposed population and its appropriate control.

Carcinogen—A chemical capable of inducing cancer.

Case-Control Study—A type of epidemiological study that examines the relationship between a particular outcome (disease or condition) and a variety of potential causative agents (such as toxic chemicals). In a case-controlled study, a group of people with a specified and well-defined outcome is identified and compared to a similar group of people without outcome.

Case Report—Describes a single individual with a particular disease or exposure. These may suggest some potential topics for scientific research, but are not actual research studies.

Case Series—Describes the experience of a small number of individuals with the same disease or exposure. These may suggest potential topics for scientific research, but are not actual research studies.

Ceiling Value—A concentration of a substance that should not be exceeded, even instantaneously.

Chronic Exposure—Exposure to a chemical for 365 days or more, as specified in the Toxicological Profiles.

Cohort Study—A type of epidemiological study of a specific group or groups of people who have had a common insult (e.g., exposure to an agent suspected of causing disease or a common disease) and are followed forward from exposure to outcome. At least one exposed group is compared to one unexposed group.

Cross-sectional Study—A type of epidemiological study of a group or groups of people that examines the relationship between exposure and outcome to a chemical or to chemicals at one point in time.

Data Needs—Substance-specific informational needs that if met would reduce the uncertainties of human health assessment.

Developmental Toxicity—The occurrence of adverse effects on the developing organism that may result from exposure to a chemical prior to conception (either parent), during prenatal development, or postnatally to the time of sexual maturation. Adverse developmental effects may be detected at any point in the life span of the organism.

Dose-Response Relationship—The quantitative relationship between the amount of exposure to a toxicant and the incidence of the adverse effects.

Embryotoxicity and Fetotoxicity—Any toxic effect on the conceptus as a result of prenatal exposure to a chemical; the distinguishing feature between the two terms is the stage of development during which the insult occurs. The terms, as used here, include malformations and variations, altered growth, and *in utero* death.

Environmental Protection Agency (EPA) Health Advisory—An estimate of acceptable drinking water levels for a chemical substance based on health effects information. A health advisory is not a legally enforceable federal standard, but serves as technical guidance to assist federal, state, and local officials.

Epidemiology—Refers to the investigation of factors that determine the frequency and distribution of disease or other health-related conditions within a defined human population during a specified period.

Genotoxicity—A specific adverse effect on the genome of living cells that, upon the duplication of affected cells, can be expressed as a mutagenic, clastogenic, or carcinogenic event because of specific alteration of the molecular structure of the genome.

Half-life—A measure of rate for the time required to eliminate one half of a quantity of a chemical from the body or environmental media.

Immediately Dangerous to Life or Health (IDLH)—The maximum environmental concentration of a contaminant from which one could escape within 30 minutes without any escape-impairing symptoms or irreversible health effects.

Immunologic Toxicity—The occurrence of adverse effects on the immune system that may result from exposure to environmental agents such as chemicals.

Immunological Effects—Functional changes in the immune response.

Incidence—The ratio of individuals in a population who develop a specified condition to the total number of individuals in that population who could have developed that condition in a specified time period.

Intermediate Exposure—Exposure to a chemical for a duration of 15–364 days, as specified in the Toxicological Profiles.

In Vitro—Isolated from the living organism and artificially maintained, as in a test tube.

In Vivo—Occurring within the living organism.

Lethal Concentration_(LO) (LC_{LO})—The lowest concentration of a chemical in air that has been reported to have caused death in humans or animals.

Lethal Concentration₍₅₀₎ (LC_{50})—A calculated concentration of a chemical in air to which exposure for a specific length of time is expected to cause death in 50% of a defined experimental animal population.

Lethal Dose_(LO) (LD_{Lo})—The lowest dose of a chemical introduced by a route other than inhalation that has been reported to have caused death in humans or animals.

Lethal Dose₍₅₀₎ (LD_{50})—The dose of a chemical that has been calculated to cause death in 50% of a defined experimental animal population.

Lethal Time $_{(50)}$ (LT₅₀)—A calculated period of time within which a specific concentration of a chemical is expected to cause death in 50% of a defined experimental animal population.

Lowest-Observed-Adverse-Effect Level (LOAEL)—The lowest exposure level of chemical in a study, or group of studies, that produces statistically or biologically significant increases in frequency or severity of adverse effects between the exposed population and its appropriate control.

Lymphoreticular Effects—Represent morphological effects involving lymphatic tissues such as the lymph nodes, spleen, and thymus.

Malformations—Permanent structural changes that may adversely affect survival, development, or function.

Minimal Risk Level (MRL)—An estimate of daily human exposure to a hazardous substance that is likely to be without an appreciable risk of adverse noncancer health effects over a specified route and duration of exposure.

Modifying Factor (MF)—A value (greater than zero) that is applied to the derivation of a Minimal Risk Level (MRL) to reflect additional concerns about the database that are not covered by the uncertainty factors. The default value for a MF is 1.

Morbidity—State of being diseased; morbidity rate is the incidence or prevalence of disease in a specific population.

Mortality—Death; mortality rate is a measure of the number of deaths in a population during a specified interval of time.

Mutagen—A substance that causes mutations. A mutation is a change in the DNA sequence of a cell's DNA. Mutations can lead to birth defects, miscarriages, or cancer.

Necropsy—The gross examination of the organs and tissues of a dead body to determine the cause of death or pathological conditions.

Neurotoxicity—The occurrence of adverse effects on the nervous system following exposure to a chemical.

No-Observed-Adverse-Effect Level (NOAEL)—The dose of a chemical at which there were no statistically or biologically significant increases in frequency or severity of adverse effects seen between the exposed population and its appropriate control. Effects may be produced at this dose, but they are not considered to be adverse.

Octanol-Water Partition Coefficient (K_{ow})—The equilibrium ratio of the concentrations of a chemical in n-octanol and water, in dilute solution.

Odds Ratio (OR)—A means of measuring the association between an exposure (such as toxic substances and a disease or condition) that represents the best estimate of relative risk (risk as a ratio of the incidence among subjects exposed to a particular risk factor divided by the incidence among subjects who were not exposed to the risk factor). An OR of greater than 1 is considered to indicate greater risk of disease in the exposed group compared to the unexposed group.

Organophosphate or Organophosphorus Compound—A phosphorus-containing organic compound and especially a pesticide that acts by inhibiting cholinesterase.

Permissible Exposure Limit (PEL)—An Occupational Safety and Health Administration (OSHA) allowable exposure level in workplace air averaged over an 8-hour shift of a 40-hour workweek.

Pesticide—General classification of chemicals specifically developed and produced for use in the control of agricultural and public health pests.

Pharmacokinetics—The dynamic behavior of a material in the body, used to predict the fate (disposition) of an exogenous substance in an organism. Utilizing computational techniques, it provides the means of studying the absorption, distribution, metabolism, and excretion of chemicals by the body.

Pharmacokinetic Model—A set of equations that can be used to describe the time course of a parent chemical or metabolite in an animal system. There are two types of pharmacokinetic models: data-based and physiologically-based. A data-based model divides the animal system into a series of compartments, which, in general, do not represent real, identifiable anatomic regions of the body, whereas the physiologically-based model compartments represent real anatomic regions of the body.

Physiologically Based Pharmacodynamic (PBPD) Model—A type of physiologically based dose-response model that quantitatively describes the relationship between target tissue dose and toxic end points. These models advance the importance of physiologically based models in that they clearly describe the biological effect (response) produced by the system following exposure to an exogenous substance.

Physiologically Based Pharmacokinetic (PBPK) Model—Comprised of a series of compartments representing organs or tissue groups with realistic weights and blood flows. These models require a variety of physiological information: tissue volumes, blood flow rates to tissues, cardiac output, alveolar ventilation rates, and possibly membrane permeabilities. The models also utilize biochemical information, such as air/blood partition coefficients, and metabolic parameters. PBPK models are also called biologically based tissue dosimetry models.

Prevalence—The number of cases of a disease or condition in a population at one point in time.

Prospective Study—A type of cohort study in which the pertinent observations are made on events occurring after the start of the study. A group is followed over time.

 q_1^* —The upper-bound estimate of the low-dose slope of the dose-response curve as determined by the multistage procedure. The q_1^* can be used to calculate an estimate of carcinogenic potency, the incremental excess cancer risk per unit of exposure (usually $\mu g/L$ for water, mg/kg/day for food, and $\mu g/m^3$ for air).

Recommended Exposure Limit (REL)—A National Institute for Occupational Safety and Health (NIOSH) time-weighted average (TWA) concentration for up to a 10-hour workday during a 40-hour workweek.

Reference Concentration (RfC)—An estimate (with uncertainty spanning perhaps an order of magnitude) of a continuous inhalation exposure to the human population (including sensitive subgroups) that is likely to be without an appreciable risk of deleterious noncancer health effects during a lifetime. The inhalation reference concentration is for continuous inhalation exposures and is appropriately expressed in units of mg/m³ or ppm.

Reference Dose (RfD)—An estimate (with uncertainty spanning perhaps an order of magnitude) of the daily exposure of the human population to a potential hazard that is likely to be without risk of deleterious effects during a lifetime. The RfD is operationally derived from the no-observed-adverse-effect level (NOAEL, from animal and human studies) by a consistent application of uncertainty factors that reflect various types of data used to estimate RfDs and an additional modifying factor, which is based on a professional judgment of the entire database on the chemical. The RfDs are not applicable to nonthreshold effects such as cancer.

Reportable Quantity (RQ)—The quantity of a hazardous substance that is considered reportable under the Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA). Reportable quantities are (1) 1 pound or greater or (2) for selected substances, an amount established by regulation either under CERCLA or under Section 311 of the Clean Water Act. Quantities are measured over a 24-hour period.

Reproductive Toxicity—The occurrence of adverse effects on the reproductive system that may result from exposure to a chemical. The toxicity may be directed to the reproductive organs and/or the related endocrine system. The manifestation of such toxicity may be noted as alterations in sexual behavior, fertility, pregnancy outcomes, or modifications in other functions that are dependent on the integrity of this system.

Retrospective Study—A type of cohort study based on a group of persons known to have been exposed at some time in the past. Data are collected from routinely recorded events, up to the time the study is undertaken. Retrospective studies are limited to causal factors that can be ascertained from existing records and/or examining survivors of the cohort.

Risk—The possibility or chance that some adverse effect will result from a given exposure to a chemical.

Risk Factor—An aspect of personal behavior or lifestyle, an environmental exposure, or an inborn or inherited characteristic that is associated with an increased occurrence of disease or other health-related event or condition.

Risk Ratio—The ratio of the risk among persons with specific risk factors compared to the risk among persons without risk factors. A risk ratio greater than 1 indicates greater risk of disease in the exposed group compared to the unexposed group.

Short-Term Exposure Limit (STEL)—The American Conference of Governmental Industrial Hygienists (ACGIH) maximum concentration to which workers can be exposed for up to 15 minutes continually. No more than four excursions are allowed per day, and there must be at least 60 minutes between exposure periods. The daily Threshold Limit Value-Time Weighted Average (TLV-TWA) may not be exceeded.

Standardized Mortality Ratio (SMR)—A ratio of the observed number of deaths and the expected number of deaths in a specific standard population.

Target Organ Toxicity—This term covers a broad range of adverse effects on target organs or physiological systems (e.g., renal, cardiovascular) extending from those arising through a single limited exposure to those assumed over a lifetime of exposure to a chemical.

Teratogen—A chemical that causes structural defects that affect the development of an organism.

Threshold Limit Value (TLV)—An American Conference of Governmental Industrial Hygienists (ACGIH) concentration of a substance to which most workers can be exposed without adverse effect. The TLV may be expressed as a Time Weighted Average (TWA), as a Short-Term Exposure Limit (STEL), or as a ceiling limit (CL).

Time-Weighted Average (TWA)—An allowable exposure concentration averaged over a normal 8-hour workday or 40-hour workweek.

Toxic Dose₍₅₀₎ (**TD**₅₀)—A calculated dose of a chemical, introduced by a route other than inhalation, which is expected to cause a specific toxic effect in 50% of a defined experimental animal population.

Toxicokinetic—The absorption, distribution, and elimination of toxic compounds in the living organism.

Uncertainty Factor (UF)—A factor used in operationally deriving the Minimal Risk Level (MRL) or Reference Dose (RfD) or Reference Concentration (RfC) from experimental data. UFs are intended to account for (1) the variation in sensitivity among the members of the human population, (2) the uncertainty in extrapolating animal data to the case of human, (3) the uncertainty in extrapolating from data obtained in a study that is of less than lifetime exposure, and (4) the uncertainty in using lowest-observed-adverse-effect level (LOAEL) data rather than no-observed-adverse-effect level (NOAEL) data. A default for each individual UF is 10; if complete certainty in data exists, a value of 1 can be used; however, a reduced UF of 3 may be used on a case-by-case basis, 3 being the approximate logarithmic average of 10 and 1.

Xenobiotic—Any chemical that is foreign to the biological system.

CHLORINE A-1

APPENDIX A. ATSDR MINIMAL RISK LEVELS AND WORKSHEETS

The Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) [42 U.S.C. 9601 et seq.], as amended by the Superfund Amendments and Reauthorization Act (SARA) [Pub. L. 99–499], requires that the Agency for Toxic Substances and Disease Registry (ATSDR) develop jointly with the U.S. Environmental Protection Agency (EPA), in order of priority, a list of hazardous substances most commonly found at facilities on the CERCLA National Priorities List (NPL); prepare toxicological profiles for each substance included on the priority list of hazardous substances; and assure the initiation of a research program to fill identified data needs associated with the substances.

The toxicological profiles include an examination, summary, and interpretation of available toxicological information and epidemiologic evaluations of a hazardous substance. During the development of toxicological profiles, Minimal Risk Levels (MRLs) are derived when reliable and sufficient data exist to identify the target organ(s) of effect or the most sensitive health effect(s) for a specific duration for a given route of exposure. An MRL is an estimate of the daily human exposure to a hazardous substance that is likely to be without appreciable risk of adverse noncancer health effects over a specified duration of exposure. MRLs are based on noncancer health effects only and are not based on a consideration of cancer effects. These substance-specific estimates, which are intended to serve as screening levels, are used by ATSDR health assessors to identify contaminants and potential health effects that may be of concern at hazardous waste sites. It is important to note that MRLs are not intended to define clean-up or action levels.

MRLs are derived for hazardous substances using the no-observed-adverse-effect level/uncertainty factor approach. They are below levels that might cause adverse health effects in the people most sensitive to such chemical-induced effects. MRLs are derived for acute (1–14 days), intermediate (15–364 days), and chronic (365 days and longer) durations and for the oral and inhalation routes of exposure. Currently, MRLs for the dermal route of exposure are not derived because ATSDR has not yet identified a method suitable for this route of exposure. MRLs are generally based on the most sensitive chemical-induced end point considered to be of relevance to humans. Serious health effects (such as irreparable damage to the liver or kidneys, or birth defects) are not used as a basis for establishing MRLs. Exposure to a level above the MRL does not mean that adverse health effects will occur.

MRLs are intended only to serve as a screening tool to help public health professionals decide where to look more closely. They may also be viewed as a mechanism to identify those hazardous waste sites that

are not expected to cause adverse health effects. Most MRLs contain a degree of uncertainty because of the lack of precise toxicological information on the people who might be most sensitive (e.g., infants, elderly, nutritionally or immunologically compromised) to the effects of hazardous substances. ATSDR uses a conservative (i.e., protective) approach to address this uncertainty consistent with the public health principle of prevention. Although human data are preferred, MRLs often must be based on animal studies because relevant human studies are lacking. In the absence of evidence to the contrary, ATSDR assumes that humans are more sensitive to the effects of hazardous substance than animals and that certain persons may be particularly sensitive. Thus, the resulting MRL may be as much as 100-fold below levels that have been shown to be nontoxic in laboratory animals.

Proposed MRLs undergo a rigorous review process: Health Effects/MRL Workgroup reviews within the Division of Toxicology and Environmental Medicine, expert panel peer reviews, and agency-wide MRL Workgroup reviews, with participation from other federal agencies and comments from the public. They are subject to change as new information becomes available concomitant with updating the toxicological profiles. Thus, MRLs in the most recent toxicological profiles supersede previously published levels. For additional information regarding MRLs, please contact the Division of Toxicology and Environmental Medicine, Agency for Toxic Substances and Disease Registry, 1600 Clifton Road NE, Mailstop F-62, Atlanta, Georgia 30333.

MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical Name: Chlorine gas CAS Numbers: 7782-50-5 Date: June 2010

Profile Status: Final Draft Post-Public Comment

Route: [X] Inhalation [] Oral

Duration: [X] Acute [] Intermediate [] Chronic

Graph Key: 7, 9, 10, 11 Species: Human

Minimal Risk Level: 0.06 [] mg/kg/day [X] ppm

<u>References</u>: Anglen DA. 1981. Sensory response of human subjects to chlorine in air. Ann Arbor, MI: University of Michigan.

D'Alessandro A, Kuschner W, Wong H, et al. 1996. Exaggerated responses to chlorine inhalation among persons with nonspecific airway hyperreactivity. Chest 109:331-337.

Rotman HH, Fliegelman MJ, Moore T, et al. 1983. Effects of low concentrations of chlorine on pulmonary function in humans. J Appl Physiol 54:1120-1124.

Schins RPF, Emmen H, Hoogendijk L, et al. 2000. Nasal inflammatory and respiratory parameters in human volunteers during and after repeated exposure to chlorine. Eur Respir J 16:626-632.

Shusterman D, Murphy, MA, Balmes J. 1998. Subjects with seasonal allergic rhinitis and nonrhinitic subjects react differentially to nasal provocation with chlorine gas. J Allergy Clin Immunol 101:732-740.

Shusterman D, Murphy, MA, Balmes J. 2003b. Influence of age, gender, and allergy status on nasal reactivity to inhaled chlorine. Inhal Toxicol 15:1179-1189.

Experimental design and effects noted in each study: Anglen (1981) exposed up to 29 male and female volunteers to 0, 0.5, 1, or 2 ppm chlorine for either 4 or 8 hours. Sensations were recorded before and during exposure, and pulmonary function was monitored by measuring FVC and FEV₁ before and at various times during exposure. Itching and burning of the throat were the highest responses and were most prevalent by the end of an 8-hour exposure to 1 ppm chlorine. Responses for sensations of itching or burning of the nose and eyes were also prevalent at 1 ppm chlorine. In general, males provided stronger irritation responses than females. Exposure to 1 or 2 ppm chlorine for 8 hours produced significant changes in pulmonary function, but similar exposures to 0.5 ppm did not. Exposure to 2 ppm for up to 30 minutes produced no increase in subjective irritation and exposure to 2 ppm for 2 hours did not alter pulmonary function.

Rotman et al. (1983) studied eight healthy male volunteers exposed to target concentrations of 0, 0.5, or 1 ppm chlorine (Rotman et al. 1983). Pulmonary tests were conducted before exposure, after a 4- and 8-hour exposure period and again 2 and 24 hours after exposure ceased. During exposure, the subjects exercised on a treadmill for 15 minutes of each hour to simulate light-to-moderate work that raised the heart rate to 100 beats per minute. Specific respiratory parameters measured included FVC, FEV₁, FEV₁% forced expired volume in 1 second as %FVC (FEV1%), peak expiratory flow rate (PEFR), FEF₅₀ and FEF₂₅, TLC, expiratory reserve volume (ERV), functional residual capacity (FRC), residual volume, airway resistance (Raw), single-breath DL_{CO}, closing volume, and difference in nitrogen concentrations between 750 and 1,250 mL of inhaled vital capacity (Δ N₂). Exposure to 1 ppm chlorine caused runny

nose and mild burning in the throat, but no such effects were reported at 0.5 ppm. Significant changes in pulmonary function tests were mostly restricted to the 1 ppm exposure level and were evident after 4 hours of exposure. Changes were observed in FEV₁, PEFR, FEF₅₀, FEF₂₅, TLC, Raw, and ΔN_2 . Greater changes in some of these parameters were seen after 8 hours of exposure. Few changes were still evident 24 hours after exposure, but most parameters had returned to pre-exposure values by that time. It should be noted that one volunteer who was atopic experienced severe distress during exposure to 1 ppm and was forced to exit the chamber before the full 8-hour period due to shortness of breath and wheezing.

D'Alessandro et al. (1996) evaluated pulmonary function in subjects with (n=10) and without (n=5) airway hyperreactivity (HR, defined by baseline methacholine hyperresponsiveness). The HR subjects were exposed to 0.4 or 1.0 ppm chlorine, whereas the healthy subjects were exposed to 1.0 ppm chlorine. All exposures lasted 60 minutes. Airflow and airway resistance were measured immediately before and immediately after exposure. Also, lung volumes, airflow, diffusing capacity, airway resistance, and responsiveness to methacholine were measured 24 hours before and 24 hours after exposure. Exposure of the HR group to 0.4 ppm chlorine resulted in no significant change in airflow or resistance either immediately or 24 hours after exposure. Exposure to 1.0 ppm chlorine resulted in an immediate decrease in FEV₁ and FEF_{25-75%} and increase in airway resistance among normal and HR subjects, but the magnitude of the effects among HR subjects was significantly greater than in healthy subjects. Twenty-four hours after exposure, there were no significant changes for healthy or HR subjects in airflow, lung volumes, diffusing capacity, resistance, or methacholine responsiveness. Comparing relative changes from baseline immediately after exposure between normal and HR subjects showed that HR subjects had much greater changes in pulmonary function tests.

Schins et al. (2000) studied eight volunteers exposed to chlorine 6 hours/day on 3 consecutive days to each of the four exposure conditions, 0, 0.1, 0.3, and 0.5 ppm chlorine (Schins et al. 2000). Pulmonary function including effort-dependent parameters and effort-independent parameters were evaluated before and after exposures. In addition, nasal lavage measurements were performed before and after each exposure and 1 and 4 days after each exposure. The nasal lavage fluid was examined for total cells, epithelial cells, neutrophils, lymphocytes, eosinophils, monocytes, albumin (an indicator of epithelial permeability), and interleukin-8 (indicator of inflammatory response). Subjective complaints by the subjects were judged to be not treatment-related. Examination of the nasal lavages gave no indication of an inflammatory response or irritant effects on the nasal epithelium. The results of the pulmonary function tests showed that the only significant effect related to chlorine exposure was a difference in maximal mid expiratory flow (MMEF) between 0 and 0.5 ppm exposure; however, this was attributed to an unexplained shift in baseline values during control exposure (0 ppm).

Shusterman et al. (2003b) measured nasal airway resistance in 52 healthy adults (24 males and 28 females) before and after exposure to 0 or 1 ppm chlorine for 15 minutes. Subjects were stratified on age (18–34, 35–51, 52–69 years), gender, and allergic rhinitis status (27 were positive). Nasal airway resistance was measured by active posterior rhinomanometry. Exposures to air and chlorine were a week apart. Subjects with allergic rhinitis showed a significantly greater increase in nasal airway resistance (49% increase from baseline) than healthy subjects (10% increase from baseline) 15 minutes after exposure. The increase in nasal airway resistance was most pronounced in older subjects and least pronounced in the youngest group. No significant differences were seen between males and females. In an earlier study, the same group of investigators had reported that subjects with SAR (n=8) exposed to 0.5 ppm chlorine for 15 minutes experienced a much greater increase in nasal airway resistance than subjects without SAR (n=8), as measured by active posterior rhinomanometry (Shusterman et al. 1998). However, when subjective responses to odor, nasal irritation, and nasal congestion were analyzed separately by rhinitis status, no significant exposure-related changes were observed for rhinorrhea, postnasal drip, or headache either on a pool or stratified basis. In addition, within either the SAR or non-SAR group, there was no relationship between subjective and objective congestion after chlorine

exposure. Pulmonary peak flow tests showed that none of the subjects exhibited clinically significant changes in peak flow, nor did they complain of cough, wheezing, or chest tightness on chlorine exposure days. The increased nasal airway resistance instrumentally detected in subjects with SAR is not considered an adverse effect since the subjects did not perceive it as such.

Collectively, this group of studies provides evidence of sensory irritation and transient pulmonary changes occurring in humans exposed to 1 ppm chlorine for up to 8 hours/day. The pulmonary changes indicated increased airway resistance and reduced air flow. No such changes were reported in volunteers exposed to 0.5 ppm chlorine

<u>Dose and end point used for MRL derivation</u>: 0.5 ppm is a NOAEL for sensory irritation and pulmonary function

The MRL is derived by adjusting for continuous exposure based on the fact that Rotman et al. (1983) reported that exposure to 1 ppm for 8 hours induced greater changes in pulmonary function tests than exposure to the same concentration for 4 hours, suggesting that the response was related to some function of concentration and duration rather than to concentration alone.

MRL = 0.5 ppm (8 hours/24 hours) = 0.167 ppm

8 hours was the longest period of exposure for which there is information.

[X] NOAEL [] LOAEL

Uncertainty Factors used in MRL derivation:

[] 10 for use of a LOAEL[] 10 for extrapolation from animals to humans[X] 3 for human variability

Although sensitive individuals were tested in some of these studies, the number of individuals tested at the region of the NOAEL (0.4–0.5 ppm) was small. Therefore, an uncertainty factor of 3 is used to account for sensitive populations.

Was a conversion factor used from ppm in food or water to a mg/body weight dose? Not applicable.

If an inhalation study in animals, list conversion factors used in determining human equivalent dose: Not applicable.

Was a conversion used from intermittent to continuous exposure? Yes, see above.

Other additional studies or pertinent information that lend support to this MRL: Results from the five studies summarized above are supportive of each other.

Agency Contacts (Chemical Managers): G. Daniel Todd

MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical Name: Chlorine gas CAS Numbers: 7782-50-5 Date: June 2010

Profile Status: Final Draft Post-Public Comment

Route: [X] Inhalation [] Oral

Duration: [] Acute [X] Intermediate [] Chronic

Graph Key: 30 Species: Rat

Minimal Risk Level: 0.002 [] mg/kg/day [X] ppm

<u>Reference</u>: Kutzman RS. 1983. A study of Fisher-344 rats subchronically exposed to 0, 0.5, 1.5, or 5.0 ppm chlorine. Upton, NY: Brookhaven National Laboratory. BNL 32710.

Experimental design: Groups of F344 rats (24 males, 10 females) were exposed to 0, 0.5, 1.5, or 5 ppm chlorine 6 hours/day, 5 days/week for 62 days (Kutzman 1983). Pulmonary function tests (plethysmograph-based assessment of multiple end points, including lung and tidal volumes, breathing frequency, transpulmonary pressure, lung compliance, N₂ washout, diffusing capacity for CO₂, maximum expiratory flow volume, peak expiratory flow, and airway resistance) were conducted in 21–24 anesthetized males 6 hours after the last exposure. Respiratory tissues from these rats were prepared for histopathology. The lung from some of these rats was also examined for collagen, elastin, total protein, and DNA. Histopathology of selected organs (nasal turbinates, lungs, peribronchial lymph node, brain, kidney, liver, spleen, testes, and heart) was evaluated in eight males per group. Also, 8 males were mated with untreated females and 10 exposed females were mated with untreated males for reproductive studies on females sacrificed on GD 19. In addition, 1 day after the last exposure, samples of blood and bone marrow from 10 males per group were prepared for analysis of chromosomal aberrations and sister chromatid exchanges.

Effects noted in study and corresponding doses: Exposure to 5 ppm cause severe eye and upper respiratory irritation, whereas rats exposed to 1.5 ppm showed occasionally less severe signs of irritation, and exposure to 0.5 ppm caused no obvious signs of irritation or discomfort. Female rats exposed to 5 ppm lost weight. Final weight was approximately 32% lower than controls; at 1.5 and 0.5 ppm, final weight was approximately 15 and 11% lower than in control rats, respectively. In males exposed to 5 ppm, final weight was approximately 15% lower than controls. No information was provided regarding food and water intake. Changes in organ weights were unremarkable. The tests of pulmonary function did not reveal marked abnormalities. The most significant effect was a reduction in airflow at 25% vital capacity in all exposed groups, indicating some degree of small airway involvement. An electrocardiogram did not reveal any significant cardiac alterations due to chlorine exposure. The lung biochemistry only showed an increased collagen concentration at 1.5 and 5 ppm. The cytogenetic studies showed no increased incidence of sister chromatid exchange or cellular proliferation in bone marrow and no increase in sister chromatid exchanges or chromosomal aberrations in peripheral lymphocytes. Analysis of sperm morphology was unremarkable. Results of the reproductive studies showed no effects on fertility, number of corpora lutea, viable embryos, early or late deaths, or preimplantation loses. There were no significant exposure-related increases in the incidences of animals with histological lesions in any of the examined tissues with the exception of a loss of cilia in the trachea. The incidences of slight to moderate loss of tracheal cilia were 1/23, 12/23, 4/23, and 13/23 in the 0, 0.5, 1.5, and 5 ppm exposure groups, respectively. Although the incidence for this lesion in the mid-exposure group was not significantly different from the control incidence, a statistically significant (p=0.0055) Cochran-Armitage trend test for these data can be demonstrated. However, when attempts were made to apply dose-response models to the data, no adequate fits of EPA Benchmark Dose Software models to the data were obtained (p-values for chi-square goodness of fit statistics were <0.1).

<u>Dose and end point used for MRL derivation</u>: 0.5 ppm is a minimal LOAEL for tracheal lesions.

Uncertainty Factors used in MRL derivation:

- [X] 3 for use of a minimal LOAEL
- [X] 3 for extrapolation from animals to humans with dosimetric adjustment
- [X] 10 for human variability

Was a conversion factor used from ppm in food or water to a mg/body weight dose? Not applicable.

<u>If an inhalation study in animals, list conversion factors used in determining human equivalent dose</u>: The intermediate-duration inhalation MRL was calculated using EPA's methodology (EPA 1994a) for a category 1 gas.

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\begin{aligned} &LOAEL_{[HEC]} = \ LOAEL_{[ADJ]} \ x \ RGDR_{TB} \\ &where: \\ &LOAEL_{[ADJ]} = \ 0.5 \ ppm \ x \ 6/24 \ hours \ x \ 5/7 \ days = 0.09 \ ppm \ and \\ &RGDR_{TB} = ratio \ of \ the \ regional \ gas \ dose \ in \ rats \ to \ that \ of \ humans \ for \ the \ tracheobronchial \ region \\ &RGDR_{TB} = \ (VE/SA_{TB})_A \ / \ (VE/SA_{TB})_H \\ &where: \\ &VE = minute \ volume \ (0.137 \ L/minute \ for \ rats, \ 13.8 \ L/minute \ for \ humans \ [EPA \ 1994a]) \ and \\ &SA_{TB} = surface \ area \ of \ the \ tracheobronchial \ region \ (22.5 \ cm^2 \ for \ rats \ and \ 3,200 \ cm^2 \ for \ humans \ [EPA \ 1994a]) \end{aligned}
```

 $LOAEL_{\rm [HEC]} = 0.09~ppm~x~(0.137~L/minute/22.5~cm^2)~/~(13.8~L/minute/3,200~cm^2) = ~0.14~ppm$

Was a conversion used from intermittent to continuous exposure? Yes, see above.

Other additional studies or pertinent information that lend support to this MRL: In a similar study, Barrow et al. (1979) evaluated the respiratory response in F344 rats exposed to 0, 1, 3, or 9 ppm chlorine 6 hours/day, 5 days/week for 6 weeks. Nasal discharge was seen occasionally in rats exposed to 1 ppm, but was common in rats exposed to 3 and 9 ppm. Respiratory difficulty was also apparent in some rats exposed to 9 ppm. At termination, gross necropsy revealed accumulation of inflammatory reactions in the upper nasal passages in rats exposed to 3 and 9 ppm chlorine. Microscopic evaluations showed indications of inflammatory reactions in the upper and lower respiratory tract of high-dose males and females. The nasal turbinates showed mucopurulent inflammation with secretory material and erosions of the mucosal epithelium. Changes in the trachea and bronchi consisted mostly of hyperplasia of the epithelial lining and inflammatory reactions. The alveolar sacs contained macrophages and secretory material and epithelial cells showed necrosis, hypertrophy, and hyperplasia. Alterations in rats exposed to 1 and 3 ppm were less extensive and were limited to focal mucopurulent inflammation of the nasal turbinates in females. Males exposed to 1 or 3 ppm showed deeper pulmonary changes consisting of slight to moderate inflammatory reaction around the respiratory bronchioles and alveolar ducts, increased alveolar macrophages, and isolated areas of atelectasis (incomplete expansion). A LOAEL of 1 ppm for respiratory effects can be defined in this study based on the presence of inflammatory changes in the nasal

turbinates of females and in the lungs of males; no NOAEL was established. Incidences of animals with respiratory lesions were not presented in this study.

Agency Contacts (Chemical Managers): G. Daniel Todd

MINIMAL RISK LEVEL (MRL) WORKSHEET

Chemical Name: Chlorine gas CAS Numbers: 7782-50-5 Date: June 2010

Profile Status: Final Draft Post-Public Comment

Route: [X] Inhalation [] Oral

Duration: [] Acute [] Intermediate [X] Chronic

Graph Key: 34 Species: Monkey

Minimal Risk Level: 0.00005 [] mg/kg/day [X] ppm

<u>Reference</u>: Klonne DR, Ulrich CE, Riley MG, et al. 1987. One-year inhalation toxicity study of chlorine in Rhesus monkeys (*Macaca mulatta*). Fundam Appl Toxicol 9:557-572.

Experimental design: Male and female Rhesus monkeys (4/sex/exposure level) were exposed to 0, 0.1, 0.5, or 2.3 ppm chlorine 6 hours/day, 5 days/week for 1 year (Klonne et al. 1987). Pulmonary diffusing capacity of CO and distribution of ventilation, body weights, urinalysis, EKG, hematology, and clinical chemistry were evaluated monthly during the study. At termination, the heart, lungs plus trachea, liver, gonads, kidneys, spleen, and brain were weighed. Histological evaluations were done on all major tissues and organs. The nasal tissues (at the first palatine ridge and just posterior to the third, fifth, and seventh palatine ridges), trachea, and lungs were also examined.

Effects noted in study and corresponding doses: Exposure to chlorine did not significantly affect body weight, hematology and clinical chemistry parameters, urinalysis, or the EKG. At approximately week 6 of exposure, monkeys in the 2.3 ppm group showed overt signs of ocular irritation (tearing, reddened eyes, rubbing the eyes) during the daily exposures; no signs of irritation were seen in the other exposure groups. Examination of the eyes at termination showed irritation of the conjunctiva at 2.3 ppm, but no evidence of gross changes; the corneas were not affected. During the study, there was a statistically significant trend in each group, including controls, for increasing pulmonary diffusing capacity and distribution of ventilation. However, there was no evidence of treatment-related effects at any interval during the study. The only treatment-related histopathological effects consisted of focal epithelial hyperplasia characterized by increased cell numbers and loss of cilia and goblet cells in the respiratory epithelium of the nose and trachea. The affected areas of the nasal passages showed hypercellularity with loss of goblet cells and cilia. In some of these areas, the nuclei showed altered polarity. Lesions were more frequent on the angular margins of the turbinates and less frequent on the lateral wall or septum adjacent to these margins. In some cases, the respiratory epithelial hyperplasia was associated with mild suppurative inflammatory response. Lesions in the trachea resembled those in the nose, but were less severe and involved only a small circumferential section of the ventral and ventrolateral trachea. The combined incidences of hyperplasia in the nasal epithelium with loss of goblet cells and cilia, characterized as trace and mild in males and females, were 1/8, 3/8, 6/8, and 8/8 in the control, 0.1, 0.5, and 2.3 ppm exposure groups, respectively. The exposure concentration of 0.1 ppm is considered a LOAEL for nasal lesions in monkeys.

Incidence data for nasal lesions in male and female monkeys exposed to chlorine gas (Klonne et al. 1987) were analyzed using the BMD approach for MRL derivation. Models in the EPA BMDS (version 1.4.1) (i.e., Gamma, Logistic, Log-logistic, Multi-stage, Probit, Log-probit, Quantal linear, Weibull) were fit to the nasal lesion data to determine potential points of departure for the MRL. A Quantal linear model provided the best fit to the data. The predicted exposure concentration associated with a 10% extra risk

(BMC $_{10}$) for nasal lesions in monkeys was 0.04 ppm; the lower 95% confidence limit on this concentration (BMCL $_{10}$) was 0.02 ppm.

Dose and end point used for MRL derivation: BMCL₁₀ of 0.02 ppm for nasal lesions in monkeys.

[] NOAEL [] LOAEL [X] BMCL₁₀

Uncertainty Factors used in MRL derivation:

[] 10 for use of a LOAEL

[X] 3 for extrapolation from animals to humans with dosimetric adjustment

[X] 10 for human variability

Was a conversion factor used from ppm in food or water to a mg/body weight dose? Not applicable.

<u>If an inhalation study in animals, list conversion factors used in determining human equivalent dose</u>: The intermediate-duration inhalation MRL was calculated using EPA's methodology (EPA 1994a) for a category 1 gas.

$$BMCL_{10[HEC]} = BMCL_{10[ADJ]} \times RGDR_{ET}$$

where:

BMCL_{10[ADJ]} = 0.02 ppm x 6/24 hours x 5/7 days = 0.004 ppm and RGDR_{ET} = ratio of the regional gas dose in rats to that of humans for the extrathoracic region

$$RGDR_{ET} = (VE/SA_{ET})_A / (VE/SA_{ET})_H$$

where:

VE = minute volume 2.1 m³/day for monkeys, calculated using the allometric equation for monkeys in EPA (1988) assuming a body weight of 7 kg for Rhesus monkeys with nasal cavity surface area of 62 cm² (Gross and Morgan 1991); 20 m³/day for humans (EPA 1994a) and

 $SA_{ET} = 62 \text{ cm}^2$ surface area of the nasal cavity in Rhesus monkeys weighing 7 kg (Gross and Morgan 1991); 200 cm² for humans (EPA 1994a)

$$RGDR_{ET} = (2.1 \text{ m}^3/\text{day} / 62 \text{ cm}^2) / (20 \text{ m}^3/\text{day} / 200 \text{ cm}^2) = 0.34$$

$$BMCL_{10[HEC]} = 0.004 \text{ ppm x } 0.34 = 0.00136 \text{ ppm}$$

Was a conversion used from intermittent to continuous exposure? Yes, see above.

Other additional studies or pertinent information that lend support to this MRL: Wolf et al. (1995) exposed groups of F344 rats and B6C3F₁ mice (approximately 70/sex/exposure level) to 0, 0.4, 1, or 2.5 ppm chlorine gas for 2 years. Males from both species and female mice were exposed 6 hours/day, 5 days/week, whereas female rats were exposed 6 hours/day, 3 days/week. The reduced exposure of female rats was based on unpublished data from the investigators that showed female rats to have a greater sensitivity to repeated long-term exposure to chlorine. End points evaluated included gross and microscopic examination of the respiratory tract; the nasal passages were examined microscopically at

five different levels. Both in rats and mice, there were no gross lesions attributable to exposure to chlorine, and microscopic evaluation of the respiratory tract showed that chlorine-related effects were restricted to the nasal passages. In the study, the incidences were presented as percentages of all animals for which the nasal passages were adequate for microscopic examination, but the number of animals examined was not provided. No lesions were seen in the larynx, trachea, bronchi, or bronchioles. In general, rats and mice exhibited similar types of lesions. For the most part, the nasal lesions were sitespecific, but the severity and/or incidence were not always concentration-dependent. The majority of the nasal responses exhibited a rostral-to-caudal severity gradient. The lesions rarely extended to the nasopharyngeal meatus. Lesions observed included respiratory and olfactory epithelial degeneration, septal fenestration, mucosal inflammation, respiratory epithelial hyperplasia, squamous metaplasia, and goblet cell (only rats) hypertrophy and hyperplasia, and secretory metaplasia of the transitional epithelium of the lateral meatus. Also observed was intracellular accumulation of eosinophilic proteinaceous material involving the respiratory, transitional, and olfactory epithelia. Lesions were also observed in controls, but the incidences were significantly lower than in the treated groups. One of the lesions with the lowest incidence in controls was Goblet cell hyperplasia in female rats (4%); the respective incidences in the 0.4, 1, and 2.5 ppm group were 71, 90, and 91%. In mice, olfactory epithelium atrophy exhibited one of the lowest incidences in controls (3%); the respective incidences in the 0.4, 1, and 2.5 ppm group were 20, 21, and 39%. In both cases, severity also was concentration-related. Based on the increased incidence of various types of lesions in the nasal passages, the exposure level of 0.4 ppm constitutes a LOAEL for respiratory effects in rats and mice; a NOAEL was not defined.

Agency Contacts (Chemical Managers): G. Daniel Todd

BENCHMARK MODELING OF NASAL LESIONS IN MONKEYS

Incidence data for nasal lesions in male and female monkeys exposed to chlorine gas (Klonne et al. 1987) were analyzed using the BMD approach for MRL derivation (Table A-1). Models in the EPA BMDS (version 1.4.1) (i.e., Gamma, Logistic, Log-logistic, Multi-stage, Probit, Log-probit, Quantal linear, Weibull) were fit to the nasal lesion data to determine potential points of departure for the MRL. A Quantal linear model provided the best fit to the data (Table A-2).

Table A-1. Incidence of Nasal Lesions Observed in Monkeys Exposed to Chlorine for 1 Year

Dose (ppm) Total number of monkeys		Number of monkeys with lesions	
0	8	1	
0.1	8	3	
0.5	8	6	
2.3	8	8	

Source: Klonne et al. 1987

Table A-2. Modeling Predictions for the Incidence of Nasal Lesions Observed in Monkeys Exposed to Chlorine for 1 Year

Model	BMC ₁₀ (ppm)	BMCL ₁₀ (ppm)	x² p-value	AIC
Gamma ^a	0.04	0.02	0.95	29.72
Logistic	0.09	0.05	0.76	30.17
Log-logistic ^b	0.05	0.009	0.53	32.21
Multi-stage ^c	0.04	0.02	0.95	29.72
Probit	0.08	0.05	0.77	30.14
Log-probit ^b	0.06	0.03	0.81	30.09
Quantal linear	0.04	0.02	0.95	29.72
Weibull ^a	0.04	0.02	0.95	29.72

^aRestrict power ≥1.

Source: Klonne et al. 1987

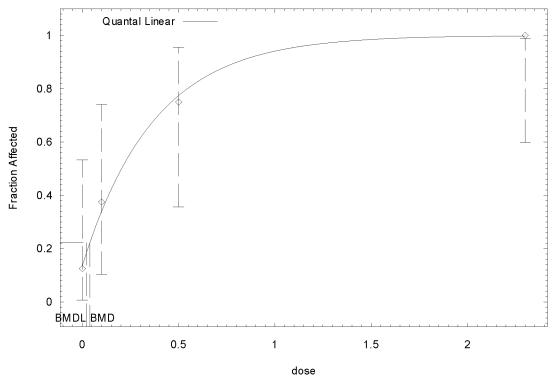
^bSlope restricted to >1.

[°]Restrict betas ≥0; lowest degree polynomial with an adequate fit is reported; degree of polynomial = 1.

From this model, the predicted exposure concentration associated with a 10% extra risk (BMC₁₀) for nasal lesions in monkeys was 0.04 ppm; the lower 95% confidence limit on this concentration (BMCL₁₀) was 0.02 ppm (Figure A-1).

Figure A-1. Predicted and Observed incidence of Nasal Mucosal Lesions in Monkeys Exposed to Chlorine for 1 Year

Quantal Linear Model with 0.95 Confidence Level



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The form of the probability function is:

P[response] = background + (1-background)*[1-EXP(-slope*dose)]

Background = 0.135767

Slope = 2.67521

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CHLORINE B-1

APPENDIX B. USER'S GUIDE

Chapter 1

Public Health Statement

This chapter of the profile is a health effects summary written in non-technical language. Its intended audience is the general public, especially people living in the vicinity of a hazardous waste site or chemical release. If the Public Health Statement were removed from the rest of the document, it would still communicate to the lay public essential information about the chemical.

The major headings in the Public Health Statement are useful to find specific topics of concern. The topics are written in a question and answer format. The answer to each question includes a sentence that will direct the reader to chapters in the profile that will provide more information on the given topic.

Chapter 2

Relevance to Public Health

This chapter provides a health effects summary based on evaluations of existing toxicologic, epidemiologic, and toxicokinetic information. This summary is designed to present interpretive, weight-of-evidence discussions for human health end points by addressing the following questions:

- 1. What effects are known to occur in humans?
- 2. What effects observed in animals are likely to be of concern to humans?
- 3. What exposure conditions are likely to be of concern to humans, especially around hazardous waste sites?

The chapter covers end points in the same order that they appear within the Discussion of Health Effects by Route of Exposure section, by route (inhalation, oral, and dermal) and within route by effect. Human data are presented first, then animal data. Both are organized by duration (acute, intermediate, chronic). *In vitro* data and data from parenteral routes (intramuscular, intravenous, subcutaneous, etc.) are also considered in this chapter.

The carcinogenic potential of the profiled substance is qualitatively evaluated, when appropriate, using existing toxicokinetic, genotoxic, and carcinogenic data. ATSDR does not currently assess cancer potency or perform cancer risk assessments. Minimal Risk Levels (MRLs) for noncancer end points (if derived) and the end points from which they were derived are indicated and discussed.

Limitations to existing scientific literature that prevent a satisfactory evaluation of the relevance to public health are identified in the Chapter 3 Data Needs section.

Interpretation of Minimal Risk Levels

Where sufficient toxicologic information is available, ATSDR has derived MRLs for inhalation and oral routes of entry at each duration of exposure (acute, intermediate, and chronic). These MRLs are not meant to support regulatory action, but to acquaint health professionals with exposure levels at which adverse health effects are not expected to occur in humans.

MRLs should help physicians and public health officials determine the safety of a community living near a chemical emission, given the concentration of a contaminant in air or the estimated daily dose in water. MRLs are based largely on toxicological studies in animals and on reports of human occupational exposure.

MRL users should be familiar with the toxicologic information on which the number is based. Chapter 2, "Relevance to Public Health," contains basic information known about the substance. Other sections such as Chapter 3 Section 3.9, "Interactions with Other Substances," and Section 3.10, "Populations that are Unusually Susceptible" provide important supplemental information.

MRL users should also understand the MRL derivation methodology. MRLs are derived using a modified version of the risk assessment methodology that the Environmental Protection Agency (EPA) provides (Barnes and Dourson 1988) to determine reference doses (RfDs) for lifetime exposure.

To derive an MRL, ATSDR generally selects the most sensitive end point which, in its best judgement, represents the most sensitive human health effect for a given exposure route and duration. ATSDR cannot make this judgement or derive an MRL unless information (quantitative or qualitative) is available for all potential systemic, neurological, and developmental effects. If this information and reliable quantitative data on the chosen end point are available, ATSDR derives an MRL using the most sensitive species (when information from multiple species is available) with the highest no-observed-adverse-effect level (NOAEL) that does not exceed any adverse effect levels. When a NOAEL is not available, a lowest-observed-adverse-effect level (LOAEL) can be used to derive an MRL, and an uncertainty factor (UF) of 10 must be employed. Additional uncertainty factors of 10 must be used both for human variability to protect sensitive subpopulations (people who are most susceptible to the health effects caused by the substance) and for interspecies variability (extrapolation from animals to humans). In deriving an MRL, these individual uncertainty factors are multiplied together. The product is then divided into the inhalation concentration or oral dosage selected from the study. Uncertainty factors used in developing a substance-specific MRL are provided in the footnotes of the levels of significant exposure (LSE) tables.

Chapter 3

Health Effects

Tables and Figures for Levels of Significant Exposure (LSE)

Tables and figures are used to summarize health effects and illustrate graphically levels of exposure associated with those effects. These levels cover health effects observed at increasing dose concentrations and durations, differences in response by species, MRLs to humans for noncancer end points, and EPA's estimated range associated with an upper-bound individual lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. Use the LSE tables and figures for a quick review of the health effects and to locate data for a specific exposure scenario. The LSE tables and figures should always be used in conjunction with the text. All entries in these tables and figures represent studies that provide reliable, quantitative estimates of NOAELs, LOAELs, or Cancer Effect Levels (CELs).

The legends presented below demonstrate the application of these tables and figures. Representative examples of LSE Table 3-1 and Figure 3-1 are shown. The numbers in the left column of the legends correspond to the numbers in the example table and figure.

LEGEND

See Sample LSE Table 3-1 (page B-6)

- (1) Route of Exposure. One of the first considerations when reviewing the toxicity of a substance using these tables and figures should be the relevant and appropriate route of exposure. Typically when sufficient data exist, three LSE tables and two LSE figures are presented in the document. The three LSE tables present data on the three principal routes of exposure, i.e., inhalation, oral, and dermal (LSE Tables 3-1, 3-2, and 3-3, respectively). LSE figures are limited to the inhalation (LSE Figure 3-1) and oral (LSE Figure 3-2) routes. Not all substances will have data on each route of exposure and will not, therefore, have all five of the tables and figures.
- (2) Exposure Period. Three exposure periods—acute (less than 15 days), intermediate (15–364 days), and chronic (365 days or more)—are presented within each relevant route of exposure. In this example, an inhalation study of intermediate exposure duration is reported. For quick reference to health effects occurring from a known length of exposure, locate the applicable exposure period within the LSE table and figure.
- (3) <u>Health Effect</u>. The major categories of health effects included in LSE tables and figures are death, systemic, immunological, neurological, developmental, reproductive, and cancer. NOAELs and LOAELs can be reported in the tables and figures for all effects but cancer. Systemic effects are further defined in the "System" column of the LSE table (see key number 18).
- (4) <u>Key to Figure</u>. Each key number in the LSE table links study information to one or more data points using the same key number in the corresponding LSE figure. In this example, the study represented by key number 18 has been used to derive a NOAEL and a Less Serious LOAEL (also see the two "18r" data points in sample Figure 3-1).
- (5) Species. The test species, whether animal or human, are identified in this column. Chapter 2, "Relevance to Public Health," covers the relevance of animal data to human toxicity and Section 3.4, "Toxicokinetics," contains any available information on comparative toxicokinetics. Although NOAELs and LOAELs are species specific, the levels are extrapolated to equivalent human doses to derive an MRL.
- (6) Exposure Frequency/Duration. The duration of the study and the weekly and daily exposure regimens are provided in this column. This permits comparison of NOAELs and LOAELs from different studies. In this case (key number 18), rats were exposed to "Chemical x" via inhalation for 6 hours/day, 5 days/week, for 13 weeks. For a more complete review of the dosing regimen, refer to the appropriate sections of the text or the original reference paper (i.e., Nitschke et al. 1981).
- (7) System. This column further defines the systemic effects. These systems include respiratory, cardiovascular, gastrointestinal, hematological, musculoskeletal, hepatic, renal, and dermal/ocular. "Other" refers to any systemic effect (e.g., a decrease in body weight) not covered in these systems. In the example of key number 18, one systemic effect (respiratory) was investigated.
- (8) <u>NOAEL</u>. A NOAEL is the highest exposure level at which no harmful effects were seen in the organ system studied. Key number 18 reports a NOAEL of 3 ppm for the respiratory system, which was used to derive an intermediate exposure, inhalation MRL of 0.005 ppm (see footnote "b").

- (9) <u>LOAEL</u>. A LOAEL is the lowest dose used in the study that caused a harmful health effect. LOAELs have been classified into "Less Serious" and "Serious" effects. These distinctions help readers identify the levels of exposure at which adverse health effects first appear and the gradation of effects with increasing dose. A brief description of the specific end point used to quantify the adverse effect accompanies the LOAEL. The respiratory effect reported in key number 18 (hyperplasia) is a Less Serious LOAEL of 10 ppm. MRLs are not derived from Serious LOAELs.
- (10) <u>Reference</u>. The complete reference citation is given in Chapter 9 of the profile.
- (11) <u>CEL</u>. A CEL is the lowest exposure level associated with the onset of carcinogenesis in experimental or epidemiologic studies. CELs are always considered serious effects. The LSE tables and figures do not contain NOAELs for cancer, but the text may report doses not causing measurable cancer increases.
- (12) <u>Footnotes</u>. Explanations of abbreviations or reference notes for data in the LSE tables are found in the footnotes. Footnote "b" indicates that the NOAEL of 3 ppm in key number 18 was used to derive an MRL of 0.005 ppm.

LEGEND

See Sample Figure 3-1 (page B-7)

LSE figures graphically illustrate the data presented in the corresponding LSE tables. Figures help the reader quickly compare health effects according to exposure concentrations for particular exposure periods.

- (13) <u>Exposure Period</u>. The same exposure periods appear as in the LSE table. In this example, health effects observed within the acute and intermediate exposure periods are illustrated.
- (14) <u>Health Effect</u>. These are the categories of health effects for which reliable quantitative data exists. The same health effects appear in the LSE table.
- (15) <u>Levels of Exposure</u>. Concentrations or doses for each health effect in the LSE tables are graphically displayed in the LSE figures. Exposure concentration or dose is measured on the log scale "y" axis. Inhalation exposure is reported in mg/m³ or ppm and oral exposure is reported in mg/kg/day.
- (16) <u>NOAEL</u>. In this example, the open circle designated 18r identifies a NOAEL critical end point in the rat upon which an intermediate inhalation exposure MRL is based. The key number 18 corresponds to the entry in the LSE table. The dashed descending arrow indicates the extrapolation from the exposure level of 3 ppm (see entry 18 in the table) to the MRL of 0.005 ppm (see footnote "b" in the LSE table).
- (17) <u>CEL</u>. Key number 38m is one of three studies for which CELs were derived. The diamond symbol refers to a CEL for the test species-mouse. The number 38 corresponds to the entry in the LSE table.

- (18) <u>Estimated Upper-Bound Human Cancer Risk Levels</u>. This is the range associated with the upper-bound for lifetime cancer risk of 1 in 10,000 to 1 in 10,000,000. These risk levels are derived from the EPA's Human Health Assessment Group's upper-bound estimates of the slope of the cancer dose response curve at low dose levels (q₁*).
- (19) Key to LSE Figure. The Key explains the abbreviations and symbols used in the figure.

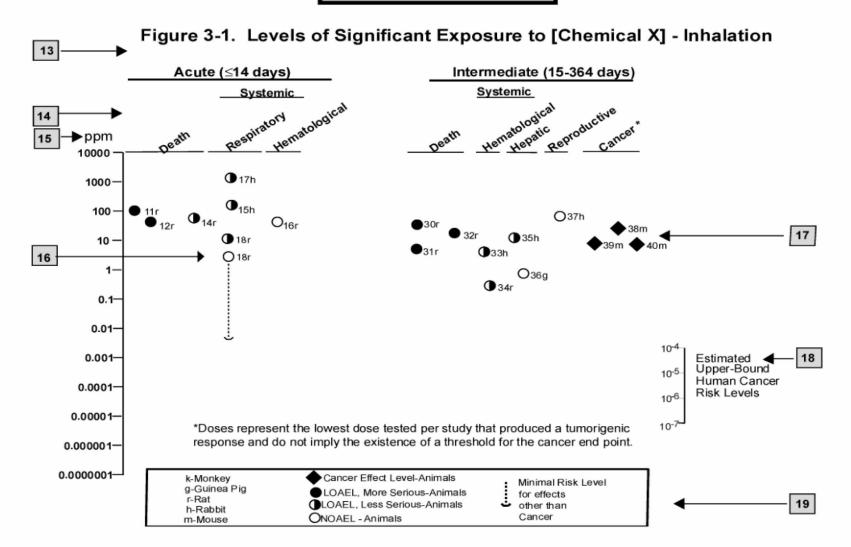
SAMPLE

Table 3-1. Levels of Significant Exposure to [Chemical x] – Inhalation

				Exposure			LOAEL (effect)			
		Key to figure ^a	Species	frequency/ duration	System	NOAEL (ppm)	Less seriou (ppm)	ıs	Serious (ppm)	Reference
2 →		INTERMEDIA	ATE EXPO	SURE						
			5	6	7	8	9			10
3	\rightarrow	Systemic	\downarrow	\downarrow	\downarrow	\downarrow	\downarrow			\
4	\rightarrow	18	Rat	13 wk 5 d/wk 6 hr/d	Resp	3 ^b	10 (hyperplasia)			Nitschke et al. 1981
		CHRONIC EXPOSURE								
		Cancer						11		
								\downarrow	-	
		38	Rat	18 mo 5 d/wk 7 hr/d				20	(CEL, multiple organs)	Wong et al. 1982
		39	Rat	89–104 wk 5 d/wk 6 hr/d				10	(CEL, lung tumors, nasal tumors)	NTP 1982
		40	Mouse	79–103 wk 5 d/wk 6 hr/d				10	(CEL, lung tumors, hemangiosarcomas)	NTP 1982

^a The number corresponds to entries in Figure 3-1.
^b Used to derive an intermediate inhalation Minimal Risk Level (MRL) of 5x10⁻³ ppm; dose adjusted for intermittent exposure and divided by an uncertainty factor of 100 (10 for extrapolation from animal to humans, 10 for human variability).

SAMPLE



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APPENDIX C. ACRONYMS, ABBREVIATIONS, AND SYMBOLS

ACGIH American Conference of Governmental Industrial Hygienists
ACOEM American College of Occupational and Environmental Medicine

ADI acceptable daily intake

ADME absorption, distribution, metabolism, and excretion

AED atomic emission detection
AFID alkali flame ionization detector
AFOSH Air Force Office of Safety and Health

ALT alanine aminotransferase AML acute myeloid leukemia

AOAC Association of Official Analytical Chemists

AOEC Association of Occupational and Environmental Clinics

AP alkaline phosphatase

APHA American Public Health Association

AST aspartate aminotransferase

atm atmosphere

ATSDR Agency for Toxic Substances and Disease Registry

AWQC Ambient Water Quality Criteria
BAT best available technology
BCF bioconcentration factor
BEI Biological Exposure Index

BMD/C benchmark dose or benchmark concentration

BMD_x dose that produces a X% change in response rate of an adverse effect

BMDL_x 95% lower confidence limit on the BMD_x

BMDS Benchmark Dose Software benchmark response

BSC Board of Scientific Counselors

C centigrade CAA Clean Air Act

CAG Cancer Assessment Group of the U.S. Environmental Protection Agency

CAS Chemical Abstract Services

CDC Centers for Disease Control and Prevention

CEL cancer effect level

CELDS Computer-Environmental Legislative Data System

CERCLA Comprehensive Environmental Response, Compensation, and Liability Act

CFR Code of Federal Regulations

Ci curie

CI confidence interval CL ceiling limit value

CLP Contract Laboratory Program

cm centimeter

CML chronic myeloid leukemia

CPSC Consumer Products Safety Commission

CWA Clean Water Act

DHEW Department of Health, Education, and Welfare DHHS Department of Health and Human Services

DNA deoxyribonucleic acid
DOD Department of Defense
DOE Department of Energy
DOL Department of Labor

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DOT Department of Transportation

DOT/UN/ Department of Transportation/United Nations/

NA/IMDG North America/Intergovernmental Maritime Dangerous Goods Code

DWEL drinking water exposure level ECD electron capture detection

ECG/EKG electrocardiogram electroencephalogram

EEGL Emergency Exposure Guidance Level EPA Environmental Protection Agency

F Fahrenheit

F₁ first-filial generation

FAO Food and Agricultural Organization of the United Nations

FDA Food and Drug Administration

FEMA Federal Emergency Management Agency

FIFRA Federal Insecticide, Fungicide, and Rodenticide Act

FPD flame photometric detection

fpm feet per minute FR Federal Register

FSH follicle stimulating hormone

g gram

GC gas chromatography gd gestational day

GLC gas liquid chromatography
GPC gel permeation chromatography

HPLC high-performance liquid chromatography
HRGC high resolution gas chromatography
HSDB Hazardous Substance Data Bank

IARC International Agency for Research on Cancer IDLH immediately dangerous to life and health

ILO International Labor Organization
IRIS Integrated Risk Information System

Kd adsorption ratio kg kilogram kkg metric ton

 K_{oc} organic carbon partition coefficient K_{ow} octanol-water partition coefficient

L liter

 $\begin{array}{lll} LC & liquid chromatography \\ LC_{50} & lethal concentration, 50\% \ kill \\ LC_{Lo} & lethal concentration, low \\ LD_{50} & lethal dose, 50\% \ kill \\ LD_{Lo} & lethal dose, low \\ LDH & lactic dehydrogenase \\ LH & luteinizing hormone \\ \end{array}$

LOAEL lowest-observed-adverse-effect level LSE Levels of Significant Exposure

LT₅₀ lethal time, 50% kill

m meter

MA trans,trans-muconic acid MAL maximum allowable level

mCi millicurie

CHLORINE C-3 APPENDIX C

MCL maximum contaminant level MCLG maximum contaminant level goal

MF modifying factor MFO mixed function oxidase

mg milligram
mL milliliter
mm millimeter

mmHg millimeters of mercury

mmol millimole

mppcf millions of particles per cubic foot

MRL Minimal Risk Level MS mass spectrometry

NAAQS National Ambient Air Quality Standard

NAS National Academy of Science

NATICH National Air Toxics Information Clearinghouse

NATO North Atlantic Treaty Organization NCE normochromatic erythrocytes

NCEH National Center for Environmental Health

NCI National Cancer Institute

ND not detected

NFPA National Fire Protection Association

ng nanogram

NHANES
NIEHS
NIOSH
NIOSH
NIOSH'S Computerized Information Retrieval System

NLM National Library of Medicine

nm nanometer nmol nanomole

NOAEL no-observed-adverse-effect level NOES National Occupational Exposure Survey NOHS National Occupational Hazard Survey

NPD nitrogen phosphorus detection

NPDES National Pollutant Discharge Elimination System

NPL National Priorities List

NR not reported

NRC National Research Council

NS not specified

NSPS New Source Performance Standards NTIS National Technical Information Service

NTP National Toxicology Program ODW Office of Drinking Water, EPA

OERR Office of Emergency and Remedial Response, EPA

OHM/TADS Oil and Hazardous Materials/Technical Assistance Data System

OPP Office of Pesticide Programs, EPA

OPPT Office of Pollution Prevention and Toxics, EPA

OPPTS Office of Prevention, Pesticides and Toxic Substances, EPA

OR odds ratio

OSHA Occupational Safety and Health Administration

OSW Office of Solid Waste, EPA OTS Office of Toxic Substances

C-4

OW Office of Water

OWRS Office of Water Regulations and Standards, EPA

PAH polycyclic aromatic hydrocarbon

PBPD physiologically based pharmacodynamic PBPK physiologically based pharmacokinetic

PCE polychromatic erythrocytes PEL permissible exposure limit

pg picogram

PHS Public Health Service
PID photo ionization detector

pmol picomole

PMR proportionate mortality ratio

ppb parts per billion ppm parts per million ppt parts per trillion

PSNS pretreatment standards for new sources

RBC red blood cell

REL recommended exposure level/limit

RfC reference concentration

RfD reference dose RNA ribonucleic acid RQ reportable quantity

RTECS Registry of Toxic Effects of Chemical Substances SARA Superfund Amendments and Reauthorization Act

SCE sister chromatid exchange

SGOT serum glutamic oxaloacetic transaminase
SGPT serum glutamic pyruvic transaminase
SIC standard industrial classification

SIM selected ion monitoring

SMCL secondary maximum contaminant level

SMR standardized mortality ratio

SNARL suggested no adverse response level

SPEGL Short-Term Public Emergency Guidance Level

STEL short term exposure limit STORET Storage and Retrieval

TD₅₀ toxic dose, 50% specific toxic effect

TLV threshold limit value TOC total organic carbon

TPQ threshold planning quantity
TRI Toxics Release Inventory
TSCA Toxic Substances Control Act

TWA time-weighted average UF uncertainty factor U.S. United States

USDA United States Department of Agriculture

USGS United States Geological Survey VOC volatile organic compound

WBC white blood cell

WHO World Health Organization

CHLORINE C-5 APPENDIX C

greater than >

≥ = greater than or equal to equal to

< less than

less than or equal to

≤ % percent alpha α beta β gamma $_{\delta}^{\gamma}$ delta micrometer μm microgram cancer slope factor μg_{*} q_1^*

negative positive +

weakly positive result weakly negative result (+) (-)

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CHLORINE D-1

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