

December, 1991

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PURPOSE OF THE ARBOVIRUS INFORMATION EXCHANGE: To exchange information on a timely basis. The recipients are those who study various aspects of arboviruses. The Exchange contains preliminary reports, summaries, observations, and comments submitted voluntarily by qualified agencies and individual investigators. The appearance in the Exchange of any information, data, opinions, or views does not constitute formal publication and should not be referred to in "Reference" sections of papers or included in lists of publications. The Exchange is not a "peer reviewed" publication; in fact, it is not a publication at all. Any reference to or quotation of any part of the Exchange must be authorized directly by the agency or person submitting the text.

DENGUE

Dengue-2 outbreak in Araguaina, Tocantins State, Brazil. (P.F.C. Vasconcelos, E.S. Travassos da Rosa, J.F.S. Travassos da Rosa, R.B. Freitas, A.P.A. Travassos da Rosa)

A 10 μ L ultramicroELISA for detection of human IgM immunoglobulin against dengue virus. (J.L. Pelegrino, J. Laferté, M.G. Guzmán, S. Vázquez, G. González, A. Valdivia)

Purification of specific human IgG and IgM for the detection of dengue antibody levels in human serum. (A. Otero, J.L. Pelegrino, S. Vázquez, J. Laferté)

Monoclonal antibodies that recognize a Cuban dengue strain. (C. Hermida, G. Guzmán, M. Pupo, M. González, R. Marcet, S. Vázquez, Y. Caballero, I. Delgado)

Pathological changes of liver in dengue infections (S. Hotta)

Nuclear localization of the core protein of dengue virus. (R. Bulich, J. Haig, J. Aaskov)

Dengue in French Guiana. (letter from J.M. Reynes)

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Experimental transmission of Venezuelan equine 1 encephalomyelitis virus by Aedes albopictus (Diptera: Culicidae) from New Orleans, Louisiana. (M.J. Turell, J.R. Beaman)

Effect of environmental temperature on the vector competence of 1 Aedes taeniorhynchus for Venezuelan equine encephalomyelitis and Rift Valley fever viruses. (M.J. Turell)

First confirmation of breeding populations of Aedes albopictus 1 in Continental Africa. (H.M. Savage, B.R. Miller, V.I. Ezike, A.C.N. Nwankwo, R. Spiegel)

Culicoides (Diptera: Ceratopogonidae) vector species in Israel 1 trapped from burrows of rodents that might be potential reservoirs of bluetongue and Akabane viruses. (Y. Braverman, N. Messaddeq, and M Kremer)

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Expression of the bacterial gene, Chloramphenicol 1 acetyltransferase (CAT) in mosquitoes and mosquito cells using recombinant Sindbis virus vectors. (K. Olson, S. Higgs, J. Carlson, B. Beaty) Detection of genetic variation between populations of 2 <u>Aedes aegypti</u> mosquitoes by amplification of genomic DNA using random primers. (M.E. Ballinger-Crabtree, B.R. Miller)

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Epizootiological aspects of hantavirus infection in Belgium, The Netherlands, and Germany: Of Mice and Men. (J.P. Clement, A. Lefevre, J. Groen, A. Osterhaus, R. Verhagen, H. Leirs, G. van der Groen)

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Study on the experimental infection of tree shrew with chikungunya virus. (H. Zhang, Z. Mi, H. Shi, D. Zi)

The natural infection rate of mosquitoes by Japanese encephalitis B virus in Yunnan Province, China. (H. Zhang, D. Zi, H. Shi, Z. Mi, Z. Gong, J Zhang, Z. Hou)

Interferon studies in Mayaro virus-infected TC7 cells. (M.C.S. Rebello, J.L. Oliveira, M.E.F. Fonseca, M.A. Rebello)

Inability of Barmah Forest virus to cross the placenta in mice. (P. Birt, J. Haig, J. Aaskov)

Effect of pH on infectivity of Eyach virus in Vero cells. (G. Dobler, H. Meier-Ewert)

PATHOLOGY

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LYME DISEASE

Serological studies on the infection of dogs in Ontario, Canada, with *Borrelia burgdorferi*. (H. Artsob, I. Barker, R. Fister, G. Sephton)

A serosurvey for antibody to *Borrelia burgdorferi* in an "at risk" population of southern Portugal. (A.R. Filipe)

Ixodes pacificus and Borrelia burgdorferi in Arizona. (C.A. Olson, E.W. Cupp, S. Luckhart, J.M.C. Ribeiro, C. Levy)

Borreliae in culicine mosquitoes. (J. Halouzka)

Sample controlled seroprevalence study of Lyme infection in Belgium. (J. Clement, A. Ramon, W. Vranckx, A. Lefevre, A. Fain, G.R.F. Krueger, R. Ackermann)

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NOTE: As noted in "PURPOSE OF THE ARBOVIRUS INFORMATION EXCHANGE", which is found on the front page of each issue, you are encouraged to submit a brief summary of your work. The summary need not be in manuscript style, the results do not have to be definitive, you need not include tables (unless you want to). This is not a peer-reviewed publication. The intent is to communicate among ourselves and to let each other know what we are doing.

The next issue will likely be mailed June 1, 1992 (probable deadline for submissions: May 15, 1992). There is nothing that requires you to wait until the last minute. If you have something to communicate in January, February, March or April, please send it. Also, there is nothing that prevents you from submitting a report to <u>every issue</u>. There are no pages charges either but, then again, this is not a publication.

PLEASE !!!

Follow the directions for submitting reports. Double-spaced pages take twice as much space as single-spaced pages. Do not double-space or number pages. Single-space them and leave them unnumbered. Do not staple pages together.

Charles H. Calisher, Ph.D. DVBID/NCID/CDC P.O. Box 2087 Ft. Collins, CO 80522

GUIDELINES FOR SUBMITTING REPORTS

We want to keep this mechanism timely and viable. Therefore, submit only recent news and summaries of your work. **PLEASE** limit the submission to 1 or a very few sheets (21.59 cm x 27.94 cm = 8.5×11 inches) plus a table or two; condense as much as you can (single space the text); do not staple pages together; do not number pages.

Editor's comments

This issue of the Arbovirus Information Exchange is of considerable size. Thanks to a multitude of contributors around the world, we are able to print 56 reports and a poem. Reports on the discovery of Aedes albopictus in Africa, expression of a bacterial gene in mosquitoes, detection of genetic variation in mosquitoes using random primers, and Venezuelan hemorrhagic fever, plus books reviews, announcements, and other pieces of information demonstrates the viability of arbovirology. The poem demonstrates its romance.

Thanks to the continued succor of the Centers for Disease Control (specifically, the financial support and enthusiasm of the staff of the Division of Vector-Borne Infectious Diseases and its Director, Duane Gubler) and the enthusiasm of the American Committee on Arthropod-borne Viruses (ACAV), the Arbovirus Information Exchange continues to allow arbovirologists to swap information, let others know what each group is doing, stay in touch personally, and help stay professionally abreast. Since this newsletter was established, a revolution in information exchange has taken place, one that is far beyond what anyone could have imagined 30 years ago. Nevertheless, it is personal interchange that has served as a foundation of arbovirology. I trust that this will continue.

For the time being, and at the request of the Chairman of the Executive Council of the ACAV (Joel Dalrymple, re-elected to the post during the annual meeting earlier this month), I will remain as Editor. However, there are so many competent, enthusiastic, and hardworking people active on the ACAV that I cannot imagine there would be a problem replacing me very quickly, should the situation change. I am gratified that so many have responded to my requests and reminders to submit reports; the Arbovirus Information Exchange exists for and because of you.

ANNOUNCEMENT

The Third International Symposium on Arboviruses and Hantaviruses in <u>Mediterranean Countries</u> will be held in Cortina, Italy 25-29 March, 1992. Here is an opportunity to discuss the latest information about these viruses and to do a bit of skiing. For further information, write: Organizing Secretariat, Third International Symposium on Arboviruses, MACI, P.O. Box 6164, Roma Prati 00100, Italy (telephone: 06-589-7084; fax 06-574-1762 or 06-445-3904.



AMERICAN SOCIETY OF TROPICAL MEDICINE AND HYGIENE

September 23, 1991

Mr. Thomas M. Yuill

Assoc. Dean and Professor

Madison, Wisocnsin 53706

2015 Linden Drive West

University of Wisconsin-Madison School of Veterinary Medicine RICHARD J. BURK, JR., Executive Director 8000 Westpark Drive, Suite 130

McLean, VA 22102 Telephone: (703) 790-1745 Telex: 90-1811 FAX: (703) 790-9063

OCT 2 1991 School of Veturinary Modicine

Dear Mr. Yuill:

Below please find the breakdown for the ACAV Treasury, Scherer Memorial, and the Young Memorial. I hope that these figures will help you with your report to the committee.

BALANCES AS OF 8/31/91

ACCOUNT	BALANCE 12/31/89	AC	TIVITY 1990	ACTIVITY JAN-AUG 91	CURRENT BALANCE
ACAV	6722.91	TIES PURCH INT	2120.00 (3278.38) 403.37	1125.00 .00 369.83	7462.73
SCHERER MEMORIAL	3325.91	INT	199.55	183.32	3708.78
YOUNG MEMORIAL	812.31	INT	48.74	44.77	905.82

Please feel free to call me concerning any of the breakdown figures.

Sincerely,

Deborah A. Reitz – Fiscal Administrator

cc: R. Burk J. Ravdin

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Review

Classification and Nomenclature of Viruses: Fifth Report of the International Committee on Taxonomy of Viruses

Archives of Virology/Supplementum 2

R.I.B. Francki, C.M. Fauquet, D.L. Knudson, and F. Brown (Eds) Springer-Verlag, Wien, New York, 1991, 450 p., DM 110

With ever increasing number of viruses discovered and characterized every year it is an obvious necessity that they are given appropriate names and classified in a timely fashion. The International Committee on Taxonomy of Viruses (ICTV) is charged with the immense task of updating virus taxonomy every three years based on evolving knowledge on virus structure, genetic organization, replication strategies, epidemiological data, etc. The last official ICTV report was published in 1982, nearly a decade ago in <u>Intervirology</u>. Understandably, the present report has been much awaited by the community of virologists as a single current authentic source of information on viral taxonomy.

The 5th report has been published as a supplementum to <u>Archives of</u> <u>Virology</u>, the newly designated official journal of the virology division of the International Union of Microbial Societies. While it generally follows similar format as that of its predecessor, several improvements including stylistic were made in the page layouts, sub-headings, etc. In addition to taxonomic description of viruses, the book's beginning sections contain relevant information about the rules for virus classification, mechanistic aspects of the workings of the sub-committees, format for submission of new taxonomic proposals, names of members of ICTV, etc. This information is very educative and useful for any virologist who is considering placing a newly discovered virus into its proper taxonomic position.

Classified are 2430 different viruses into 73 families and groups. Of these, 19 are new families or groups which encompass viruses of plants, animals, fungi and bacteria. The descriptions for each virus family/group under the subheadings "properties of the virus particle, replication and biological aspects" have been greatly expanded and some were completely rewritten. The presentations are concise, up to the point and current. Where available, English vernacular names were provided for each virus group and genera. Each genus is typified by a well characterized prototype virus, and where appropriate each genus is subdivided into subgroups. List of other members of the genera are included in addition to possible members. Overall, the data are well organized and logically presented.

Animal virologists will quickly realize many changes, additions, and deletions in the present report. A few examples: A new order Mononegavirales was created and a new family Hepadnaviridae came into existence. Subfamilies Paramyxovirinae and Pneumovirinae were established under the family Paramyxoviridae. <u>Pestivirus</u> genus was transferred to the Flaviviridae family. A new genus <u>Tospovirus</u> which infects plants via thrips has been established and an old genus, <u>Ukuvirus</u> was consolidated into the <u>Phlebovirus</u> in the family Bunyaviridae. Based on the new information about the genomic organization of hepatitis C virus, a new genus was created under the Flaviviridae family. Similarly, Dhori and Thogoto viruses were formally initiated into the Orthomyxoviridae family. In the family Retroviridae, the three previously existing subfamilies Onocovirinae, Lentivirinae, and Spumavirinae were eliminated, and instead, the existing viruses were grouped into seven genera.

The individual viruses and virus families are well indexed. For the first time, names of contributors responsible for each family are indicated. There are numerous typographical errors and occasional not-so-minor errors: for example, in the virus index on page 424, bovine immunodeficiency and leukemia viruses are listed under Reoviridae when they should be listed as belonging to the family Retroviridae. Overall, the fifth report presents the classification of viruses in a systematic manner with a lot of useful information. The book is moderately priced and deserves shelf space in every virologist's office.

> Ramesh Akkina Dept. Microbiology Colorado State University Fort Collins, Colorado

Book: "Studies on the natural history of yellow fever in Trinidad", 1991, edited by Elisha S. Tikasingh.

This 188 page summary is the first in a series of monographs to be published by the Caribbean Epidemiology Centre. Dedicated to the late Dr. Wilbur G. Downs, with a Preface by Franklin White and an Introduction by the Editor, the book is divided into six parts: History, The 1954 Outbreak, The 1978-1980 Outbreak, The Interepidemic/ Interepizootic Years 1980-1988, The 1988-1989 Epizootic, and The Future.

From the titles of these sections one can begin to understand the remarkable and well-studied epidemiology of this disease in Trinidad. Epidemic yellow fever was first described in Trinidad in 1793, with periodic outbreaks since then. Beginning with a brief history of yellow fever in Trinidad this book methodically and succinctly takes the reader from then to now, stopping along the way to provide pertinent data, references, and summaries. For those of us interesting in this fascinating and important disease of the Americas and Africa, this is a well edited and useful publication, if not handbook.

Cost of the soft cover edition is a nominal \$12.00 U.S. (checks made payable to CAREC) and the book can be obtained through the CAREC Librarian, CAREC (PAHO/WHO), P.O. Box 164, Port of Spain, Trinidad, West Indies.

(reviewed by C.H. Calisher)

ROYAL SOCIETY OF TROPICAL MEDICINE AND HYGIENE

Objectives of the Society

The objectives of the Society are to promote health and to advance the study, control and prevention of disease in man and other animals in warm climates, to facilitate discussion and the exchange of information among those who are interested in tropical diseases, and generally to promote the work of those interested in these objectives.

Fellowship

All registered medical and veterinary practitioners, scientists and others interested in the objectives of the Society whose qualifications are deemed satisfactory by the Council, are eligible for election as Fellows. The names of candidates should be submitted on the form provided for the purpose, which must be signed by two Fellows, one of whom must have personal knowledge of the nominee.

Anyone who desires to become a candidate for the Fellowship of the Society should write to the Honorary Secretaries at the address below, or to the Editor, <u>Arbovirus Information Exchange</u>.

Subscriptions

The annual subscription by Fellows becomes due in advance on lst April each year. There is a reduced subscription for <u>bone fide</u> students which is limited to 3 years. The annual subscripton for 1992 is $\pounds40.00$ and for students is $\pounds25.00$.

Payment of the subscription may be made by:

- a) cheque in sterling (including European Community cheques);
- b) cheque in equivalent U.S. or Canadian dollars (but <u>not</u> sterling cheques drawn on US banks);

c) draft direct to our bank (sterling or dollars):

National Westminster Bank plc, 1 Cavendish Square, London W1A 4NU.

Quote Sortcode 60-40-02, Account No. 24814237;

- d) standing order (please write to us for a form);
- e) credit card: Access, Eurocard, Mastercard, Visa.

Meetings

Meetings of the Society are normally held on the third Thursday of every month (except July, August and September) usually at Manson House unless otherwise notified.

Publications

The Society has two regular publications - the <u>Transactions</u> which is issued 6 times annually (No.1 of each volume in February) and the "Yearbook" issued annually in October. These are sent without charge to each Fellow of the Society whose annual subscription is not in arrears. Both publications are available for purchase by non-Fellows, libraries and departments.

Manson House

The headquarters and offices of the Society are at Manson House, 26 Portland Place, London W1N 4EY (Tel: 071 580 2127: Fax: 071 436 1389), where any Fellow may obtain information from the staff who are in attendance from 9.00 a.m. to 4.00 p.m. Monday to Friday.

Local Secretaries

Local Secretaries are appointed by Council to act on behalf of the Society in various parts of the world; the Editor of <u>Arbovirus Information Exchange</u> is the Local Secretary for Colorado, USA.

DENGUE 2 OUTBREAK IN ARAGUAINA, TOCANTINS STATE, BRAZIL.

Pedro F.C. VASCONCELOS, ELIZABETH S. TRAVASSOS DA ROSA, Jorge F. S. TRAVASSOS DA ROSA, Ronaldo B. FREITAS and Amélia P.A. TRAVASSOS DA ROSA. Seção de Virus, Instituto Evandro Chagas. Fundação Nacional de Saúde-MS. Av. Almirante Barroso, 492, 66065, Belém-PA.

report the first outbreak of dengue fever caused by dengue 2 ωe (DEN 2) in Araguaina, Tocantins State (Figure 1). Four hundred people of 74 families, living at S. João, Araguaina Sul and Neblina districts were questioned and then bled, in order to obtain sera to test for anti-dengue antibodies. If a person was sick, a small quantity of blood was collected for virus isolation. The main clinical picture of disease was characterized by fever, headache, myalgias, arthralgias and skin rash. The diagnosis of infection was made by both virus isolation into Aedes albopictus (C6/36) cells and serology, by Hemagglutination-inhibition (HI) and IgM capture enzyme immuno assay (MAC ELISA). Five strains of DEN 2 were isolated, and another 111 infections were diagnosed serologically (IgM positive). The positivity rate of the samples was 29% (116/400), while that of the families was 66.2% (45/72), where at least one member of the each family was infected. It was also detected 10.2% assymptomatic infections. With regard to age all groups were affected. By other hand it was detected further significative differences forward the female sex. Assuming that the sample of 400 persons is representative of the entire population of Araguaina, it may be concluded that there were 90,000 infections among the 300,000 inhabitants of Araguaina, between march 15 and may 31, 1991.

This is the first epidemic of DEN 2 in the Brazilian Amazonian region, as well as the first evidence of the spread of the serotype outside Rio de Janeiro State.

Note: In addition, seventeen more strains of DEN 2 were isolated from people not included in the familiar survey. Aedes aegypti was widely distributed in the usual domestic breeding sites.

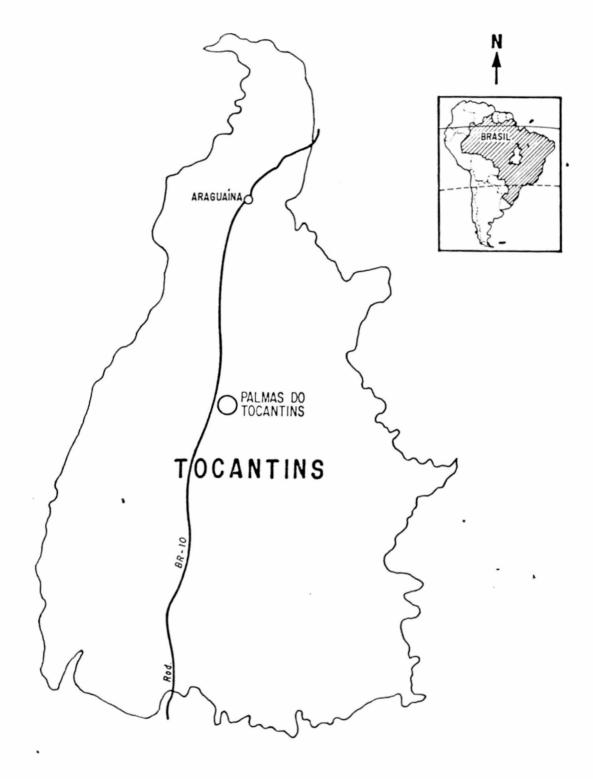


Figure 1: Map of Tocantins State, showing Araguaina Citv Dengue 2 outbreak occurred.

A 10 µL ULTRAMICROELISA FOR DETECTION OF HUMAN IgM IMMUNOGLOBULIN AGAINST DENGUE VIRUS

Pelegrino, J.L., Laferté. J., Guzmán, M.G., Vázquez, S. González, G. and Valdivia, A.

PEDRO KOURI INSTITUTE OF TROPICAL MEDICINE. P.O. Box 601, LA HABANA, CUBA.

During last years an increasing in Dengue virus infection cases have been observed even in areas under previous efficient control.

Dengue serological diagnosis have been focused on the classic haemagglutination inhibition technique, complement fixation and viral neutralization; nevertheless, the use the third generation immunoassays as ELISA have played an important role due a remarkable easy performance and sensitivity. On the other hand, assays based on IgM capture have the advantage of detection of recently acquired infections.

We have set up a capture 10 μ L ultramicroELISA in automated microtitration system using a 96 (25 μ L) well PVC plates coated with goat affinity purified polyclonal anti human IgM antibody. A blocking step consisting of bovine albumin 4% in carbonate/bicarbonate buffer was introduced.

The antigen preparation was obtained from suckling mouse brains and extracted by the sucrose-acetone method and adjusted to 16 haemagglutination units. As conjugate we used a monoclonal anti Dengue complex antibody bound to alkaline phosphatase. A fluorigenic substrate was used for developing the reaction (4 methyl umbelypheryl phosphate).

The automatic fluorescence readings were analyzed by a coupled microcomputer in which a parallel statistic quality control processing have been done. The test positive criteria have been established 260 human sera from Dengue patiens and blood donors were studied. Results were compared with MAC ELISA².A 100% of coincidence in both tests was obtained. The UltramicroElisa has the advantage of massive screening using ultra micro volumes of reaction.

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- 1.-Otero, A., Sarracent, J., Fernandez Yero, J.L. and Rodriguez, I. (1984) A 10 µL ultramicroELISA for the detection of monoclonal antibodies against human alphafetoprotein. Hybridoma 3: 391.
- 2.-Fernández, R.J. and Vázquez, S. (1990) Serological diagnosis of Dengue by an ELISA inhibition method (EIM). Mem. Inst. Oswaldo Cruz, Rio de Janeiro vol 85 (3): 347.

PURIFICATION OF SPECIFIC HUMAN IGG AND IGM FOR THE DETECTION OF DENGUE ANTIBODY LEVELS IN HUMAN SERUM.

ANSELMO OTERO, JOSE L. PELEGRINO, SUSANA VAZQUEZ AND JOSE LAFERTE. PEDRO KOURI INSTITUTE OF TROPICAL MEDICINE P.O. Box 601, LA HABANA, CUBA.

The use of human antibodies in immunoassay systems has the advantage of utilize the specificity reached in natural outbreaks against the native antigen both in direct or indirect assays for detecting antibody levels including subclasses¹. We have used purified antibodies from high titre human serum mixtures against Dengue viruses determined by Hemagglutination Inhibition test as a conjugate for the last step of an indirect microELISA. The separation was started from ammonium sulfate precipitate which was loaded onto a Mono Q HR 5/5 column (FPLC System). We found a remarkable lost of specific activity of both IgG and IgM peaks when run conditions were set up as (KH2/K2H)PO4 10 mM/L pH 6.8 in the sample application (IgG) and the same buffer plus NaCl 1 M/L for the elution (IgMD. The flow rate was 1 mL/min and the sensitivity of 1 AUFS. After this, we changed conditions to (Na2H/NaHz)PO4 10 mM/L pH 6.5 in the sample application and the same buffer plus NaCl 1 M/L for the elution, keeping the temperature all the time at 4° C. The IgG peak, when pooled, shown about 95% of its original activity with a high degree of purity as revealed by SDS-PAGE (two bands) and was successfully used as conjugated antibody to enzyme peroxidase in an indirect microELISA. The IgM fraction was used as positive control in a lgM capture assay.

We also evaluated the purification conditions for separating IgG and IgM directly from human serum² and we found a very high specific activity recovery but the yield was, of course, strongly related with the ionic interchange capacity of the column (analytical scale). On the other hand the high concentration of serum loaded onto the column definitively affects the working life of the matrix. In this case would be more reasonable the use of a matrix having a larger capacity as Q Sepharose Fast Flow.

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MONOCLONAL ANTIBODIES THAT RECOGNIZE A CUBAN DENGUE STRAIN.

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In May 1981, Cuba report a great Dengue Hemorrhagic Fever (DHF) epidemic caused by Dengue 2 virus. It was the first Dengue Hemorrhagic outbreak in the America Region. Since May to October 1981, some Dengue 2 strains were isolated in Tropical Medicine Institute of Havana, Cuba, from Dengue patients and infected mosquitoes.

Taking into account the usefulness of monoclonal antibodies (Mabs) for diagnosis and molecular characterization, we have generated two different Mabs against A35 strain of Dengue 2 virus which was isolated from a patient with DHF grade II using newborn mice.

For the immunization step female Balb/c mice were injected with 200 μ l of macerated mice brain (20 % in Hank's saline solution) from infected mice with A35 strain. The first immunization was done with complete Freund's adjuvant, while the other ones were done with incomplete Freund's adjuvant. The three immunizations were carried out by intraperitoneal route and every 15 days. Monoclonal antibodies - secreting hybridomas have been obtained from the fusion between murine myeloma cell line P3/ X63 -Ag8 6.5.3 and spleen cells from an immunized Balb/c mice.

For the characterization of the specificity of Mabs the following strains were used in our research: DEN-1 (Hawaii), DEN-2 (New Guinea), DEN-2 (A35), DEN-3 (H-87), DEN-4 (H-247), Saint Louis Encephalitis (SLE) and Yellow fever (YF). The specific antibodies produced by those hybridomas were simultaneously detected by an indirect inmunofluorescence (IFI) and the Enzyme Linked Immunosorbent Assay (ELISA). In the IFI test, fixed cells from Aedes albopictus (C6/36) line previously infected with different Dengue virus serotypes were used.

For the ELISA "sandwich" test we used a human polyclonal against Dengue also isolated by us and the different Dengue virus serotypes. Two positive hybrids were chosen by their strong recognition in both methods: A35/1 and A35/2. Mab A35/1 specifically recognized the Dengue 2 virus standard serotype and the cuban A35 strain, while did not recognize any other serotypes. On the other hand, Mab A35/2 showed an extensive recognition of all virus serotypes tested.

Our results suggests that the recognition of Mab A35/1 is strongly directed to serotype 2 of Dengue virus, while Mab A35/2 seems to recognize a group antigen of Dengue virus. At this moment these Mabs are been used in molecular biology experiments for the characterization of Dengue strains and in the standardization for diagnostic purposes. .'

REPORT FROM VIRUS LABORATORY, DEPARTMENT OF TROPICAL MEDICINE, MEDICAL RESEARCH INSTITUTE, KANAZAWA MEDICAL UNIVERSITY, UCHINADA, ISHIKAWA, JAPAN 920-02

Pathological Changes of Liver in Dengue Infections

SUSUMU HOTTA

Histopathological findings of the liver in DV infection of experimental animals (monkeys and mice) and humans (fatal cases) will be reviewed and discussed.

Monkeys, especially Japanese monkeys <u>(Macacus fuscatus</u>), infected intracutan. or subcutan. with DV show pyrexia and viremia which however are variable and which therefore cannot necessarily be used as markers of infection. Histopathological alterations of the liver are inevitable and constant; changes of varying degrees such as fatty degeneration and cellular infiltration are manifest. It seems that the cellular damages are parallel with virulence of infecting virus; when an "attenuated" strain of DV (e.g., Mochizuki strain) is inoculated, the pathological pictures manifested are mild, whereas the changes shown in monkeys inoculated with a newly isolated (humanvirulent) virus are much severer. The liver damages, when evaluated by a "scoring" method, may be a "marker of virulence" of DV.

Tissues from DHF/DSS patients (fatal cases) exhibit pathological changes of varying sorts and degrees. Particularly, liver tissues show marked changes. Pictures to be called "midzonal necrosis" are manifest. Liver cells undergo degeneration and/or necrosis. Kupffer cells contain eosinophilic granular structures which cannot be distinguished from Councilman bodies seen in the liver of yellow fever cases. DEN antigen in the Kupffer cells can be detected by applying immunoenzyme antibody stain method. By use of anti-DEN typespecific monoclonal antibodies, types of infecting viruses can be determined. Pictures of "macrophage activation" are revealed in various tissues.

Mice succumb to encephalitis after being inoculated with DEN virus intracerebrally. This is one of the DEN virus isolation methods. However, liver damages of ic-infected mice are not marked. Moreover, an extraneural (e.g. intraperitoneal) inoculation of DV can hardly infect ordinary mice. Contrarily "nude mice" can be infected with DV through ip route, although such infectivity may differ from a particular virus strain to another. The active virus can be detected in various organs and tissues. The liver cells reveal degeneration and Kupffer cells contain eosinophilic granular structures which resemble the Councilman bodies found in the liver of humans. Specific DV antigen is detected in those cells by applying immunofluorescent or immunoenzyme stain techniques. It is to be added that similar changes are seen in liver tissues of nude mice inoculated ip with YF virus 17D strain.

In summary, the liver damage is one of the characteristic signs of DV infection commonly in monkeys, humans and nude mice. Hence DV may be called a "hepatotropic" agent and the damage of liver is perhaps a key phenomenon underlying the pathogenesis of dengue. The hepatotropism is shared by DV and YFV from the histopathological viewpoint. It has already been known that both viruses are near in their immunological properties as well as in molecular-biological characteristics such as nucleotides and/or amino acids sequences of genomic RNA.

(The human specimens were taken from collection of USA-AFIP, Washington,D.C. during the writer's visit there. He is grateful to Dr. Wear, Mr. Duckett and Mr. Bratthauer for their kindness in providing the materials. His deep thanks are also due to Dr. Russell and the scientific staff of WRAIR for their helpful suggestions and favorable arrangements.)

QUEENSLAND HEALTH - QUEENSLAND UNIVERSITY OF TECHNOLOGY CENTER FOR ARBOVIRUS REFERENCE AND RESEARCH 63 GEORGE STREET, BRISBANE, 4000 AUSTRALIA

Nuclear localization of the core protein of dengue virus.

As part of a vaccine project we developed a panel of monoclonal antibodies against the core protein of dengue 2. These react with dengue 2, dengue 4 and weakly with dengue 1 in western blots but they do not react at all with dengue 3. In indirect IFA and ELISA a similar pattern of reactivity was seen - a strong reaction with dengue 2 infected cells, a weak reaction will dengue 4, and no detectable reaction with dengue 1, dengue 3, Murray Valley encephalitis or Kunjin virus infected cells.

A noticeable feature in the IFA studies was the intense staining in the perinuclear region and in, or on, the nuclear membrane. This nuclear staining was even more distinct in CV-1 cells infected with a vaccinia construct containing cDNA coding for the dengue 2 core protein.

Another noticeable feature of the histological studies was the distinct staining patterns produced by anti-envelope and anti-core protein antibodies. While the anti-core protein antibodies produced some cytoplasmic staining and a strong granular pattern over the nucleus, the anti-envelope protein antibody stained only the cytoplasm.

A PEPSCAN assay of the reactivity of the above monoclonals with octapeptides composed of overlapping amino acid sequences from the dengue 2 core protein indicated they all reacted with the region ⁹RNTPFNMLKRE¹⁹. Although the sequence NMLKR is common to all the viruses used in the indirect IFA discussed above, Chou and Fassman and "Surface Plot" predictions suggest that secondary structure and surface accessibility in this region varies between dengue 2 and the other flaviviruses studied.

We have not been able to demonstrate, convincingly, core protein <u>in</u> the nucleus of infected cells. Although gold labelled antibody could be seen, by electron microscopy, localised, apparently specifically, to the nucleus of dengue 2 infected cells, the pattern of the deposition was unlike that seen by light microscopy.

The different patterns of localisation of the dengue envelope and core proteins raises a number of questions as to the possible role - particularly of the nuclear associated core protein - of proteins not incorporated into the virion.

R. Bulich, J. Haig, J. Aaskov

Cayenne, 1991 October 24h

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Docteur REYNES J-Marc Laboratoire d'Entomologie

> Charles H CALISHER, Ph. D. Editor Arbovirus Information Exchange

Dear Sir,

A dengue outbreak occured in July 1991 and still continues in French Guiana. The serotype DEN-2 was isolated (indirect immunofluorescence with serotype specific. MAb on serum - infected AP 61 cell cultures). The dengue activity is restricted to Cayenne, chief town of the department. For the first time, few hemorrhagic cases were reported and confirmed by virus isolation or serologically (commu. in press). An epidemiologic survey is in progress.

Sincerely yours

EXPERIMENTAL TRANSMISSION OF VENEZUELAN EQUINE ENCEPHALOMYELITIS VIRUS BY <u>AEDES</u> <u>ALBOPICTUS</u> (DIPTERA: CULICIDAE) FROM NEW ORLEANS, LOUISIANA

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Experimental studies were undertaken to ascertain the vector competence of a strain of <u>Aedes</u> <u>albopictus</u> (Skuse) collected in New Orleans, LA (Gentilly strain) for an epizootic (Trinidad donkey) strain of Venezuelan equine encephalomyelitis (VEE) virus. This strain of Ae. albopictus was significantly more susceptible to infection with VEE virus than were any of the four strains tested previously (Table 1), including two from North America and two from South America. Likewise, dissemination and transmission rates were significantly higher in the Gentilly strain. This strain was also more susceptible to infection with a second alphavirus, chikungunya (CHIK) virus, than were any of the other strains of Ae. albopictus tested. Although all three strains of Ae. albopictus tested were more susceptible to VEE virus than to CHIK virus, susceptibility to infection and dissemination with one alphavirus appeared to be directly related to susceptibility to infection and dissemination with the other virus, and may indicate shared receptor sites for the two alphaviruses in Ae. albopictus.

		Day of extrinsic incubation						
Criteria	7	14	21	<u>></u> 28	Totals			
Number tested	40	40	40	60	180			
Infection	78%	95%	83%	85%	85%			
Dissemination [*]	73%	93%	78%	85%	82%			
Dissemination (I) $^+$	94%	97%	94%	100%	97%			
Transmission	50% (8) ⁺	70% (20)	38% (26)	35% (34)	45% (88)			

Table 1. Infection, dissemination, and transmission rates by day of extrinsic incubation in the Gentilly strain of <u>Aedes</u> <u>albopictus</u> after ingestion of 10^{4.5} PFU of VEE virus.

Percentage of all mosquitoes with virus in their legs Percentage of infected mosquitoes with virus in their legs Percentage transmitting (total no. feeding) Table 1. Effect of environmental temperature on the susceptibility of <u>Aedes</u> <u>taeniorhynchus</u> to Rift Valley fever virus after ingesting 10^{5.5} plaque-forming units of virus.

Temper	rature of			
Rearing	Incubation	Number tested	Infection rate [*]	Dissemination Rate ⁺
19 ⁰ C	19 ⁰ C	119	72%	42%
19 ⁰ C	26 ⁰ C	107	67%	60%
26 ⁰ C	19 ⁰ C	140	51%	18%
26 ⁰ C	26 ⁰ C	140	45%	37%

*Percentage of mosquitoes containing virus

⁺Percentage of mosquitoes with virus recovered from their legs

Table 2. Effect of environmental temperature on the susceptibility of <u>Aedes</u> <u>taeniorhynchus</u> to Venezuelan equine encephalomyelitis virus after ingesting $10^{4.5}$ plaque-forming units of virus.

Temper	rature of			
Rearing	Incubation	Number tested	Infection rate*	Dissemination Rate ⁺
19 ⁰ C	19 ⁰ C	128	97%	76%
19 ⁰ C	26 ⁰ C	115	97%	87%
26 ⁰ C	19 ⁰ C	140	80%	41%
26 ⁰ C	26 ⁰ C	140	75%	44%

*Percentage of mosquitoes containing virus

⁺Percentage of mosquitoes with virus recovered from their legs

EFFECT OF ENVIRONMENTAL TEMPERATURE ON THE VECTOR COMPETENCE OF <u>AEDES</u> <u>TAENIORHYNCHUS</u> FOR VENEZUELAN EQUINE ENCEPHALOMYELITIS AND RIFT VALLEY FEVER VIRUSES

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Environmental temperature has long been known to affect the ability of arthropods to transmit an arbovirus, with most studies indicating decreased periods of extrinsic incubation and increased transmission rates when mosquitoes are held at warmer temperatures after an infectious blood meal. However, recent studies indicate that the temperature at which mosquito larvae are reared may influence their susceptibility to infection as adults. Therefore, we evaluated the effect of environmental temperature, both during mosquito rearing and after virus exposure, on the vector competence of Aedes taeniorhynchus mosquitoes for Venezuelan equine encephalomyelitis (VEE) and Rift Valley fever (RVF) viruses. Infection rates were significantly higher in Ae. taeniorhynchus that were exposed to VEE virus than in those exposed to RVF virus, regardless of temperature, even though mosquitoes were exposed to a 10-fold higher infectious dose with RVF virus.

Environmental temperature had a dramatic effect on the vector competence of <u>Ae</u>. <u>taeniorhynchus</u> for both viruses. Mosquitoes reared at low temperature (19°C) were significantly ($\underline{P} < 0.001$) more susceptible to infection with either virus than were those mosquitoes reared at standard temperature (26°C), regardless of the temperature at which mosquitoes were held after virus exposure (19 or 26°C) (Tables 1 and 2). In contrast, in infected mosquitoes, virus disseminated from the midgut to the hemocoel more rapidly in those mosquitoes held at 26°C than in those held at

19°C, regardless of the rearing temperature. However, once a disseminated infection was attained, environmental temperature did not appear to affect subsequent transmission of virus by bite to a susceptible hamster. Thus, a combination of low rearing temperature and warm holding temperature produced the most efficient vectors for both viruses.

FIRST CONFIRMATION OF BREEDING POPULATIONS OF AEDES ALBOPICTUS IN CONTINENTAL AFRICA

Delta State, Nigeria experienced a yellow fever (YF) epidemic from April 15 to July 20, 1991. A rapid assessment team estimated that 600-1200 cases of YF occurred in Ika Local Government Area (LGA), Delta State, with human fatalities of 300-600 (1).

In September, 1991, mosquito oviposition cups were placed in four rural communities of Ika and Aniocha LGAs, Delta State, that had experienced YF activity. The vegetation of the study area is deciduous forest referred to as derived savanna. In the four surveyed communities, areas immediately surrounding human dwellings, typically at a radius of 5-10 meters, are cleared and farmed as large gardens. Secondary forests still exist as strips stretching from the edge of each village to extensive farmlands located 2-3 kilometers away from the village. These forests are traversed by foot paths and jeep roads and include economically important trees such as kola nut, pear, breadfruit and oil-palm trees.

Mosquito oviposition cups with cloth liners were placed on the ground in the forest edge at an average distance of 200 m from human dwellings. Liners were collected after 48 hours and those with eggs of Aedes mosquitoes were sent to the Division of Vector-Borne Infectious Diseases (DVBID), Centers for Disease Control (CDC). Eggs were hatched and larvae mass reared, by locality, to the adult stage for specific identification. Ae. albopictus were present in collections from three communities: Igbodo, Owa-Alero, and Egbudu-Akah. The composition of the 271 specimens in combined collections from Igbodo and Owa-Alero follows: Ae. aegypti, 73.8 %; Ae. albopictus, 18.1 %; Ae. apicoargenteus, 4.0 %; Ae. lilii, 2.6 %; and Ae. simpsoni subgroup, 1.5 %. The 14 specimens from Eqbudu-Akah represented only two taxa: Ae. albopictus, 64%; and Ae. africanus, 36%. An additional eight adults were reared from Mbiri and all were identified as Ae. africanus.

Egg collections of <u>Ae</u>. <u>albopictus</u> from three separate localities, none of which are associated with harbors or with tire dumps, indicate that this species is well established in this area. These collections represent the first record of breeding populations of <u>Ae</u>. <u>albopictus</u> in continental Africa.

Field and/or laboratory data indicate that <u>Ae</u>. <u>albopictus</u> is susceptible to or an efficient vector of a number of viruses that currently cause human disease in Africa including dengue (DEN), YF, chikungunya, Rift Valley fever and West Nile viruses (2,3). <u>Ae</u>. <u>albopictus</u> is highly anthropophilic, able to utilize both artificial and natural containers for oviposition, and is an aggressive colonizer as demonstrated by its rapid spread and establishment in North America, Brazil and various islands of the South Pacific (4). The further spread and establishment of <u>Ae</u>. <u>albopictus</u> in Africa from established populations in Nigeria and other undetected populations seems likely based on the previous colonization history of this species. Indeed, <u>Ae</u>. <u>albopictus</u> may displace <u>Ae</u>. <u>aegypti</u> in some areas of the southern U.S. (5) and alter established arbovirus transmission cycles.

Of particular concern to public health officials is the potential role of <u>Ae</u>. <u>albopictus</u> in DEN and YF transmission. Strains of <u>Ae</u>. <u>albopictus</u> from Asia and strains introduced into North America are efficient vectors of DEN (6,7,3). In addition to its role in epidemic transmission, <u>Ae</u>. <u>albopictus</u> may act as a reservoir host and facilitate endemic transmission due to its ability to transmit dengue viruses vertically (8).

The potential role of Ae. albopictus in the more complex urban and sylvan YF transmission cycles is of particular concern. Vector competence studies on different strains of Ae. albopictus introduced into North and South America indicate that Ae. albopictus is a competent vector of YF virus (9,10). Virus transmission studies with African strains of YF virus and Ae. albopictus from Nigeria are in progress. Due to its ability to utilize a wide variety of oviposition sites, its biting habits, and its competence as a YF virus vector in the laboratory, Ae. albopictus may link the sylvan and urban YF transmission cycles. Our data indicate that Ae. albopictus already has colonized both urban and rural forested areas in Delta State. This. in conjunction with the recent YF epidemic in Delta State, suggests that the components of the above scenario are already in place in some areas of Nigeria.

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Reported by H.M. SAVAGE & B.R. MILLER (Medical Entomology-Ecology Branch, DVBID, NCID, CDC, P.O. Box 2087, Ft. Collins, CO 80522), V.I. EZIKE, & A.C.N. NWANKWO (National Arbovirus and Vectors Research Division, Federal Ministry of Health, P.O. Box 104, Enugu, Nigeria) and R. SPIEGEL (International Health Programs Office, CDC, and Combatting Childhood Communicable Diseases, USAID, 1601 Adeola Hopewell Street, Lagos, Nigeria CULICOIDES (DIPTERA: CERATOPOGONIDAE) VECTOR SPECIES IN ISR TRAPPED FROM BURROWS OF RODENTS THAT MIGHT BE POTENTIAL RESERVOIRS OF BLUETONGUE AND AKABANE VIRUSES

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The cyclic occurence of bluetongue in Israel is short, when that of Akabane is longer. Both viruses, especially blueton cause considerable economic loss, mainly by restriction breeding material exportation.

Bluetongue and probably Akabane are endemic diseases in the Mediterranean. So far their reservoir animals were not evided established, despite the long time that these pathogens known. In Israel and other Mediterranean countries the taxof of <u>C</u>. schultzei - <u>oxystoma</u> gp. was not studied in deta: Therefore it is not known if the species which exist in Mediterranean is identical to <u>C</u>. <u>oxystoma</u> Kieffer in Japan, whis a known vector of Akabane. This study was undertaken a first step in order to elucidate the role of <u>C</u>. <u>schultze</u> <u>oxystoma</u> in the epidemiology of these diseases.

Emergence traps put over rodent holes in June, July 1979 al Pezael in the Jordan Valley yielded several male <u>Culico:</u> <u>schultzei</u> - <u>oxystoma</u> gp. (subgenus Remmia), females <u>C. mosuler</u> and also other Ceratopogonidae. Suggesting the burrows are u as breeding or resting sites. <u>Culicoides oxystoma</u> Kieffer proven vector of Akabane, ephemeral fever and Kagoshima virus and a suspected vector of bluetongue. The close associat reported here, between a member of <u>Culicoides schultzei</u> <u>oxystoma</u> gp. and rodents suggest a possible cycle of the viru between a reservoir, a vector and susceptible animals such livestock. THE USE OF NUCLEIC ACID PROBES FOR THE DETECTION OF BLUETONGUE VIRUS

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The <u>Orbiviruses</u> constitute a large and diverse genus of the <u>Reoviridae</u> family. They all have a genome of segmented doublestranded RNA (Gonzalez and Knudson, 1988).

Viruses belonging to the <u>Orbivirus</u> genus infect a wide range of domestic and wild ruminants including sheep, cattle, deer, antelope,goats, buffalo, horses and others (Squire <u>et al</u>.,1987). Consequently orbivirus infections could lead to the loss of domestic animals adversely affecting the agricultural industry.

Currently diagnosis of orbivirus infections is based on a variety of serological techniques as well as on the isolation of the virus in tissue culture and embryonated eggs (Osburn <u>et al</u>.,1981, Squire <u>et al</u>.,1987). These techniques are very time consuming.

Little progress has been made, however in the application of new technology to identify orbiviruses in tissue culture cells, blood, animal tissue or insect vectors.

The emphasis of research in our laboratory is on the <u>orbivirus</u> group of organisms and my personal research is more on bluetongue virus (BTV), the prototype of the <u>Orbivirus</u> genus. Recently all the segments of BTV have been cloned and compared with respect to their use as either serogroup-specific or serotype-specific probes (Huismans and Cloete, 1987).

A dot spot assay of ds RNA of all 24 serotypes of BTV were hybridized to 6 cloned segments of BTV. Segment 5 detected almost all the serotypes and looked as a possible group specific probe. Segments 1, 3,4 and 8 are also suitable candidates.

These investigations have shown that genome segments that encode non-structural proteins as well as proteins on the inside of the core particle are generally the most highly conserved within a particular serogroup and are therefore the most suitable candidates for a group specific probe. Roy <u>et al</u> (1985) have used segment 3, that codes for a major core protein, as probe to identify BTV in infected cells.

During BTV replication m-RNA, coded by the different genome segments, are not synthesized in equimolar amounts (Huismans and Verwoerd,1973). It can therefore be predicted that the different probes will detect virus-specified RNA with a sensitivity that reflects the relative amount of target m-RNA in the infected cell. We have used segment 5 of BTV serotype 4, coding for the nonstuctural protein NS1, as a probe to detect BTV in infected tissue culture cells. Since during replication, m-RNA of this segment is transcribed in large excess compare to all the other genome segments of the virus.

This segment was compared to other cloned BTV segments in the detection of BTV m-RNA during replication of the virus in a synchronised tissue culture system. Segment 5 detected the m-RNA at 4h p.i.in a pool of 1×10^5 infected cells and in only 600 infected cells after 24h replication. Segments 3 and 8 detected m-RNA from 8h p.i. and all the other probes from 12h p.i. This result suggests that segment 5 is the probe of choice.

The probe was applied further in the detection of BTV in infected tissue culture cells. Cells were infected with a series of different pfu and harvested after certain times p.i. Segment 5 was used as probe and at the lowest moi (1x10⁻⁵ pfu/cell) the RNA was first detected at 48h p.i.

The serogroup-specificity of the NS1 gene probe was investigated by cross-hybridization to cells infected with heterologous serogroups. It was found that the probe is group-specific under conditions of high stringency and should be able to distinguish between the different viruses even in the case of mixed infections.

At present we are using the probe in In situ hybridization studies both on infected tissue culture cells and on infected mouse brain sections. Good results are obtained and will soon be published.

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REPORT FROM THE ARTHROPOD-BORNE AND INFECTIOUS DISEASES LABORATORY, COLORADO STATE UNIVERSITY, FORT COLLINS, COLORADO.

EXPRESSION OF THE BACTERIAL GENE, CHLORAMPHENICOL ACETYL-TRANSFERASE (CAT) IN MOSQUITOES AND MOSQUITO CELLS USING RECOMBINANT SINDBIS VIRUS VECTORS.

Sindbis (SIN; Togaviridae) virus expression vectors, developed by C.M. Rice et al. (Washington University, St. Louis, MO), have been used to express CAT protein in mosquito cells (C6/36; Aedes albopictus) and adult female mosquitoes (Aedes triseriatus and Aedes aegypti). The pTRCAT expression vector is non-infectious and transcribes a self-replicating RNA containing the non-structural coding sequences of SIN infectious clone TOTO 1002. The structural genes were removed and replaced with the CAT gene. Expression vector pTE/3'2J/CAT transcribes RNA containing both SIN virus structural and nonstructural genes as well as the CAT gene. The genome of this vector contains two subgenomic RNA promoters which transcribe mRNA for the SIN structural genes (26S RNA) and the CAT gene. Genomic RNA from pTE/3'2J/CAT generates infectious recombinant virus within the appropriate cell lines.

TRCAT RNA was transfected into C6/36 cells using lipofectin; CAT protein within the cells was detected by an indirect immunofluorescence assay. Approximately one in 300 cells were shown to express the CAT polypeptide. Genomic RNA transcribed from pTE/3'2J/CAT was transfected into BHK-21 (hamster) cells to generate recombinant virus. The virus in turn has been used to infect C6/36 cells and Aedes triseriatus adult female mosquitoes. C6/36 cells were infected with an MOI of 1.0 and peak virus titer (7-8 log₁₀TCID₅₀) was detected within 24 h. Virtually all cells expressed CAT protein by indirect immunofluorescence. SIN Virus titers and CAT protein levels remained constant at 72 h postinfection. Approximately 3.0 log₁₀ TCID₅₀ of recombinant virus was intrathoracically inoculated into adult female **Aedes triseriatus** mosquitoes. Titers greater than 6.0 log₁₀ TCID₅₀ could be detected within four days post-infection. CAT polypeptide was detected at this time by indirect immunofluorescence of mosquito squashes. Virus and CAT polypeptide can be detected in mosquitoes up to 16 days after infection. CAT polypeptide production has been further confirmed by assaying CAT enzyme activity. We are currently quantifying the levels of CAT expression in C6/36 cells and adult mosquitoes. These expression vectors should be useful for introducing novel genes into mosquito cell lines as well as adult mosquitoes.

(Report from K. Olson, S. Higgs, J. Carlson, and B. Beaty)

DETECTION OF GENETIC VARIATION BETWEEN POPULATIONS OF <u>Aedes aegypti</u> MOSQUITOES BY AMPLIFICATION OF GENOMIC DNA USING RANDOM PRIMERS

Reported by: M. E. Ballinger-Crabtree and B. R. Miller, Medical Entomology-Ecology Branch, Division of Vector-Borne Infectious Diseases, Centers for Disease Control, P. O. Box 2087, Fort Collins, Colorado, USA, 80522.

Amplification of mosquito genomic DNA with random 10-base primers was used to detect genetic differences between two subspecies and eleven geographic populations of <u>Aedes aegypti</u> mosquitoes.¹(Table 1)

TABLE 1

Ae. aegypti MOSQUITOES USED IN THIS STUDY

aumanna tha	DODUL ATTON ODICIN	COUNCE	DESCRIPTION	CODE
SUBSPECIES	POPULATION ORIGIN	SOURCE	DESCRIPTION	CODE
Formosus	Ogbomosho, Nigeria	V.I. Ezike	Colonized 1990, F15	OG
	Enugu, Nigeria	V.I. Ezike	Colonized 1989	S2
	Enugu, Nigeria	V.I. Ezike	Ugbo-Owa rural area; colonized 1989	S3
	Enugu, Nigeria	V.I. Ezike	Colonized 1989	S4
	Agbor, Nigeria	V.I. Ezike	Field collected, rural township	AG
	Igbodo, Nigeria	V.I. Ezike	Field collected, deciduous forest	IG
	Entebbe, Uganda	L.G. Mukwaya	Colonized 1991, F3	EN
Aegypti	San Juan, Puerto Rico (Rexville)	D.J. Gubler	Colonized 1985	RX
	San Antonio, Texas	D.A. Eliason	Colonized 1984	SA
	Chachoengsao Province, Thailand (Village 6)	J. Edman	Colonized 1991, F1	V 6
	Townsville, Australia	I. Fanning	Colonized	TN

* Mosquitoes were originally collected from urban areas unless otherwised noted. Unless specified, year of colonization or number of colonized generations unknown.

Forty primers (Operon Technologies, Inc., Alameda, CA) were screened for usefulness in distinguishing between <u>Ae. aegypti aegypti</u> and <u>Ae. aegypti formosus</u> subspecies. Each primer was used to amplify genomic DNA from three individual mosquitoes each from the Ogbomosho and Rexville populations. Primers which generated fragments unique to all three individuals of only one of these populations were then used to amplify DNA from 10 individuals from each of the populations listed in Table 1. Certain useful fragments were selected and base pair size ranges were determined to allow for measurement error. The presence or absense of fragments within these size ranges for each individual was compared using nearest neighbor statistical analysis, a nonparametric multivariate discriminant analysis (PROC NEIGHBOR IDENTITY in SAS²).

Analysis of the frequency of occurence of 16 DNA fragments resulting from amplification of each individual with three different primers uncovered enough genetic diversity to allow 100% discrimination between the <u>aegypti</u> and <u>formosus</u> subspecies (Table 2) and 89% discrimination between the populations listed above (Table 3). This technique will be applied in the future to studies involving molecular taxonomy where sibling and cryptic species/subspecies are difficult to discriminate using existing techniques and in determination of genetic similarity between mosquito populations which will be useful in studies of the geographic movement of vector populations. Table 2 Nearest neighbor analysis: Classification of known individual mosquitoes based on 16 RAPD bands generated by amplification with primers A2, B3, and B13

	<pre># CLASSIFIED INTO GROUPS/TOTAL</pre>				
FROM	A	F			
A	40/40	0/40			
	(100)	(0)			
F	0/70	70/70			
	(0)	(100)			

A=Ae. aegypti aegypti; F=Ae. aegypti formosus

Table 3

Nearest neighbor analysis: Percent classification of known individual mosquitoes into populations based on the presence or absence of 16 RAPD bands generated by amplification with primers A2, B3, and B13

	PERCENT CLASSIFIED INTO POPULATION											
FROM	OG	S2	S 3	S4	AG	IG	EN	RX	SA	V 6	TN	OTHER
OG	90	0	0	0	0	0	0	0	0	0	0	10
S2	0	90	0	0	0	0	0	0	0	0	0	10
S3	0	0	80	0	0	0	0	0	0	0	0	20
S4	0	0	0	100	0	0	0	0	0	0	0	0
AG	0	0	0	0	90	0	0	0	0	0	0	10
IG	0	0	0	0	0	90	0	0	0	0	0	10
EN	0	0	0	0	0	0	100	0	0	0	0	0
RX	0	0	0	0	0	0	0	100	0	0	0	0
SA	0	0	0	0	0	0	0	0	70	0	0	30
V 6	0	0	0	0	0	0	0	0	0	100	0	0
TN	0	0	0	0	0	0	0	0	0	0	70	30

 Ten individuals from each population were tested. Individuals with unclassifiable band patterns were classified as other.

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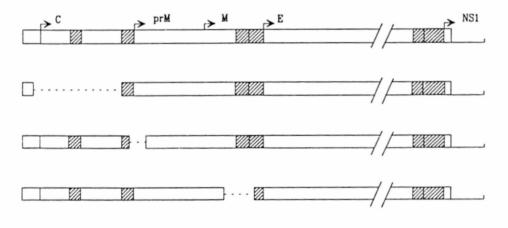
 SAS Institute. 1989. <u>SAS User's Guide: Statistics</u>. SAS Institute, Cary, NC.

The Synthesis of Dengue Virus Type 2 Structural Proteins Containing Deletions in Hydrophobic Domains

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We investigated the role of the hydrophobic domains of C and prM in the processing of DEN-2 structural proteins in intact cells. The 5' end of the genome of DEN-2 encoding the structural proteins was expressed using recombinant vaccinia virus. Three additional recombinants derived by deletion of selected dengue sequences were also expressed.



encoded hydrophobic domains

The first construct contained a deletion of nucleotides encoding most of the C protein; nucleotides encoding the hydrophobic domain at the carboxy terminus were retained. The second and third constructs contained smaller deletions of 72 bp and 129 bp encoding hydrophobic domains at the carboxy termini of C and prM respectively. We used indirect immunofluorescence and radioimmunoprecipitation to detect prM and E in cells infected with the recombinant viruses. We found that deletion of 90% of C had no apparent effect on the processing of prM and E, and that the signal sequence for E at the carboxy terminus of prM was active in the absence of 65% of the upstream signal sequence for prM located at the carboxy terminus of C. Deletion of the hydrophobic sequences preceding the amino terminus of E prevented cleavage at the prM/E junction. The results demonstrated the importance of membrane association in the cleavage of structural proteins from the flavivirus polyprotein. Cells infected with the parental recombinant virus or with the construct containing the large C deletion released the E glycoprotein into the culture medium.

Nucleotide and encoded amino acid sequences of the structural and nonstructural NS1 protein genes of a Malaysian dengue-2 virus from *Aedes albopictus*

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We report here the nucleotide and encoded amino acid sequences of the capsid (C), membrane precursor (prM), membrane (M), envelope (E) and nonstructural NS1 protein genes of a Malaysian dengue-2 (DEN-2) virus, strain P8-377. This virus was isolated in 1968 by the Hooper Foundation from a pool of *Aedes albopictus* mosquitoes from Malacca.

V/e determined the nucleotide sequences by direct RNA sequencing achieved by primer extension with synthetic oligonucleotide primers and the Sanger dideoxyribonucleotide chain termination method. Total viral RNA was isolated from infected A. albopictus C6/36 cells.

The 15-mer synthetic oligonucleotide primers used for RNA sequencing were based on the sequence of DEN-2 JAM (Deubel <u>et al.</u>, 1986) for the structural region and on the sequence of the Puerto Rican Vaccine Strain, PR 159/S1 (Hahn <u>et al.</u>, 1988) for the NS1 region.

However, because certain primers were not able to hybridize with the template RNA, three new primers were synthesized to enable the completion of the sequencing of the structural and NS1 protein genes of P8-377.

A comparative analysis was carried out at nucleotide and encoded amino acid levels between P8-377 and other DEN-2 viruses which included three Malaysian human isolates (M1, M2 and M3), NGC (New Guinea C), JAM, and S1.

At the nucleotide level, P8-377 - S1 had 95.6% similarity in the C region; P8-377 - NGC and P8-377 - JAM had 94.5% similarity in the prM region; P8-377 - NGC had 97.3% similarity in the M region; P8-377 - JAM had 94.2% similarity in the E region; and P8-377 - S1 showed 94.5% similarity in the NS1 region.

At the amino acid level, P8-377 - JAM had 100% similarity in the C region; P8-377 - S1 had 92.3% similarity in the prM region; P8-377 - JAM had more than 96.0% similarity in the M region and the E region; and P8-377 - NGC had 95.6% similarity in the NS1 region.

From the data generated, we observed that most of the base changes were transitions rather than transversions, and that codon usage was non-random. Cysteine residues and glycosylation sites were conserved.

Results also indicated that the Malaysian mosquito isolate showed greater similarity with other DEN-2 viruses at both the nucleotide and amino acid levels than with the three Malaysian human isolates.

The differences that exist between the Malaysian mosquito isolate and the Malaysian human isolates (M1, M2 and M3) could be attributed to a number of factors including host difference (arthropod and human), time of isolation [P8-377 (1968) and M1, M2 and M3 (1987)] and laboratory passage level.

Further studies will be carried out if we can isolate DEN-2 virus from mosquitoes during the current outbreak in order to explain the genetic variabilities among species of DEN-2 viruses at the level of host difference and temporal variability.

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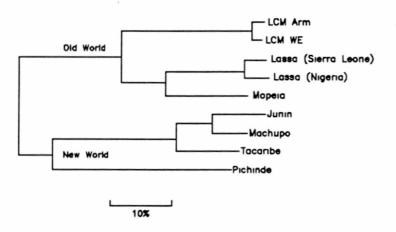
Close Relationship of Machupo Virus to Another Pathogenic South American Arenavirus, Junín

Report from the Special Pathogens Group, Public Health Laboratory Service Centre for Applied Microbiology and Research, Porton Down, Salisbury SP4 0JG, U.K.

Machupo virus is the aetiological agent of Bolivian haemorrhagic fever, a severe disease presently confined to a sparsely populated agricultural area in the northeast of the country. It is identified as a member of the New World group of the *Arenaviridae* by its serological cross-reactivity in immunofluorescence and complement fixation tests, but is clearly distinguishable from the other viruses in neutralization assays. We have now cloned and sequenced the nucleocapsid (N) protein gene of Machupo virus, and have defined more precisely its relationship to other members of the family by comparison of the derived amino acid sequences.

A cDNA library was made in the phage λ gt11 using as template RNA extracted from CV-1 cells infected 4 days previously with Machupo virus (strain AA288-77) at a multiplicity of 0.1. Reverse transcription was primed with a 19 base oligonucleotide complementary to the conserved 3'-terminus of the arenavirus S RNA segment (5'CGCACAGTGGATCCTAGGC). The library was screened at low stringency using a ³²P-labelled PCR product of 1.7 kb containing most of the sequence of the N gene of Junín virus. Hybridizing recombinant viruses were plaque-purified, and insert DNA transferred into phagemid vectors for dideoxy sequencing. The phylogenetic relationship of Machupo virus to other arenaviruses was investigated using the progressive alignment tree-building programs of Feng and Doolittle.

The cloned sequence contains 1814 nt (EMBL Accession no. X62616). In the viruscomplementary sense an open reading frame starts at nt68, coding for a protein of 564 amino acid residues which shows amino acid sequence similarities to the N proteins of other arenaviruses. The figure shows a phylogenetic tree for the arenavirus N proteins constructed by the progressive alignment method. The scale bar indicates the horizontal distance corresponding to a 10% divergence in amino acid sequence. Machupo virus N protein is most closely related to that of Junín virus, the aetiological agent of Argentine haemorrhagic fever. The Tacaribe virus N protein also appears on this branch of the tree, while that of Pichinde virus appears to be considerably more divergent.



The tree mirrors relationships among the New World viruses defined by patterns of cross-reactivity with monoclonal antibodies as well as heterologous cross-protection of experimentally infected animals. It has long been known that prior infection of guinea pigs or primates with Tacaribe virus protects against subsequent challenge with Junín virus. It has also been reported that guinea pigs which survive experimental Machupo virus infection are protected against

Junín virus challenge, while previous Pichinde virus infection confers no resistance.

Two interesting possibilities arise from the relationships among Machupo, Junín and Tacaribe viruses which are apparent when the molecular, serological, and cross-protection data are correlated. First, it seems likely that Tacaribe virus infection would protect experimental animals against Machupo virus as well as against Junín virus challenge. Secondly, a vaccine strain of Junín virus may well also confer immunity to Machupo virus challenge. An attenuated vaccine strain (Candid-1) developed for human use is currently undergoing large-scale clinical trial in the endemic region of Argentina. There would be important implications for the control of any future outbreak of Bolivian haemorrhagic fever if it could be confirmed that a vaccine at this advanced stage of development was indeed effective in protecting against Machupo virus infection.

Caroline Griffiths, Stuart Wilson, Christopher Clegg

VENEZUELAN HEMORRHAGIC FEVER

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In September 1989, an outbreak of a severe hemorrhagic illness, initially thought to be dengue hemorrhagic fever, was first recognized by local physicians in the Municipality of Guanarito, Portuguesa State, Venezuela. Cases of the disease have continued to be present, with a few patients seen each month. In the period from May 1, 1990 until March 30, 1991, a total of 104 presumed cases of the disease with 26 deaths were reported to the Ministry of Health. Most of the patients have been adults; to date, all have come from rural areas in the Municipality of Guanarito or from adjacent regions of neighboring Barinas State.

The Muncipality of Guanarito is located in the central plains (<u>llanos</u>) of Venezuela, is about 2,481 km in total area, and comprises most of the southeastern portion of Portuguesa State. The climate is tropical with a mean annual temperature of 28°C and annual rainfall of 1,300mm. Most of the 24,000 inhabitants of the municipality live in rural zones and are involved in agriculture or cattle raising.

Sera from acutely ill patients and tissue samples taken at autopsy from fatal cases were submitted to the National Institute of Hygiene in Caracas for virus isolation. Some of these samples were also examined at the Yale Arbovirus Research Unit in New Haven. Clinical specimens for virus isolation were inoculated into cultures of mosquito (C6/36) and Vero cells. After 7 to 10 days, spot slides of the cultures were prepared and examined for viral antigen by indirect fluorescent antibody test (IFAT), using dengue serotype-specific mouse monoclonal antibodies and a polyclonal hyperimmune mouse ascitic fluid made against Guanarito virus. Guanarito virus, the presumed etiologic agent of this newly recognized disease, was originally isolated by us from a fatal case in September 1990. Serological studies (CF, IFAT and PRNT) done at the Yale Arbovirus Research Unit and at the United States Army Medical Research Institute of Infectious Diseases (P.B. Jahrling, personal communication) indicate that Guanarito virus is a new member of the Tacaribe complex of the family Arenaviridae; it will be more fully described in a future publication.

Paired acute and convalescent phase sera from surviving patients were tested for antibodies to dengue and Guanarito viruses by IFAT and by IgM antibody capture enzyme immunoassay (MACEIA).

Fifteen cases of Guanarito virus infection were confirmed by virus isolation and/or antibody conversion between September 1990 and April 1991. All of the patients were rural residents of the Municipality of Guanarito. The patients ranged in age from 6 to 54 years; there were 8 males and 7 females. Fourteen of the cases were hospitalized; the duration of their illness at the time of admission ranged from 3 to 12 days (medium = 6). The most common presenting symptoms were fever, prostration, headache, arthralgia, cough, sore throat, nausea/vomiting, diarrhea, epistaxis, bleeding gums, menorrhagia (in females) and melena. In general, the initial physical findings were compatible with a viral illness and usually revealed an acutely ill patient. often dehydrated and somnolent, with one or more of the following signs: pharvngitis/tonsilitis, marked conjunctival injection, cervical lymphadenopathy, facial edema, scattered pulmonary rales and petechiae. Most of the patients had thrombocytopenia (86%) and leucopenia (79%) on admission, but their initial laboratory tests were otherwise not particularly revealing. The initial clinical impressions of the examining physicians of 14 patients were: viral syndrome (5), dengue hemorrhagic fever (4), classical dengue fever (1), viral hemorrhagic syndrome (2), lobas pneumonia (1), and convulsive syndrome (1).

Nine (60%) of the 15 confirmed cases ultimately died, despite vigorous treatment with blood, fresh plasma, concentrated platelets, fibrinogen, vitamin K, intravenous fluids and electrolytes, antibiotics, oxygen and other supportive measures. Death occurred within 1 to 6 days after hospitalization. Autopies were done on 6 of the fatal cases. Although their clinical courses varied, the gross and histopathological autopsy findings on these patients were remarkably similar and generally showed the following: (1) diffuse pulmonary edema and congestion with intraparenchymal and subpleural hemorrhages; (2) the liver was congested with focal hemorrhages and was yellow in color; (3) the heart was enlarged and showed punctiform epicardial hemorrhages; (4) the spleen was enlarged and congested; (5) the kidneys were edematous with loss of the corticomedular border; and (6) blood was present in the stomach, large and small intestines, rectum, bladder and uterus.

The 6 surviving patients recovered without serious sequellae. One reported temporary alopecia and another had a mild hearing loss. Convalescence was prolonged; one survivor reported that he was very weak and unable to work for 2 months.

Guanarito virus was recovered from the sera and/or spleens from all of the fatal cases and from 2 of the surviving patients. Specimens for culture were not obtained from the other 4 survivors; however, specific antibodies to the virus were detected in their convalescent sera and were absent in their acute phase sera. None of the patients showed cultural or serological evidence of recent dengue virus infection.

Based on the epidemiology characteristics of these cases, their clinical and laboratory features, and their association with a previously unrecognized arenavirus (Guanarito), the 15 patients appear to represent a new disease entity which we have designated "Venezuelan hemorrhagic fever" (VHF). Although VHF is quite similar in its clinical manifestations, laboratory findings and pathology to Lassa fever and to Argentine and Bolivian hemorrhagic fevers, its etiology and currently recognized geographic distribution are distinct. To date, all recognized cases of the disease have come from rural areas of 2 states (Portuguesa and Barinas) in Central Venezuela. Epidemiologic studies are now in progress to determine the prevalence, full geographic distribution, mode of transmission and risk factors for Guanarito virus infection in the Republic.

ARBOVIRUSES IN THE BRAZILIAN AMAZON REGION

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Studies in the Amazon region of Brazil carried out between 1954 and June 1991 revealed the presence of 181 different November types of arboviruses, belonging to several antigenic and taxonomic groups, some of which include certain viruses not transmitted bч arthropods. Of these arboviruses, 157 were isolated for the first in Brazil and 87 of them have been confirmed to be new to the time Thirty-three of these viruses are known to cause illness in world. man. Clinical manifestations range from mild febrile, which may 01 may not be accompagnied by skin rash and by arthralgia to severe and often fatal haemorrhagic fever. Four arboviral disease considered at the most important in public health are discussed: present as (ORD), Mayaro (MAY), Yellow fever (YF) and Dengue (DEN). Oropouche has caused extensive outbreaks involving thousands of ORO virus people. Although some patients were severely ill, there Were no fatalities. MAY virus has caused fever with arthralgia and skin Cases of jungle Yellow fever are recorded almost every year; rash. they may occur sporadically or in outbreaks. DEN outbreaks have been Boa Vista, Roraima, involving the serotypes 1 and 4 reported in this the first time for 50 years that DEN has been (Note was confirmed in Brazil) and in Araguaína, Tocantins where the serotype 2 was incriminated. It is important to emphasize that DEN virus type was first isolated in Brazil from a human travelling from Luanda, 2 february 1989. Antibodies to eastern equine Angola, in encephalomyelitis (EEE), Western equine encephalomyelitis (WEE), and Louis encephalitis (SLE), have been found in inhabitants of the St. region but only SLE has been isolated from humans. without encephalitis.

The maintenance cycles of arboviruses in the Amazon region complex, involving wild vertebrates such as primates, often are birds, rodents, bats, marsupials, sloths as well as reptiles and genera Haemagogus, Culex, amphibians. The mosquitoes Aedes, and *Wycomyia* are important Sabethes vectors of Psorophora, in the region. Haemagogus janthinomys is the principal arboviruses vector of both MAY and YF. *Culicoides paraensis* is the primary of ORO virus during urban epidemics. The only means to solve vector the problem of epidemic dengue is actually the control of its vector Aedes aegypti, since a vaccine is not yet disponible.

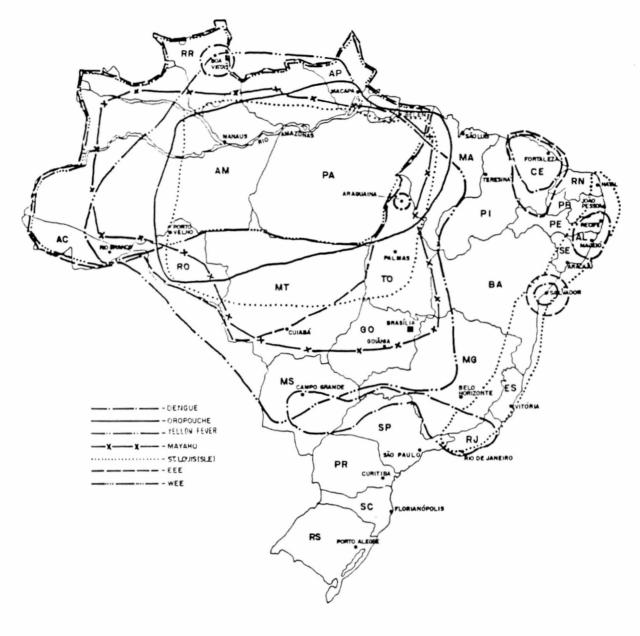


Fig. 1: Areas of YF, MAY, ORO, DEN, SLE, EEE & WEE viruses distribution in the Brazilian Amazon according to data from virological and serological studies at Instituto Evandro Chagas.

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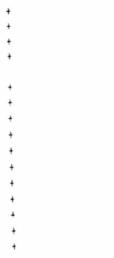
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OROPOUCHE VIRUS OUTBREAKS IN RONDONIA STATE, BRAZIL, 1991.

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An extensive outbreak of Oropouche (ORO) virus disease broke out in the Rondonia State, Brazil, during the first quarter of 1991. The was recognized in early february, in the villages of epidemic Ariquemes (9º 56' 18''S, 63º 1' 24''W) and Duro Freto do Deste (10º 30''s, 62º 16' 24''W). Clinical manifestations were 39' characterized by fever, chills, headache, myalgias, arthralgias, dizziness and nausea. The cases in general were mild, although, a patients became severely ill and were hospitalized. No few fatalities have been recorded. From 15 to 28 march a serological survey for ORO was carried out in those two localities. A total of 229 people were bled, 152 of them reported an ORO-like illness. The diagnosis procedures used were virus isolation in suckling mice and serologic tests. Both Hémagglutination inhibition (HI) as screening and an enzyme immuno assay (MAC ELISA) as a confirmatory were used. ORD was isolated from 19 febrile patients (16 from Ariquemes and 3 from Ouro Freto do Oeste). The survey revealed and HI antibody prevalence rate of 52.8% (121/229) to ORD. When tested by MAC ELISA there was a concordance of 96.2% with the HI results. THe incidence among females was 46.3% (56/121) and among males it reached 61.1% (65/108). All age groups were affected. A randomized survey carried out in Ariquemes by Health Secretary of Rondonia estimate in 58,574 cases of ORO infections. This is first outbreak of ORO disease reported in Rondonia State.

Note - Between 1961 and 1988, ORO virus, has caused extensive outbreaks in Pará, Amazonas, Amapá, Maranhão and Goiás states, Brazil, involving thousands of people. CONCOMITANT INFECTIONS BY MALARIA AND ARBOVIRUSES IN THE BRAZILIAN AMAZON REGION.

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The authors describe six cases, in which infections of arboviruses and malaria were observed in the same patient and at the same time, in the Amazon region of Brazil. The arboviruses isolated are included in family Bunyaviridae, genus Bunyavirus. Flasmodium falciparum (diagnosed by thick and thin smears) was associated with the following arboviruses: Guaroa (California serogroup) 3 times; Tacaiuma (Anopheles A serogroup) twice; Catu (Guama serogroup) once. The latter was also infected by F. vivax.

Five patients were male and one female. All were seventeen years old or more. None were born in Pará State, although all were living there. The female was a domestic help, while four men were agricultural workers and one was a commercial traveller.

The main clinical history of disease was fever with headache, chills, myalgia and arthralgia. Sometimes we noted abdominal pain, nausea, vomiting and dizziness. Jaundice was recorded in two cases of *F. falciparum* in association with Tacaiuma. The typical periodic fever associated with malaria was not observed. It was continuous. Patients were treated by SUCAM with chloroquine, primaquine, quinine, or other drugs when necessary. Five recovered quickly, but one died.

These cases are important because in Amazonia thousands of people are diagnosed and treated for malaria. About 10% of the strains of \mathcal{F} . falciparum are considered to be drug resistant. The possibility (probably much underestimated) of concomitant infections with arboviruses may obscure the effectiveness of the treatment, or may lead to an erroneus diagnosis.

It is supposed that the joint infections with malaria and Guaroa or Tacaiuma viruses were due to the fact that in Amazonia, mosquito vectors may be in either cases *Anopheles nuneztovari* or *An*. *triannulatus*.

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In mid-July, Nigerian Federal Ministry of Health (FMOH) officials were alerted to an outbreak of severe hemorrhagic fever centered in the village of Mbiri in Bendel State (now Delta State), Nigeria, approximately 300 km east of Lagos. On July 24-27, a rapid assessment team travelled to Bendel State to determine the extent and the etiology of the outbreak. This is a preliminary report of the findings.

The outbreak occurred between April 15 and July 20, 1991. Patients presented with a history sudden onset of prefrontal headache, fever, nausea, vomiting and abdominal pain. These early symptoms were often treated with chloroquine for suspected malaria. Over the next two to six days of illness, patients commonly developed marked jaundice, oliguric kidney failure, hematemesis and melena. Many severly ill patients lapsed into coma and developed intractable myoclonic seizures before death. Because of the prominence of jaundice among the patients, yellow fever (YF) virus was considered to be the most likely cause of the outbreak.

During the rapid assessment, 11 hospitals and health centers were visited. From May 1 to July 27, 169 patients with jaundice were admitted to these 11 health centers; 69 died (case fatality ratio=40%). However, only a fraction of the health centers in the affected area were visited during this initial assessment and many moribund patients reportedly left hospital and died elsewhere. These patients were not counted among the 69 deaths. A suvey of village chiefs in the area suggested that as many as 300-400 deaths and 800-1200 cases may have occurred during this outbreak.

Initial studies performed on serum samples from acutely ill patients were not diagnostic for yellow fever infection because of a high level of heterotypic flaviviral antibodies; both IgMcapture ELISA and complement-fixation tests for antibody to YF virus showed high levels of cross-reactivity and were not interpretable. Inoculations of sera into C6/36 and Vero cells failed to grow any detectable viruses. However, YF virus was cultured and identified from the sera of three acutely ill patients after intracerebral inoculation into newborn mice. Identification of these viruses was confirmed by YF-specific monclonal antibody IFA and by polymerase chain-reaction.

The areas of Bendel State affected by this outbreak were primarily agricultural. There was no evidence of transmission in nearby cities, nor was this outbreak characteristic for a sylvatic outbreak of YF: most affected villages were not located near forests and monkeys were not reported to occur in the effected area. A serosurvey of an affected village was performed and an analysis of risk factors for YF infection is ongoing. The unusual ecology of this YF outbreak suggests that YF transmission in Nigeria may differ from that of classical sylvatic YF.

(Reported by the rapid assessment team--University of Ibadan: Oyewole Tomori, A. Anoja; Nigerian Federal Ministry of Health: Abdul Nasidi, Femi Oyewole, Y. Saka, V. Eziki; USAID/CCCD, Lagos: Richard Spiegel, U.S. Centers for Disease Control, Division of Vector-Borne Infectious Diseases: Patrick Moore, Duane Gubler)

Arbovirus Surveillance Using Multiple Small Sentinel Chicken Flocks

Report from the Arbovirus Research Unit, Department of Biomedical and Environmental Health Sciences, School of Public Health, University of California, Berkeley, CA 94720

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A major component of the California Encephalitis Virus Surveillance Program is the monthly testing of serum samples from flocks of 20-25 sentinel chickens located throughout the state. Recent outbreaks of St. Louis encephalitis (SLE) in urban areas indicated the need to find alternate methods to detect transmission of arboviruses in ecological settings where large sentinel chicken flocks are not practical. In 1989 we began to evaluate the usefulness of smaller sentinel flocks of 5-10 chickens as a means to detect more accurately the occurrence and spread of virus activity in a populated region. Although initial efforts were only marginally successful, due to very low levels of virus activity, our experience in 1991 has demonstrated the effectiveness of small sentinel flocks in tracing the spread of both SLE and western equine encephalomyelitis (WEE) viruses from an enzootic transmission focus in the Coachella Valley of southeastern California.

Three "regular" sentinel flocks (20 chickens each) at Mecca, Thermal and Indio in Riverside County were augmented this year by 12 "miniflocks" (10 chickens each) placed along the north shore of the Salton Sea and extending outward to surround the city of Mecca. WEE virus was detected initially in a pool of <u>Culex tarsalis</u> mosquitoes collected at a miniflock site in Salton Sea State Park, a known focus of WEE and SLE virus activity. By late June, chickens in 3 neighboring flocks within 13 km of the initial WEE virus detection had become seropositive for WEE virus. In the July bleeding, WEE virus infection was detected at 3 additional sites at distances of up to 28 km. The spread of WEE virus activity appeared to follow the lake shore and extend northward along the Whitewater River channel, and encompassed 12 flocks at distances of up to 34 km by late August; 85 chickens in 13 of the total 15 flocks had seroconverted to WEE virus by late September.

A similar pattern of spread was observed for SLE virus, which was first detected in July at the Salton Sea State Park site. Three neighboring flocks were infected by August, and another 4 by September, for a total of 60 SLEpositive chickens within 20 km of the Park.

Most seropositive chickens were replaced with uninfected chickens the month after they tested positive, thus allowing 2 consecutive samples to confirm the test results. In a year of widespread virus activity, replacement chickens which have not been exposed to virus infection must be brought in from distant sources, adding to the cost of the program. However, if the goal of the surveillance program is simply to detect the onset of virus activity in a known focus of enzootic transmission, routine replacement of positive chickens would not be necessary.

Arbovirus surveillance in NSW, Australia, 1989-91.

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The annual arbovirus surveillance programme for the state of NSW in southeastern Australia monitors mosquito populations and activity of arboviruses through virus isolation from mosquitoes, serological testing of sentinel chickens, and collation of data on human cases. The field and laboratory programme covers both inland and coastal regions (although the sentinel chickens are confined to the inland localities) from October through April.

Weekly collections of mosquitoes from 2 sites at each of up to 30 localities (see map) are received alive for identification, counting and processing for viruses. Population profiles of the dominant species are constructed for each site.

The mosquito collections are pooled for species, sex, date and collection site. The pools are tested for the presence of flavivirus antigen by ELISA, and processed for virus isolation through mosquito and vertebrate cell culture. The identification of isolates is undertaken by ELISA and neutralisation tests for alpha- (Barmah Forest, Ross River, Sindbis), bunya- (Gan Gan, Mapputta, Termeil, Trubanaman), and flavi- (Alfuy, Edge Hill, Kokobera, Kunjin, Murray Valley Encephalitis, Stratford) viruses.

Sentinel chickens at up to 14 inland sites are bled weekly or monthly (depending on site). The sera are tested for the presence of flavivirus antibodies by indirect ELISA; seroconversions are confirmed and the eliciting virus is identified by neutralisation tests. In 1990/91 the chickens were also tested for antibodies to Sindbis and Barmah Forest viruses, although not for Ross River virus infection because chickens do not exhibit an appropriate antibody response to this virus.

Human serological diagnosis is based on detection of specific IgM antibodies by ELISA; data relating to locality, age, sex, and symptoms are collected from various laboratories and collated for epidemiological analysis.

The virus isolates from mosquitoes, and the chicken and human infection data for the past 3 years according to locality are summarised in Table 1.

Ross River virus has been active throughout the state each year since 1988/89 as evidenced by both human serology and mosquito isolations. Human infections were recorded in all surveillance areas, although the north coast and the southwest of the state (e.g. Griffith) were the regions with greatest incidence. Sindbis virus was recovered from the mosquitoes each year and was widely distributed in the inland of the state although there was only a single isolation from the coast (at Maclean on the far north coast) during the 3 years; the chicken sentinels were tested for antibody to this virus in 1990/91 only and seroconversions were shown in the Darling (Menindee), Lachlan (Condobolin, Lake Cargelligo), Murrumbidgee (Griffith, Hay, Leeton) and Murray (Wentworth) River basins, confirming the mosquito results. Sindbis virus infection in clinically-derived human sera was not reported in any year.

Barmah Forest virus infection in humans was confirmed from the Byron Bay (Brunswick Heads), Wee Waa (Narrabri), Leeton (Wagga Wagga) and Walgett (Lightning Ridge) localities; no isolates were obtained from mosquitoes and no sentinel chickens seroconverted to this virus.

Kunjin virus was detected during the 1990/91 period only, when it was active in the Darling, Lachlan, Murrumbidgee and Murray River regions as evidenced by its isolation from or detection in mosquitoes (Bourke, Condoblin, Lake Cargelligo, Leeton and Wentworth), the detection of antibodies in sentinel chickens (Bourke, Hay, Leeton, Wentworth), and confirmed human cases from the Wentworth and Griffith (Coleambally) areas. This is the first time that Kunjin virus has been detected in NSW since a seroconversion in sentinel chickens in early 1982.

There was also an unspecified flavivirus detected in mosquitoes from Wentworth and a human infection with unspecified flavivirus from Deniliquin.

Trubanaman virus activity was detected once only, it was isolated from mosquitoes collected at Wentworth in 1989/90. No human infections were reported.

A number of virus isolates that could not be identified as any of the above-mentioned alpha-, flavi-, and bunyaviruses were obtained. These are undergoing investigation by Dr C. Calisher at the CDC arbovirus reference laboratory in Fort Collins, USA.

The mosquitoes <u>Aedes vigilax</u> and <u>Culex annulirostris</u> continue to be implicated as the most important arbovirus vectors in the coastal and inland regions, respectively, of NSW. Ross River, Sindbis and Kunjin viruses were most often obtained from <u>Cx</u> <u>annulirostris</u> for the inland collection areas, while <u>Ae vigilax</u> was the most common source of Ross River virus in coastal areas.

Mosquito populations over the past 3 years have not been unusually high in regional terms but it is difficult to generalise for the whole state. Overall numbers and species composition vary with locality. Both <u>Ae vigilax</u> and <u>Cx</u> <u>annulirostris</u> exhibit seasonal activity peaking generally from mid-summer through early-autumn. A sample of the collection data reflecting relative abundance of the principal vector species is presented in Table 2. <u>Cx annulirostris</u> populations in inland areas with extensive irrigated agriculture can be relatively high in drier summers when mosquito numbers elsewhere are relatively low. Thus, notwithstanding adverse environmental conditions, some localities have substantial numbers of vectors during mid-summer each year. The prime example is Griffith, in the southwest, where nightly trap samples of <u>Cx annulirostris</u> during summer were consistently much higher than at other localities (one trap at Griffith collected >6000 female <u>Cx annulirostris</u> in one night in February 1991); the incidence of Ross River virus cases in the Griffith region appears to be higher than in other inland localities, probably as a consequence of the substantial vector populations generated by local irrigation practices.

<u>Ae vigilax</u> populations in coastal areas are primarily determined by tidal inundation of estuarine mudflats and associated marshlands (although rainfall can be an influential factor in some localities), and monthly peaks related to tide cycles can be detected within the seasonal pattern of activity. Trap numbers from individual sites following emergence of a new generation can be substantial (one trap at Batemans Bay collected >800 female <u>Ae</u> <u>vigilax</u> in one night in February 1991) although this may not be reflected in monthly trap averages.

Many people contribute to this programme. I wish to acknowledge the Environmental Health Officers of the NSW Health Department's Public Health Units who organise the regional activities, and those various local personnel who collect the mosquitoes and chicken sera at the surveillance sites; the success of the programme is dependent upon these field workers as well as the following who are involved in the laboratories: John Clancy and Peter Wells in my laboratory at Westmead Hospital identify the mosquitoes, and isolate and culture the viruses; Helen Naim and Royle Hawkes at the University of New South Wales identify the flaviviruses from mosquitoes; Michael Fennell, Ian Carter and Michael Cloonan at the Prince Henry Hospital undertake human serology for Ross River and Barmah Forest viruses and identify the alphaviruses and bunyaviruses from mosquitoes; Linda Hueston and Tony Cunningham at Westmead Hospital undertake the sentinel chicken serology, human serology for Ross River virus and flaviviruses, and collect human case data for epidemiology; we are indebted to the laboratories of Barrett & Smith, Douglass Pathology, Gribbles Pathology, Hampson and Partners, IMVS, Macquarie Pathology, Mansfield Pathology, Queensland State Health Department, Sullivan and Nicolaides, VDRL, the Victorian Department of Health, and many local practitioners for providing human case data.

<u>TABLE</u> 1: Arbovirus surveillance in NSW, 1988/89 - 90/91, by locality and method (detection of virus/antigen in mosquitoes, detection of IgG antibody in sentinel chickens, detection of IgM antibody in humans).

| LOCALITY | | | QUITOES | ANTIBODY | | | ANTIBO | | IUMANS |
|----------------|-------|-------|-----------|----------|------|-------|--------|-------|--------|
| | 88/89 | 89/90 | 90/91 | 88/89 8 | 9/90 | 90/91 | 88/89 | 89/90 | 90/9 |
| Inland | | | | | | | | | |
| Albury | -* | - | S** | nc*** | nc | nc | R | R | R |
| Balranald | nc | - | nc | nc | nc | nc | R | R | R |
| Bourke | - | R | к | - | - | ĸ | R | R | R |
| Condobolin | nc | - | K,S | nc | nc | S | R | R | R |
| Deniliquin | R,S | Х | - | - | - | - | R | R | F,R |
| Forbes | nc | - | - | nc | nc | - | R | R | R |
| Briffith | R,S,X | s,x | s,x | - | - | - | B,R | R | K,R |
| Hay , | - | - | S | - | - | K,S | R | R | R |
| Lk. Cargelligo | nc | - | K,S | nc | nc | S | R | R | R |
| Leeton | s,x | - | к,х | - | - | ĸ,s | R | R | B,R |
| Menindee | - | S | s,x | - | - | S | - | - | R |
| Moree | - | - | - | - | - | - | B,R | R | R |
| Walgett | nc | nc | - | nc | nc | - | R | R | B,R |
| Warren | - | - | - | - | - | - | R | R | R |
| Wee Waa | S | - | Х | - | - | - | R | R | B,R |
| Wentworth | X | S,T,X | K,F,R,S,) | - | - | ĸ,s | R | R | K,R |
| Coastal | | | | | | | | | |
| Ballina | nc | Х | х | nc | nc | nc | R | R | R |
| Batemans Bay | - | - | R | nc | nc | nc | R | R | R |
| Bellingen | nc | nc | - | nc | nc | n c | R | R | R |
| Byron Bay | - | - | - | nc | nc | nc | R | R | B,R |
| Coffs Harbour | - | nc | nc | nc | nc | nc | R | R | R |
| Eden | nc | - | х | nc | лc | nc | R | R | R |
| Evans Head | - | nc | nc | nc | nc | nc | R | R | R |
| Forster | - | - | - | nc | ΠC | nc | R | R | R |
| Gosford | - | - | - | nc | nc | nc | R | R | R |
| (empsey | - | - | - | nc | nc | nc | R | R | R |
| lacksville | - | - | - | nc | nc | nc | R | R | R |
| laclean | - | s,x | - | nc | лc | nc | R | R | R |
| ferimbula | nc | nc | R | nc | nc | BC | R | R | R |
| °t Macquarie | nc | nc | - | nc | nc | nc | R | R | R |
| ot Stephens | R | R | - | nc | nc | nc | R | R | R |
| Shoalhaven | - | - | - | nc | nc | nc | R | R | R |
| aree | - | R | - | nc | nc | nc | R | R | R |
| Tathra | - | - | х | nc | nc | nc | R | R | R |
| weed Heads | - | - | - | nc | nc | nc | R | R | R |
| | | | | ne | | | | | п |

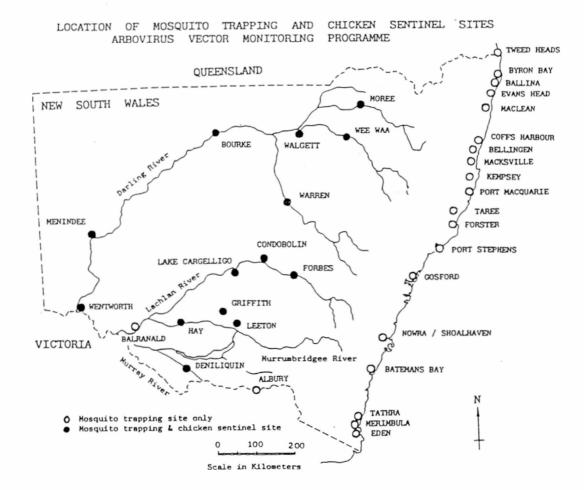
B = Barmah Forest virus, K = Kunjin virus, R = Ross River virus,

S = Sindbis virus, T = Trubanamov, virus, F = flavivirus unspecified,

X = unidentified virus not flavivirus nor Ross River, Sindbis,

Barmah Forest, Getah, Gan Gan, Mapputta, Termeil, nor Trubanaman.

*** nc = no collections



<u>TABLE 2</u>: Average number of female <u>Culex annulirostris</u> and <u>Aedes vigilax</u> in samples per night at some of the surveillance localities in the inland and coastal regions for Jan-Mar 1988/89, 1989/90 and 1990/91.

| Locality | 1988/89 | 1989/90 | 1990/91 |
|-----------------------------------|---------|---------|---------|
| LOCALICY | 1700/07 | 1707/70 | 1770/91 |
| | | | |
| <u> Inland - Cx annulirostris</u> | | | |
| Bourke | 58 | 110 | 69 |
| Deniliquin | 502 | 439 | 128 |
| Griffith | 1522 | 1037 | 1952 |
| Leeton | 669 | 171 | 386 |
| Moree | 34 | 10 | 105 |
| Wee Waa | 28 | 36 | 45 |
| Wentworth | 169 | 107 | 213 |
| | | | |
| <u>Coastal - Ae vigilax</u> | | | |
| Batemans Bay | 163 | 39 | 250 |
| Port Stephens | 17 | 84 | 22 |
| Maclean | | 11 | 73 |
| | 44 | | |
| | | | |

Central European encephalitis virus activity in Styria, Austria D. Stünzner, W. Sixl, R. Schäffler Institute of Hygiene, University of Graz, Graz, Austria M. Labuda, O. Kožuch, E. Kociánová Institute of Virology, Slovak Academy of Sciences, Bratislava V. Výrosteková Institute of Epidemiology, Faculty of Medicine, Comenius University, Bratislava, Czechoslovakia

[Report from the WHO Collaborating Centre for Arbovirus Reference and Research, Bratislava, Czechoslovakia]

In collaboration between Institute of Virology, Bratislava and Institute of Hygiene, Graz ticks and mammals were collected in May, 1990 for the detection of tick-borne pathogens.

For a survey of Central European encephalitis [CEE] virus, Ixodes ricinus adults and nymphs were collected by flagging the vegetation with a white cotton blanket. In total, 3,404 Ixodes ricinus ticks were collected in 12 localities of Styria. From 2,445 nymphs, 9 CEE virus isolates were recovered. From 451 tick females, 4 isolates and from 508 males, 2 isolates were obtained. Minimal field infection rate reached 4.4 virus containing ticks in 1,000 collected ticks [Table 1].

A serosurvey was performed based on virus neutralizing antibodies detected in small mammals collected in two localities [Grambach and Wagnitz] of Styria, Austria. A high prevalence of CEE virus was demonstrated in Apodemus flavicollis [47.9%], the most abundant collected mammal species. The second abundant rodent species, Clethrionomys glareolus was also frequently in contact with CEE virus [29.4%]. Of 83 mammals tested, 57.8% were Apodemus flavicollis and 41.0% were Clethrionomys glareolus [Table 2].

These results show a much higher CEE virus activity in Styria, Austria than previously demostrated in natural foci of CEE virus in Slovakia, Czecho-slovakia.

Table 1

Central European encephalitis virus isolations from Ixodes ricinus ticks collected in Styria, Austria [May 14-18, 1990]

| Locality | Nymphs | Females | Males | Total | MFIR* |
|------------------|---------|---------|-------|----------|-------|
| Grambach | 1/501 | 0/ 84 | 1/117 | 2/702+ | 2.9 |
| Fernitz | 1/ 10 | 0/ 1 | 0/ 5 | 1/ 16 | 62.5 |
| Keinach | 1/344 | 0/ 72 | 0/ 73 | 1/489 | 2.0 |
| Preding | 0/159 | 0/ 29 | 0/ 28 | 0/216 | - |
| Zwaring | 1/154 | 0/ 13 | 0/ 21 | 1/188 | 5.3 |
| Unterpremstatten | 0/ 9 | 0/ 27 | 0/ 19 | 0/ 55 | - |
| Wagnitz | 0/ 50 | 0/ 15 | 0/ 31 | 0/ 96 | - |
| Fasslberg | 0/245 | 0/ 12 | 0/ 11 | 0/268 | - |
| Not | 0/136 | 0/ 10 | 0/ 12 | 0/158 | - |
| Kleinsemmering | 3/412 | 1/106 | 0/106 | 4/624 | 6.4 |
| Mortantsch | 2/310 | 2/ 45 | 1/ 55 | 5/410 | 12.2 |
| Kirchdorf | 0/115 | 1/ 37 | 0/ 30 | 1/182 | 5.5 |
| Grand total | 9/2,445 | 4/451 | 2/508 | 15/3,404 | 4.4 |

* Number of isolated strains / number of examined ticks

^{*} MFIR - minimal field infection rate; minimal number of infected ticks calculated for 1,000 collected ticks

Table 2

Central European encephalitis virus neutralizing antibodies in small mammals collected in two localities of Styria, Austria [May 14-18, 1990]

| Locality | Gramt | bach | Wagn | itz |
|----------------------------|-------------------------|------------|---------------------------|----------|
| Species | Animals
posit/tested | % positive | Animals %
posit/tested | positive |
| Clethrionomys
glareolus | 1/ 4 | 25.0 | 9/30 | 30.0 |
| Apodemus
flavicollis | 6/16 | 37.5 | 17/32 | 51.3 |
| Crocidura
leucodon | - | | 0/ 1 | 0 |
| In total | 7/20 | 35.0 | 26/63 | 41.3 |

Arboviral Surveillance Guidelines

The Division of Vector-Borne Infectious Diseases, NCID, CDC, is developing a document tentatively titled "Guidelines for Arboviral Surveillance Programs." The Guidelines will partially replace CDC's Vector Topics series, written in the mid-1970's. That series has not been revised, and some sections are badly out of date.

In the new Guidelines, which deal with arboviruses of major public health importance in the United States, we describe surveillance methods for arboviruses, for vectors, and for vertebrate hosts. There are brief descriptions of the ecology of each disease, the biology and behavior of the major vectors, and recommendations of appropriate trapping and sampling schemes. The target date for distribution is spring, 1992.

Report submitted by C.G. Moore, R.G. McLean, T.F. Tsai, C.J. Mitchell, C.H. Calisher, and D.J. Gubler, Division of Vector-Borne Infectious Diseases, National Center for Infectious Diseases, Centers for Disease Control, Fort Collins, Colorado, USA.

Progress Report: Central American and Caribbean Bluetongue Epidemiology Project

The Interamerican Bluetongue Team*

A regional study of bluetongue (BLU) epidemiology in the Caribbean Basin has been in progress since 1986 with the cooperation of the Central American Animal and Plant Health Organization (OIRSA), the Interamerican Institute for Cooperation in Agriculture, the Universities of Wisconsin-Madsion and Florida, the Arthropod-borne Animal Disease Research Laboratory of USDA-ARS, and the Office of International Cooperation and Development of USDA and the Ministries of Agriculture of Barbados, Costa Rica, the Dominican Republic El Salvador, Guatemala, Honduras, Jamaica, Nicaragua, Panama, Puerto Rico, Trinidad and Tobago.

In addition to elucidating the epidemiology of BLU virus in this endemic area, a long term objective is to provide information to assist regional and national veterinary services in development of procedures and policies to reduce the economic impact of bluetongue in the hemisphere.

The project design revolves around a sentinel herd system. Sentinel herds are located in all participating countries. Young ruminants are selected to join sentinel groups of 25 animals at approximately 3 months of age. Blood samples are taken monthly for serologic and virologic testing. Samples are shipped from each country via air freight to the central project laboratory in Costa Rica, where group specific antibody testing is done. Serologic results are communicated back to field personnel in time for the next monthly sampling. Samples from animals in which recent BLU virus infection is detected serologically are processed for virus isolation. BLU virus isolate positive samples are shipped to USDA ABADRL for serotyping. Vector populations are monitored on sentinel farms by use of light traps. Virus isolation is attempted from potential *Culicoides* vectors. Surveillance for suspected cases of disease is coordinated with field personnel supervising sentinel herds and with vesicular disease surveillance in the Central American countries.

As previously reported to the Arbovirus Information Exchange, BLU virus serotypes isolated from the Caribbean islands in the period 1987-1989 included 1, 3, and 12. BLU virus serotypes isolated from the Central American isthmus in the same period included 1, 3, and 6. Since 1989 we have isolated BLU virus serotypes 3, 4, 6, 8, and 17 in the Caribbean Islands and BLU serotypes 1, 3, 6, and 17 in Central America.

No confirmed cases of BLU disease have been observed and no clinical signs attributable to BLU infection have been observed in animals with lifetime residence in the region. However BLU virus has been isolated from animals coincidently with VSV virus, as well as from two groups of cattle experiencing abortions following importation from North America.

Isolates of BLU virus serotype 6 have been made from parous *Culicoides insignis* and *C. pusillus* at sites where BLU is actively circulating in sentinel animals. Additional isolates have come from from blood fed *C. filarifer*.

BLU viruses appear to have a stable endemic distribution in the region with their distribution dependent on the availability of vector habitat rather than that of ruminant animal populations.

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ARBOVIRUS LABORATORY TESTING IN DELAWARE 1990

A result of rainfall patterns with humid spring and early summer, Delaware provided favorable environment for multiplication of mosquitoes in the year of 1990. The arbovirus surveillance for 1990 season was highlighted by extensive Eastern Equine Encephalitis (EEE) virus activity which was reflected from the results of our laboratory testing below.

In all 3 counties in Delaware where significant numbers of EEE cases were detected, from early July through late October, by either virus isolations from mosquito pools or seroconversions of sentinel birds.

Eighteen of 101 mosquito pools were positive for EEE virus isolation represented 3 of 4 locations which were designated for mosquito collection. Three of the 12 sites throughout the state where sentinel chicken were bled biweekly for testing antibody to the virus, showed seroconversion, indicating a current infection during this surveillance season.

Activity of EEE virus of the year 1990 was at the highest level since the Delaware Public Health Laboratory started the testing program with the collaboration of the Mosquito Control Agent of Delaware in 1981. For years EEE viral activities were detected almost at the same areas such as Cedar Neck, Cool Spring, Wooden Hawk, Cypress Swamp, etc. The observations suggested that indigenous virus likely overwintered and maintained in nature by cycles involving wild birds, or rural mosquitoes.

Results of the study clearly suggest that the threat of an EEE virus activity in Delaware. Our results help define the area where special care of mosquito control should be focussed.

> Jong-Ho Jean, Patricia Randolph, Sugan Dee and Margaret Willey

Virology Section Delaware Public Health Laboratory Smyrna, DE. TICK-BORNE ENCEPHALITIS IN HUNGARY IN THE LAST TEN YEARS

E. Ferenczi and E. Molnár

Department of Virology, "B. Johan" National Institute of Public Health, Budapest, Hungary

About 300 patients have been hospitalized because of tick-borne encephalitis /TBE/ virus infection every year in Hungary. Nine tenths of the infections occured in the vicinity of endemic forests. /Figure 1/ The number of serologically verified cases between 1969 and 1978 was 1391. Between 1981 and 1990 2838 cases have been registered. Nevertheless, the increase in number is probably due to the improved diagnostic procedures. There are no indications which would suggest a modification in the geographical distribution. Incidence or case fatality rate of the disease. Minor changes in the frequency of TBE in different counties were observed. The highest prevalence was observed in the sounties of Zala, Somogy and Vas in both decades. In fourth place was Nógrád county which has replaced Komárom in the last 10 years.

The long-term character of the clinical disease, its prolonged convalescent period, the possible sequalae and the 1.0 to 1.5 per cent fatality rate of cases render TBE to one of the important social and national economic problems in the country. Vaccination campaigns have been organized since 1977. Between 1977 and 1990 about 147,240 doses of FSME-Immun and FSME-Immun Inject have been used for the vaccination of professionally endangered inhabitants of the endemic areas. Two week long vaccination campaigns were carried out every year in the spring. These were organized and controlled by the local sanitary-epidemiological stations. The amount of the distributed doses was proportional to the incidence of TBE in the counties. Nineteen per cent of the total amount was distributed in Zala county, 12 per cent in Somogy whereas Vas and Nógrád obtained only 8 and 4 per cent respectively.

The epidemiological data up to now are insufficient for the evaluation of vaccination effectiveness. It has to be mentioned, however, that 50 per cent of the Hungarian cases between 1969 and 1978 were diagnosed in Zala county, but this county only accounted for 22 per cent of all cases in the last ten years. In addition to other factors such as improvement of diagnostics in regions of lower prevalence, post-vaccination immunity may play a role in the relative reduction of the frequency of TBE in this county.

The seroconversion rate after vaccination with FSME-Immun was 75 per cent between 1977 and 1979. It has to be emphasized, however, that no clinical TBE was observed among individuals who had been vaccinated at that time. In contrast, several verified TBE cases could be registered among individuals who were vaccinated between 1980 and 1989 with the complete series of three vaccine doses. The number of such cases is 20 according to our vaccination data obtained from physicians of these patients or territorial sanitary-epidemiological stations, or both. Twelve of them contracted TBE within 3 years after the 3rd dose of the vaccine, eight of these were vaccinated between 1979 and 1983. This fact is consonant with the results of a retrospective serological study made in 1987 and 1988, involving a group of people vaccinated between 1980 and 1983. This group only showed a 25 per cent seropositivity. These observations raise the need of regular control of vaccination efficacy, examinations of the actual antib**go**y responses and additional tests for maintenance of antibody levels upon revaccination.

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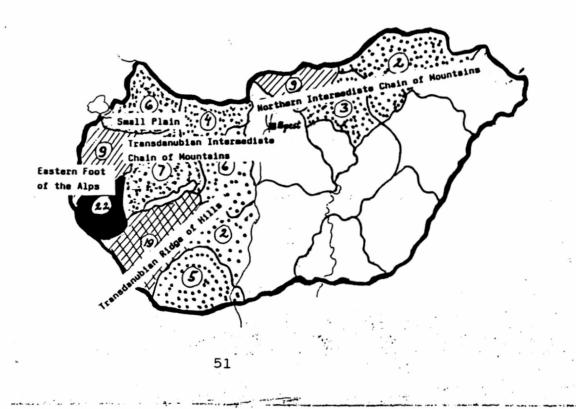
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Figure 1. SEROLOGICALLY VERIFIED TICK-BORNE ENCEPHALITIS CASES IN HUNGARY BETWEEN 1981 and 1990 (N=2838)

(Numbers indicate the per cent of cases in different geographical areas)



REPORT OF THE ARBOVIRUS RESEARCH UNIT

INSTITUT PASTEUR DE MADAGASCAR. BP 1274. TANANARIVE

RIFT VALLEY FEVER IN CENTRAL HIGHLANDS OF MADAGASCAR

(Jacques Morvan, Jean Roux)

Rift Valley fever virus (RVFV) was isolated in Madagascar in 1979 from pools of mosquitoes captured in a primary rain forest. Serological investigations in man and cattle showed a low viral circulation (prevalence of IgG antibody to RVFV inferior to 1 percent). In 1990 we described the first epizootic outbreak observed in Madagascar, among zebu cattle in the district of Fenerive on the East-coast. Since February 1991, several abortion foci in bovines were observed around Tananarive in Central Highlands (abortion rate average: 12 percent of pregnant females). For virus isolation 40 samples (organs of still-born calf, products of abortion) were inoculated into tissue culture (Vero E6, Aedes pseudoscutellaris cells, Aedes albopictus cells) and intracerebrally into 1-2 day old suckling mice. They were also tested for ELISA antigene detection using an IgM trapping. A total of 460 human sera and 873 bovine sera were collected between March and April 1991 and tested for IqM specific RVFV antibody by an ELISA IgM capture assay, and for IgG antibody by an indirect immunofluorescence assay (IFA).

22 samples were confirmed positive for Rift Valley fever by virus isolation, and viral antigene was detected from 25 samples. No virus was detected from the 120 pools of mosquitoes captured in the same areas. The results of serological investigation in man and cattle are presented in the Table.

Since the viral isolations in 1979, the RVFV circulation was low. Epizootic outbreaks appeared during 1990 with an extension to breeding countries of the Central Highlands. In the same time beside we observed a constant increase in the prevalence of RVFV specific antibodies.

The reasons of this recent and intensive spread of RVFV infection among livestock are still not well explained. Further studies are needed to assess the extent of the affected area, and to identify the cause of the epizootic. **Table:** RVFV specific antibodies in man and cattle in Central Highlands of Madagascar during epidemic outbreaks. February-April 1991.

| | IgG
(IFA) | | IgM
(ELISA) | |
|-----------------------------|--------------|------|----------------|-------|
| HUMAN SERA | | | | |
| Tananarive. March 1991 | | | | |
| Hospital patients | | | 2/135 | (1.5) |
| healthy contacts | 19/220 | (8) | 11/220 | (5) |
| Mangamila, March 1991 | 11/55 | (20) | 4/55 | (7) |
| Tsiroanomandidy, March 1991 | 2/31 | (6) | -/31 | (0) |
| Antsirabe. April 1991 | | | 3/19 | (15) |
| BOVINE SERA | | | | |
| Tananarive. March 1991 | | | | |
| Aborted females | 23/28 | (82) | 21/28 | (75) |
| Other bovines | 119/295 | (40) | 111/295 | (37) |
| Mangamila. March 1991 | | | 41/180 | (22) |
| Manjakandrina. March 1991 | 3/23 | (13) | 9/23 | (39) |
| Tsiroanomandidy, March 1991 | 3/10 | (30) | 4/10 | (40) |
| Antsirabe, April 1991 | | | 71/337 | (21) |

no/n positive/total sera

() percentage of positive sera

A REPORT FROM THE VIRUS REFERENCE LABORATORY, INC 7540 Louis Pasteur, Suite 205 San Antonio TX 78229 Tel. (512) 614 7350 FAX (512) 614 7355

The Virus Reference Laboratory, Inc. was established approximately 3 years ago in order to continue the activities of the WHO Collaborating Center for Reference and Research in Simian Viruses and the NIH Simian Virus Reference Laboratory. The principal activity of VRL was to provide a viral diagnostic service to those nonhuman primate facilities without this capability. The greatest need was testing both human and non-human primates for evidence of B virus (Herpesvirus simiae). However, over the years a multitude of other zoonotic conditions occurred and VRL provided diagnostic testing to determine the etiology. Accordingly, in addition to B virus and other herpesvirus infections (SA8, H.tamarinus, H. saimiri, CMV, varicella/zoster, lymphotrophic herpes, etc.) there were outbreaks of hepatitis, Marburg, poxvirus and the more recently retrovirus (simian AIDS) infection, simian hemorrhagic fever and filovirus infection. Although arboviruses in general have never been a major source of concern among several human-nonhuman investigators using monkeys and apes, arbovirus diseases have been recognized: yellow fever, Kyasanur forest, etc. In addition to the loss of animal life, downtime because of illness, enhanced costs due to therapy and disease control, etc., there was the continuing concern over human infection. Compounding the direct effect of infection/disease on animal colony was the environmental impact, conservation the concerns and the marked increase in costs per animal.

In an attempt to ascertain the potential threat of newly captured animals to other animals and possibly to humans, VRL has developed a test kit for the detection of antibody at the time of animal capture in the field. This kit now permits detecting the presence of antibody to B virus, retroviruses and measles virus on blood collected on provided filter paper and may be performed in the non-laboratory setting. Preliminary studies with such viruses as SLE, WEE, EEE and VEE indicates the suitability of this system for detecting the presence of arbovirus antibody. The kit is selfcontained requiring only a liter of water, forceps and a discard Individuals testing the animals should be aware of the pan. potential danger to themselves principally from B virus and take the precautions recommended for all individuals working with non human primates. Tests are completed in approximately 60 minutes and read visually. An insert supplied with the kit provides directions for performing the test along with interpretation of the results. A kit for filovirus testing will be available soon.

For information regarding availability of these kits, please contact VRL.

(Submitted by S.S. Kalter, PhD.)

Preliminary Summary of Surveillance for Mosquito-borne Encephalitis Virus Activity in California, 1991

As usual, an extensive surveillance program was conducted, involving cooperative efforts by many groups and individuals from local mosquito control agencies; the Arbovirus Research Program at the University of California at Berkeley; the California Mosquito and Vector Control Association (CMVCA) and the CMVCA Research Foundation; county and local public health departments; the California Department of Food and Agriculture; physicians and veterinarians throughout California; and three branches of the California Department of Health Services - the Infectious Disease Branch, the Environmental Management Branch, and the Viral and Rickettsial Disease Laboratory of the Division of Laboratories. A complete summary will be published in the proceedings of the California Mosquito and Vector Control Association annual conference. The CMVCA is now providing a large portion of the funding for this effort, to replace the loss of a significant part of state-funded resources.

Clinical and laboratory surveillance for human and equine cases of arboviral encephalitis, meningoencephalitis and meningitis has so far detected l human case of St. Louis encephalitis (SLE): a 39 year old man from Valinda, Los Angeles County, with onset 10/11/91 and place of exposure most probably the Sante Fe flood control basin area near Irwindale, where 14 of a 15-bird sentinel chicken flock have converted to SLE antibody-positive during September/October. There have been no confirmed cases of WEE in equines or humans.

The Arbovirus Research Program, UCB, detected numerous WEE and SLE isolates from <u>Culex tarsalis</u> during July in the Seeley area of Imperial county, and subsequent seroconversions to SLE in Seeley, Derrick, Scurlock, and Gracey in the Imperial Valley, and the Salton Sea area; and seroconversions to WEE at Adohr Farms, Thermal, Artesian Acres, Gonzalez, and Jessup in the Coachella Valley, have been confirmed by the ARPUCB and the VRDL. The VRDL also detected 3 WEE in <u>Culex quinquefasciatus</u>, 4 WEE in <u>C. tarsalis</u>, and 1 SLE in <u>Culex stigmatosoma</u>, from Los Angeles County sites. There has been no evidence thus far of activity in central or northern California.

Laboratory and field research this winter will seek even more cost-effective and rapid, early detection methods, such as filter-paper blood collections, using the best combination of mosquito pools and sentinel chicken bleedings that can be devised.

Richard W. Emmons, M.D., Ph.D. Viral and Rickettsial Disease Laboratory

REPORT FROM DEPARTMENT OF VIRUSES ECOLOGY, THE D.I.IVANOVSKY INSTITUTE OF VIROLOGY, ACADEMY OF MEDICAL SCIENCES, MOSCOW, USSR.

Accoding to current antigenic classification of the California (CAL) serogroup viruses (family Bunyaviridae, genus Bunyavirus), Inkoo (INK) virus has been placed in the California encephalitis complex within the serogroup as a subtype of the first main type of the complex, California encephalitis type, together with other four subtypes: California encephalitis (CE) virus, La Cross (LC) virus, San Angelo (SA) virus and Tahyna (TAH) virus. The second main type (virus) within the complex, Melao, is composed of 4 subtypes: Melao (MEL) virus, Jamestown Canyon (JC) virus, Keyston (KEY) virus, and Serra do Navio (SN) virus (Calisher, 1983).

An attempt was made to investigate antigenic relationship of INK virus (strain KN-3641) with other members of California encephalitis type: TAH virus (strain 92) and snowshoe hare (SSH) virus (original strain) and with members of Melao type: KEY virus (strain C 14031-33) and JC virus (strain C 14001). For this purpose double immunodiffusion precipitation (Ouhterlony) test in agar was used and considered to be succesful for distinguishing within the complex. This test was performed with antigens prepared from infected cell culture suppernatant fluid (Pig Kidney cell line) by precipitation of virus with polyethilene glycol (PEG m. w. 6000) and hyperimmune mouse ascitic fluids (HMAF). Cross-titration of two-fluid dilututions of antigens and HMAF was used Microtiter Serum Dilution Neutralization (SDN) tests with HMAF for subtyping and with selected sera from patients were performed in the same cell culture using inhibition of cytopatic effect (CPE) as indicator. Brirect sandwich Enzyme-Linked

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Immunosorbent Assay (ELISA) typing system to TAH and INK viruses was also used. ELISA typing method was performed as previously discribed (Artsob et al., 1984).

When CAL serogroup viruses were assayed by precipitation and ELISA tests (Fig. 1, 2, 3, 4), INK virus exhibited patterns of crossreactivity obvious different from other members of California encephalitis type (TAH and SSH viruses) and more similar to members of Melao type (KEY and JC viruses). When tested by SDN test, HMAF to INK virus had heterologous titer against JC virus the same as its homologous was (Tabl. 1). Results of SDN test with TAH, INK, JC and SSH viruses using sera taken from Inkoo virus infected individuals, supported the above data (Tabl. 2).

These preliminary results suggest that INK virus is probably a member of Melao type and it is antigenically closely related to JC virus.

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(M.S. Nedyalkova, A. M. Butenko, D.K. Lvov).

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TABLE 1. COMPARISON OF TAHYNA (TAH), INKOO (INK), JAMESTOWN CANYON (JC) AND KEYSTONE VIRUSES BY SERUM DILUTION NEUTRALIZATION (SDN) TEST. (approximate viruses dose in test was 100 TCID 50)

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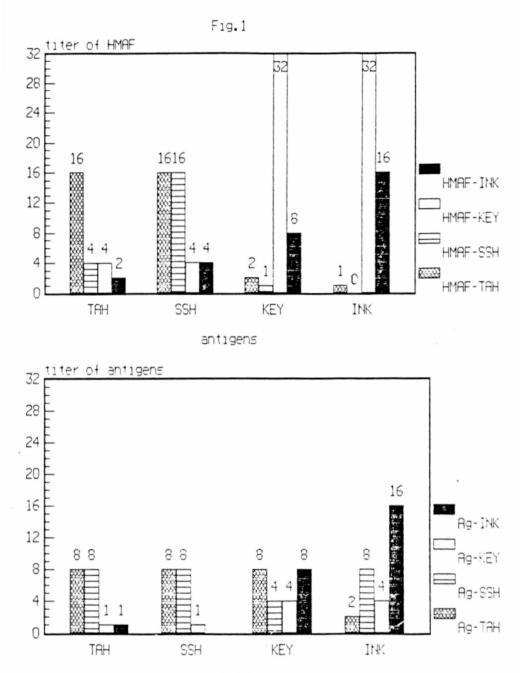
70

| Virus | ТАН | INK | JC | KEY |
|---|------|------------|------|------------|
| ŤAHYNA
(strain 92) | 2560 | 160 | 80 | nd |
| INKOO
(strain
KN-3641) | 20 | 1280 | 320 | 20 |
| JAMESTOWN
CANYON
(strain
61V-2235) | <10 | 1280 | 5120 | nd |
| KEYSTONE
(strain
B64-5587) | nd | 4 0 | nd | <u>320</u> |

Titer of hyperimmune mouse ascitic fluids to:

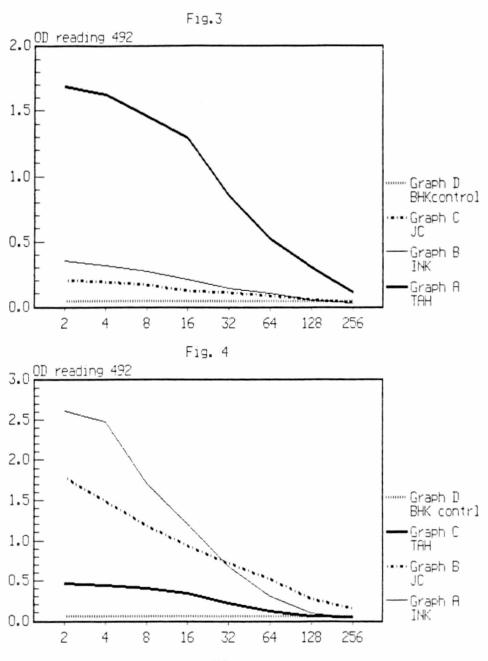
nd - not done.

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HMAF

COMPARISON OF THE CALIFORNIA SEROGROUP TAH, SSH, KEY AND INK VIRUSES BY DOUBLE IMMUNODIFFUSION PRECIPITATION TEST IN AGAR. Fig. 1. Comumns show reciprocals of highest HMAF dilution giving a visible precipitin line with homologous and heterologous antigens. Fig. 2. Columns show reciprocals of highest antigen dilution giving a visible precipitin line with homologous and heterologous antigens. 1 - only undiluted HMAF or antigen gives a visible line. 0 - no reaction.



antigen dilution

ELISA TYPING OF CALIFORNIA SEROGROUP TAHYNA (TAH), INKOO (INK)

AND JAMSTOWN CALIFORNIA SERVERVOLP TAHINA (TAH), INKOU (TNK) AND JAMSTOWN CANYON (JC) VIRUSES. Homologous and heterologous reaction of TAH, INK and JC viruses by ELISA typing system to TAH (Fig. 3) and INK (Fig. 4) viruses. Plates were coated with mouse IgG prepared from anti-TAH or anti -INK hyperimmune mouse ascitic fluid. The enzyme conjugate was horseradish peroxidasae -conjugated the same anti-TAH or anti-INK mouse IgG. Table 2. Antibody titers to four California serogroup Tahyna(TAH), Inkoo(INK), Jamestown Canyon(JC) and snowshoe hare (SSH) viruses in sera from selected patients suspected to be Inkoo virus infection by serum dilution neutralization (SDN) test.

| NN | Serum
code | | | Days
post- | a
SDN titer | | | |
|----|---------------|--------|----|---------------|----------------|-----|-----|-----|
| | No | nogran | | onset | TAH | INK | JC | SSH |
| 1. | 1150 | Moscow | 19 | 7 | 20 | 40 | 40 | 0 |
| | | | | 12 | 20 | 160 | 160 | 0 |
| 2. | 1346 | - " - | 13 | З | 40 | 160 | 80 | 20 |
| | | | | 11 | 40 | 320 | 160 | 20 |
| | | | | 22 | 40 | 80 | 160 | 10 |
| З. | 1357 | _ " _ | 17 | 6 | 40 | 320 | 160 | 10 |
| | | | | 14 | 40 | 160 | 160 | 10 |
| 4. | 1391 | _ " _ | 17 | 5 | 20 | 30 | 40 | 0 |
| | | | | 13 | 20 | 320 | 160 | 10 |
| 5. | 1412 | - " - | 30 | 5 | 80 | 80 | 80 | 10 |
| | | | | 10 | 80 | 160 | 160 | 0 |
| | | | | | | | | |

a - approximate virus dose in test was 100 TCID 50

Patients N 1, 3, 4 and 5 had an infuenza-like disease. Patient N 2 had meningitis.

A SURVEY OF SUSPECTED MOSQUITO VECTORS OF JAPANESE ENCEPHALITIS ON SAIPAN

A discrete outbreak of Japanese encephalitis (JE) occurred on Saipan, Commonwealth of Northern Mariana Islands (CNMI), during October, 1990. This is the first documented incursion of JE into Oceania since an outbreak on Guam during 1947-48. Eleven human cases were identified, all with onset within a period of four weeks (William S. Paul, personal communication). The prevalence of JE antibody in swine (96%, n=52) suggests that swine were important amplifying hosts (Wm. S. Paul, per. comm.)

We collected adult mosquitoes during September 21-27, 1991, at the same time of year that virus amplification could be expected to have occurred last year. Our collection sites are located in the general areas of case occurrence and swine seroconversions. The mosquito collection data are summarized in Table 1. Almost all of our specimens came from CDC light traps baited with dry ice (CO₂) and collections made from swine from sunset until midnight. Among the known vectors of JE from other areas where epidemics have occurred. Culex tritaeniorhynchus was the predominant species in our collections. Culex annulirostris, the probable vector on Guam during the 1947-48 outbreak made up a negligible proportion (<1%) of our collection. Almost 11% of our total collection was identified only as Culex sitiens group which includes both Cx. annulirostris and Cx. tritaeniorhynchus. An additional 35% was identified only as <u>Culex</u> (<u>Culex</u>) species because of the poor condition of the specimens. However, there is no reason to believe that the proportion of Cx. annulirostris would have been greater in these groups than in the group that was identifiable to species. Subsequent collections, currently being processed, support this conclusion.

Virus was not isolated from 119 pools of mosquitoes collected during 1991 (Table 1). We may never be certain which mosquito species was the principal vector of JE on Saipan during 1990, but results from our retrospective studies strongly suggest that it was \underline{Cx} . tritaeniorhynchus. To our knowledge, the only previous reference to the presence of this species on Saipan, is in an unpublished trip report for July 6-17, 1981, by Dr. K. Ando, an Associate Entomologist assigned to the Pacific Trust Territories by VBC/WHO. Our results indicate that \underline{Cx} . tritaeniorhynchus currently is an abundant and widely distributed species on Saipan, at least on the southern half of the island.

Report submitted by Carl J. Mitchell, Harry M. Savage, and Gordon C. Smith, Medical Entomology-Ecology Branch, Division of Vector-Borne Infectious Diseases, National Center for Infectious Diseases, Centers for Disease Control, Fort Collins; Sean P. Flood, Medical Director, Public Health, CNMI, Saipan; and Luis T. Castro and Mario Roppul, Sanitarians, CNMI, Saipan. Table 1. Summary of adult mosquito collections on Saipan, September 21-27, 1991.

| Collection Method | | | | | | | | | | | | |
|--------------------------------------|--------------|-------------|------------|--------------|------|---------------------------------------|-----------|-------------|-------------------------------|--|--|--|
| Species or group | Human
No. | bait
(%) | Pig
No. | bait*
(%) | | ght trap
CO ₂ **
(%) | To
No. | tals
(%) | No. pools tested
for virus | | | |
| <u>Aedes</u> <u>albopictus</u> | | | 1 | (<0.1) | 7 | (0.1) | 8 | (0.1) | 3 | | | |
| <u>Ae. oakleyi</u> | | | 5 | (0.2) | 32 | (0.7) | 37 | (0.5) | 5 | | | |
| <u>Ae. saipanensis</u> | | | | | 37 | (0.8) | 37 | (0.5) | 2 | | | |
| <u>Ae. vexans</u> | 123 | (100) | 338 | (13.9) | 583 | (12.4) | 1044 | (14.4) | 16 | | | |
| <u>Culex</u> annulirostris | | | 4 | (0.2) | 9 | (0.2) | 13 | (0.2) | 5 | | | |
| <u>C</u> x. <u>tritaeniorhynchus</u> | | | 995 | (40.8) | 1820 | (38.8) | 2815 | (38.8) | 36 | | | |
| <u>Cx. sitiens</u> group | | | 467 | (19.1) | 308 | (6.6) | 775 | (10.7) | 11 | | | |
| <u>Cx</u> . (<u>Cux</u> .) spp. | | | 630 | (25.8) | 1889 | (40.3) | 2519 | (34.7) | 39 | | | |
| <u>Cx. fuscanus</u> | | | | | 2 | (<0.1) | 2 | (<0.1) | 2 | | | |
| Totals | 123 | (100) | 2440 | (100) | 4687 | (100) | 7250 | (100) | 119 | | | |

*Principally night-time biting/resting collections in pig sties; a few specimens collected immediately adjacent to sties with a sweep net.

**In addition to mosquitoes, 181 Ceratopogonidae were collected and tested for virus in 2 pools with negative results.

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Arbovirus Activity in Colorado During 1991

During the summer of 1991 mosquitoes were collected in northern Colorado using CDC light traps baited with CO₂. Weekly collections were made between July 26 and August 29, 1991. Each collection was sorted and identified for species composition. The laboratory work load did not allow for testing of all the mosquitoes collected, therefore, only species of <u>Culex</u> and <u>Culiseta</u> were routinely pooled and tested for virus in vero cell culture (table 1).

A total of 62 virus strains were isolated during the season including 21 WEE, 4 TUR, 35 HPV, 1 TUR/HPV, and a CAL serogroup virus. All 62 virus isolates were from pools identified as <u>Cx.</u> <u>tarsalis</u>. The seasonal distribution of virus isolations is shown in table 2.

No suspected WEE human or horse cases occurred in the area where mosquitoes were collected and tested. A clinically diagnosed horse case of WEE occurred in the metropolitan Denver area.

Report submitted by Gordon C. Smith, Carl J. Mitchell, Chester G. Moore, Harry M. Savage, Medical Entomology-Ecology Branch, Division of Vector-Borne Infectious Diseases, National Center for Infectious Diseases, Centers for Disease Control, Fort Collins, Colorado and Arun Bhadra Thapa and Shambhu Lall Shrestha, Division of Epidemiology, Ministry of Health, Kathmandu, Nepal. Table 1.

| Species | Mosq.
tested | Pools
tested |
|----------------------|-----------------|-----------------|
| <u>Cx.p.pipiens</u> | 49 | 4 |
| <u>Cx.tarsalis</u> | 4,537 | 146 |
| <u>Cx.(Cux)</u> spp. | 256 | 18 |
| <u>Cs.inornata</u> | 140 | 19 |
| Total | 4,982 | 187 |

Summary of Colorado mosquitoes tested for arbovirus during 1991.

Table 2.

Seasonal distribution of virus isolations from <u>Cx. tarsalis</u> collected in Colorado during 1991.

| | Nege | Deele | Vi | ated | d d | | |
|--------------------|-----------------|-----------------|--------|------|-----|-----|------|
| Collection
date | Mosq.
tested | Pools
tested | CAL gr | HPV | TUR | WEE | *mix |
| June 26 | 205 | 5 | | 2 | | | |
| July 2 | 341 | 7 | | 2 | | 1 | |
| July 9 | 364 | 9 | | 6 | | 1 | |
| July 16 | 645 | 14 | | 5 | | 3 | 1 |
| July 23 | 313 | 10 | | 2 | 1 | 2 | |
| July 30 | 1,065 | 42 | | 10 | | 7 | |
| Aug. 8 | 412 | 11 | 1 | 2 | 1 | 2 | |
| Aug. 13 | 254 | 11 | | 3 | | 1 | |
| Aug. 20 | 754 | 30 | | 2 | 2 | 3 | |
| Aug. 29 | 184 | 7 | | 1 | | 1 | |
| Total | 4,537 | 146 | 1 | 35 | 4 | 21 | 1 |

*mix--TUR/HPV isolated in same pool.

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Birds as arboviruses' hosts in Brazilian Amazonia¹.

Nicolas Dégallier², Amélia P. A. Travassos da Rosa³, José-Maria C. da Silva⁴, Suely Guerreiro Rodrigues³, Pedro F. C. Vasconcelos³, Jorge F. S. Travassos da Rosa³, Geraldo Pereira da Silva³ e Raimundo Pereira da Silva³

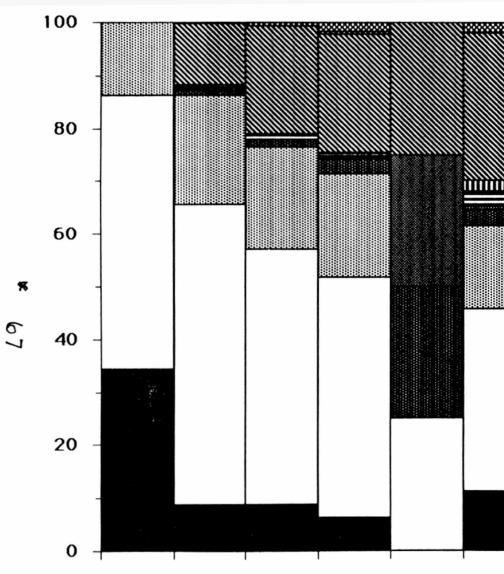
12423 samples of birds' sera or viscera, representing 40 families, 193 genera and 304 species, were collected in 18 localities in brazilian Amazonia. 70 strains of 14 distinct types of arboviruses were isolated and 1743 sera were found with antibodies against at least one arbovirus type. Formicariidae family furnished most strains (1,20 %) and positive sera (30,21 %). The most prevalent arboviruses in birds were Oropouche (3,86 %), Western Equine Encephalitis (3,06 %), Saint Louis Encephalitis (2,80 %), Turlock (1,31 %), Itaporanga (1,00 %), Tacaiuma (0,73 %), Mayaro (0,49 %) and Eastern Equine Encephalitis (0,48 %). Arboviruses were classified according to the preferred vegetation types and strates of their birds' hosts.

The ecological distribution of the arboviruses seems to localize along a continuum from O-15 m levels in terra firme forest (Rocio, Utinga, Kwatta, Gamboa and Icoaraci), intermediate levels of the same forest and secondary growth areas (Oropouche, Turlock, Itaporanga, Guaroa, Triniti, Caraparu, Jurona, Una, Encefalite de Saint Louis and Encefalite equina do oeste), canopy of the forest and secondary growth vegetation (Cacipacore, Mayaro, Ilheus, Candiru and Tacaiuma), the latter vegetation type and inundated forest, and finally extremely versatile viruses like Eastern Equine Encephalitis which were encountered in birds from every vegetation types and levels (fig. 1).

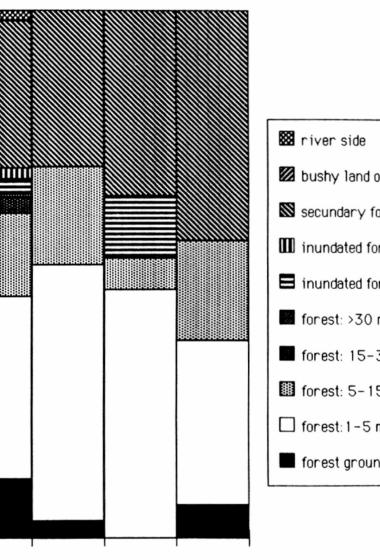
The important role of birds in the sylvatic cycles of viruses Jurona, Itaporanga, Mayaro, Oropouche, Belem, Pixuna, Una and Tacaiuma is confirmed. Bird sera positive for Rocio virus is the first indication of the presence of this agent in the amazonian region. Birds are as yet the only known hosts for the viruses Cacipacore, Candiru and Pacoralike.

The ecological niche concept is discussed in relation with ecology of arboviruses. The adopted definition is that of a hyperdimensioned volume, of which each dimension represents one ecological variable. One of the most limiting factors in the evolution of arboviruses' cycles may be the group-reacting antibodies produced by the vertebrate host and resulting in the impossibility for the same individual to be viremic for more than one virus type in the same serological group, at least during some time. However, more studies are needed to quantify the relative importance of numerous vectors, and determine the width and superposition of the econiches of neotropical arboviruses.

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SEROLOGIC EVIDENCE FOR TICK-BORNE ENCEPHALITIS (TBE) IN NORTH-AMERICAN MILITARY STATIONED IN GERMANY.

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INTRODUCTION

Tick-borne Encephalitis (TBE), also known as Früh-Sommer-Meningo-Encephalitis (FSME) is a tick-borne viral disease endemic in many parts of Central Europe, Sweden, Finland and the USSR. In Western Europe, the spread of the virus is poorly documented, except for South-Germany and Austria. The disease is unknown however in North-America. This paper presents preliminary data of a serosurvey conducted in North-American military stationed in South-Germany.

MATERIAL AND METHODS

A total of 1,125 serum samples of U.S. soldiers, selected for training in outdoor environments, were collected in 1989 from different localities in Bayern and Rhein and-Pfalz in Germany. The majority of these sera were palred, taken from the same 511 individuals with an interval of about six months. Additionally, 89 samples were collected from Canadian military, newly arrived in Baden-Württemberg. Serum samples were sent to the Military Hospital in Brussels (Belgium), the Institute of Public Health in Bilthoven (Netherlands) and the University of Vienna (Austria).

Presence of anti-TBE IgG antibodies was tested first in a home-made ELISA in Bilthoven using a Central European TBE-strain as antigen. Confirmation of positives was performed in the same ELISA system comparing O.D. values on TBE-infected and uninfected Vero E6 cells. All positives, borderline positives and a part of the negatives were retested in Brussels, using the commercially available IMMUNOZYM FSME IgG ELISA kit (Immune AG). The same sera were sent to the TBE reference laboratory in Vienna for confirmation with a home-made more specific ELISA kit, using a highly purified antigen of a Western subtype TBE-strain (Neudorfl prototype).

RESULTS

All sera negative in the Bilthoven test were also negative in Brussels and in Vienna. The first Bilthoven ELISA test however showed 26 positives, but only 17 of them were confirmed in the Bilthoven TBE/Vero system. All these sera were clearly positive in the Brussels and Vienna tests, whereas the other 9 were negative or borderline. In addition, 35 of the 109 Bilthoven borderlines proved to be clearly positive in the Vienna confirmation tests, giving a total of 52 (17+35) confirmed positives.

Based on the final Vienna results, a total seroorevalence of **4.62**% clearly positives (52/1,125) was thus registered; if borderline positives are also included, the overall TBE-seroprevalence increases even up to 5.69% (64/1,125). Five of the positive sera were not paired. Of the others, one individual showed a positive and another one

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a borderline positive result in the first sample, whereas a negative result was found in the second serum of the pair. 4 U.S. individuals seroconverted from negative to positive, 5 others to borderline positive. All other sera were positive in the first sample and remained so during the studied interval of \pm 6 months. Of the Canadian sera, only 2.25% (2/89) were positive.

DISCUSSION

A TBE seroprevalence of **4.62%** is relatively low, when compared to the contiguous local populations : a study from an adjacent area (Thuringia) reported seroprevalences up to 25% (Storch et al. 1991). The seroprevalence in an endemic focus in Sweden was 12% (Gustafson et al. 1991). In contrast, a study in Spain revealed only one seropositive case of 700 tested (Juste 1991) and in Switzerland, seroprevalence for TBE was only 0.6% (de Marvai 1991).

This seroprevalence is unexpectedly high however, taking into account that the studied population consists of healthy young military, residing in South-Germany for a limited period of time (months to years), and originating from the U.S. or from Canada, where TBE has not been documented so far,

The figures found in this study are comparable with another tick-borne disease assessed in the same study group, namely Lyme borreliosis (7.8%), whereas another rodent-borne zoonosis namely Hantavirus disease, scores much lower (1.4%) (unpublished own data).

The three different laboratory tests used showed a slightly different specificity. There is a need for a narrower "grey zone" of ELISA borderline positives. It is suggested that TBE-serology should be performed more routinely when clinical neurological symptoms are preceded by open air activities. The possibly increased risk for active military or other outdoor professions must be evaluated.

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Report from the Vertebrate Ecology Section, Medical Entomology and Ecology Branch, Division of Vector-borne Infectious Diseases, NCID, CDC, Fort Collins, Colorado.

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AVIAN HOSTS OF ST. LOUIS ENCEPHALITIS IN PINE BLUFF, ARKANSAS, 1991

An investigation of the avian hosts of St. Louis encephalitis (SLE) virus was conducted in Pine Bluff, Arkansas during August 30-September 5, 1991 following an SLE epidemic which resulted in 24 confirmed or probable human cases (Centers for Disease Control, 1991). A total of 363 birds of 33 species were captured with ground-level mist nets at 4 sites along the northern edge of the city. No viruses were isolated from the serum of these birds but 91 birds (25%) of 11 species had detectable neutralizing antibody against the TBH-28 strain of SLE virus in the constant-virus serumdilution plague reduction test in VERO cell culture. The antibody prevalence varied among the sites from 11% to 44% (Table 1) but the prevalence at each site was influenced by the avian species composition. The two most abundant species captured and present in the city were also the species with the highest prevalence of antibody, American robin (43%) and house sparrow (42%). Therefore, the sites containing either of these two species had the highest prevalences of antibody (Table 1). Nine other species were infected but at significantly lower rates for the abundant species.

| AVIAN SPECIES | TOTAL | PARK | S I T
GRANARY | E S
CEMETERY | STOCKYARD |
|----------------------------|--------|-------|------------------|-----------------|----------------------|
| House Sparrow | 129* | 0 | 108 | 0 | 21 |
| | (42%) | | (44%) | - | (33%) |
| American Robin | 54 | 51 | 0 | 3 | 0 |
| | (43%) | (45%) | | | |
| Cardinal | 54 | 46 | 0 | 6 | 2 |
| | (6%) | (2%) | _ | (17%) | (50%) |
| Carolina Wren | 29 | 21 | 0 | 8 | 0 |
| | (7%) | (10%) | | | |
| Blue Jay | 14 | 11 | 0 | 3 | 0 |
| | (14%) | (9%) | | (33%) | |
| Brown-headed Cowbird | 9 | 9 | 0 | 0 | 0 |
| Tufted Titmouse | 9 | 3 | 0 | 6 | 0 |
| | (11%) | | | (17%) | |
| Starling | 7 | 5 | 0 | 0 | 2 |
| 2 | (14%) | (20%) | | | |
| Mockingbird | 6 | 4 | 0 | 0 | 2 |
| 5 | (33%) | (25%) | | | (50%) |
| Downy Woodpecker | ົ 5໌ | 2 | 0 | 3 | Ó |
| 1 1 | (20%) | | | (33%) | |
| Red-bellied Woodpecker | 5 | 5 | 0 | 0 | 0 |
| Free Proventier Proventier | (20%) | (20%) | - | - | - |
| Gray catbird | 1 | 0 | 0 | 0 | 1 |
| , | (100%) | - | | - | (100 ⁻ 8) |
| Other species (21) | 41 | 35 | 0 | 6 | 0 |
| TOTAL (33 Species) | 363 | 192 | 108 | 35 | 28 |
| | (25%) | (16%) | (44%) | (11%) | (36%) |

Table 1. Prevalence of neutralizing antibody against SLE virus in birds by species captured and sites sampled in Pine Bluff, Arkansas, 1991.

*Total number tested (percentage positive)

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QUEENSLAND HEALTH - QUEENSLAND UNIVERSITY OF TECHNOLOGY CENTER FOR ARBOVIRUS REFERENCE AND RESEARCH 63 GEORGE STREET, BRISBANE, 4000 AUSTRALIA

Epidemiology

Two children from Arakun (North Queensland) were diagnosed with Murray Valley encephalitis virus infections. Both children suffered a period of encephalitis, with one child having residual paralysis. These are the first cases of MVE infection in Queensland since 1982 which have demonstrated CNS involvement.

During the period July 1990 - June 1991, 11 cases of Kunjin infection have been diagnosed serologically. The most common symptoms were fever and polyarthralgia. Encephalitis was not recorded in any of these patients. Kunjin virus was isolated from the acute phase serum of one patient. The patient developed fever, rigors, headache, photophobia, nausea, muscle weakness, fatigue and lethargy. A convalescent serum sample collected 14 days after the first specimen, contained IgM antibodies to Kunjin only. This is the first isolation of Kunjin from a patient with a natural infection.

Other isolations recorded from acute phase serum samples include 16 Dengue 1 and 12 Ross River virus isolates.

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D. Phillips, C. Atkins, M. Wiemers

Evidence for persisting high level enzootic activity of certain orbiviruses and Malpais Spring virus in wild ungulates of central New Mexico.

The White Sands Missile Range (WSMR), in south central New Mexico, has a high population of Mule Deer (Odocoileus hemionus), Pronghorn (Antilocapra americana), and Oryx (Oryx gazella); feral horses, coyotes, and other species compose the remainder of the large The indigenous and feral wildlife at WSMR are mammal fauna. monitored for population densities and overt disease but are allowed to range freely, public access to the area is limited but periodic hunts are used to cull the burgeoning populations of wild ungulates indigenous to the area, and habitat is protected. A description of WSMR has been published.' In previous publications we reported the isolation of a newly recognized vesiculovirus, Malpais Spring (MPS) virus, from mosquitoes collected in south central New Mexico, high prevalences of antibodies to MPS virus and to orbiviruses bluetongue (BLU)-10, BLU-11, BLU-13, BLU-17, epizootic hemorrhagic disease (EHD) New Jersey, and EHD Alberta, and lower prevalences of antibodies to vesiculoviruses vesicular stomatitis (VS) Indiana and VS New Jersey viruses and to the togavirus western equine encephalitis (WEE) virus.^{1, 2} As a follow-up we determined whether high antibody prevalences to the orbiviruses, to MPS virus, and to WEE virus remain at remarkably high levels in the populations indigenous to the area. Serum samples from Deer, Pronghorn, and Oryx were collected in the period 1987-1991 and tested for neutralizing antibody to eight of the viruses with which the 1985-1986 collection was tested.

Prevalences of antibody in Oryx, Pronghorn, and Deer remained high during the 4 years of the study. For example, as shown in Table 1, antibody to BLU viruses in Oryx differed somewhat by virus but ranged from 40.1% to as high as 94.7%, antibody to EHD viruses ranged from about 14.0% to 63.2%, antibody to MPS ranged from 55.3% to 78.9%, and antibody to WEE virus usually was, with the exception of 1989, relatively low. Similar high antibody prevalence rates (low for WEE virus) in Pronghorn and Deer, age, and other data indicate that the orbiviruses for which we tested, and MPS virus, infect most of the large wild mammals at WSMR and infect them as young, essentially as soon as maternal antibody wanes. Given that antibody from prior infection protects against superinfection with the homologous virus and that neutralizing antibody persists for the life of most animals, it is unlikely that these results are due to recurrent epizootics. Type-specific antibody titers indicate that BLU-10, -11, -13, and -17, EHD New Jersey, and EHD Alberta viruses are all present at WSMR and that the broad reactivity of antibody detected is not due to cross-reactivities between closely related viruses. Complete results are available and are intended for publication.

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C.H. Calisher (Division of Vector-Borne Infectious Diseases, C.D.C., Ft. Collins, Colorado 80522.) and D.E. Taylor (WSMR, Environmental Office, Department of the Army, N.M. 88002-5076.)

| Virus | 1985² | 1986 | 1987 | 1988 | 1989 | 1990 |
|------------------|-------|------|------|------|------|------|
| bluetongue-10 | 55.3 | 78.0 | 49.6 | 41.1 | 57.8 | 68.4 |
| bluetongue-11 | 84.2 | 65.9 | 51.1 | 40.1 | 59.2 | 78.9 |
| bluetongue-13 | 65.8 | 81.7 | 71.9 | 85.6 | 80.3 | 94.7 |
| bluetongue-17 | 78.9 | 75.6 | 54.7 | 72.3 | 46.3 | 87.7 |
| EHD New Jersey | 63.2 | 35.4 | 20.1 | 46.5 | 26.5 | 42.1 |
| EHD Alberta | 42.1 | 36.6 | 44.6 | 62.4 | 20.4 | 14.0 |
| Malpais Spring | 55.3 | 70.7 | 71.9 | 65.8 | 78.9 | 64.9 |
| western eq. enc. | 2.6 | 6.1 | 6.5 | 6.9 | 10.2 | 1.8 |
| Number tested | 38 | 82 | 139 | 202 | 147 | 57 |

Report from Department of Veterinary Science and School of Veterinary Medicine, University of Wisconsin, Madison, Wisconsin 53706.¹

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HANTAVIRUSES IN WISCONSIN AND DULUTH, MINNESOTA

Hantaviruses cause a complex of human diseases known as hemorrhagic fever with renal syndrome (HFRS) throughout the Eurasian continent. These viruses are carried persistently in their rodent hosts and are transmitted to people by aerosols of contaminated urine, feces and saliva. The discovery of pathogenic hantaviruses in rats in maritime ports of Korea and Japan raised the concern for international dispersal of rat-associated hantaviruses. Global serologic and virologic studies uncovered evidence for rat-associated hantaviruses world-wide, including the United States. During the course of this hantavirus search, two other members of this genus were isolated in the United States; Prospect Hill virus (PH) from <u>Microtus pennsylvanicus</u> and Leaky virus from <u>Mus musculus</u>.

The purpose of the present study was to determine if a hantavirus is present in Wisconsin and Duluth, MN. Rodents were trapped in the Great Lakes port cities of Wisconsin including Green Bay, Milwaukee, and Superior and Duluth, Minnesota, all of which are ports involved in international shipping. A total of 675 wild rodents were trapped, including 310 meadow voles (Microtus pennsylvanicus), 173 norway rats (Rattus norvegicus), 179 deer mice (Peromyscus spp., including P. leucopus, and two subspecies of P. maniculatus), and 13 house mice (Mus musculus). Twenty percent of the rats, 17% of the field voles, 8% of the house mice, and 3% Peromyscus spp. were seropositive to a hantavirus by immunofluorescent antibody assay (IFA). Βv the plague reduction neutralization test (PRNT), sera from 9 field voles, 1 Peromyscus leucopus, and 1 rat were positive with highest titer to PH virus. All of the PRNT seropositive individuals were from the twin cities of Superior, WI and Duluth, MN. Presence of hantavirus antigen was detected in lung tissue by IFA in M. pennsylvanicus and P. spp., but not in rats. Two hantaviruses, designated SD-1 and SD-2, were isolated from M. pennsylvanicus captured in Duluth. MN and found to be very similar to prototype PH virus by cross-IFA and cross-PRNT. SD-1 can be differentiated from prototype PH virus by monoclonal antibody analysis. There was no convincing evidence for the presence of a typical rat-associated hantavirus since IFA serology was not supported by PRNT, nor was antigen detected, nor virus isolated from rat tissues. In summary, we have serologic and virologic evidence of a PH-like hantavirus, designated SD virus in Superior, WI and Duluth, MN. The human disease potential is unknown although in other studies, people with antibody to PH virus have been found with no history of HFRS-like disease.

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Serological ver fied Hantavirus infections in Hungary.

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The Haemorrhagic Fever with Renal Syndrome (HFRS) has been a well known syndrome in Hungary from the early 50's. Formerly both epidemic and sporadic cases were mainly diagnosed on the basis of clinical symptoms and epidemiological characteristics. The laboratory testing was prevented by the lack of the biohazard level working conditions (P4). Since 1989 138 sera of suspect cases have been selected fro routine samples in our laboratory. The samples were tested by indirect immunofluorescence antibody test (IFAT) using infected rat splenocyte antigen provided and specified by the Military Medical Academy of Bulgarian Army/Sofia.Recently tests are carried out by "Hantavirus Antibody IF Assay Kit" of Progen/Heidelberg of infected Vero E6 cell origin.During the last three years altogether 17 cases (of 138) have been verified as Hantavirus infection. Five cases were positive both for Hantaan-virus specific IgG and IgM antibodies. The others were diagnosed as Puumala-virus positiv cases. The results supposed our working hypothesis that there is cocirculation of a few different Hantavirus strains in Hungary.

E Falser Falledi

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Endemic foci of hemorrhagic fever with renal syndrome (HFRS)

in Germany

We had started our search for Hemorrhagic Fever with Renal Syndrome (HFRS) in Germany with seroepidemiological surveys in humans and different species of free-living and laboratory-bred rodents in 1984. The indirect immunofluorescence antibody (IFA) technique was applied starting with spot slides kindly supplied to us by Dr. J. LE DUC, Fort Detrick/USA. These slides contained vero E6 cells infected with hantaan virus strain 76-118. Subsequently we could prepare our own spot slides when Dr. G. VAN DER GROEN, Antwerpe/ Belgium, had kindly supplied us with different hantavirus strains.

Different seroepidemiogical surveys in humans revealed the presence of hantavirus circulating in Germany:

- a) 7 out of 303 (2.3 %) sera from soldiers,
- b) 10 out of 820 (1.3 %) sera from in-door patients of 3 different hospitals at Düsseldorf,
- c) 27 out of 989 (2.7 %) sera from in-door patients of hospitals at Würzburg,
- d) 12 out of 202 (5.9 %) sera from patients of a nephrology department at Munich, and
- e) 41 out of 1106 (3.7 %) sera from wood-cutters collected within a distance of 150 km around UIm were found positive by IFA.

A total of 23 out of 308 (7.5 %) sera from laboratory rats collected from different breeding stocks in Germany and other European countries was found positive by IFA, titers ranging between 1 : 10 and 1 : 1280. Dr. H.W. LEE, Seoul/Korea, could demonstrate an IgM antibody titer of 1 : 32 against hantavirus strains Seoul and 76-118 in one serum from a laboratory rat collected in July 1986 in a German breeding colony. So far no laboratory-associated clinical cases of HFRS have been observed in our country.

We have demonstrated hantavirus antibodies in sera from 4 different species of wild-living rodents: Rattus norvegicus, Clethrionomys glareolus, Microtus arvalis and Mus musculus (Table 1). A hantavirus strain, named "Polle 18", was isolated from Rattus norvegicus captured in Lower Saxony.

The first HFRS case in Germany was described by ZEIER and coworkers. In 1986 we had informations about 3 clinical cases. However this figure increased every year: 1987 to 8 cases, 1988 to 16, 1989 to 18, 1990 to 24, and 1991 to 46 cases, consisting out of 33 cases which had been serologically confirmed by IFA, IgM ELISA and Western blot analysis mainly at Düsseldorf and additional 13 cases which had been diagnosed at Göttingen (data of 7 cases kindly supplied by Dr. B. KRONE) and at Reutlingen (data of 6 cases kindly supplied by Dr. U. SCHUBERT). The homes of all 46 patients are mapped in Fig. 1.

Thus it can be demonstrated that different HFRS foci exist in Germany. The "Schwäbische Alb", a low-mountain area in the south-western part of Germany, seems to be the most important endemic focus. Sero-survey data from wood-cutters in this area revealed that the hantavirus antibody prevalence rates in wood-cutters differed in 32 forest districts. The highest prevalence rate of 26 % was found in the district Geislingen in the center of the natural focus (PILASKI et al., 1991). The HFRS outbreak in February 1990 in US soldiers camping in the "Schwäbische Alb" area stresses the active role of this focus (UNDERWOOD et al., 1991).

Table 1:Results of IFA serosurveys in 494 wild-living rodents (77 Rattus
norvegicus; 129 Apodemus sylvaticus; 88 Apodemus flavicollis;
30 Mus musculus; 2 Microtus agrestis; 20 Microtus arvalis;
145 Clethrionomys glareolus ; 3 Arvicola terrestris) trapped at
different locations in Western Germany

| Location | Species | Date of
sample
(month.
year) | No. pos./
No. test. | Total
No. positive/
No. tested | % positive |
|-------------|-----------------|---------------------------------------|------------------------|--------------------------------------|------------|
| Polle | R. norvegicus | 7.85 | 7/22 | | |
| Stade | R. norvegicus | 7.85 | 4/25 | | |
| Polle | R. norvegicus | 6.86 | 10/29 | 21/77 | 27.3 |
| Hannover | R. norvegicus | 1.86 | 0/1 | | 27.0 |
| Münster zoo | Ap. sylvaticus | 2.78-9.80 | 0/21 | | |
| Münster | Ap. sylvaticus | 7.85 | 0/20 | | |
| Daun/Eifel | Ap. sylvaticus | 910.85 | 0/53 | 0/129 | 0 |
| Celle | Ap. sylvaticus | 10.85 | 0/3 | | - |
| Lonsee | Ap. sylvaticus | 7.86-10.88 | 0/5 | | |
| Amstetten | Ap. sylvaticus | 7.88-10.88 | 0/27 | | |
| Daun/Eifel | Ap. flavicollis | 910.85 | 0/52 | | |
| Lonsee | Ap. flavicollis | 7.85 | 0/3 | 0/88 | 0 |
| Amstetten | Ap. flavicollis | 7.,10.88 | 0/23 | | |
| Eppingen | Ap. flavicollis | 8.86 | 0/10 | | |
| Münster zoo | | 2.78-9.80 | 3/13 | | |
| Celle | Mus musculus | 7.,10.85 | 1/7 | 4/30 | 13.3 |
| Amstetten | Mus musculus | 7.,10.88 | 0/10 | | |
| Eppingen | Micr. agrestis | 8.86 | 0/2 | 0/2 | 0 |
| Oldenbrok | Micr. arvalis | 9.86 | 0/8 | 6/20 | 30.0 |
| Oldenbrok | Micr. arvalis | 11.86 | 6/12 | | |
| Daun/Eifel | CI. glareolus | 910.85 | 0/98 | | |
| | n Cl. glareolus | 9.85 | 0/5 | | |
| Celle | Cl. glareolus | 10.85 | 0/1 | 1/145 | 0.7 |
| Lonsee | CI. glareolus | 7.,8.86 | 1/22 | | |
| Amstetten | Cl. glareolus | 7.,10.88 | 0/10 | | |
| Eppingen | Cl. glareolus | 8.86 | 0/9 | 0/2 | • |
| Lonsee | Arv. terrestris | 10.88 | 0/3 | 0/3 | 0 |
| T I | | | | 00/404 | |

32/494 6.5



Fig. 1: Map of Germany indicating homes of 46 HFRS patients (• serologically confirmed at Düsseldorf; o diagnosed at Göttingen and Reutlingen) and the location of the HFRS outbreak in US soldiers in February 1990 described by UNDERWOOD et al. (*).

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14/11 91 17:05 2687249

EPIZOOTIOLOGICAL ASPECTS OF HANTAVIRUS INFECTION IN BELGIUM, THE NETHERLANDS AND GERMANY : OF MICE AND MEN

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Except for Scandinavia, little is known concerning epizootiology of Hantavirus (HV)-infection in European rodent populations, and the ways of transmission of Puumala-HV, the etiologic agent of Nephropathla epidemica (NE), to humans through infected wild rodents.

On a total of 5,038 small mammals captured in Belgium, the Netherlands and Germany, 6.88% (153/2,225) showed IFA or ELISA presence of Puumala-like antigen in the lungs, and 5.22% (194/3,718) had serological evidence of IFA IgG Hantaviral antibodies. Clethrionomys glareolus (CI. gl.) (the red bank vole) was the second most abundant (total N captured : 2,012), and in each country by far the most infected rodent species, showing a Puumala-like antigen presence in 14.7% (129/872) for Belgium, 12.5% (4/32) for the Netherlands and 12.2% (11/90) for Germany. Gl. gl. captured around habitats of recent human NE-cases in these 3 countries appeared however more abundant and significantly more infected with HV than the average in each country, showing Puumala-like antigen in up to 35.5%.

High density regions (HDR) for CI. gl. in Belgium were defined as areas where the incidence of CI. gl. skulls in owl pellets (total N of examined preys : 268,423) exceeded the average for Belgium (2.1%). Habitats of 81 asymptomatic human HV-seropositives of 2 previously published Belgian epidemiological studies were equally distributed inside (54%) and outside (46%) these HDR. In contrast, 73% of a total of 22 serologically confirmed recent human NE-cases in Belgium appeared to live and work inside these HDR vs only 27% (p<0.05) cutside. The majority of HVpositive rodents in Belgium were also found in these regions.

CI. gl. is confirmed as the main rodent resevoir for Puumala-like virus in Belgium, the Netherlands and Germany. Sporadic or clustered human cases of NE may appear when the local CI. gl. population is abundant and/or highly infected with Puumala. A low virulemce strain of JBE virus isolated from Culex Tritaeniorhynchus mosquitoes in Indonesia

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A study on virulence and immunogenicity of 5 strain of JBE virus isolated from Culex Tritaeniorhynchus mosquitoes by Dr. Olson in Indonesia was carried out .

| 1. Viral strain
Jak – l | :
Lot 974 | BHK 21 | P-2 | SMB2 |
|----------------------------|--------------|--------|-----|------|
| Jak - 2 | Lot 2380 | Vero | P-3 | SMB1 |
| Jak - 3 | Lot451 | Vero | P-3 | SMB1 |
| Jak - 4 | Lot6577 | Vero | P-3 | SMB1 |
| Jak - 5 | Lot788 | Vero | P-2 | SMB1 |

The virulence of JBE virus Table 1. The virulence of JBE virus strain isolated in Indonesia

| strain | PFU/ml.
LoglO | I.C.titer
LD50 | PFU/LD50 | Sub titer
LD50 | PFU /LD50 |
|--------|------------------|-------------------|----------|-------------------|-----------|
| Jak-1 | 9.0 | 6.67 | 2.33 | 3.3 | 5.7 |
| Jak-2 | 9.0 | 5.67 | 3.23 | 3.3 | 5.7 |
| Jak-3 | 8.0 | 6.50 | 1.50 | 5.0 | 3.0 |
| Jak-4 | 8.5 | 7.33 | 1.17 | 5.0 | 3.5 |
| Jak-6 | 7.5 | 6.67 | 0.83 | 4.3 | 3.2 |

Table 1 showed two Of them Jak-1 and Jak-2)to have low virulence. Thier ratioes of PFU/LD50 titer were 2.33 and 3.23 respectively,.

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| | n by plaque se | | n seleted fr |
|--------|----------------|---------|--------------|
| No. of | PFU/ml | LD50/ml | PFU/LD50 |
| strain | Logl0 | LoglO | |
| 1 | 7.00 | 3.50 | 3.50 |
| 2 | 7.00 | 3.50 | 3.50 |
| · J | 7.56 | 3.50 | 4.06 |
| 4 | 7.19 | 3.50 | 3.69 |
| 5 | 6.95 | 3.50 | 3.45 |
| 6 | 7.13 | 1.50 | 5.63 |
| 7 | 7.13 | 2.00 | 5.13 |
| 8 | 7.12 | 2.00 | 5.12 |
| 9 | 7.45 | 2.00 | 5.45 |
| 10 | 7.16 | 3.50 | 3.66 |

Table 2. The virulence of JBE virus strain seleted from

Table 2 showed strain 6 ,7, 8, 9, to have low virulence than others. The ratio of PFU/LD50 were greater than 5.00

Secondary cloning was carried out by plaque seleted method from plaque 6, 7, 8, 9, . Avirulence strian 6-2 was obtained. 6-2 strain did not kill the 3 week-old mice by i.c. inoculation, but the titer of PFU was 6.0-7.0 The minimum dose of 50% protection rate of 6-2 strain against Beijing A2 strain challenging is 0.00001 and 0.000001 when viral titer is PFU/ml log10 4.0 and 7.0 respectively.

The relatoinship between a high rate of low virulent strain existed in nature and the occurrence of inapparant infection was discussed...

STUDY ON THE EXPERIMENTAL INFECTION OF TREE SHREW WITH CHIKUNGUNYA VIRUS

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Adult Tree shrews (Tupaia belangeri chinensis) have been tested by inoculation with three strains of Chikungunya (CHIK) virus isolated from Yunnan, China and Africa, for the susceptibility. These animals showed viremia for 2-6 days after injection. Specific hemagglutination inhibition (HI) antibody appeared on the 6th day following infection and reached peak value on the 30th to 40th day. Neutralization antibody response occured 10 days after infection and reached peak value on the 30th day, coexisted with HI antibody. Complement fixation antibody response occured on the 21th day and reached peak value on the 40th to 50th day after infection. CHIK viruses were detected in the brain, lung, liver, kidney and spleen tissues 4-12 days after infection and disapeared on the 14th day. Pathological examination could find changes of inflammation and hemorrhage in the organs of these animals. The results indicated that Tree shrew is a susceptible animal to CHIK virus, and this animal might be used in some studies of CHIK virus.

THE NATURAL INFECTION RATE OF MOSQUITOES BY JAPANESE ENCEPHALITIS B VIRUS IN YUNNAN PROVINCE, CHINA

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In July-September 1983-1987, 57898 adult female mosquitoes belonging to 29 species of 8 genera were collected from Japanese B encephalitis(JE) epidemic areas in Yunnan, China, and were examined by C6/36 cell method and suckling mouse method. Twenty-eight strains of JE virus were isolated from 12 species of mosquitoes(table). The positive isolation rates by the two methods were 6.19% and 3.33% respectively. The highest positive isolation rate 12.3% (per 100 pools), mosiquito body virus carrier rate 1:208 or mosquito body virus carrier rate 4.81% (per 1000mosquitoes) were found in the middle ten days of August, corresponding to epidemic peak of JE, Nine strains of JE virus were isolated from <u>Culex tritaeniorhynchus</u> with an isolation rate of 7.44%, natural isolation ratio was 1:584 and mosquito body virus carrier rate 1.71‰. These results indicated that <u>C. tritaeniorhynchus</u> might be the main vector of JE virus in Yunnan, while <u>C. whitmorei</u>, <u>C. pseudovishnui</u> and <u>Anophees sinensis</u> are also unportant vectors of JE virus in Yunnan.

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| Species of mosq. | Mosq.
pools | | No.po-
sitive
pools | Positive
rates of
pools | Isolation
ratioes
(%) | Isolation
rates(%。) |
|--------------------------|----------------|-------|---------------------------|-------------------------------|-----------------------------|------------------------|
| Culex tritaeniorhychus | 121 | 5260 | 9 | 7.44 | 1:584 | 1.71 |
| C. whitmorei | 88 | 3517 | 5 | 5.68 | 1:703 | 1.42 |
| <u>C</u> . pseudovishnui | 80 | 2609 | 3 | 3.75 | 1:869 | 1.15 |
| C. fuscocephala | 149 | 6622 | 1 | 0.67 | 1:6622 | 0.15 |
| C. annulus | 21 | 467 | 1 | 4.76 | 1:467 | 2.14 |
| C. gelidus | 19 | 502 | 1 | 5.26 | 1:502 | 1.99 |
| Anophees sinensis | 81 | 3830 | 3 | 3.70 | 1:1276 | 0.78 |
| Mansonia uniformis | 35 | 1195 | 1 | 2.86 | 1:1195 | 0.84 |
| Aedes albopictus | 176 | 6138 | 1 | 0.57 | 1:6138 | 0.16 |
| Ae. vexans | 61 | 1920 | 1 | 1.64 | 1:1920 | 0.52 |
| Ae. lineatopennis | 19 | 683 | 1 | 5.26 | 1:683 | 1.46 |
| Ae. assamensis | 22 | 456 | 1 | 4.55 | 1:456 | 2.19 |
| Other species | 656 | 24699 | 0 | | | |
| Total | 1528 | 57898 | 28 | 1.83 | 1:2068 | 0.48 |

TABLE Isolation of Japanese B Encephalitis Virus from Mosquitoes Collected in Yunnan, China, 1983-1987

Interferon studies in Mayaro virus-infected TC7 cells

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Treatment of TC7 cells with interferon (IFN) drastically reduced the yield of infectious Mayaro virus under experimental conditions which did not detect modification of virus attachment and penetration into the cells. In IFN-treated cells, synthesis of Mayaro virus proteins was inhibited and cellular protein synthesis restored. This phenomenon is dependent on IFN concentration and multiplicity of infection. Electron microscopy of these cells revealed anomalous viral particles and normal ones inside cytoplasmic vacuoles. This suggests that IFN also interferes with Mayaro virus morphogenesis and inhibits the release of virions from treated cells. Studies on second infection cycles with virions obtained from IFN-treated cells indicated that both viral populations inhibit host protein synthesis and synthesized the same spectrum of viral proteins. However, replication of this second viral population is impaired when cells are treated with much lower doses of IFN.

QUEENSLAND HEALTH - QUEENSLAND UNIVERSITY OF TECHNOLOGY CENTER FOR ARBOVIRUS REFERENCE AND RESEARCH 63 GEORGE STREET, BRISBANE, 4000 AUSTRALIA

Inability of Barmah Forest virus to cross the placenta in mice.

A number of alphaviruses including two found in Australia (Ross River virus and Getah) cross the placenta in pregnant mice. Ross River virus also crosses the placenta in pregnant women but at a lower rate that seen in mice.

Barmah Forest virus, an alphavirus serologically related to Sindbis, has been isolated from patients in Australia with rash, pyrexia and polyarthralgia. There is also convincing serological evidence (Barmah Forest virus specific IgM in acute phase sera) that this virus causes disease in humans.

The prototype strain of BFV prepared as a suckling mouse carcass and brain pool failed to produce any viraemia in either pregnant CBA or Balb C mice infected iv., although these animals did produce a classical primary immune response (IgM antibody production beginning about day 3 and IgG about day 6). No live virus could be detected in maternal muscle or in placentas or foetuses using either C6-36 Aedes albopictus mosquito cells or veros.

Similar results were obtained with two other strains of BFV which had been prepared as separate suckling mouse brain and carcass pools. These consisted of one strain (Murweh) isolated from mosquitoes in Charleville in central Australia and one of the human isolates of BFV.

Although these latter four virus pools all grew well in the C6-36 cells, none produced cytopathic effects in vero cells – in contrast to the prototype strain which produced clear CPE in veros.

Since humans certainly are viraemic following infection with Barmah Forest virus we conclude that there remains a possibility that BFV could cross the placenta in humans but that mice are probably not the experimental model in which to study this.

P. Birt, J. Haig, J. Aaskov

5.

Report from the Abt. für Virologie, Technische Universität München

Effect of pH on infectivity of EYACH virus in Vero cells

EYACH virus is a member of the newly created genus <u>Coltivirus</u> in the family Reoviridae. It was isolated in Germany and up to date is the only European virus, serologically related to Colorado tick fever virus. Due to difficulties of cultivation of Eyach virus in cell culture, little information is available concerning conditions of virus infection and replication in cell culture.

We succeeded in establishing a plaque test assay for EYACH virus in Vero cells. Using this system we investigated the effect of pH on virus stability and plaque formation.

Medium from infected Vero cells used as stock virus with a titer of 5×10^5 pfu/ml was diluted in buffer with various pH values. Confluent monolayers of Vero cells in Petri dishes were infected for plaque titration. After virus adsorption of two hours at 37° C virus inoculum was removed and cells were washed once with buffer of the appropriate pH. For plaque formation cells were overlayed with 1% agarose and incubated as usual in a 5% CO₂ atmosphere. Dally microscopic examination showed no signs of adverse effects in cells inoculated with inocula of unphysiologic pH. After 8 days of incubation, cells were fixed with formaldehyde and stained with 0,5% Coomasie Blue.

The inocula pH values and the resulting titers were as follows:

| pH 6,0 | no plaques |
|--------|--------------------------|
| pH 6,5 | 2x10 ⁴ pfu/ml |
| pH 7,0 | 1x105 pfu/ml |
| pH 7,5 | 4x10 ⁵ pfu/ml |
| pH 8,0 | 1x10 ⁵ pfu/ml |

The results show that maximal EYACH virus infectivity is dependent on alkaline pH.

In addition we tested whether the effect of low pH on virus infectivity was irreversible.

Virus was kept at pH 6.0 for one hour at 37°C and then pH was readjusted to pH 7.4. Infectivity of the inoculum was assayed in the same manner as described above.

Titer of the virus control showed 6×10^5 pfu/ml. Exposure to pH 6.0 resulted in an infectivity reduction of three logs to 6×10^2 pfu/ml. Reising the pH to pH 7,4, had no effect on infectivity.

The experiment shows that infectivity of EYACH virus in Vero cells is sensitive to acidic pH values, and also that infectivity could not be restored by raising the pH to normal. Acidic pH seemed to irreversibly destroy infectivity of this coltivirus.

(G. Dobler, H. Meier-Ewert)

AXOPLASMIC RETROGRADE TRANSPORT OF JURONA VIRUS (BE AR 40578)

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The prototype strain of Jurona (JUR) virus, BE AR 40578, was isolated from a pool of 89 females *Haemagogus sp.* captured by hand off human bait in Belém-Brasília highway, km 87, Pará, Brazil on 9 March 1962. Optical and electromycroscopy, of experimentally inoculated animals has shown an encephalitis with neuronal necrosis 48h F.I. Virus particles bullet shaped (typical Rabdhovirus) were founded in cytoplasmic budding from cell membranes. Serological studies has classified JUR as *Vesiculovirus*.

Identification of transport mechanisms of arbovirus in the Central Nervous System is important for pathogeny and pathology studies. Using immunocytochemical procedures we have been able to pathways of the virus particles in visual cortex of trace the newborn swiss mice. Injections of 0.5ul of a virus suspension were inoculated in the Area 17 of the brain mice (Wagor et al., 1981). Animals were perfused 24 and 48 hours later with saline, followed by 4% paraformaldebyde diluted in 0.1M phosphate buffer, PH and cryoprotector solutions (25% and 50%). 150 um thick 7.2-7.4. from freezing microtome were washed 20min three times in sections 0.05M Tris buffer saline, 0.1% triton (TBST), pH 7.4. Sections were incubated 48 hours in primary antibody (1:1000) in TBST and serum (4%). After 20 min three washes in TBST sections were block incubated with biotin labed secondary antibody in TBST, pH 7.4. A overnight incubation with ABC complex in TBST was followed new bч histochemical reaction with Ni enhanced glucosean et oxidase/diaminobenzidin technique (Shu al, 1989). In extrastriated visual areas projecting to Area 17 we were able to find labeled degenerating neurons in supragranular area. Axons were also labeled in white cortical matter. The label thopographic specificity is compatible with the view that retrograde axoplasmic transport can be one of the paths virus use to disseminate in central nervous system.

Note: JUR virus since that time has not been isolated in our laboratory until 1989, when we get the second strain (BE H 493533) from the blood of a patient living in Costa Marques, Rondonia, Brazil.

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SEROLOGICAL STUDIES ON THE INFECTION OF DOGS IN ONTARIO, CANADA, WITH BORRELIA BURGDORFERI

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Lyme disease has been documented in Canada but the true extent of its occurrence is presently unclear. To date the only area in Canada where Borrelia burgdorferi has been definitively shown is at Long Point in southern Ontario¹. Between January 1988 and August 1989 a serosurvey was undertaken on 1,095 Ontario dogs to determine whether the results might provide further insight into the distribution of B. burgdorferi in the province. The sera were arbitrarily chosen from specimens that passed through a veterinary clinic with no selection bias based on age, sex, breed, geographic location or clinical symptoms of the dogs.

All dog sera were tested by enzyme-linked immunosorbent assay (ELISA) with all ELISA positives subsequently tested by the immunofluorescent antibody (IFA) test. Western blot (WB) tests were undertaken on reactive sera.

Sixty-five (5.9%) of the 1,095 sera tested were positive by ELISA. IFA reactivity of the ELISA positive sera was as follows: \leq 1:32-43 sera, 1:32-6 sera, 1:64-8 sera, 1:128-2 sera, 1:256-4 sera, 1:512-1 serum and 1:1024-1 serum. Twenty-one of the 22 ELISA positive, IFA reactive and ten ELISA positive, IFA negative sera were tested by WB with eight WB positive and three WB equivocal reactors obtained (Table 1).

Two of the eight WB confirmed reactors had been to known Lyme endemic areas of Long Point, Ontario and the northeastern United States. Travel histories for five of the remaining six dogs were not available while the sixth dog, from North Bay (46020'N, 79025'W), had lived in this part of northern Ontario for its entire life. Three of the six dogs resided in communities in southern Ontario within approximately 20 miles of each other (43055'N, 8000'N). Given the fact that a positive serological test can be expected to have a low predictive value in areas where Lyme disease does not occur² and that the only conclusive evidence to date for the occurrence of B. burgdorferi in Ontario is at Long Point, it was felt that WB confirmatory tests were necessary. Using WB as the standard, this serosurvey did not provide evidence to suggest that B. burgdorferi is widespread in Ontario. These findings are consistent with field studies undertaken in Ontario from 1987-1990.¹

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Table 1.

. Pattern of reactivity to Borrelia burgdorferi by immunofluorescent antibody and western blot assays of FLISA positive serum from Ontario dogs.

| BERUH MULIBER | IFA TITRE | MEDIERO BLOT RESUL |
|---------------|-----------|--------------------|
| DL6 | 1:256 | positive |
| DLS78 | 1:64 | negative |
| DLS203 | _ 1 | negative |
| DLS286 | - | positive |
| DLS289 | - | negative |
| DLS307 | 1:64 | equivocal |
| DLS337 | 1:64 | negative |
| DLS345 | - | negative |
| DLS355 | - | negative |
| DLS377 | 1:64 | negative |
| DLS384 | 1:1024 | positive |
| DLS456 | 1:256 | positive |
| DLS464 | 1:512 | positive |
| DLS485 | 1:256 | negative |
| DLS1137 | 1:128 | negative |
| DLS1170 | 1:32 | negative |
| DLS1244 | 1:32 | positive |
| DLS1269 | 1:32 | negative |
| DLS1273 | - | negative |
| DLS1368 | 1:32 | positive |
| DLS1373 | 1:64 | positive |
| DLS1393 | - | negative |
| DLS1416 | - | equivocal |
| DLS1447 | 1:256 | negative |
| DLS1496 | 1:128 | negative |
| DLS1551 | 1:32 | negative |
| DLS1573 | 1:64 | negative |
| DLS1600 | - | equivocal |
| DLS1610 | 1:32 | negative |
| DLS1651 | - | negative |
| DLS1724 | 1:64 | negative |

1 - = IFA titre <1:32

A SEROSURVEY FOR ANTIBODY TO Borrelia burgdorferi IN AN "AT RISK" POPULATION OF SOUTHERN PORTUGAL.

Having found the first clinical case of Lyme disease in Portugal, it was decided to organize a serologic survey of a sample of the human population living in the southern part of the country (Portalegre, Evora, Beja, and Setúbal). Samples were from people who farm; physicians collecting these samples asked whether the patient remembered being bitten by ticks and whether the patient hunted (usually for recreation). People who reported being bitten by ticks were considered "at risk".

Serologic tests were done using an immunofluorescence test that is commercially available, considering titers ≥256 positive; positive and negative controls were included in all tests. All positive sera were screened by the suggested VDRL test.

Three of 9 people who had been clinically diagnosed as having Lyme disease had antibody to *Borrelia burgdorferi*; two of the 9 were considered questionably positive serologically.

In people with a known history of tick contact 11/45 (24.4%) had antibody to Borrelia burgdorferi. Thus, there is clinical and epidemiologic evidence for the presence of Borrelia burgdorferi, or a closely related organism, in southern Portugal.

REPORT FROM THE CENTER FOR ZOONOSES RESEARCH, NATIONAL INSTITUTE OF HEALTH, AGUAS DE MOURA, PORTUGAL (Armindo R. Filipe)

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Ixodes pacificus and Bonelia burgdorferi in Arizona

8-91 FRI 11:43 VEISCIENCE/MICROPICLOS,

C.A. Olson, E.W. Cupp, S. Luckhart, J.M.C. Ribeiro (Department of Entomology, University of Arizona; Tucson) and C. Levy (Division of Disease Prevention, Arizona Department of Health Services; Phoenix)

On 10 February 1991, two I. pacificus adults were collected by two hikers from their legs after painful bites alerted them to the presence of these ticks. The incident occurred on the Aspen Peak trail in the Hualapai Mountain County Park south of Kingman, Mohave County, Arizona, at an altitude of approximately 7,200 feet. On 17-19 April 1991, an examination of the trail area was made, resulting in the collection of 65 adult I. pacificus from along the south-facing slopes in a scrub oak zone. The altitude in this area of the park was approximately 7,700 feet. Adult ticks were swept from the dry grass understory along the edges of the trail. Temperatures during the second collection were in the high 50's F. The gut/salivary gland extracts were examined by indirect immunofluorescence for B. burgdorferi using a monoclonal antibody (H5332) that recognizes a 31 kDa outer surface protein of this spirochete followed by anti-mouse sera conjugated with fluorescein isothiocyanate. Of the 48 extracts examined, two were strongly positive, indicating a minimal adult tick infection rate of 4% along the Aspen Peak trail.

Two subsequent trips to the area in May and June also resulted in collection of both adults and immatures in other locations. In May, seven adult ticks were collected from vegetation in a narrow glen located at \geq 7,000 feet near Flag Mine, outside the park. Some adults (n=3) were also found questing along a deer trail which was located on a hillside above the mine in a more open and sparsely vegetated zone. In early June, immatures (n=27) were collected from noosed fence lizards and adults (n=9) were again taken along the hiking trail in the park.

This is the first record of both <u>I. pacificus</u> and <u>B. burgdorferi</u> in Arizona. The infection rate of the single field-collected sample (4%) is within the range of those reported for this tick in northern California. However, much more survey works is required before the ecology of <u>I. pacificus/B. burgdorferi</u> in Arizona can be validly compared with other locations in the western USA.

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Report from the Institute of Systematical and Ecological Biology, Czechoslovak Academy of Sciences, 603 65 Brno, Czechoslovakia

Borreliae in culicine mosquitoes

Borrelia burgdorferi (the Lyme disease spirochete) is considered to be transmitted primarily by ticks of the *Ixodes ricinus* complex (Burgdorfer et al. 1982: Science 216: 1317-1319). However, biting insects (deer flies, black flies and mosquitoes) have been found to be infected with borreliae occasionally (Magnarelli et al. 1986: J.Inf.Dis. 154: 355-358, Magnarelli and Anderson 1988: J.Clin.Microbiol. 26: 1482-1486, M. Pejčoch – personal communication). Unidentified spirochetes were also observed in the digestive tract of anopheline mosquitoes (Sinton and Shute 1939: J.Trop.Med.Hyg. 42: 125-126, Laird 1959: Ecology 40: 206-221). This paper demonstrates the presence of borreliae in mosquitoes of the genera *Aedes* Meig. and *Culex* L.

Female mosquitoes were sampled with an aspirator in two localities in southern Moravia (CSFR). Locality I. A lowland forest near the town Lednice: 98 Aedes vexans Meig., 21 Aedes cantans (Meig.) and 21 Aedes spp. mosquito specimens were collected in August and September 1990. Locality II. Basement rooms of the buildings in the town Valtice. Two samples were collected here: 480 females of *Culex pipiens molestus* Forsk. (in September and October 1990) and 400 females of the same species in March and April 1991. All specimens were kept in the laboratory at 18 C alive until examined. Digestive tract was examined by dark-field microscopy at 200x (400x) magnification for the presence of borreliae.

Borreliae were detected in 4.1% of Ae. vexans collected in the locality I, and in 4.3% and 3.5% of C. pipiens molestus mosquitoes collected in the locality II in autumn 1990 and spring 1991, respectively. Borreliae were found in numbers from two spirochetes up to several thousands in the positive mosquitoes (Table).

The borreliae detected in the mosquitoes were morphologically similar or identical to those observed in *I.ricinus* L. ticks collected in the same areas which were identified as *Borrelia burgdorferi* (Hubálek et al. 1990: Folia parasitol. 37: 359-362, Kryuchechnikov et al. 1990: Zh.Mikrobiol.Epid.Immunobiol.(Moskva) 6: 10-13). The detection of borreliae in naturally infected *Ae. vexans* mosquitoes is in accordance with literature references as mentioned above. The presence of borreliae in the mosquito head and digestive tract observed after experimental infection (in *Ae*. aegypti, Ae. atropalpus and Ae. triseriatus mosquitoes) seems to be ephemeral (Magnarelli et. al. 1987: J.Inf.Dis. 156: 694-695). However, our results indicate persistence of borreliae in C. pipiens molestus mosquitoes over the winter season. Findings of motile borreliae (occasionally in large numbers) in hibernating anthropophilic C. pipiens molestus mosquitoes show one possible way of overwintering borreliae under natural conditions.

| Mosquito species | Ae.
vexans | Ae.
cantans | Aedes
spp. | C. pipiens
molestus | |
|---|--------------------|----------------|---------------|------------------------|--------------|
| Season of collection | А | А | А | А | S |
| Infection rate | ⁺ 4/98 | 0/21 | 0/21 | 21/480 | 14/400 |
| No. of borreliae
per mosquito
1 - 10
10 - 100
100 - 1 000
>> 1 000 | **2
1
1
- | | | 14
4
2
1 | 10
4
- |

*No. infected/No. examined
**No. of individual mosquitoes
 A - August-October 1990, S - March-April 1991

(J. Halouzka)

A-2 SAMPLE CONTROLLED SEROPREVALENCE STUDY OF LYME INFECTION IN BELGIUM

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AIM OF THE STUDY

Little is known concerning epidemiology and geographical spread of Lyme infection in large Belgian standard population groups, the clinical relevance of seropositivity and its possible correlation with frequent open air activities and/or history of a previous tick bite. We tried to answer these questions with a two-sample prospective serosurvey performed in Belgium between 1987 and 1988 in the Belgian Army.

MATERIAL AND METHODS

A/ COMPOSITION OF THE STUDY GROUPS

A group of Belgian paratroopers ("Para")(total N° of sera : 1,126) with frequent open field activities was considered as a high-risk group and compared with an age- and sex-matched control group of the Belgian Medical Service ("MS") (total N° of sera : 1,569) with less or no open air activities. All study subjects were male asymptomatic draftees between 18 and 28 years (mean age 20.9 years \pm 4.2 years) considered at good health after having passed initial conscription medical screening procedures. In 200 "Para", respectively 401 "MS", a double scrum sampling with a mean interval period of 10 months could be carried out.

B/ SEROLOGICAL TECHNIQUES

Screening of the seta for IgG antibodies against Borrelia burgdorferi (Bb) was performed with a home-made ELISA kit and confirmed with IFA. The antigen used in both kits was gained from an European N34 Bb-strain. For ELISA, the antigen was further sonicated, centrifuged and filtrated, before addition of conjugate (Sigma anti-human IgG) and substrate (Sigma Phosphatase n° 104). O.D. reading was done at 405 nM. Results were expressed as the ratio of the observed O.D. value divided by the cut-off O.D. value. ELISA ratio values >1.5 were considered as borderline positive, and >2.0 as clearly positive.

C/ TICK COLLECTION TECHNIQUES

Ticks were obtained by "flagging" different grass or low brush areas in Belgium during summer months.

RESULTS

On a total of 2,695 sera, 95 (3.25%) appeared clearly positive (ELISA ratio >2.0) whereas 76 appeared borderline positive (2.0 < ELISA ratio > 1.5), giving an overall seroprevalence of 6.35% (171 out of 2,695). There was no statistically significant difference between seroprevalence in the risk group "Para" (2.9%) vs the control group "MS" (3.9%) ($X^2 = 2.16 \text{ p} > 0.10$). However, a higher rate of seroconversions was noted in the risk group "Para" (3 out of 200 or 1.49%) vs the control group "MS" (1 out of 401 or 0.25%). None of the seropostive nor of the seroconverted study subjects presented clinical symptoms or a suggestive illness history.

When seroprevalences were calculated according to the provincial distribution of the subjects' habitat and/or the finding places of the vector <u>IXODES RICINUS</u> in Belgium, southern provinces appeared slightly more preponderant for Bb-seroprevalence and tick density (Fig 1).

No correlations were found between scropositivity and recent (3 years) open field activities, the presence or not of house pets etc. in a total of 1,469 study subjects who fully and adequately completed their computer questionnaires. Of those, 116 (7.9%) mentioned a tick bite in their recent history (Table 1). However, Lyme scropositivity (ELISA ratio >2.0) was NOT significantly higher in the group with a previous tick bite (5/116 or 4.3%) vs the group without (51/1,353 or 3.8%), nor was there any significant difference between the 2 subgroups "Para" and "MS" of the study (Table 1) or between those with a tick bite, followed by Erythema chronicum migrans (ECM) or not (data not shown).

CONCLUSIONS

1. In Belgium, seroprevalence of IgG Borrelia burgdorferi Ab (overall positivity 6.35 %, clear positivity [ELISA ratio > 2.0] 3.52%) in asymptomatic young Belgian males was correlated with the geographical localisation of their habitats, but not with limited periods of open air activities before (3 years) or during (10 months) the study. 2. Clear seropositivity (ELISA ration > 2.0) nor even seroconversion (2-sample study) had any clinical relevance, nor could a suggestive illness history be elicited in either of these 2 groups (total N° of subjects 95, respectively 4).

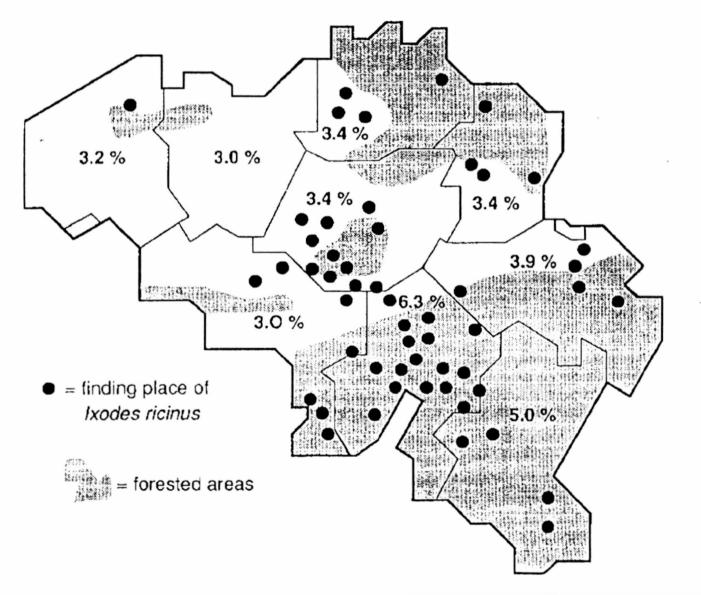
3. A previous history of a tick bite could NOT be confirmed as an important factor for developing Bb-seropositivity.

A MOIO II

| | | Total
N° | With
Tick bite
N° % | | ELISA
positive
N° % | | Without
Tick bite
N° % | | ELISA
positive
N° % | |
|----|-------|-------------|---------------------------|------|---------------------------|-----|------------------------------|------|---------------------------|-----|
| | Para | 563 | 59 | 10.5 | 3 | 5.1 | 504 | 89.5 | 23 | 4.6 |
| | MS | 906 | 57 | 6.3 | 2 | 3.5 | 849 | 93.7 | 28 | 3.3 |
| 66 | Total | 1,469 | 116 | 7.9 | 5 | 4.3 | 1,353 | 92.1 | 51 | 3.8 |

LEGEND : Comparison between a high-risk group "Para" and a control group "MS", with a previous tick bite history or not, and the presence in their sera of Bb-antibodies (ELISA ratio > 2.0).

Fig. 1. Borrelia burgdorferi IgG ELISA prevalence in Belgium according to the study subjects' habitat per province and the finding places of the vector Ixodes Ricinus. Data from a total of 2,695 sera of healthy young male Belgian military examined between 1987 and 1988.



CVIND

CTISH

BUNY, VIRIDAE

Dre.Norma Evangelina Mettler #

Mi nombre viene de Uganda,donde se sisló el prototipo de una familia que es grande,la mas grande de los virus y sunque cesi todos tienen el rotulo de arbovirus hay un genero que es nuevo,sin artropodo atribuido.

Mis viriones son esfericos, de capside nelicoidal con genoma de ARN que en tres segmentos están que son monocatenarios en negativo ademas que acompañan proteinas que pueden ser tres o mas.

Como mis miembros son mucnos,mas de un cuarto de millar varios tamaños registro de acuerdo a especie viral pero en promedio les aigo que mis viriones ostentan de 90 a cientoveinte nanimetros con cubierta.

Cinco generos componen a esta iamilia viral los BUNYA,NAIRO y los UUMI que con el PHIEBO irán todos ellos arbovirus que acoplaron ademas al genero de los HANTA y a otros sin agrupar

a algún genero asignado y a putativos nomas aunque algunos ya poseen un antigeno grupal y esperan ser emplazados,mas tarde,sí,lo serán si no en uno de estos cinco,otros géneros crearán.

Los que no son arbovirus los lidera el HANTAAN que es el que causó en Corea esa epidemia fatal que fué fiebre hemorragica con un sindrome renal y para desgracia nuestra tiene distribución mundial.

Fues estos H4NTAVIRUS que ya tienen varios miembros dos de ellos de Corea,uno de India,otro europeo tiene en nuestro continente al Prospect mill que fué aislado de unos ratones silvestres que en prados fueron hallados.

Los que están como arbovirus que ya les he mencionado son 135 del BUNYA; cien menos al PHIEBO dados 24 para el NAIRO, seis a UUAU le pusieron y 42 quedaron en familia sin un género.

Pero lo mas importante que debo comunicar es aquello que interesa al nombre o al animal causando las epidemias, las zoonosis o quizas algún caso que esporadico es casuística nomas.

Y siendo tantos los miembros para poder estudiar se agruparon por antigenos comunes para encontrar y se pusieron en grupos antigenicos que ya por simple serologia los puede catalogar.

Ln el genero primero,me refiero al BURYAVIRUS son dieciocno estos grupos que les debo mencionar lo encabeza el ANOPHELES,tiene B y tiene a son todos americanos,LAS MAYOLAS es de acá.

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Le siguen los BUNYAMWERA, que agrupa unos 26 el nombre suena africano, la membresía no lo es Kairi y Maguari son de ellos y en la Argentina tenes aislados y de mosquitos en ambiente coribbes.

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Los BWAMBA dos africanos.Los grupo C de estas tierras sels de ellos brasileños y seis del resto de America afectana los humanos con enfermedad sistemica al igual que CalifORNIA que son once de estas tierras.

y otros dos miembros del grupo la Europa tiene con ella y estos virus CALIFORNIA son importantes colegas pues a veces nos producen enfermedad con secuelas encefalitis llamadas pues al encefalo arectan

siendo el virus de LA CROSSE el que mas nos da problemas los transmiten los mosquitos y de estos bichos alados ya se na dislado en Santa Fé el que llamamos MELAO y contra el virus GUAROA anticuerpos ne encontrado.

Hay otros grupos del genero:los GUAMA que son docena los CAFIE que ya son ocho,los GAMBOA ya son tres los PATOIS llegan a siete,son todos americanos y el SAN JUAN de los GAMBOA ya fué aislado en Santa Fé.

OLIFANTSVLEI y NYANEO son grupos de otras regiones como TURLOCK, BAKAU, el KOONGOL y los TETE y de los tres que sin grupo se encuentran en BUNYAVIRUS me interesa MOJU DOS CAMPOS, de Brasil, en portugués.

Y haolando de brasileños,epidemias importantes produce el CROPOUCHE que pertenece al SIEBU fué nallado primeramente en isla del mar Caribe y a los hermanos vecinos les duebranta su salud.

El SIMBU es grupo grande, tiene 21 individuos pero es bueno que recuerden de este conjunto a dos uno es el OROPOUCHE que mencioné en epidemias y otro es el AKABANE que en el Jagón surgió.

El AKABANE produce perdida para el ganadero por suerte no en este suelo,pero de él se habló y otro que quiero traerles al recuerdo de la mente es el PARA que en el Cnaco de la Argentina se aisló

Y al final en BUNYAVIRUS les nablo del MINATITLAN otro grupo de este género que en Aéjico y Ecuador están y del genero UURUVIRUS solo les diré al paser que son seis de garrapatas y eso es todo a mencionar.

Los NAIROVIRUS contienen seis complejos agrupados dieron el nombre a este género los NAIROBI que en ovejas producen enfermedad que es zoonosis y que las deja con glomerulonefritis,leucopenia y otras quejas

Por suerte se describieron en Africa y en la India al igual que los WALYUB y los DERA GHAZI KHAN que están en esos lugares y también en Pakistán. utros son los SAKHALIN que están hasta en Janadá pero al igual que en USA solo en Ixodes se da. En cambio el grupo que es HUGHES tiene virus en America uno de ellos en Perú que es del sur y que está cerca. Pero el que importa en la ciencia es el grupo de CRIME.

que causa en esos lugares enfermedad y que es seria es una fiebre nemorragica llamada la de Crimea cientos de casos anuales produce en areas rurales aunque es también de temer infección en nospitales.

Nos quedan los PHLIBOVIRUS que les dije cuantos son y de estos treinta y ocno y como el nombre es razón son moscas de las arenas las que actúan de vector y mas de veinte en America recuerdo como lector

En Italia es que producen en Napoles y en Sicilia las fiebres de flebotomos,cada una con su estigma pero lo mas importante del genero a conocer es FIEBRE DEL VALLE DE RIFT,donde suele aparecer

Esta es una gran zoonosis que debes de retener pues tiene equivalentes entre nosotros también mata ovinos y bovinos y a los caprinos también y los cuadros proteiformes en el nombre puedes ver

y si miras el listado de este género final encontrarás que del hombre se pudo recuperar el ALENQUER y el CANDIRU,el CHAGRES y el PUNTA TURO todos ellos en America,de Brasil al norte todo.

Ademas depo informar que la Argentina agregó un grupo de virus nuevos que esta familia reunió con el nombre RESISTENCIA del lugar en que se halló BARRANQUERA y ANTEQUERA, son del Checo, lo integró.

Y doy por finalizado este relato colega pues me llevó mucho tiempo nacerlo de esta manera que es muy facil de leer de esta forma presentada ¡Si vieras los documentos usados para extractarla;

DP:Dra.Norma E.Mettier M.Larumbe 571 Martinez 1640 Buenos Aires ARGENTINA

Nota: - Investigadore de la Comisión de Investigaciones de la Pcia.de Buenos Aires y Docente Autorizada,U

LETTER TO THE EDITOR

The Institute of Tropical Medicine "Pedro Kouri" is pleased to announce to all professionals vinculated to Arthropod Borne Viral Diseases the celebration of its IVth International Course on Dengue Virus and Dengue hemorrhagic fever which is due to be held in the cadre of the IVth Cuban Congress on Microbiology and Parasitology during the second quarter of march, 1992.

Sponsored by the Cuban Ministry of Health, the Institute of Tropical Medicine "Pedro Kouri" and the Pan American Health Organization, this congress will gather around 600 delegates from Cuba and abroad and will have the participation of distinguished investigators from Europe, Latin America and the United States.

The scientific program will include lectures to be delivered by outstanding personalities, oral presentations, and posters as well as round tables covering Infectious and Tropical diseases, AIDS, Immunological Diagnosis and Molecular Biology and the cuban experience in the production, development and application of the BC anti-meningococcal vaccine. A social, cultural and sigthseeing programme will also be arranged for the delegates and accompanying persons.

The Program of the International Course of Dengue will include such topics Virological Diagnosis, Vector Control, Clinical Manifestations, Epidemiology and Molecular Biology applied to the study/diagnosis of Dengue strains; having in this opportunity the valuable participation of Prof V. Deubel CPasteur Institute of Paris). As in previously celebrated workshops, registrations fees are affordable and include transportation and accommodation facilities.

Those interested should contact to: Dr. Nereyda Cantelar de Francisco. Instituto de Medicina Tropical "Pedro Kouri" Apartado 601, Marianao 13. Ciudad de La Habana, Cuba. Telex: CU IPK 511902, FAX CU 215957. Medical Doctors, Biologist or Technologists can participate. We will meet you all in Havana with our traditional warm welcome.

Professor Dr. Gustavo Kourí Director Pedro Kourí Institute of Tropical Medicine 15th Avenue at 200 street, Siboney, La Habana, CUBA

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BULLETIN No 1

The Chinese Academy of Agricultural Sciences and the Australian Centre for International Agricultural Research are co-sponsoring the first international review of the arboviral disease, bovine ephemeral fever. Although this disease is usually not fatal it has a severe effect on milk production, draught cattle and buffalo performance and loss of meat production. In recent years it has become apparent that this virus has antigenic relatives and probably related diseases. Its impact is felt throughout Asia, Australia and Africa.

THE VENUE

The Symposium will be held in Beijing, China at the Shangri-La hotel which is where delegates will be lodged. Beijing is a focal point for international airlines. It is also a major centre for domestic airlines.

PROGRAM CONTRIBUTIONS

The symposium will cover bovine ephemeral fever in the broadest sense, to bring together the various facets of epidemiology, pathogenesis, diagnostic methods, entomology and the emerging detail of the virion. Ephemeral feverlike diseases are poorly understood but include Kotonkan and Puchong. The for international contributors and to:-

First International BEF Symposium Harbin Veterinary Research Institute 101 Maduan Street Harbin People's Republic of China

for Chinese contributors.

Two copies of full manuscripts must be in the hands of the Organisers by 15 April 1992. There is a formidable problem of translation to be done by the Organising Committee. This lead time is essential. Where possible it should be accompanied by a 5 1/4 or 3 1/2 inch floppy diskette in Microsoft Word, ASCII or WordPerfect 5.0 or 5.1. Manuscripts will be limited to 6-8 pages of single spaced typing equivalent, including all illustrations and references.

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POSTERS

These are very welcome. Dimensions will be circulated to anyone who expresses an interest in presenting a poster.

LANGUAGE

In general terms, the symposium will be based on English. The abstracts will be made available in Chinese and English. Experience has shown the difficulty of attempting simultaneous translation of presentations involving

MORE QUOTES

Isaac Bashevis Singer: "We must believe in free will. We have no choice."

Russian proverb: "A promised horse will not pull the cart."

Chinese proverb: "In multitudes of words, surely some mistakes."

Edward Abbey: "What do the coyotes mean when they yodel at the moon? What are the dolphins trying so patiently to tell us? Precisely what did those two enraptured gopher snakes have in mind when they came gliding toward my eyes over the naked sandstone? If I had been as capable of trust as I am susceptible to fear I might have learned something new or some truth so very old we have all forgotten it. They do not sweat and whine about their condition, they do not lie awake in the dark and weep for their sins."

Woody Allen: "I took a course in speed reading and was able to read 'War and Peace' in twenty minutes. It's about Russia."

Woody Allen: "If only God would give me some clear sign! Like making a large deposit in my name in a Swiss bank."

"The Girls Next Door": "Love'll get you through times with no money better'n money will get you through times with no love."

Bill (The Winery), describing two cabernets: "The difference between the two wines is like the difference between having a conversation with Broderick Crawford and having one with Leslie Howard."

Melvin Gilliam of Oklahoma State University, about free throws: "I think the trouble is the name. If they're free, they ought to give them to you."

Arthur M. Schlesinger on Aaron Burr: "A man of undoubted talents who, however, was trusted by no one in the long course of American history except for his daughter Theodosia and Gore Vidal."

Ashley Montagu: "Science has proof without any certainty. Creationists have certainty without any proof."

Robert A. Heinlein ("The Notebooks of Lazarus Long"): "A human being should be able to change a diaper, plan an invasion, butcher a hog, con a ship, design a building, write a sonnet, balance accounts, build a wall, set a bone, comfort the dying, take orders, give orders, cooperate, act alone, solve equations, analyze a new problem, pitch manure, program a computer, cook a tasty meal, fight efficiently, die gallantly. Specialization is for insects."

Jay Leno: "The Suzuki Samurai is on its back so often its a wonder Jimmy Swaggert doesn't buy one."

Homer D. King: "Outlawing all atomic weapons could be a magnificent gesture. However, it should be remembered that Gettysburg had a local ordinance forbidding the discharge of firearms."

tenebrific (adj.) causing or producing darkness

A. Bartlett Giamatti: "Baseball is about homecoming. It is a journey by theft and strength, guile and speed, out around first to the far island of second, where foes lurk in the reeds and the green sea suddenly grows deeper, then to turn sharply, skimming the shallows, making for a shore that will show a friendly face, a color, a familiar language and at third, to proceed, no longer by paths indirect but straight to home."

A. Bartlett Giamatti: "The appeal of baseball is intimately wrapped up with one's youth. Baseball is very much about being young again in a harmless way. And one of its core appeals is to remind America of a time when it was young. You fly over a major city at night in the summer and suddenly you'll see that green oasis that reminds everybody of baseball's basic mythology: We come from a rural, simpler America. What's home? Home is longing for when you were happy because you were younger."

A. Bartlett Giamatti: "The hunger for home makes the green geometry of the baseball field more than simply a metaphor for the American experience and character. The baseball field and the game that sanctifies boundaries, rules, and law, and appreciates cunning, theft, and guile; that exalts energy, opportunism, and execution, while paying lip service to management, strategy, and long-range planning, is finally closer to an embodiment of American life than the mere sporting image of it."

A. Bartlett Giamatti: "Baseball began in a bright green field with an ancient name when this country was new and raw and without shape, and it has shaped America by linking every summer from 1846 to this one, through wars and depressions and seasons of rain. Baseball is one of the few enduring institutions in America that has been continuous and adaptable and in touch with its origins. As a result, baseball is not simply an essential part of this country; it is a living memory of what American culture at its best wishes to be."

Gabriel García Márquez (Chronicle of a Death Foretold): "In that smile, for the first time since her birth, Angela Vicario saw her as she was: a poor woman devoted to the cult of her defects."

Oscar Wilde: "Horse sense is what keeps horses from betting on what people will do."

"How many New Yorkers does it take to screw in a light bulb?" "None of your damned business."

Marshall Lumsden: "There is only one thing that can keep growing without nourishment: the human ego."

Alex Comfort: "We may eventually come to realize that chastity is no more a virtue than malnutrition."

Beilby Porteus: "One murder makes a villain, millions a hero."

Edward Abbey: "We need more predators. The sheepmen complain, it is true, that the coyotes eat some of their lambs. This is true but do they eat enough? I mean, enough lambs to keep the coyotes sleek, healthy and well fed. That is my concern. As for the sacrifice of an occasional lamb, that seems to me a small price to pay for the support of the coyote population. The lambs, accustomed by tradition to their role, do not complain; and the sheepmen, who run their hooved locusts on the public lands and are heavily subsidized, most of them as hog-rich as they are pigheaded, can easily afford these trifling losses."

H.L. Mencken: "A cynic is a man who, when he smells flowers, looks around for a coffin."

Oscar Wilde: "A cynic is a man who knows the price of everything and the value of nothing."

Woody Allen: "I'm astounded by people who want to "know" the universe when they can't find their way around Chinatown."

Ed Sanders: "The writing of more than 75 poems in any fiscal year should be punishable by a fine of \$500."

Unknown: "The difference between genius and stupidity is that genius has its limits."

Unknown: "There's nothing wrong with teenagers that reasoning with them won't aggravate."

Unknown: "A manuscript, like a foetus, is never improved by showing it to somebody before it is completed."

Unknown: "They were such a progressive couple they tried to adopt a gay baby."

Mark Twain: "I would rather be with Lillian Russell stark naked than Ulysses S. Grant in full military regalia."

Jean Giraudoux: "Only the mediocre are always at their best."

Oscar Wilde: "Seriousness is the only refuge of the shallow."

Ambrose Bierce: "Man: An animal whose chief occupation is extermination of other animals and his own species, which, however, multiplies with such insistent rapidity as to infect the whole habitable earth and Canada." (For Harvey and Leslie ©)

David Steinberg: "My father never lived to see his dream come true of an all Yiddish-speaking Canada." (For Harvey and Leslie ©)

Elayne Boosler: "I'm just a person trapped inside a woman's body."

David Letterman: "Fall is my favorite season in Los Angeles, watching the birds change color and fall from the trees." Samuel Butler: "Man is the only animal that laughs and has a state legislature."

Frank Layden: "We got a pretty good reading on one player when we looked at the information sheet he had filled out for the public relations department. On the line for church preference he had written 'red brick'."

Henry Miller: "To enter life by way of the vagina is as good a way as any."

Russian proverb: "Since when does the fiddle pick the tune?"

When Pastor Niemoeller was released, he was asked, "How did the world let this happen?" He responded: "In Germany, the Nazis first came for the Communists, and I didn't speak up because I wasn't a Communist. Then they came for the Jews, and I didn't speak up because I wasn't a Jew. Then they came for the trade unionists, and I didn't speak up because I wasn't a trade unionist. Then they came for the Catholics, and I didn't speak up because I was a Protestant. Then they came for me, and by that time there was no one left to speak for me."

Percy Bysshe Shelley:

"I met a traveler from an antique land Who said, 'Two vast and trunkless legs of stone Stand in the desert. Near them, on the sand, Half sunk, a shattered visage lies, whose frown, And wrinkled lip, and sneer of cold command, Tell that its sculptor well those passions read Which yet survive. Stamped on those lifeless things, The hand that mocked them and the heart that fed. And on the pedestal these words appear: "My name is Ozymandias, King of kings: Look on my works, ye mighty, and despair." Nothing beside remains. Round the decay Of the colossal wreck, boundless and bare, The lone and level sands stretch far away.'"

Russian proverb: "Lie down with dogs and you will get up with fleas."

(A Canticle for Leibowitz) Non cogitamus, ergo nihil sumus: We do not know, therefore we are not.

Harry the Horse: "A guy must pay his bookmaker, no matter what."

--- and by the way, the thoughtful Patricia Nuttall has given me a copy of "The Oxford Dictionary of Modern Quotations".