## RESEARCH ARTICLE



# Repeated rectal application of a hyperosmolar lubricant is associated with microbiota shifts but does not affect PrEP drug concentrations: results from a randomized trial in men who have sex with men

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## Abstract

**Introduction:** Oral pre-exposure prophylaxis (PrEP) with tenofovir disoproxil fumarate (TDF) and emtricitabine (FTC) is highly effective in preventing HIV infection among men who have sex with men (MSM). The effects of consistent personal lubricant use in the rectum on tissue PrEP drug concentrations and the rectal microbiota are unknown. We investigated rectal PrEP drug concentrations and after repeated rectal application of a hyperosmolar lubricant.

**Methods:** We randomized 60 HIV-negative MSM to apply 4 mL of hyperosmolar rectal lubricant daily (n = 20), take daily oral TDF/FTC (n = 19), or both (n = 21) for seven days. Blood, rectal biopsies and rectal secretions were collected via rigid sigmoidoscopy before and on day 8 after product use. Tenofovir (TFV) and FTC as well as their intracellular metabolites tenofovir-diphosphate (TFV-DP), FTC-triphosphate (FTC-TP) were measured by HPLC-mass spectrometry. Rectal mucosal microbiota was sequenced with 16S rRNA sequencing using Illumina MiSeq.

**Results:** Seven days of lubricant application was not associated with differences in PrEP drug concentrations in rectal tissue or secretions. Lubricant use was associated with a decrease in the relative abundance of the *Bacteroides* genus (p = 0.01) and a non-significant increase in the *Prevotella* genus (p = 0.09) in the rectum. PrEP drug concentrations in rectal tissue and secretions were not associated with microbiota composition or diversity either before or after lubricant use.

**Conclusions:** Repeated rectal application of a hyperosmolar lubricant does not affect mucosal PrEP drug concentrations but is associated with changes in the rectal microbiome.

Keywords: lubricant; men who have sex with men; pre-exposure prophylaxis; anti-retroviral agents; HIV; gut microbiota

Additional Supporting Information may be found online in the Supporting information tab for this article.

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## 1 | INTRODUCTION

Daily oral pre-exposure prophylaxis (PrEP) with tenofovir disoproxil fumarate (TDF) and emtricitabine (FTC) is highly effective in preventing HIV infection, particularly among men who have sex with men (MSM) [1,2]. Receptive anal intercourse (RAI) remains the primary route of infection for MSM, yet the potential for biological and/or sexual factors to reduce PrEP efficacy remains largely unexplored. Personal lubricants are commonly used by MSM engaging in RAI [3]. While lubricants are typically made with ingredients from the "generally recognized as safe" list and do not contain pharmacologically active ingredients, there is a wide range of variability in the composition and chemical properties of personal lubricants [4]. In particular, water-based lubricants with hyperosmolar properties have been shown to induce epithelial tissue damage and cytotoxicity when applied in a single dose to rectal explants and epithelial cell cultures, or applied intermittently to the rectum of rhesus macaques *in vivo* [4-9]. Inflammation caused by tissue damage alters expression of drug metabolizing enzymes and transporters that can affect the absorption, distribution and clearance of antiretroviral (ARV) drugs [10]. However, single dose *ex vivo* tissue and intermittent dose *in vivo* animal models did not report lubricant-associated increases of susceptibility to HIV or SHIV infection [4,8]. In contrast, consistent use of personal lubricants for RAI among

a cohort of MSM in the United States was linked to increased acquisition of non-HIV sexually transmitted infections [11].

Recent reports show that when lubricants were used for nearly 90% of sex acts [3] of MSM engaging in condomless RAI, the rectal mucosal microbiota was enriched for the Prevotellaceae family as compared to Bacteroidaceae for controls. The ratio of Prevotella compared to Bacteroides in the gut microbiota has received significant attention as these taxa appear to demonstrate an antagonistic relationship and Prevotella has been associated with gut markers of inflammation [12-14]. Interestingly, differences in the composition of vaginal mucosal microbiota have been associated with topical PrEP efficacy in the CAPRISA 004 trial [15] but not with oral PrEP efficacy in the Partners PrEP study [16]. It is unknown if changes occurring in the rectal mucosa microbiota after repeated use of a hyperosmolar lubricant might affect PrEP drug concentrations in the rectal mucosa. Therefore, we conducted a clinical trial to measure rectal mucosal PrEP drug concentrations and microbiota among MSM who received daily oral TDF/FTC or repeated rectal application of hyperosmolar lubricant, or both in combination.

## 2 | METHODS

### 2.1 Ethics statement

The study protocol was funded by the US Centers for Disease Control and Prevention (CDC) and approved by Institutional Review Boards at Emory University and CDC and written informed consent was obtained from all study participants. The trial is registered at the ClinicalTrials.gov registry (NCT02401230) and conforms to the US Federal Policy for the Protection of Human Subjects.

## 2.2 Study participants and specimen collection

Sixty HIV-negative MSM in Atlanta, GA were enrolled in a non-blinded, randomized clinical trial to apply 4 mL of commercially available, hyperosmolar personal lubricant to the rectum daily with a study-supplied pre-filled applicator (n = 20), take oral TDF/FTC (Truvada®) daily (Gilead Pharmaceuticals, Foster City, CA) (n = 19), or both (n = 21) for seven consecutive days from March 2015 to July 2016 (Figure S1). Participants were instructed to apply the lube and take TDF/FTC near bedtime at approximately the same time each day and product adherence was evaluated by self-report. Sample size was estimated to be able to detect a fivefold reduction in tissue drug concentrations among participants using the lubricant. Participants who self-identified as male aged 19 to 46 years, reported RAI with a man in the last six months, and were in good general health as assessed by the study clinician were eligible to enrol. Men were allocated 1:1:1 to each study arm and randomized in blocks of three by random number generation. Randomization assignments were generated by study staff and concealed in sequentially numbered envelopes until after the study participant provided informed consent and study staff revealed study group assignment.

Participants were asked to refrain from RAI during the intervention, and only men who completed all study visits were included in analyses. The properties of the hyperosmolar lubricant have been described previously (osmolarity: 8064

mmol/kg, pH 4.44) [5]. Blood, rectal biopsies, and rectal secretions were collected at two study visits, the first at least 21 days prior to initiating study product and the second on day 8 following seven consecutive days of study product use. Seven days of TDF/FTC is estimated to be sufficient to achieve steady-state concentrations in rectal mononuclear cells based on a previous study [17]. An anorectal swab to test for *Neisseria gonorrhoeae* and *Chlamydia trachomatis* by nucleic acid amplification was collected at each biopsy visit.

Peripheral blood was collected in sodium citrate cell preparation tubes and separated into plasma and peripheral blood mononuclear cell (PBMC) fractions by centrifugation. Biopsies were collected from the mucosa approximately 8 to 10 cm above the external anal aperture, using a rigid sigmoidoscope and flexible sigmoidoscopic forceps (Olympus America, Center Valley, PA) mounted on a semi-flexible rod. Rectal secretions were collected via rigid sigmoidoscopy using Weckcel spears (Merocel, Mystic, CT). An enema was not used prior to collection of biopsies or rectal secretions.

### 2.3 | ARV drug measurement

The primary outcome for this trial was the difference in PrEP drug concentrations with and without lubricant use after seven days of study product use. Laboratory staff members were blinded to study arm assignment. Tenofovir (TFV) and FTC were eluted from Weckcel spears using a previously established method prior to measurement [18]. TFV and FTC in rectal fluid eluates were measured by HPLC-mass spectrometry as previously described with a lower limit of quantification of 10 ng/specimen [19]. TFV and FTC concentrations are presented in ng/swab for rectal secretions. Intracellular metabolites tenofovir-diphosphate (TFV-DP), emtricitabinetriphosphate (FTC-TP) and their corresponding natural substrates, dATP and dCTP in biopsies and PBMCs, were also measured by HPLC-mass spectrometry as previously described with a lower limit of quantification of 100 fmol/sample (500 fmol/sample for FTC-TP) for PBMC (5  $\times$  10<sup>6</sup> PBMC/ specimen) and biopsy [20]. Intracellular TFV-DP and FTC-TP concentrations in PBMC and biopsies were normalized to concentrations per 10<sup>6</sup> cells and mg tissue weight respectively. Specimens from one participant in the study arm receiving TDF/FTC and lubricant contained no detectable TFV, FTC or their respective intracellular metabolites TFV-DP or FTC-TP, suggesting the participant had not taken oral TDF/FTC. This participant was excluded from further analyses.

## 2.4 | Microbiome sequencing

Microbiota sequencing before and after study product use was performed as an exploratory outcome for this trial. Rectal mucosal swabs collected approximately 8 to 10 cm from the anal verge were stored in lysis buffer at -80°C. DNA was extracted, amplified and sequenced using methods described previously [21]. Microbiomes were unable to be sequenced in specimens collected from one participant in the study arm receiving TDF/FTC and a second participant in the study arm receiving TDF/FTC and lubricant. Reads were trimmed (Q20) and primer sequences removed by means of FastQ-mcf (https://code.google.com/archive/p/ea-utils/wikis/FastqMcf. wiki). Paired-end reads were joined using PANDASeq [22] and

analysed with QIIME 1.9.1 [23]. The 16S V1 to V2 contigs were clustered into operational taxonomic units (OTUs) by means of closed-reference OTU picking at 97% sequence similarity. OTU centroids were aligned using PyNAST [24] and subsequent phylogenetic trees were inferred by FastTree [25]. Centroid sequences from each OTU were aligned against the Greengenes database (version 13-8) for taxonomy assignment [26]. Finally, OTUs were summarized at the Genus level, resulting in 340 genera for the following statistical analyses. Raw 16s rRNA sequences were placed in the NCBI Sequence Read Archive under BioProject SRP148821.

### 2.5 Statistical analyses

Distributions of demographics and behavioural characteristics were compared across the three study arms with the nonparametric Kruskal-Wallis test. Drug concentration distributions were compared between men who took PrEP for seven days and men who took PrEP and used lubricant for seven days with the non-parametric Wilcoxon rank-sum test. For microbiome analyses, alpha diversity was measured using the Shannon index. Shannon index and the top 10 most abundant taxa were modelled according to the change in mean diversity and bacterial genus and mean relative abundance before and after study product use across study arms using linear regression models with an indicator for use of lubricant and an indicator for use of PrEP as two covariates with no intercept. We examined associations between the Shannon index and the relative abundance of specific genera of interest, Bacteroides, Prevotella and Prevotella unspecified before product use and PrEP drug concentrations after product use with the Spearman rank correlation coefficient. Finally, we examined the overall as well as individual influence of all bacterial taxa, including low abundant taxa, measured before study product use on PrEP drug concentrations by applying the linear decomposition model (LDM) method [27] using the Bray-Curtis and Jaccard distance measures; for Jaccard, we rarefied each specimen to 2500 total number of reads (with three out of thirty-seven specimens collected from men receiving TDF/ FTC removed due to insufficient reads), which is sufficient to cover almost full genera based on rarefaction curves, and we obtained multiple rarefied OTU tables to confirm the conclusion. The overall influence of all bacteria was confirmed by the permutational multivariate analysis of variance (PERMANOVA) test [28]. We visualized the overall association by the principal coordinates analysis (PCoA) plot (i.e. using the first two principal components of the distance matrix) which coloured the specimens differently for high versus low drug concentration defined as above and below the median.

## 3 | RESULTS

Demographic and risk behaviour data for men in the study are reported in Table 1. Median age, race and the number of reported RAI episodes in the last six months were similar between the three study arms.

The proportion of specimens with detectable ARVs in rectal secretions and tissues were similar among men applying a hyperosmolar lubricant (rectal secretions TFV 16/19, FTC 15/ 19; rectal tissue TFV-DP 20/20, FTC-TP 17/20) compared to men receiving TDF/FTC alone (rectal secretions TFV 17/19. FTC 17/19; rectal tissue TFV-DP 19/19, FTC-TP 15/19). Specimens with undetectable ARV concentrations were recoded to a value of 0 in subsequent analyses. Concentrations of TFV and FTC in rectal secretions were similar ( $p \ge 0.25$ ) between men who applied the hyperosmolar lubricant (median, range TFV: 540 ng/swab, 0 to 10,180 ng/swab; FTC: 141 ng/swab, 0 to 4815 ng/swab) and men who did not (median, range TFV: 772 ng/swab, 0 to 12,450 ng/swab; FTC: 261 ng/swab, 0 to 3895 ng/swab) (Figure 1). Similarly, concentrations of TFV-DP and FTC-TP detected in rectal tissues were not significantly different among men applying lubricant (median, range TFV-DP: 280 fmol/mg, 19 to 4536 fmol/mg; FTC-TP 108 fmol/mg, 0 to 674 fmol/mg) compared to men who did not (median, range TFV-DP: 510 fmol/mg, 0 to 4488 fmol/mg; FTC-TP: 75 fmol/mg, 4 to 489 fmol/mg); and no difference was observed in PBMC drug concentrations between men who applied lubricant (median, range TFV-DP: 38 fmol/10<sup>6</sup> cells, 0 to 89 fmol/10<sup>6</sup> cells; FTC-TP: 4020 fmol/10<sup>6</sup> cells, 1400 to 9330 fmol/10<sup>6</sup> cells) and those who did not (median, range TFV-DP: 44 fmol/10<sup>6</sup> cells, 0 to 79 fmol/10<sup>6</sup> cells; FTC-TP: 4980 fmol/ $10^6$  cells 1554 to 9470 fmol/ $10^6$  cells) (Figure 1). One participant who applied lubricant had a rectal N. gonorrhoeae infection at the time of specimen collection, but TFV (170 ng/swab) and FTC (141 ng/swab) in rectal secretions as well as TFV-DP (151 fmol/mg) and FTC-TP (65 fmol/mg) in rectal tissues remained within the range of concentrations observed in other study participants who applied lubricant. Median TFV-DP:dATP and FTC-TP:dCTP ratios were similar

Table 1. Demographic and behavioural characteristics of 60 HIV-negative MSM enrolled in a three-arm clinical trial to examine the mucosal effects of lube and PrEP on the rectal mucosal microbiota and PrEP drug concentrations

Characteristic	PrEP only (n = 19)	Lube only (n = 20)	PrEP+Lube (n = 21)	p-value
Median age (range)	28 (22 to 46)	28.5 (19 to 46)	26 (21 to 49)	0.64
Race/ethnicity, n (%)				
White	8 (42)	9 (45)	10 (48)	
Black	9 (47)	11 (55)	11 (52)	0.57
Other	2 (11)	0	0	
Median RAI acts with condom in last 6 months (range)	1 (0 to 26)	1.5 (0 to 20)	2 (1 to 50)	0.87
Median RAI acts without condom in last 6 months (range)	2 (0 to 30)	4 (0 to 30)	5 (1 to 50)	0.07

MSM, men who have sex with men; PrEP, pre-exposure prophylaxis; RAI, receptive anal intercourse.

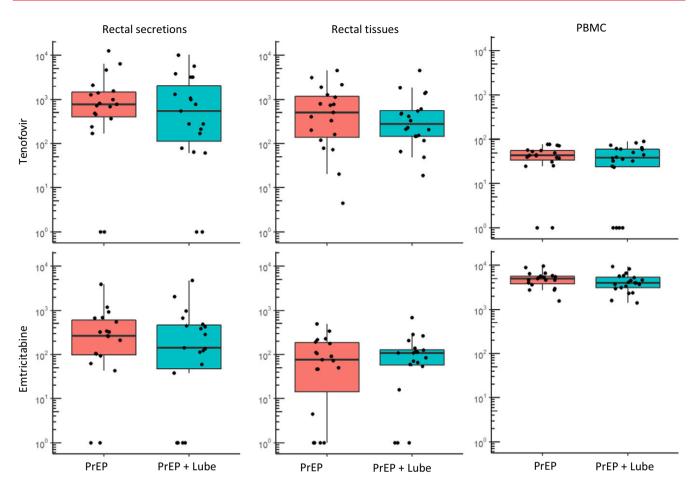


Figure 1. PrEP drug concentrations in rectal secretions, rectal tissue and PBMC in study participants who took oral PrEP stratified by lubricant use.

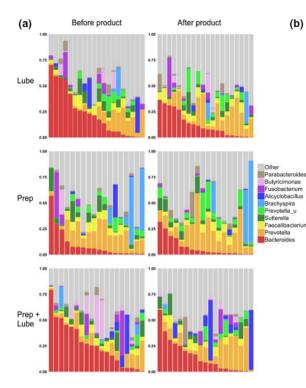
TFV and FTC concentrations were measured by HPLC-mass spectrometry and are presented in ng/swab for rectal secretions. Intracellular metabolites tenofovir-diphosphate (TFV-DP) and emtricitabine-triphosphate (FTC-TP) in biopsies and PBMCs were also measured by HPLC-mass spectrometry and were normalized to concentrations per mg of tissue and per 10<sup>6</sup> cells respectively. All *p*-values for median differences between study arms by Wilcoxon rank sum test >0.25. Drug levels below the limit of quantification are displayed at 10°. FTC, emtricitabine; PBMC, peripheral blood mononuclear cell; PrEP, pre-exposure prophylaxis; TFV, tenofovir; TFV-DP, tenofovir-diphosphate.

among men who applied the hyperosmolar lubricant (TFV-DP: dATP 5.1; FTC-TP:dCTP 1.4) when compared to those who did not (TFV-DP:dATP 4.4; FTC-TP:dCTP 2.1). Intracellular concentrations of dATP and dCTP in rectal tissues remained unchanged following lubricant application (data not shown).

Summary data for the relative abundance of the top 10 genera across the three study arms and between study visits are presented in Figure 2. Overall, there was a significant (p = 0.01) decrease in the relative abundance of Bacteroides and a non-significant (p = 0.086) increase in the relative abundance of the Prevotella genus associated with lubricant use. Mean Bacteroides abundance in the lube arms before study product use was 23.2% before and 12.8% after study product use (Figure 2b). Mean Prevotella abundance in the lube arms before study product use was 9% before and 14.25% after study product use. There were no differences in less abundant taxa associated with lubricant use. Lubricant use was also associated with an increase in microbiota diversity as measured by Shannon index (mean Shannon index before product = 2.44 vs. after product = 2.78; p = 0.006). TDF/FTC use was associated with a small, but significant, increase in

the mean relative abundance of an unspecified genus of Prevotellaceae (mean before 3.7% vs. after 7.1%; p = 0.02), but no other changes in the composition or diversity of the microbiota were associated with TDF/FTC use. There was evidence (p = 0.039) for statistical interaction between oral PrEP and repeated application of lubricant only for the relative abundance of the genus *Alicyclobacillus*.

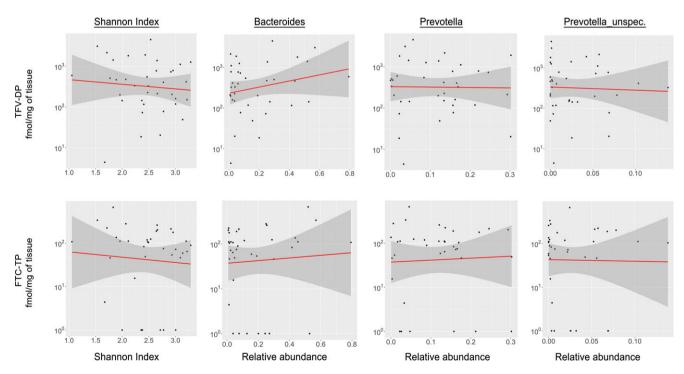
Finally, we examined the effect of microbiota diversity and composition on TDF/FTC drug concentrations in PBMCs, rectal secretions and rectal tissue (representative data shown for rectal tissue in Figure 3). We found no significant associations between the Shannon index, relative abundance of *Bacteroides* or relative abundance of *Prevotella* (including an unspecified genus of Prevotellaceae), measured before study product use with TDF or FTC drug concentrations in biopsies or secretions after product use. To examine if all taxa, including lower abundant taxa, were overall associated with differences in PrEP drug concentrations, PCoA plots using the Bray–Curtis and Jaccard distance measures were constructed and did not show separation between high and low drug concentrations (representative data for rectal tissue shown in Figure 4).



Study Arm Taxonomy	Lube		PrEP		PrEP+Lube		Model based p-value		
	Before	After	Before	After	Before	After	Overall Lube effect	Overall PrEP effect	Interac- tion
Bacteroides	0.232	0.128	0.098	0.072	0.264	0.148	0.01	0.56	0.85
Prevotella	0.09	0.142	0.15	0.185	0.076	0.142	0.09	0.28	0.71
Faecalibacterium	0.042	0.051	0.05	0.042	0.038	0.044	0.28	0.36	0.75
Sutterella	0.045	0.034	0.058	0.045	0.042	0.012	0.34	0.28	0.25
Prevotella_u	0.232	0.045	0.037	0.071	0.018	0.031	0.31	0.02	0.14
Brachyspira	0.016	0023	0.084	0.089	0.017	0.004	0.86	0.79	0.60
Alicyclobacillus	0.036	0.003	0.021	0.006	0.037	0.084	0.55	0.25	0.04
Fusobacterium	0.03	0.036	0.035	0.008	0.05	0.03	0.23	0.79	0.89
Butyricimonas	0.022	0.016	0.014	0.002	0.063	0.015	0.54	0.75	0.40
Parabacteroides	0.02	0.021	0.014	0.03	0.017	0.021	0.23	0.92	0.55

#### Figure 2. Lubricant use is associated with shifts in rectal microbiota composition.

Mean relative abundance of the top 10 most abundant genera in the rectal mucosa before and after study product use stratified by study arm: Lube only, PrEP only, or PrEP and Lube use for seven days (**a**). Change in mean relative abundance before and after study product use in terms of lubricant and PrEP use for the top 10 genera were examined by linear regression modelling (**b**). PrEP, pre-exposure prophylaxis.



#### Figure 3. Microbiota species are not associated with differences in tissue PrEP drug concentrations.

Associations by Spearman correlation between gut microbial diversity (Shannon index), the relative abundance of *Bacteroides*, *Prevotella* or an unspecified genus of Prevotellaceae as measured before study product use, and PrEP drug concentrations in rectal tissues following seven days of oral TDF/FTC dosing. All *p*-values for correlations >0.25. Similar results were seen for PBMC and rectal secretions specimens. FTC, emtricitabine; PBMC, peripheral blood mononuclear cell; PrEP, pre-exposure prophylaxis; TDF, tenofovir disoproxil fumarate.

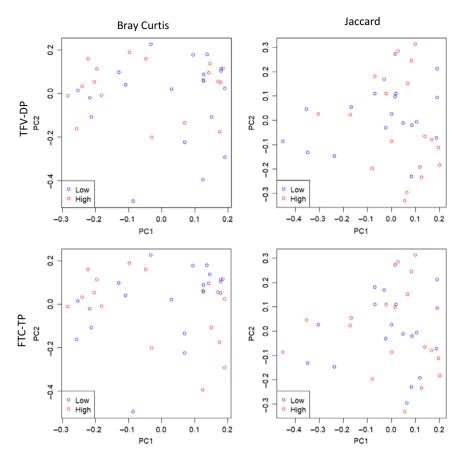


Figure 4. Microbiota composition are not associated with changes in PrEP drug concentrations.

Representative data for TFV-DP and FTC-TP drug concentrations in rectal tissues following seven days of oral TDF/FTC dosing were stratified at the median into low (blue circles) and high (red circles) drug level groups. Bray–Curtis and Jaccard distance-based principal coordinates analysis (PCoA) plots are shown for rectal tissues demonstrating no separation between specimens with low and high drug levels. Similar results were seen for PBMC and rectal secretions specimens. FTC, emtricitabine; PBMC, peripheral blood mononuclear cell; PrEP, pre-exposure prophylaxis; TDF, tenofovir disoproxil fumarate; TFV, tenofovir; TFV-DP, tenofovir-diphosphate.

When considering drug concentrations as continuous variables, LDM generated a near statistically significant (p = 0.053) global association using Bray–Curtis but detected no associated genera, while PERMANOVA yielded a slightly larger p-value (0.056), suggesting a possible association between the overall composition of the microbiota and PBMC FTC-TP drug concentrations, but no other drug concentrations examined. Both LDM and PERMANOVA generated non-significant global associations using Jaccard, indicating no association between the overall presence–absence of the microbiota and any drug concentration.

## 4 DISCUSSION

Hyperosmolar lubricants are commonly used during RAI, but their use may exacerbate tissue damage during RAI raising concerns that cytotoxic effects could potentially reduce the efficacy of biomedical interventions to prevent HIV. This study examined potential changes in the rectal mucosal microbiota and PrEP drug concentrations that may occur with repeated rectal application of a representative product of hyperosmolar water-based personal lubricants. The results presented here indicate that despite alterations in microbiota diversity and composition associated with repeated rectal application of hyperosmolar lubricant, PrEP drug concentrations in blood, rectal secretions and tissue remain similar among MSM who apply lubricant and those who do not.

Studies in non-human primates and tissue explants indicate cytotoxic effects of hyperosmolar lubricants do not increase susceptibility to SHIV or HIV infection respectively [4,8]. These results combined with the observation that lubricant application does not significantly affect the concentration and metabolism of PrEP ARVs indicate ARVs are likely to retain their protective efficacy even as lubricants are used during RAI. Certain personal lubricants may also contain anti-HIV properties as has been suggested previously [29,30]. Furthermore, systemic drug concentrations predict protection from oral dosing regimens suggesting that despite localized tissue damage caused by hyperosmolar personal lubricants, systemic drug concentrations will remain protective in persons adherent to daily oral PrEP [31,32]. These observations are consistent with results of clinical trials indicating daily TDF/FTC remains highly protective among adherent study participants even in the presence of additional biological and behavioural factors [1,2].

The concentrations of TFV-DP and FTC-TP in rectal tissue as well as TFV and FTC in rectal secretions presented here

are within ranges of previously reported tissue concentrations among MSM receiving oral TDF/FTC for PrEP [33]. The maintenance of intracellular ARV metabolite concentrations in rectal tissue and extracellular ARV concentrations in rectal secretions indicates HIV target cells within mucosal tissues are likely to maintain ARV concentrations consistent with protection even in the presence of tissue damage caused by hyperosmolar lubricants. Hyperosmolar lubricant properties could induce transient release of drug-containing fluid from tissues. However, as lubricant application and oral TDF/FTC dosing were concurrent, it is likely the lube mucosal effect occurred prior to oral drug reaching rectal tissue resulting in no differences in drug measured in rectal secretions. However, repeated lubricant application is likely to induce tissue damage and inflammation resulting in detectable differences in intracellular drug metabolites that accumulate with subsequent dosing, yet we were unable to detect such differences in this study. TFV-DP and FTC-TP are natural analogs of dATP and dCTP, respectively, and decreased ratios of TFV-DP to dATP have been associated with cellular activation [34-36]. However, immune activation and metabolic changes associated with inflammation did not result in a corresponding decrease in the ratio of either TFV-DP:dATP or FTC-TP:dCTP, which might have resulted in reduced efficacy.

While expression of drug metabolizing enzymes and transporters is altered during inflammation [10], our results suggest these changes may not affect ARV concentrations in mucosal tissues. It is possible that lubricant use insignificantly reduced mucosal ARV concentrations due to the number of study participants in this study. However, it is also possible that adherence to daily oral PrEP and the ability of the rectum to repair itself rapidly combine and continually replenish drug concentrations in rectal tissues even in the presence of shortlived tissue damage. It is notable that imperfect adherence to daily oral dosing PrEP regimens continues to challenge implementation efforts and our study was not designed to determine if lubricant use may exacerbate suboptimal drug concentrations in these situations.

Recent data on suboptimal efficacy of topically applied TFV in the setting of dysbiosis in the female genital tract have underlined the importance of potential interactions between the mucosal immune environment and biomedical HIV prevention interventions [15]. We showed an enrichment for Prevotella over Bacteroides in the rectal mucosa with repeated application of hyperosmolar lubricant. Previous studies, including one from members of our group, demonstrated enrichment of the rectal microbiota for Prevotella among MSM, who commonly use lubricants for RAI, and these data suggest an important mechanism for this finding [37,38]. We hypothesize Prevotella may be better able to metabolize products of mucosal injury, such as may occur with repeated application of hyperosmolar lubricant, and thus flourishes in this environment. Other factors such as microtrauma during intercourse, semen exposure or enema use may also cause mucosal injury, and contribute to Prevotella enrichment among MSM and deserve further investigation.

Nonetheless, it is still unclear that enrichment for *Prevotella* in the gut microbiota represents a pathogenic or "dysbiotic" state. *Prevotella* and *Bacteroides* both belong to the Bacteroidetes phylum, which contributes to a healthy gut microbiota; however, some have suggested enrichment for *Prevotella* 

versus Bacteroides represents a distinct "enterotype" [39], or underlying microbial community, while others have not confirmed this finding [40,41]. Prevotella has been associated with gut mucosal markers of inflammation in chronic HIV infection [13], rheumatoid arthritis and bacterial vaginosis [14], although some associations are likely confounded by sexual behaviour. However, in vitro data do not support a clear mechanism for a direct inflammatory response to Prevotella, and its association with plant-based diets suggests that enrichment for Prevotella alone is not, by itself, pathogenic [42,43]. While further research will certainly be needed to elucidate the relationship between Prevotella and mucosal inflammation, our data show there is no association of gut microbiota diversity or composition, including both majority and low abundant taxa, with mucosal PrEP drug concentrations after oral dosing. This reassuring finding is consistent with the preservation of oral PrEP efficacy seen in the setting of vaginal dysbiosis in the Partners PrEP study [16]. While our data do not support a reduction in PrEP drug concentrations with differences in the gut microbiota when dosed orally, it is still unknown whether gut microbiota diversity or composition can influence rectal mucosal HIV transmission or efficacy of other biomedical prevention interventions such as candidate rectal microbicides or vaccines and deserves further investigation.

This study has a number of limitations as it was designed to model the effects of repeat rectal application of a hyperosmolar lubricant on mucosal PrEP drug concentrations. Microbiome analyses were initially planned as future, exploratory topics; however, the recent data from the female genital tract prompted us to prioritize these analyses. Lubricant application was documented by self-report and some men may not have applied the lubricant; nonetheless, the finding of shifts in the microbiota in the lubricant study arms suggests adherence to study product. Additionally, the changes in the microbiota are assumed to result from tissue damage and inflammation caused by the lubricant; however, these direct effects of the lubricant were not tested for in this study. The small sample size and the wide range of tissue ARV concentrations may obscure subtle changes in ARV concentrations following lubricant application, and statistical power was insufficient to detect small differences in bacterial taxa abundance with study product use. This study modelled the effects of hyperosmolar lubricants on PrEP drug concentrations in the rectal mucosa in the absence of RAI. It is unclear if the combination of coitus and lubricant application might create unfavourable changes in the protection afforded by daily oral PrEP. Additionally, the lubricant studied here is one of a number of commercially available lubricants and was chosen as a representative of water-based hyperosmolar personal lubricants; therefore, the results presented here may not be applicable to other products.

## 5 | CONCLUSIONS

Repeated application of a hyperosmolar lubricant did not affect mucosal PrEP drug concentrations among MSM, but was associated with changes in the rectal microbiome. While hyperosmolar personal lubricants may exacerbate damage to the rectal mucosa during RAI and lead to shifts in the rectal mucosal microbiota to favour *Prevotella* over *Bacteroides*, protective mucosal and systemic drug concentrations in persons adherent to daily oral TDF/FTC remain unchanged and are likely to retain the ability to prevent HIV infection among MSM.

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#### COMPETING INTERESTS

The authors declare they have no competing interests.

#### AUTHORS' CONTRIBUTIONS

REH, CEH and CFK designed the research study. LH, NAP and SH recruited study participants and collected data and specimens for analysis. JF, AH and CD performed and analysed drug measurements. YH, YX and Y-J H performed and analysed microbiome data. REH and CFK wrote the manuscript with contributions and interpretation of findings from all co-authors. All authors have read and approved the final manuscript.

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#### DISCLAIMER

The findings and conclusions in this manuscript are those of the authors and do not necessarily represent the official position of the United States Centers for Disease Control and Prevention or the Department of Health and Human Services.

#### REFERENCES

1. Grant RM, Lama JR, Anderson PL, McMahan V, Liu AY, Vargas L, et al. Preexposure chemoprophylaxis for HIV prevention in men who have sex with men. N Engl J Med. 2010;363:2587–99.

2. Molina JM, Capitant C, Spire B, Pialoux G, Cotte L, Charreau I, et al. Ondemand preexposure prophylaxis in men at high risk for HIV-1 infection. N Engl J Med. 2015;373:2237–46.

3. Rolle CP, Bolton MD, Kelley CF. Use of a prospective sex diary to study anal lubricant and enema use among high risk men who have sex with men-implications for human immunodeficiency virus prevention. Sex Transm Dis. 2016;43:476–8.

4. Dezzutti CS, Brown ER, Moncla B, Russo J, Cost M, Wang L, et al. Is wetter better? An evaluation of over-the-counter personal lubricants for safety and anti-HIV-1 activity. PLoS One. 2012;7:e48328.

5. Begay O, Jean-Pierre N, Abraham CJ, Chudolij A, Seidor S, Rodriguez A, et al. Identification of personal lubricants that can cause rectal epithelial cell damage and enhance HIV type 1 replication *in vitro*. AIDS Res Hum Retro-viruses. 2011;27:1019–24.

6. Fuchs EJ, Lee LA, Torbenson MS, Parsons TL, Bakshi RP, Guidos AM, et al. Hyperosmolar sexual lubricant causes epithelial damage in the distal colon: potential implication for HIV transmission. J Infect Dis. 2007;195:703–10.

7. Adriaens E, Remon JP. Mucosal irritation potential of personal lubricants relates to product osmolality as detected by the slug mucosal irritation assay. Sex Transm Dis. 2008;35:512–6.

8. Vishwanathan SA, Morris MR, Wolitski RJ, Luo W, Rose CE, Blau DM, et al. Rectal application of a highly osmolar personal lubricant in a macaque model induces acute cytotoxicity but does not increase risk of SHIV infection. PLoS One. 2015;10:e0120021.

9. Hladik F, Burgener A, Ballweber L, Gottardo R, Vojtech L, Fourati S, et al. Mucosal effects of tenofovir 1% gel. Elife. 2015;4:e04525.

10. Seifert SM, Castillo-Mancilla JR, Erlandson KM, Anderson PL. Inflammation and pharmacokinetics: potential implications for HIV-infection. Expert Opin Drug Metab Toxicol. 2017;13:641–50.

11. Gorbach PM, Weiss RE, Fuchs E, Jeffries RA, Hezerah M, Brown S, et al. The slippery slope: lubricant use and rectal sexually transmitted infections: a newly identified risk. Sex Transm Dis. 2012;39:59–64.

12. Ley RE. Gut microbiota in 2015: Prevotella in the gut: choose carefully. Nat Rev Gastroenterol Hepatol. 2016;13:69–70.

13. Dillon SM, Lee EJ, Kotter CV, Austin GL, Dong Z, Hecht DK, et al. An altered intestinal mucosal microbiome in HIV-1 infection is associated with mucosal and systemic immune activation and endotoxemia. Mucosal Immunol. 2014;7:983–94.

14. Larsen JM. The immune response to Prevotella bacteria in chronic inflammatory disease. Immunology. 2017;151:363–74.

15. Klatt NR, Cheu R, Birse K, Zevin AS, Perner M, Noël-Romas L, et al. Vaginal bacteria modify HIV tenofovir microbicide efficacy in African women. Science. 2017;356:938–45.

16. Heffron R, McClelland RS, Balkus JE, Celum C, Cohen CR, Mugo N, et al. Efficacy of oral pre-exposure prophylaxis (PrEP) for HIV among women with abnormal vaginal microbiota: a post-hoc analysis of the randomised, placebocontrolled Partners PrEP Study. Lancet HIV. 2017;4:e449–56.

17. Seifert SM, Chen X, Meditz AL, Castillo-Mancilla JR, Gardner EM, Predhomme JA, et al. Intracellular tenofovir and emtricitabine anabolites in genital, rectal, and blood compartments from first dose to steady state. AIDS Res Hum Retroviruses. 2016;32:981–91.

18. Massud I, Martin A, Dinh C, Mitchell J, Jenkins L, Heneine W, et al. Pharmacokinetic profile of raltegravir, elvitegravir and dolutegravir in plasma and mucosal secretions in rhesus macaques. J Antimicrob Chemother. 2015;70: 1473–81.

19. Kuklenyik Z, Martin A, Pau CP, Garcia-Lerma JG, Heneine W, Pirkle JL, et al. Effect of mobile phase pH and organic content on LC-MS analysis of nucleoside and nucleotide HIV reverse transcriptase inhibitors. J Chromatogr Sci. 2009;47:365–72.

20. Kuklenyik Z, Martin A, Pau CP, Holder A, Youngpairoj AS, Zheng Q, et al. On-line coupling of anion exchange and ion-pair chromatography for measurement of intracellular triphosphate metabolites of reverse transcriptase inhibitors. J Chromatogr B Analyt Technol Biomed Life Sci. 2009;877:3659–66.

21. Halpin AL, de Man TJ, Kraft CS, Perry KA, Chan AW, Lieu S, et al. Intestinal microbiome disruption in patients in a long-term acute care hospital: a case for development of microbiome disruption indices to improve infection prevention. Am J Infect Control. 2016;44:830–6.

22. Bartram AK, Lynch MD, Stearns JC, Moreno-Hagelsieb G, Neufeld JD. Generation of multimillion-sequence 16S rRNA gene libraries from complex microbial communities by assembling paired-end illumina reads. Appl Environ Microbiol. 2011;77:3846–52.

23. Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, et al. QIIME allows analysis of high-throughput community sequencing data. Nat Methods. 2010;7:335–6.

24. Caporaso JG, Bittinger K, Bushman FD, DeSantis TZ, Andersen GL, Knight R. PyNAST: a flexible tool for aligning sequences to a template alignment. Bioinformatics. 2010;26:266–7.

25. Price MN, Dehal PS, Arkin AP. FastTree: computing large minimum evolution trees with profiles instead of a distance matrix. Mol Biol Evol. 2009;26:1641–50.

26. DeSantis TZ, Hugenholtz P, Larsen N, Rojas M, Brodie EL, Keller K, et al. Greengenes, a chimera-checked 16S rRNA gene database and workbench compatible with ARB. Appl Environ Microbiol. 2006;72:5069–72.

27. Hu YJ, Satten GA. Testing hypotheses about microbiome using an ordination-based linear decomposition model. [cited 2018 Apr 10]. Available from: https://www.biorxiv.org/content/early/2017/12/06/229831

28. McArdle BH, Anderson MJ. Fitting multivariate models to community data: a comment on distance-based redundancy analysis. Ecology. 2001;82: 290–7.

29. Baron S, Poast J, Nguyen D, Cloyd MW. Practical prevention of vaginal and rectal transmission of HIV by adapting the oral defense: use of commercial lubricants. AIDS Res Hum Retroviruses. 2001;17:997–1002.

30. Nguyen D, Lee H, Poast J, Cloyd MW, Baron S. Preventing sexual transmission of HIV: anti-HIV bioregulatory and homeostatic components of commercial sexual lubricants. J Biol Regul Homeost Agents. 2004;18:268–74.

31. Anderson PL, Glidden DV, Liu A, Buchbinder S, Lama JR, Guanira JV, et al. Emtricitabine-tenofovir concentrations and pre-exposure prophylaxis efficacy in men who have sex with men. Sci Transl Med. 2012;4:151ra25.

32. Donnell D, Baeten JM, Bumpus NN, Brantley J, Bangsberg DR, Haberer JE, et al. HIV protective efficacy and correlates of tenofovir blood concentrations in a clinical trial of PrEP for HIV prevention. J Acquir Immune Defic Syndr. 2014;66:340–8.

33. Cottrell ML, Prince HM, Allmon A, Mollan KR, Hudgens MG, Sykes C, et al. Cervicovaginal and rectal fluid as a surrogate marker of antiretroviral tissue concentration: implications for clinical trial design. J Acquir Immune Defic Syndr. 2016;72:498–506.

34. Robbins BL, Wilcox CK, Fridland A, Rodman JH. Metabolism of tenofovir and didanosine in quiescent or stimulated human peripheral blood mononuclear cells. Pharmacotherapy. 2003;23:695–701.

35. Garcia-Lerma JG, Aung W, Cong ME, Zheng Q, Youngpairoj AS, Mitchell J, et al. Natural substrate concentrations can modulate the prophylactic efficacy of nucleotide HIV reverse transcriptase inhibitors. J Virol. 2011;85:6610–7.

36. Haaland RE, Holder A, Pau CP, Swaims-Kohlmeier A, Dawson C, Smith DK, et al. Levels of intracellular phosphorylated tenofovir and emtricitabine correlate with natural substrate concentrations in peripheral blood mononuclear cells of persons prescribed daily oral truvada for HIV pre-exposure prophylaxis. J Acquir Immune Defic Syndr. 2017;75:e86–8.

37. Kelley CF, Kraft CS, de Man TJ, Duphare C, Lee HW, Yang J, et al. The rectal mucosa and condomless receptive anal intercourse in HIV-negative MSM: implications for HIV transmission and prevention. Mucosal Immunol. 2017;10:996–1007. 38. Noguera-Julian M, Rocafort M, Guillén Y, Rivera J, Casadellà M, Nowak P, et al. Gut microbiota linked to sexual preference and HIV infection. EBioMedicine. 2016;5:135–46.

39. Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, Mende DR, et al. Enterotypes of the human gut microbiome. Nature. 2011;473:174–80.

40. Gorvitovskaia A, Holmes SP, Huse SM. Interpreting Prevotella and Bacteroides as biomarkers of diet and lifestyle. Microbiome. 2016;4:15.

41. Pescatore NA, Pollak R, Kraft CS, Mulle JG, Kelley CF. Anatomic site of sampling and the rectal mucosal microbiota in HIV negative men who have sex with men engaging in condomless receptive anal intercourse. AIDS Res Hum Retroviruses. 2018;34:277–81.

42. Lozupone CA, Rhodes ME, Neff CP, Fontenot AP, Campbell TB, Palmer BE. HIV-induced alteration in gut microbiota: driving factors, consequences, and effects of antiretroviral therapy. Gut Microbes. 2014;5:562–70.

43. Ruengsomwong S, Korenori Y, Sakamoto N, Wannissorn B, Nakayama J, Nitisinprasert S. Senior Thai fecal microbiota comparison between vegetarians and non-vegetarians using PCR-DGGE and real-time PCR. J Microbiol Biotechnol. 2014;24:1026–33.

## SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article:

Figure S1. CONSORT flow diagram of study participants.