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Serum Perfluoroalkyl Substances and Cardiometabolic Consequences in Adolescents Exposed to the World Trade Center Disaster and a Matched Comparison Group

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Abstract

Background—Large amounts of various chemical contaminants, including perfluoroalkyl substances (PFASs), were released at the time of the World Trade Center (WTC) disaster.

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Acquisition of data: Tony Koshy, Joseph Gilbert, Lauren Burdine

Main analysis and interpretation of data: Leonardo Trasande, Xiaoxia Han

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Thousands of children who lived and/or attended school near the disaster site were exposed to these substances but few studies have examined the possible consequences related to these exposures.

Objectives—To examine the relationship of PFASs serum levels with cardiometabolic profile in children and adolescents enrolled in the World Trade Center Health Registry (WTCHR) and a matched comparison group.

Methods—We evaluated WTCHR enrollees who resided in New York City and were born between September 11, 1993 and September 10, 2001, and a matched comparison group consisting of individuals who were ineligible for WTCHR participation upon distance of their home, school or work from the WTC and lack of participation in rescue and recovery activities. Matching was based on date of birth, sex, race, ethnicity, and income. We assessed exposure to PFASs, as measured by serum levels and association with cardiometabolic profile as measured by arterial wall stiffness, body mass index, insulin resistance, fasting total cholesterol, HDL, LDL and triglycerides.

Results—A total of 402 participants completed the study and serum samples were analyzed from 308 participants, 123 in the WTCHR group and 185 in the comparison group. In multivariable regression analysis, after adjusting for relevant confounders, we observed a significant, positive association of perfluorooctanoic acid (PFOA) with triglycerides (beta coefficient = 0.14, 95% CI: 0.02, 0.27, 15.1 percent change), total cholesterol (beta coefficient = 0.09, 95% CI: 0.04, 0.14, 9.2 percent change), and LDL cholesterol (beta coefficient = 0.11, 95% CI: 0.03, 0.19, 11.5 percent change). Perfluorohexanesulfonic acid levels were associated with decreased insulin resistance (beta coefficient = -0.09, 95% CI: -0.18, -0.003, -8.6 percent change); PFOA and perfluorononanoic acid were associated with increased brachial artery distensibility.

Conclusions—This research adds to our knowledge of the physical health impacts in a large group of children exposed to the WTC disaster. Abnormal lipid levels in young adults might be an early marker of atherosclerosis and cardiovascular diseases and our findings highlight the importance of conducting longitudinal studies in this population.

Keywords

perfluoroalkyl substances; adolescents; World Trade Center disaster; cardiometabolic consequences

1. Introductiona

During the terrorist attack on the World Trade Center (WTC) on September 11, 2001, and in the months that followed, children in lower Manhattan were exposed to large amounts of contaminants such as particulate matter, heavy metals and persistent organic pollutants

^aBody Mass Index (BMI); HDL (High-density lipoprotein); Limits of Detection (LODs); N-methylperfluoro-1-octanesulfonamidoacetic acid (N-meFOSAA); N-methyl perfluorooctanesulfonamido acetic acid (N-meFOSAA); New York State Department of Health (NYSDOH); NYC Department of Health & Mental Hygiene (NYC DOHMH); Perfluoroalkyl substances (PFASs); Perfluorodecane sulfonate (PFDS); Perfluorodecanoic acid (PFDA); Perfluorodecanoic acid (PFDADA); Perfluoroneptanoic acid (PFHpA); Perfluorohexanesulfonic acid (PFHxS); Perfluorononanoic acid (PFNA); Perfluorooctane sulfonamide (PFOSA); Perfluoroctanesulfonic acid (PFOS); Perfluorooctanoic acid (PFOA); Perfluoroundecanoic acid (PFUnDA); World Trade Center (WTC); WTC Health Registry (WTCHR)

(POP).¹ Elevated concentrations of perfluoroalkyl substances (PFASs), a group of chemicals widely used in various building and construction material,² upholstery, carpet, and nonstick cookware,^{3,4} have been found in window films and in samples of dust, water, sediment, and sewage collected in and around the WTC site.^{5–7} The US Environmental Protection Agency (EPA) has recently established drinking water health advisories of 0.07 micrograms per liter for perfluorooctanoic acid (PFOA) and perfluorooctanesulfonic acid (PFOS), two of the most environmentally persistent PFASs. ^{8,9}

We recently documented that the children enrolled in the World Trade Center Health Registry (WTCHR) had higher levels of serum PFASs than matched comparisons years after the WTC disaster. ¹⁰ This observation is in agreement with data showing that PFASs persist in the environment and in humans, with half-lives ranging from 3–5 years to 8 years and longer. ^{11,12} It is also consistent with studies of responders that documented increases in PFASs in relationship to WTC-exposure. ¹³

The consequences of WTC-related PFASs exposure are less clear. Current evidence suggests that PFASs interfere with important biological processes, specifically activation of alpha-and gamma-peroxisome proliferator activated receptors, ¹⁴ which play key roles in lipid and carbohydrate metabolism and are also involved in lipid transport, cholesterol synthesis, cell communication, inflammation and oxidative stress. ^{15,16} Human studies have shown a positive association between levels of PFASs and total and non-high-density cholesterol in the NHANES, despite the relatively low level of exposure. ¹⁷ In addition, among PFASs, concentrations of perfluorooctanesulfonic acid (PFOS) and perfluorononanoic acid (PFNA) have been associated with lower levels of IGF-1 in boys and girls 6–9 years of age. ¹⁸ In turn, decreased levels of IGF-1 have been associated with metabolic syndrome ¹⁹ and increased risk of cardiovascular events in later life. ²⁰

The aim of the current study was therefore to examine the relationship of serum PFASs levels with cardiometabolic profile, as measured by blood lipids, insulin resistance, arterial stiffness, and body mass index (BMI) in children and adolescents enrolled in the WTCHR and a matched comparison group, while controlling for an array of possible confounding factors. Cardiovascular risk factors such as insulin resistance and hypertension do not typically emerge until adolescence, and identifying the adolescents who are at risk and intervening to modify diet, treat with medications and/or increase physical activity may help reduce the burden of subsequent adult chronic disease in this vulnerable group.

For the purpose of this analysis we combined the two study populations (WTCHR and comparison group), which allowed us to increase the range of exposures studied, but no comparisons were made between these two populations with regard to the outcomes of interest.

2. Methods

2.1 Study Population

WTCHR population—This group consisted of WTCHR enrollees who resided in New York City and were born between September 11, 1993 and September 10, 2001. Participants

were enrolled with the assistance of the New York City Department of Health (NYCDOHMH) using mail, email, phone, and in-person communication methods. Details of recruitment process are described elsewhere. ¹⁰

Comparison group—This group consisted of individuals who were not eligible for WTCHR participation due to their specific location on the morning of 9/11.²¹ We aimed to recruit a matched comparison group and utilized the WTCHR's 2011–12 survey cycle as a matching tool. We created a table of desired frequency distribution of the matching variables for comparisons using age (0–2, 3–5 or 6–8 years-old on 9/11/2001, with age 8 years being the upper bound for age restriction), sex, race (White, African-American, Asian, other), ethnicity (Hispanic, non-Hispanic) and income (<\$25,000, \$25,000). Multiple recruitment strategies were used, ¹⁰ and a screening questionnaire was used to determine individuals' eligibility based on the frequency-matching table. Individuals were excluded as matched comparisons if they otherwise could qualify for enrollment in the WTCHR due to location on 9/11.

Exclusion criteria—Participants were not considered eligible for either the WTCHR or the control group if any of the following was present: i) inability to follow study procedures for measurement of arterial stiffness; ii) serious lung or heart condition; iii) heart or lung surgery; iv) pregnancy.

Institutional Review Board Approval—The study was reviewed and approved by the NYU School of Medicine Institutional Review Board, as well as research committees at Bellevue and Gouverneur Hospital Centers. Adolescents under 18 years of age provided informed assent forms along with parental informed consent forms before undergoing study procedures. A Certificate of Confidentiality was obtained to protect participant privacy. The study was approved by New York State Department of Health (NYSDOH) for the analysis of serum samples.

2.2 Study visits

Visits took place on evenings, weekends and during school holidays to maximize convenience, either in 1 or 2 visits at the study site. Participants were instructed to fast for six hours before study visits, and to avoid food, caffeine-containing products, and sugary drinks. After providing informed consent, the following were performed: a fasting blood draw (6 hours); anthropometric measurements; and brachial artery distensibility/pulse wave velocity measurements.

Measurement of PFASs—Eleven PFASs were measured in serum using a solid phase extraction (SPE) procedure and high-performance liquid chromatograph interfaced with an electrospray tandem mass spectrometer, using the methods similar to those described elsewhere, ^{22,23} and documented in our previous manuscript. ¹⁰ For further details related to the methodology, please see Supplemental Material. The following PFASs were measured: Perfluorohexanesulfonic acid (PFHxS); n-methyl perfluorooctanesulfonamido acetic acid (N-meFOSAA); perfluorooctane sulfonamide (PFOSA); perfluorooctanesulfonate (PFOS); perfluoroheptanoic acid (PFHpA); perfluorooctanoic acid

(PFOA); perfluorononanoic acid (PFNA); perfluorodecanoic acid (PFDA); perfluoroundecanoic (PFUnDA); and perfluorododecanoic acid (PFDoDA).

2.3 Assessment of Cardiometabolic profile

Anthropometric measures—Weight and height were measured using calibrated stadiometers (Shorr Productions, Olney, MD) and scales (Seca model 881; Seca Corp., Hanover, MD). Body Mass Index Z-scores were derived from 2000 Centers for Disease Control and Prevention (CDC) norms, incorporating height, weight and sex; overweight and obese were categorized as BMI Z-score 1.036 and 1.64, ²⁴ which correspond to the 85th and 95th age- and sex-adjusted percentiles.

Dietary data and physical activity—To obtain dietary data, participants completed a web-based version of the Diet History Questionnaire II (DHQ II), a publicly available food frequency questionnaire (FFQ) developed by the National Cancer Institute, which has been previously validated. ²⁵ Participants also completed a three-day physical activity diary, based on the International Physical Activity Questionnaire-Short Last Seven Days, which is well validated. ²⁶ Physical activity data from the diary were converted into energy expenditure estimates as MET using published values. ²⁷

Blood Pressure (BP) and Brachial Artery Distensibility (BrachD)—Brachial artery distensibility (BrachD) measurement is a rapid method of accurately assessing the relative stiffness of a peripheral artery. A lower value indicates a stiffer vessel. The DynaPulse Pathway instrument derives BrachD and BP using the technique of pulse waveform analysis of arterial pressure signals obtained from a standard cuff sphygmomanometer. Following a 5 minute rest period, a BP cuff appropriate for the subject's upper arm size was applied, and three automatic recordings of systolic, diastolic, mean arterial BP and heart rate were obtained. Off-line analyses of brachial artery pressure curve data were then performed by Pulse Metric, Inc. using an automated system to derive parameters from the pulse curves to calculate BrachD. Because BP varies widely by age, sex and height, we calculated systolic/diastolic BP Z-scores from mixed-effects linear regression models derived using data from 1999–2000 National Health and Nutrition Examination Survey. Survey.

Arterial Wall Stiffness Assessment—Pulse wave velocity (PWV) reflects the speed for the pressure wave generated by cardiac ejection to reach the periphery. A higher value indicates a stiffer vessel. PWV was measured by obtaining the arterial pulse waveform at the common carotid and femoral arteries using the SphygmoCor CPV System (AtCor Medical, Sydney, Australia).³¹ Arterial waveforms gated to the R-wave on the ECG tracing are recorded from the carotid then distal artery of interest, and PWV is then calculated as the difference in the carotid-to-distal path length divided by the difference in R-wave-to-waveform foot times. The SphygmoCor CPV System was also used to measure central aortic pressure and the Augmentation Index (AIx), a vascular parameter incorporating both central stiffness and wave reflections (a higher value indicates arterial dysfunction).³²

Blood lipid profile, glucose and insulin—We measured fasting total cholesterol, triglycerides, HDL, LDL, insulin, and glucose. We examined continuous as well as categorical abnormal values for lipid levels, applying cut-off points for HDL of <40 mg/dL and 100 mg/dL for triglycerides, as recently done to assess components of the metabolic syndrome in analyses of adolescents in 2001–2006 NHANES. For insulin resistance we used the validated homeostatic model assessment of insulin resistance (HOMA-IR), calculated by dividing the product of insulin (μ U/mL) and glucose (mMol/L) by 22.5. $^{34-36}$

2.4 Covariates

Information on other covariates including race/ethnicity (White, African American, Asian, Other and Hispanic) and sex (male or female) was obtained through questionnaire. Exposure to tobacco smoke was evaluated by saliva cotinine concentration and questionnaire. Salivary cotinine was analyzed using a highly reliable (r=.99 compared with serum) and sensitive (limit of detection 0.05 ng/mL) test from Salimetrics, Inc. (State College, PA). Cotinine was measured as a continuous variable, and categorized into low (<0.15 ng/mL), medium (0.15 to < 2.32 ng/mL) and high (2.32 ng/mL) categories, using established conventions. For subjects without saliva cotinine concentration, we categorized using questionnaires (low: no smoker and no secondhand smoke exposure; medium: no smoker but secondhand smoke exposure; high: smoker).

2.5 Statistical Analysis

We conducted descriptive, univariate, and multivariate analyses with R Statistical Software (version 3.3.1). Chi-square test or Fisher exact test was used to compare the sociodemographic variables of two study populations. Wilcoxon Rank Sum test was used to compare caloric intake, physical activity levels, cardiometabolic markers, and serum PFASs between the two groups. PFAS concentrations were log-transformed to account for skewed distribution; following published practices, levels less than the LOD were imputed to be $LOD/\sqrt{2}$, and we limited our statistical analyses to PFASs detected in 50% of the samples. Simple linear and logistic regression was used to compare cardiometabolic profiles (blood lipids, BMI, PWV, AIx, and insulin resistance) by serum PFASs. Multiple linear regression or multiple logistic regression was used for continuous or discrete outcomes, controlling for sex, race, caloric intake, physical activity, smoke exposure, BMI category (but not for BMI related outcomes). All statistical tests were two-sided, and p values were considered significant if < 0.05.

3. Results

Figure 1 provides an overview of the enrollment process for the WTCHR study population and matched comparison group.

WTCHR individuals enrolled in this study were more likely to be older and to be from low income families (P<0.001) than those excluded. There were no differences in sex, race or ethnicity between WTCHR individuals who participated in this study and nonparticipants (Table S1, Supplemental Material). In total, 180 children from the WTCHR and 222 sociodemographically matched controls were included in the analysis; among them,

information on PFAS was available in 185 and 123 individuals, respectively (participants with consent to venous blood sampling). Characteristics of the two study populations are presented in Table 1.

Compared to the WTCHR cohort, participants in the comparison group were more likely to be female (59.9% vs. 46.1%). Caloric intake was higher in the WTCHR population (1621 calories) than the comparison group (1535 calories) but overweight and obesity status were more likely in the comparison than the WTCHR group (P=0.045). Table 2 shows the characteristics of the participants who agreed to venous sampling.

Several significant associations were detected between single chemicals and cardiometabolic outcomes in univariable analysis (Table S2, Supplemental Material), and most remained significant after adjusting for confounders in the multivariable model (Table 3).

Adjusted analyses showed consistent associations between higher serum PFASs and higher lipid levels. PFOS was associated with higher total and LDL cholesterol (beta coefficient for total cholesterol in natural log scale=0.082, 95% CI: 0.047, 0.117, corresponding to 8.5 percent change; beta coefficient for LDL=0.102, 95% CI: 0.046, 0.159, 10.7 percent change, respectively) as well as higher HDL cholesterol (beta coefficient=0.064, 95% CI: 0.003, 0.125, 6.6 percent change). PFOA was associated with higher triglycerides (beta coefficient = 0.141, 95% CI: 0.017, 0.265, 15.1 percent change), total cholesterol (beta coefficient = 0.088, 95% CI: 0.039, 0.137, 9.2 percent change), and LDL cholesterol (beta coefficient = 0.109, 95% CI: 0.031, 0.187, 11.5 percent change). Similar associations were observed with PFNA, PFDA, and PFUnDA. Higher levels of PFHxS were significantly associated with decreased insulin resistance (beta coefficient = -0.090, 95% CI: -0.176, -0.003, -8.6 percent change), and higher LDL cholesterol (beta coefficient = 0.049, 95% CI: 0.007, 0.091, 5.0 percent change). We also detected an association between higher levels of PFOA and PFNA and increased brachial artery distensibility (beta coefficient for % change/ mmHg=0.453, 95% CI 0.038, 0.868; beta coefficient= 0.343, 95% CI: 0.016, 0.670, respectively). No association was detected between serum levels of PFASs and PWV and AIx. PFUnDA was associated with lower odds of being overweight (odds ratio per unit increase in natural log PFUnDA= 0.951, 95% CI: 0.911, 0.993), but other PFASs examined in this study were not associated with BMI status. Table 4 shows the results of multivariate analyses presented as percent change in the outcome of interest for each log unit increase of the chemicals examined.

4. Discussion

This research study examined the cardiometabolic profiles of adolescents participating in the WTCHR compared to a sociodemographically-matched control group of NYC residents, to examine the potentially contributing role of PFASs exposures to cardiometabolic risks in exposed children. We have previously documented that children with subchronic dust exposure and dust cloud exposure related to the WTC disaster have higher levels of PFASs, and here we report that higher serum levels of PFASs are associated with increased blood lipid levels (triglycerides and cholesterol). ¹⁰ Since abnormal lipid levels in young adults might be an early marker of atherosclerosis and cardiovascular diseases, ⁴⁰ this population

may benefit from continuous monitoring and early interventions to prevent adverse cardiometabolic outcomes as a result of PFASs exposure.

Previous reports from cross-sectional data support the association between concentrations of PFASs and altered lipid profiles, specifically elevated plasma cholesterol and triglycerides, ^{17,41,42} although the evidence is not uniform and shows some inconsistencies in the results, also depending on the specific chemical examined. PFOS and PFOA are the two chemicals for which the evidence is the strongest, while other compounds like PFHxS and PFNA have not been studied as extensively, mainly because they are normally present in lower concentrations compared with the two PFOS and PFOA.¹⁷ Positive associations between PFOS and both HDL and LDL cholesterol have been reported among adults, 42 children and adolescents. 43,44 Similarly, positive associations between PFOA and PFOS and triglycerides have been reported in adults, ⁴⁵ as well as in children. ⁴⁶ Instead, for compounds like PFHxS the evidence is not consistent: Nelson and colleagues reported an inverse association between PFHxS and total cholesterol¹⁷ whereas others have found a significant positive association with total and LDL cholesterol. 41 In the present study we report that, in addition to PFOS and PFOA, PFHxS, PFNA and PFDA were also positively associated with increased lipid concentrations (total and LDL cholesterol). PFDA and PFUnDA, which were the two compounds with the lowest median serum concentrations among all PFASs in the WTCHR group, were both positively associated with HDL cholesterol.

With respect to insulin resistance, we detected an inverse relationship with PFHxS. Recently published data from a prospective cohort study, reporting that children with higher levels of PFASs had significantly lower insulin resistance, ⁴⁷ are consistent with our findings, and so are previous analysis from NHANES data. ¹⁷ However, other studies have reported a positive association between PFOS and insulin resistance, although this was only present in overweight children. ⁴⁸ In addition, some available data suggest that the association of PFOS with insulin resistance differs between adults and adolescents, with the former showing increased insulin resistance with higher PFOS concentrations, whereas the opposite was noted for adolescents. ⁴⁹ This sample was not large enough for stratified analysis.

The biological mechanisms underlying the associations between PFAS and lipid levels and insulin resistance is less understood. Most of our information comes from animal studies showing that PFAS have affinity for PPARa and acts as agonists to these receptors. Nonetheless, these studies also indicate that the degree of agonist effect is variable and depends on the specific compound examined. Far Affinity to PPAR γ has also been demonstrated and PPAR γ activation could potentially lead to increased insulin sensitivity, a mechanism similar to that of thiazolidinediones, which are used to in the treatment of type 2 diabetes. Despite providing valuable insight, findings of toxicological research are not directly applicable to humans and further studies are therefore warranted to elucidate the underlying mechanisms. The long-term health consequences of an increase in serum lipid levels in the ranges observed in this study are unclear. However, if confirmed in further longitudinal studies, such increments may become significant when considered at the population level, in which even small increments can result in large increases in the prevalence of hyperlipidemia, shifting the distribution of blood lipids and increasing the

number of individuals who are above the cut off points to identify hyperlipidemic individuals.

In this study we also detected an inverse association between PFOA and PFNA and increased brachial artery distensibility. To our knowledge, this is the first time that such an association is reported, since not many studies have examined the association between PFASs and vascular function. Arterial stiffness is influenced by both genetic and hormonal factors, ⁵³ and current evidence suggests an effect of PFASs on sex hormones. Increases in estradiol and decreases in testosterone with PFOA exposure have been observed in rodents ¹⁵ but the results of the few human studies conducted so far are less clear. Recently, Zhou and colleagues have reported that higher levels of PFASs are associated with lower testosterone and higher estradiol levels, and these associations seem to be more relevant in males than females. ⁵⁴ We could speculate that the associations of PFOA and PFNA with increased arterial distensibility could be partly interpreted in light of concomitant alterations in sex hormone levels which, in turn, may influence vascular stiffness.

4.1 Limitations

We cannot rule out the possibility that some of the associations could be chance findings, including the association of PFUnDA with lower odds of being overweight, since none of the other chemicals examined was associated with BMI. In addition, this study collected data at a single time point, and longitudinal studies may be more informative in assessing the extent of cardiometabolic effects related to exposure to the WTC disaster. Another limitation to interpretation is that participants in both groups experienced environmental changes in the fifteen year period following the disaster, and we cannot rule out additional factors or exposures that could contribute to explaining the associations observed here. Furthermore, PFASs increases observed in this study could be correlated with all the other chemical contaminant exposures that were associated with the WTC. We acknowledge that this is a potential confounding factor but one difficult to control in a disaster epidemiology study.

5. Conclusion

This research adds to our knowledge of the physical health impacts in a large group of children who were exposed to the WTC disaster, and pinpoints the potential high risk of atherosclerosis and cardiovascular diseases in these children as a result of PFASs exposure.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

 Landrigan PJ, Lioy PJ, Thurston G, et al. Health and environmental consequences of the world trade center disaster. Environ Health Perspect. May; 2004 112(6):731–739. [PubMed: 15121517]

- 2. Becanova J, Melymuk L, Vojta S, Komprdova K, Klanova J. Screening for perfluoroalkyl acids in consumer products, building materials and wastes. Chemosphere. Dec.2016 164:322–329. [PubMed: 27592321]
- 3. Kotthoff M, Muller J, Jurling H, Schlummer M, Fiedler D. Perfluoroalkyl and polyfluoroalkyl substances in consumer products. Environ Sci Pollut Res Int. Oct; 2015 22(19):14546–14559. [PubMed: 25854201]
- 4. Trier X, Granby K, Christensen JH. Polyfluorinated surfactants (PFS) in paper and board coatings for food packaging. Environ Sci Pollut Res Int. Aug; 2011 18(7):1108–1120. [PubMed: 21327544]
- Litten S, McChesney DJ, Hamilton MC, Fowler B. Destruction of the World Trade Center and PCBs, PBDEs, PCDD/Fs, PBDD/Fs, and chlorinated biphenylenes in water, sediment, and sewage sludge. Environ Sci Technol. Dec 15; 2003 37(24):5502–5510. [PubMed: 14717157]
- Offenberg JH, Eisenreich SJ, Gigliotti CL, et al. Persistent Organic Pollutants in Dusts That Settled at Indoor and Outdoor Locations in Lower Manhattan after September 11, 2001. Urban Aerosols and Their Impacts. 2005; 919:103–113.
- Offenberg JH, Eisenreich SJ, Gigliotti CL, et al. Persistent organic pollutants in dusts that settled indoors in lower Manhattan after September 11, 2001. J Expo Anal Environ Epidemiol. Mar; 2004 14(2):164–172. [PubMed: 15014547]
- EPA. [accessed 14 July 2017] Drinking water health advisory for PFOS. Available at https://www.epa.gov/sites/production/files/2016-05/documents/pfos_health_advisory_final_508.pdf
- EPA. [accessed 14 July 2017] Drinking water health advisory for PFOA. Available at https://www.epa.gov/sites/production/files/2016-05/documents/pfoa health advisory final 508.pdf
- Trasande L, Koshy TT, Gilbert J, et al. Serum perfluoroalkyl substances in children exposed to the world trade center disaster. Environ Res. Jan 16.2017 154:212–221. [PubMed: 28104511]
- Olsen GW, Burris JM, Ehresman DJ, et al. Half-life of serum elimination of perfluorooctanesulfonate, perfluorohexanesulfonate, and perfluorooctanoate in retired fluorochemical production workers. Environ Health Perspect. Sep; 2007 115(9):1298–1305. [PubMed: 17805419]
- Zhang Y, Beesoon S, Zhu L, Martin JW. Biomonitoring of perfluoroalkyl acids in human urine and estimates of biological half-life. Environ Sci Technol. Sep 17; 2013 47(18):10619–10627.
 [PubMed: 23980546]
- Tao L, Kannan K, Aldous KM, Mauer MP, Eadon GA. Biomonitoring of perfluorochemicals in plasma of New York State personnel responding to the World Trade Center disaster. Environ Sci Technol. May 1; 2008 42(9):3472–3478. [PubMed: 18522136]
- Zhang L, Ren XM, Wan B, Guo LH. Structure-dependent binding and activation of perfluorinated compounds on human peroxisome proliferator-activated receptor gamma. Toxicol Appl Pharmacol. Sep 15; 2014 279(3):275–283. [PubMed: 24998974]
- Lau C, Anitole K, Hodes C, Lai D, Pfahles-Hutchens A, Seed J. Perfluoroalkyl acids: a review of monitoring and toxicological findings. Toxicol Sci. Oct; 2007 99(2):366–394. [PubMed: 17519394]
- 16. Yao X, Zhong L. Genotoxic risk and oxidative DNA damage in HepG2 cells exposed to perfluorooctanoic acid. Mutat Res. Nov 10; 2005 587(1–2):38–44. [PubMed: 16219484]
- 17. Nelson JW, Hatch EE, Webster TF. Exposure to polyfluoroalkyl chemicals and cholesterol, body weight, and insulin resistance in the general U.S. population. Environ Health Perspect. Feb; 2010 118(2):197–202. [PubMed: 20123614]
- Lopez-Espinosa MJ, Mondal D, Armstrong BG, Eskenazi B, Fletcher T. Perfluoroalkyl Substances, Sex Hormones, and Insulin-like Growth Factor-1 at 6–9 Years of Age: A Cross-Sectional Analysis within the C8 Health Project. Environ Health Perspect. Aug; 2016 124(8):1269–1275. [PubMed: 26794451]
- 19. Aguirre GA, De Ita JR, de la Garza RG, Castilla-Cortazar I. Insulin-like growth factor-1 deficiency and metabolic syndrome. J Transl Med. 2016; 14:3. [PubMed: 26733412]

 Carlzon D, Svensson J, Petzold M, et al. Both low and high serum IGF-1 levels associate with increased risk of cardiovascular events in elderly men. J Clin Endocrinol Metab. Nov; 2014 99(11):E2308–2316. [PubMed: 25057875]

- 21. Friedman SM, Maslow CB, Reibman J, et al. Case-control study of lung function in World Trade Center Health Registry area residents and workers. Am J Respir Crit Care Med. Sep 1; 2011 184(5):582–589. [PubMed: 21642248]
- 22. Taniyasu S, Kannan K, So MK, et al. Analysis of fluorotelomer alcohols, fluorotelomer acids, and short- and long-chain perfluorinated acids in water and biota. Journal of Chromatography A. 2005; 1093(1–2):89–97. [PubMed: 16233874]
- Kannan K, Corsolini S, Falandysz J, et al. Perfluorooctanesulfonate and Related Fluorochemicals in Human Blood from Several Countries. Environmental science & technology. 2004; 38(17): 4489–4495. [PubMed: 15461154]
- 24. Ogden CL, Kuczmarski RJ, Flegal KM, et al. Centers for Disease Control and Prevention 2000 growth charts for the United States: improvements to the 1977 National Center for Health Statistics version. Pediatrics. Jan; 2002 109(1):45–60. [PubMed: 11773541]
- Rockett HR, Breitenbach M, Frazier AL, et al. Validation of a youth/adolescent food frequency questionnaire. Prev Med. Nov-Dec;1997 26(6):808–816. [PubMed: 9388792]
- 26. Craig CL, Marshall AL, Sjostrom M, et al. International physical activity questionnaire: 12-country reliability and validity. Med Sci Sports Exerc. Aug; 2003 35(8):1381–1395. [PubMed: 12900694]
- 27. Ainsworth BE, Haskell WL, Whitt MC, et al. Compendium of physical activities: an update of activity codes and MET intensities. Med Sci Sports Exerc. Sep; 2000 32(9 Suppl):S498–504. [PubMed: 10993420]
- Urbina EM, Brinton TJ, Elkasabany A, Berenson GS. Brachial artery distensibility and relation to cardiovascular risk factors in healthy young adults (The Bogalusa Heart Study). Am J Cardiol. 2002; 89:946–951. [PubMed: 11950433]
- Urbina EM, Dolan LM, McCoy CE, Khoury PR, Daniels SR, Kimball TR. Relationship between elevated arterial stiffness and increased left ventricular mass in adolescents and young adults. J Pediatr. May; 2011 158(5):715–721. [PubMed: 21300369]
- 30. National High Blood Pressure Education Program Working Group on High Blood Pressure in Children and Adolescents. The Fourth Report on the Diagnosis, Evaluation, and Treatment of High Blood Pressure in Children and Adolescents. Pediatrics. 2004; 114:555–576. [PubMed: 15286277]
- 31. Laurent S, Cockcroft J, Van Bortel L, et al. Expert consensus document on arterial stiffness: methodological issues and clinical applications. Eur Heart J. Nov; 2006 27(21):2588–2605. [PubMed: 17000623]
- 32. Chen CH, Nevo E, Fetics B, et al. Estimation of central aortic pressure waveform by mathematical transformation of radial tonometry pressure. Validation of generalized transfer function. Circulation. Apr 1; 1997 95(7):1827–1836. [PubMed: 9107170]
- 33. Johnson WD, Kroon JJM, Greenway FL, Bouchard C, Ryan D, Katzmarzyk PT. Prevalence of Risk Factors for Metabolic Syndrome in Adolescents: National Health and Nutrition Examination Survey (NHANES), 2001–2006. Arch Pediatr Adolesc Med. Apr 1; 2009 163(4):371–377. [PubMed: 19349567]
- 34. Bergman RN, Prager R, Volund A, Olefsky JM. Equivalence of the insulin sensitivity index in man derived by the minimal model method and the euglycemic glucose clamp. The Journal of Clinical Investigation. 1987; 79(3):790–800. [PubMed: 3546379]
- 35. Conwell LS, Trost SG, Brown WJ, Batch JA. Indexes of Insulin Resistance and Secretion in Obese Children and Adolescents. Diabetes Care. Feb 1; 2004 27(2):314–319. [PubMed: 14747206]
- 36. Keskin M, Kurtoglu S, Kendirci M, Atabek ME, Yazici C. Homeostasis Model Assessment Is More Reliable Than the Fasting Glucose/Insulin Ratio and Quantitative Insulin Sensitivity Check Index for Assessing Insulin Resistance Among Obese Children and Adolescents. Pediatrics. Apr 1; 2005 115(4):e500–e503. [PubMed: 15741351]
- 37. Strauss RS. Environmental tobacco smoke and serum vitamin C levels in children. Pediatrics. Mar; 2001 107(3):540–542. [PubMed: 11230596]

38. Wilson KM, Finkelstein JN, Blumkin AK, Best D, Klein JD. Micronutrient levels in children exposed to secondhand tobacco smoke. Nicotine Tob Res. Sep; 2011 13(9):800–808. [PubMed: 21558135]

- 39. Helsel, DR. Statistics for Censored Environmental Data Using Minitab and R. Hoboken: Wiley; 2012. p. 2
- 40. Pletcher MJ, Vittinghoff E, Thanataveerat A, Bibbins-Domingo K, Moran AE. Young Adult Exposure to Cardiovascular Risk Factors and Risk of Events Later in Life: The Framingham Offspring Study. PLoS One. 2016; 11(5):e0154288. [PubMed: 27138014]
- 41. Fisher M, Arbuckle TE, Wade M, Haines DA. Do perfluoroalkyl substances affect metabolic function and plasma lipids?--Analysis of the 2007–2009, Canadian Health Measures Survey (CHMS) Cycle 1. Environ Res. Feb.2013 121:95–103. [PubMed: 23266098]
- 42. Starling AP, Engel SM, Whitworth KW, et al. Perfluoroalkyl substances and lipid concentrations in plasma during pregnancy among women in the Norwegian Mother and Child Cohort Study. Environ Int. Jan. 2014 62:104–112. [PubMed: 24189199]
- 43. Frisbee SJ, Shankar A, Knox SS, et al. Perfluorooctanoic acid, perfluorooctanesulfonate, and serum lipids in children and adolescents: results from the C8 Health Project. Arch Pediatr Adolesc Med. Sep; 2010 164(9):860–869. [PubMed: 20819969]
- 44. Geiger SD, Xiao J, Ducatman A, Frisbee S, Innes K, Shankar A. The association between PFOA, PFOS and serum lipid levels in adolescents. Chemosphere. Mar.2014 98:78–83. [PubMed: 24238303]
- 45. Steenland K, Tinker S, Frisbee S, Ducatman A, Vaccarino V. Association of perfluorooctanoic acid and perfluorooctane sulfonate with serum lipids among adults living near a chemical plant. Am J Epidemiol. Nov 15; 2009 170(10):1268–1278. [PubMed: 19846564]
- 46. Zeng XW, Qian Z, Emo B, et al. Association of polyfluoroalkyl chemical exposure with serum lipids in children. Sci Total Environ. Apr 15.2015 512–513:364–370.
- 47. Fleisch AF, Rifas-Shiman SL, Mora AM, et al. Early Life Exposure to Perfluoroalkyl Substances and Childhood Metabolic Function. Environ Health Perspect. Sep 2.2016
- 48. Timmermann CA, Rossing LI, Grontved A, et al. Adiposity and glycemic control in children exposed to perfluorinated compounds. J Clin Endocrinol Metab. Apr; 2014 99(4):E608–614. [PubMed: 24606078]
- 49. Lin CY, Chen PC, Lin YC, Lin LY. Association among serum perfluoroalkyl chemicals, glucose homeostasis, and metabolic syndrome in adolescents and adults. Diabetes Care. Apr; 2009 32(4): 702–707. [PubMed: 19114613]
- Wolf CJ, Schmid JE, Lau C, Abbott BD. Activation of mouse and human peroxisome proliferatoractivated receptor-alpha (PPARalpha) by perfluoroalkyl acids (PFAAs): further investigation of C4–C12 compounds. Reprod Toxicol. Jul; 2012 33(4):546–551. [PubMed: 22107727]
- 51. Takacs ML, Abbott BD. Activation of mouse and human peroxisome proliferator-activated receptors (alpha, beta/delta, gamma) by perfluorooctanoic acid and perfluorooctane sulfonate. Toxicol Sci. Jan; 2007 95(1):108–117. [PubMed: 17047030]
- 52. Jiang, Q., Gao, H., Zhang, L. Metabolic Effects of PFAS. In: DeWitt, JC., editor. Toxicological Effects of Perfluoroalkyl and Polyfluoroalkyl Substances. Springer International Publishing; Switzerland: 2015.
- 53. Rossi P, Frances Y, Kingwell BA, Ahimastos AA. Gender differences in artery wall biomechanical properties throughout life. J Hypertens. Jun; 2011 29(6):1023–1033. [PubMed: 21346620]
- 54. Zhou Y, Hu LW, Qian ZM, et al. Association of perfluoroalkyl substances exposure with reproductive hormone levels in adolescents: By sex status. Environ Int. Sep.2016 94:189–195. [PubMed: 27258660]

Highlights

- WTC-related exposures to perfluoroalkyl substances (PFASs) may be associated with cardiometabolic consequences
- Cardiometabolic profiles of exposed youth were examined
- Higher serum levels of PFASs were associated with increased blood lipid levels
- These findings pinpoint the potential high risk of atherosclerosis as a result of PFASs exposure

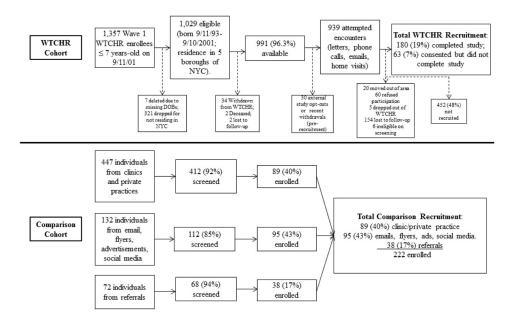


Figure 1. Recruitment Flowchart for WTCHR and Comparison Cohort

Table 1

Characteristics of the two study populations.

	Control group n=222	WTCHR group n=180	p value
Sex			
Male	89 (40.1%)	97 (53.9%)	0.008
Female	133 (59.9%)	83 (46.1%)	
Date of birth			
9/11/93–9/10/95	45 (20.3%)	47 (26.1%)	0.159
9/11/95–9/10/98	89 (40.1%)	77 (42.8%)	
9/11/98-9/10/01	88 (39.6%)	56 (31.1%)	
Income < \$25,000 ^a	49 (27.4%)	28 (19.4%)	0.126
Race/Ethnicity ^b			
Non-Hispanic White (%)	89 (40.1%)	66 (36.9%)	0.053
Non-Hispanic Black (%)	19 (8.6%)	16 (8.9%)	
Non-Hispanic Asian (%)	44 (19.8%)	49 (27.4%)	
Non-Hispanic Other (%)	10 (4.5%)	16 (8.9%)	
Hispanic (%)	60 (27.0%)	32 (17.9%)	
Caloric intake, ^C Median (IQR)	1535 (1061, 2087)	1621 (1141, 2331)	0.028
Physical activity, MET hours per week (IQR)	150 (90, 240)	180 (120, 285)	0.087
Body Mass Index Category			
Normal weight/underweight	162 (73.0%)	150 (83.3%)	0.045
Overweight	36 (16.2%)	19 (10.6%)	0.028
Obese	24 (10.8%)	11 (6.1%)	0.087
Smoking status			
Smokers	23 (10.4%)	24 (13.3%)	0.443
Median Cotinine Concentration	0.324 (0.106, 0.690)	0.412 (0.106, 0.984)	0.294
Tobacco smoke exposure d			
Low (<0.15 ng/mL)	102 (45.9)	73 (40.6)	0.353
Medium (0.15 to < 2.32 ng/mL)	95 (42.8)	79 (43.9)	0.443
High(2.32 ng/mL)	25 (11.3)	28 (15.6)	0.294
Cardiometabolic Markers, Median (IQR)			
Triglycerides (mg/dL)	66.5 (48, 95.3)	63.5 (49.8, 88.5)	0.891
High-Density Lipoprotein (mg/dL)	53 (44, 66)	52 (43.75, 60.25)	0.294
Low-Density Lipoprotein (mg/dL)	77 (66, 94)	80 (69, 96)	0.131

Control group n=222 WTCHR group n=180 p value

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Insulin Resistance (HOMA-IR) 1.54 (1.14, 2.23) 1.37 (1.05, 2.04) 0.087 Total Cholesterol (mg/dL) 148.5 (133, 166.3) 148.5 (133, 170) 0.827

 $[^]a_{\rm n=43}$ missing for comparison group; n=27 missing for WTCHR group;

b n=1 missing for race/ethnicity;

^c_{n=2} missing for caloric intake;

d Evaluated by saliva cotinine concentration and questionnaire. For subjects without saliva cotinine concentration, we categorized no smoker and no secondhand smoke exposure into "low", no smoker but secondhand smoke exposure into "medium", and smoker into "high" category.

Table 2
Serum PFASs Population Characteristics (with exclusion of those who opted out of venous blood sampling).

	Comparison (n=185)	WTCHR (n=123)	p value
Sex, n (%)			
Male	74 (40%)	69 (56.1%)	
Female	111 (60%)	54 (44.9%)	
Date of birth, n (%)			
9/11/93–9/10/95	35 (18.9%)	34 (27.6%)	
9/11/95–9/10/98	73 (39.5%)	52 (42.3%)	0.070
9/11/98-9/10/01	77 (41.6%)	37 (30.1%)	
Income < \$25,000 ^a	42 (22.7%)	19 (15.4%)	0.170
Race/Ethnicity, b n (%)			
Non-Hispanic White	72 (38.9%)	42 (34.4%)	
Non-Hispanic Black	17 (9.2%)	13 (10.7%)	
Non-Hispanic Asian	37 (20%)	30 (24.6%)	0.040
Non-Hispanic Other	6 (3.2%)	13 (10.7%)	
Hispanic	53 (28.6%)	24 (19.7%)	
Serum PFASs, Median (IQR), ng/mL			
PFHxS (n <lod= %)<="" 0="" td=""><td>0.53 (0.47)</td><td>0.67 (0.69)</td><td><0.0001</td></lod=>	0.53 (0.47)	0.67 (0.69)	<0.0001
PFOS (n <lod= %)<="" 0="" td=""><td>2.78 (2.18)</td><td>3.72 (2.82)</td><td>< 0.0001</td></lod=>	2.78 (2.18)	3.72 (2.82)	< 0.0001
PFOA (n <lod= %)<="" 0="" td=""><td>1.39 (0.75)</td><td>1.81 (0.90)</td><td>< 0.0001</td></lod=>	1.39 (0.75)	1.81 (0.90)	< 0.0001
PFNA (n <lod= %)<="" 0.3="" td=""><td>0.49 (0.33)</td><td>0.61 (0.36)</td><td>< 0.0001</td></lod=>	0.49 (0.33)	0.61 (0.36)	< 0.0001
PFDA (n <lod= %)<="" 25="" td=""><td>0.11 (0.15)</td><td>0.14 (0.12)</td><td>< 0.0001</td></lod=>	0.11 (0.15)	0.14 (0.12)	< 0.0001
PFUnDA (n <lod= %)<="" 47="" td=""><td>0.04 (0.16)</td><td>0.12 (0.21)</td><td>0.007</td></lod=>	0.04 (0.16)	0.12 (0.21)	0.007
Calories, ^C Median (IQR)	1537 (1014)	1709 (1317)	0.008
Tobacco smoke exposure			
Low	102 (45.9)	73 (40.6)	
Medium	95 (42.8)	79 (43.9)	0.353
High	25 (11.3)	28 (15.6)	
Body Mass Index Category			
Normal weight/underweight	137 (74.1)	98 (79.7)	
Obese	20 (16.3)	8 (4.3)	0.387
Overweight	28 (15.1)	17 (13.8)	

 $^{^{}a}$ n=38 missing for comparison; n=27 missing for WTCHR;

 $^{^{}b}$ _{n=1} missing for race/ethnicity;

c_{n=2} missing for caloric intake.

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Table 3

Multivariable Linear and Logistic Regression Analysis of Cardiometabolic Outcomes Associated with Serum PFASs.

	PFHxS		PFOS		PFOA		PFNA		PFDA		PFUnDA	
Cardiometabolic Outcomes (confinuous)	unit change (95% CI)	d	unit change (95% CI)	d	unit change (95% CI)	d	unit change (95% CI)	d	unit change (95% CI)	d	unit change (95% CI)	d
BMI	0.43 (-0.38, 1.23)	0.30	-0.35 (-1.432, 0.743)	0.53	0.65 (-0.841, 2.143)	0.39	-0.48 (-1.72, 0.77)	0.45	-0.42 (-1.030, 0.19)	0.18	-0.48 (-1.01, 0.06)	0.08
BMIz	0.05 (-0.15, 0.25)	0.62	-0.22 (-0.49, 0.06)	0.12	-0.02 (-0.38, 0.33)	0.90	-0.18 (-0.47, 0.10)	0.21	-0.08 (-0.23, 0.06)	0.27	-0.13 (-0.26, -0.01)	$\boldsymbol{0.03}^*$
log_trig	0.04 (-0.02, 0.11)	0.20	0.04 (-0.05, 0.13)	0.36	0.14 (0.02, 0.27)	$\boldsymbol{0.03}^*$	-0.007 (-0.11, 0.01)	0.89	0.01 (-0.047, 0.057)	0.85	-0.04 (-0.09, 0.003)	0.07
log_homair	-0.09 (-0.18, -0.003)	* 40.0	-0.06 (-0.18, 0.06)	0.31	-0.05 (-0.21, 0.12)	0.58	0.01 (-0.13, 0.14)	0.89	-0.04 (-0.11, 0.03)	0.26	-0.04 (-0.10, 0.02)	0.21
logChol	0.04 (0.01, 0.06)	$\boldsymbol{0.01}^*$	0.08 (0.05, 0.12)	<0.001	0.09 (0.04, 0.14)	<0.001	0.05 (0.01, 0.09)	$\boldsymbol{0.01}^*$	0.04 (0.02, 0.06)	<0.001	0.02 (0, 0.04)	0.00
logLDL	0.05 (0.01, 0.09)	$\boldsymbol{0.02}^*$	0.10 (0.05, 0.16)	<0.001	0.11 (0.03, 0.19)	0.006	0.07 (0.01, 0.14)	$\boldsymbol{0.03}^*$	0.04 (0.00, 0.07)	$\boldsymbol{0.03}^*$	0.01 (-0.02, 0.04)	0.49
logHDL	0.03 (-0.02, 0.07)	0.26	0.06 (0.003, 0.13)	* 40.0	0.04 (-0.04, 0.12)	0.34	0.05 (-0.02, 0.12)	0.13	0.05 (0.02, 0.09)	0.003^{**}	0.04 (0.10, 0.07)	$\boldsymbol{0.01}^*$
BrachD	0.15 (-0.07, 0.38)	69.0	0.30 (-0.01, 0.62)	90.0	0.45 (0.04, 0.87)	0.03^*	0.34 (0.02, 0.67)	0.04^*	0.11 (-0.06, 0.28)	0.10	0.11 (-0.04, 0.26)	0.97
AIx	-0.48 (-2.20, 1.25)	0.89	-0.24 (-2.02, 2.41)	0.85	-1.41 (-4.59, 1.78)	0.39	-0.51 (-2.51, 2.53)	0.70	0.08 (-1.23, 1.39)	0.14	0.37 (-0.79, 1.52)	0.35
PWV	-0.05 (-0.16, 0.07)	0.43	-0.06 (-0.23, 0.11)	0.51	0.05 (-0.17, 0.28)	0.64	-0.13 (-0.30, 0.04)	0.14	-0.04 (-0.13, 0.05)	0.39	-0.03 (-0.11, 0.05)	0.41
Cardiometabolic Outcomes (dichotomous)	OR (95% CI)	d	OR (95% CI)	д	OR (95% CI)	d	OR (95% CI)	р	OR (95% CI)	р	OR (95% CI)	р
Overweight	1.04 (0.97, 1.11)	0:30	0.98 (0.90, 1.07)	99:0	1.00 (0.90, 1.13)	76.0	1.01 (0.92, 1.13)	0.72	0.98 (0.93, 1.03)	0.49	0.95 (0.91, 0.99)	0.02

Each column represents an examination of a single exposure variable or study arm controlled for sex, race, caloric intake, physical activity, cotinine concentration and BMI category (except when the outcome examined was BMI); Beta coefficients represent the change associated with each natural log-unit increase in the PFASs examined.

** p<0.01. * p<0.05;

BMI: Body Mass Index; BMIz: BMI z score; logTrig: log-transformed triglycerides; log homair: log-transformed homeostatic model of insulin resistance; logChol: log-transformed total cholesterol; logLDL: log-transformed LDL cholesterol; logHDL: log-transformed HDL cholesterol; BrachD: brachial artery distensibility; Alx: Augmentation Index; PWV: pulse wave velocity.

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Percent Changes of Blood lipids and Insulin Resistance Outcomes Associated with Serum PFASs^a

Table 4

	Triglycerides	Insulin resistance (HOMAIR)	Total cholesterol	LDL cholesterol	HDL cholesterol
PFASs	Percent change (95% CI)	PFASs Percent change (95% CI)	Percent change (95% CI)	Percent change (95% CI)	Percent change (95% CI)
PFHxS	4.5 (-2.4, 11.7)	-8.6 (-16.1, -0.3)	3.9 (1.1, 6.6)	5 (0.7, 9.5)	2.6 (-1.9, 7.4)
PFOS	4.3 (-4.8, 14.3)	-5.8 (-16.3, 5.9)	8.5 (4.8, 12.4)	10.7 (4.7, 17.2)	6.6 (0.3, 13.3)
PFOA	15.1 (-1.7, 30.3)	-4.5 (-18.8, 12.3)	9.2 (4, 14.7)	11.5 (3.1, 20.6)	4.2 (-4.2, 13.2)
PFNA	-0.7 (-10.5, 10.2)	0.9 (-11.8, 15.4)	5.4 (1.2, 9.9)	7.4 (0.6, 14.6)	5.4 (-1.6, 13.1)
PFDA	0.5 (-4.6, 5.9)	-3.7 (-10, 2.8)	3.9 (1.8, 6.0)	3.7 (0.4, 7.1)	5.3 (1.8, 9.0)
PFUnDA	-4.1 (-8.2, 0.3)	-3.6 (-9.1, 2.1)	1.7 (0, 3.6)	1 (-1.8, 3.9)	4 (0.9, 7)

^aPercent change in the original unit of measurement for one log unit increase of the chemicals examined; HOMAIR: homeostatic model of insulin resistance

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