

## IDEA EXCHANGE

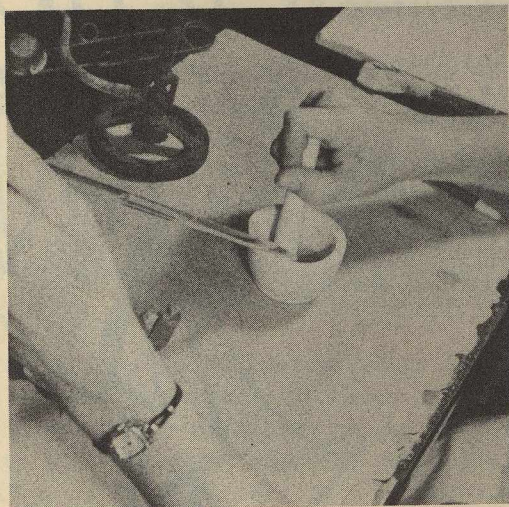
*This section of the BULLETIN is being devoted to new ideas which have proved of value in CDC activities. The purpose of this section is to exchange ideas among operating units of CDC. Contributions from the field are solicited. Any idea developed locally that can have wider application, even if not new, is welcome. Send it in!*

### MOUSE INOCULATION TEST

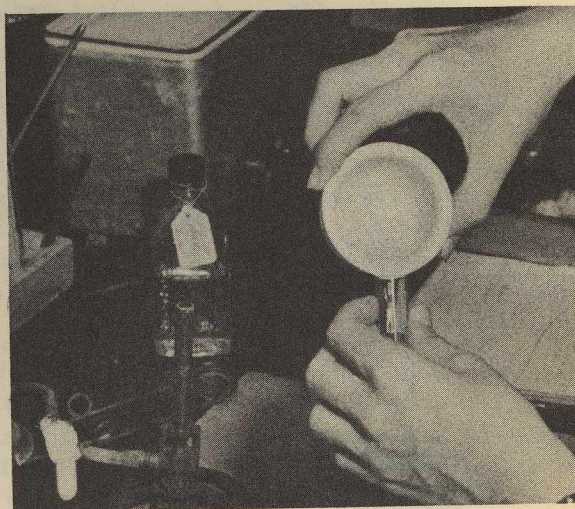
for

### RABIES DIAGNOSIS ★

Negri bodies are not always demonstrable in animals which have died of rabies. The mouse inoculation test is recommended in Negri-negative brains where the history is suggestive of rabies or when confirmation of diagnosis is indicated. The test is simple and inexpensive.



1. Brain tissue from Ammon's horn, cerebral cortex, and cerebellum of the suspected animal is pooled and ground in a mortar under aseptic conditions. Saline is added to make a 10% suspension.

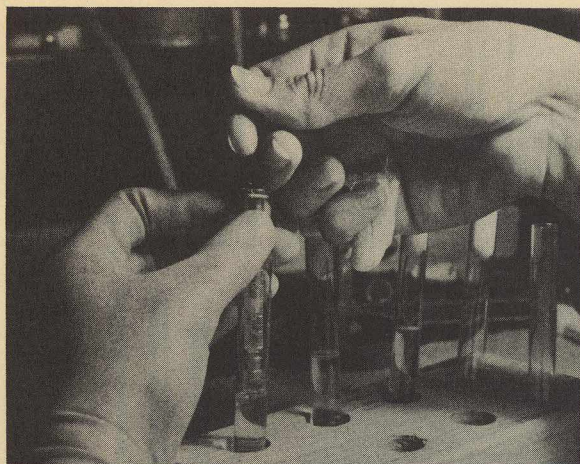


2. When tissue is ground thoroughly, the suspension is decanted into a sterile tube.

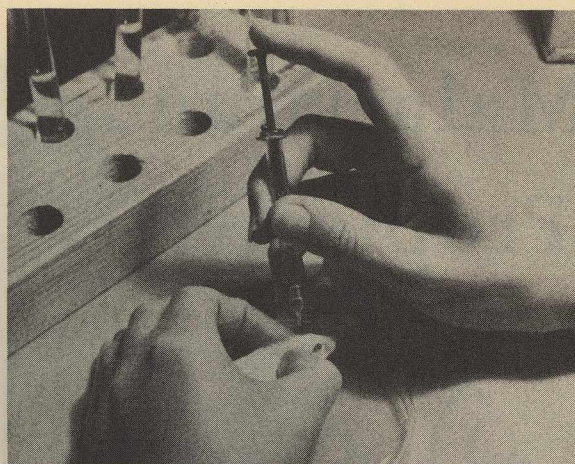
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*\*Although not "new" and not developed in CDC, this test is described in an effort to promote its wider use.*

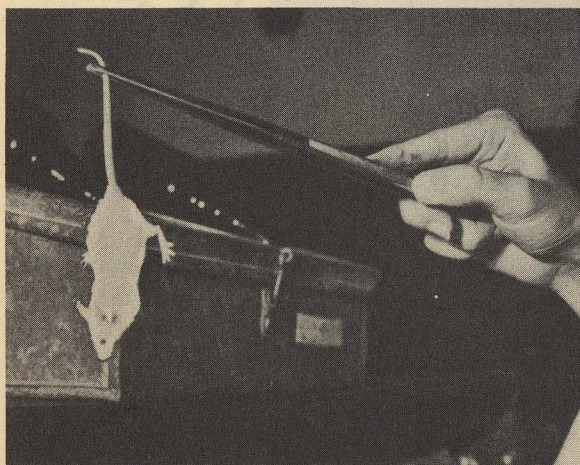




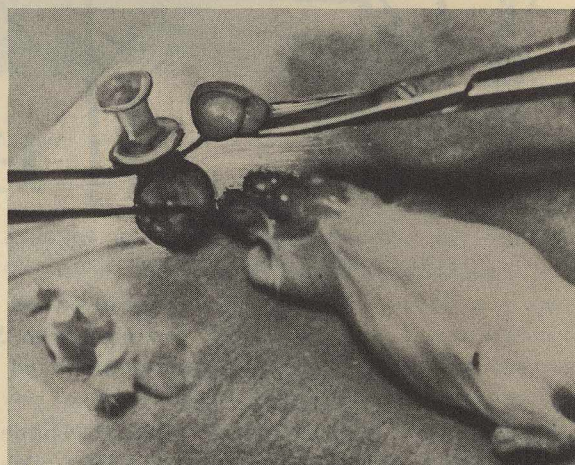
3. The saline suspension is drawn into a sterile  $\frac{1}{4}$  ml. tuberculin syringe fitted with a  $\frac{1}{4}$  inch, 27 ga. needle.



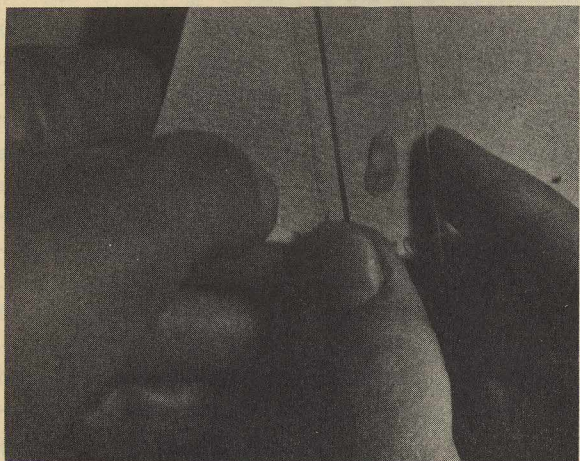
4. Each of three mice are injected intracerebrally with 0.03 ml. of the suspension.



5. Mice are kept under close observation for 14 days or until rabies symptoms develop.



6. Brains are removed from mice and lateral sections cut from the hemispheres.



7. Tissue films are prepared by the impression method.



8. Slides are stained with Sellers' stain, washed, dried, and examined for Negri bodies.