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Infectivity of attenuated poxvirus vaccine vectors and immunogenicity of a raccoonpox vectored rabies vaccine in the Brazilian Free-tailed bat (*Tadarida brasiliensis*)

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Abstract

Bats (Order Chiroptera) are an abundant group of mammals with tremendous ecological value as insectivores and plant dispersers, but their role as reservoirs of zoonotic diseases has received more attention in the last decade. With the goal of managing disease in free-ranging bats, we tested modified vaccinia Ankara (MVA) and raccoon poxvirus (RCN) as potential vaccine vectors in the Brazilian Free-tailed bat (*Tadarida brasiliensis*), using biophotonic *in vivo* imaging and immunogenicity studies. Animals were administered recombinant poxviral vectors expressing the luciferase gene (*MVA-luc*, *RCN-luc*) through oronasal (ON) or intramuscular (IM) routes and subsequently monitored for bioluminescent signal indicative of viral infection. No clinical illness was noted after exposure to any of the vectors, and limited luciferase expression was observed. Higher and longer levels of expression were observed with the *RCN-luc* construct. When given IM, luciferase expression was limited to the site of injection, while ON exposure led to initial expression in the oral cavity, often followed by secondary replication at another location, likely the gastric mucosa or gastric associated lymphatic tissue. Viral DNA was detected in oral swabs up to 7 and 9 days post infection (dpi) for MVA and RCN, respectively. While no live virus was detected in oral swabs from MVA-infected bats, titers up to 3.88×10^4 PFU/ml were recovered from oral swabs of RCN-infected bats. Viral DNA was also detected in fecal samples from two bats inoculated IM with RCN, but no live virus was recovered. Finally, we examined the immunogenicity of a RCN based rabies vaccine (RCN-G) following ON administration. Significant rabies neutralizing antibody titers were detected in the serum of immunized bats using the rapid fluorescence focus inhibition test (RFFIT). These studies highlight the safety and immunogenicity of attenuated poxviruses and their potential use as vaccine vectors in bats.

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Appendix A. Supplementary material: Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.vaccine.2016.08.088>.

1. Introduction

Over the last few decades, the importance of bats (order Chiroptera) in the maintenance and transmission of zoonotic diseases has become increasingly evident; bats are thought to harbor the most zoonotic agents per species [1]. The list of pathogens that infect bats includes the major mammalian paramyxoviruses [2], coronaviruses [3,4], filoviruses [5–7], distinct influenza lineages [8,9], hepadnaviruses [10], and hantaviruses [11], as well as lyssa-viruses such as rabies virus [12,13]. In the United States, bats are often the most common source of rabies infections in humans [14], and in Central and South America, rabies transmitted by vampire bats is a serious zoonotic and economic issue [15]. This association between bats and pathogens that significantly impacts human populations has increased public fear and misunderstanding of these animals and lead to culling campaigns [15–18]. Unfortunately, culling campaigns often lead to the death of valuable non-target bat species [16] and appear ineffective in reducing disease incidence [17]. Alternatively, vaccination of other wildlife species has been successful in mitigating the public health impact of rabies with the development of efficient and practical distribution methods for mass immunization.

Poxviral vectors have been used extensively for oral vaccines to control infectious diseases in a variety of animal species over the last 25 years [19,20]. Several advantages of poxviruses as vaccine vectors include: (1) their allowance of large insertions of foreign DNA; (2) ease of manufacturing; (3) thermal and genetic stability; (4) safety and infectivity for multiple target species; and (5) ability to infect via mucosal and dermal routes. For example, an oral rabies vaccine, constructed by inserting the rabies G glycoprotein into vaccinia virus and distributed via baits, has been used for many years to curtail rabies outbreaks in foxes, raccoons and other animals in North America and Europe [21]. Previous studies assessing a vaccinia-based rabies vaccine (VR-G) in *Desmodus* bats demonstrated the protective efficacy of that construct [22–24]. However, this vector has undesirable side-effects, especially in immunocompromised individuals [25], necessitating the development of attenuated virus strains. More recently, an oral sylvatic plague vaccine using another poxvirus (raccoon pox, RCN) was shown to protect prairie dogs and is currently being tested in large-scale field trials [26,27]. RCN was first isolated from the upper respiratory tract of apparently healthy raccoons in North America [28]. It has since been shown to be safe and effective in a variety of species, including domestic cats, piglets, dogs, raccoons, skunks, foxes, bobcats, rabbits, sheep, prairie dogs, non-human primates, and chickens, with none of the immunized animals showing clinical side effects [29–32]. Additionally, RCN has been shown to be immunogenic via non-parenteral routes in both domestic species [29] and free ranging wildlife [33,34]. Another orthopoxvirus vector, modified vaccinia Ankara (MVA), is a highly attenuated form of vaccinia [35,36] and has also been demonstrated to safely and effectively induce immunity [37,38].

Based on the success of mucosal vaccination with poxvirus vectors in many other species, we hypothesized that poxviruses could be immunogenic and safe when given mucosally in chiropteran species. To test this, we assessed the infectivity and pathogenicity of MVA and RCN in *T. brasiliensis* via *in vivo* imaging studies. The immunogenicity of RCN given

oronasally was also assessed using standard serologic techniques after vaccination with an RCN-based rabies vaccine (RCN-G).

2. Materials and methods

2.1. Ethics statement

The use of bats in this experiment was approved by (Protocol #EP111018) and conducted in accordance with the U.S. Geological Survey (USGS), National Wildlife Health Center (NWHC), Animal Care and Use Committee (ACUC).

2.2. Animals

Adult male bats (*T. brasiliensis*; $n = 22$) were caught in Brazos County, Texas under Texas Parks and Wildlife Department permit number SPR-1104-610 and Texas A&M University ACUC approval number 2012-130 (courtesy of Mike Smotherman, Texas A&M University, USA). After acclimating to captivity, the bats were transferred to NWHC (Madison, Wisconsin, USA), where all bat studies were conducted under ABSL-3 conditions. Upon transfer to the NWHC, bats were maintained in flight cages for a quarantine period of 30 days. During this time blood samples were taken and bats were treated topically for parasites. Electronic microchip identification units (Avid Identification Systems, Inc., Folsom, Louisiana, USA) were inserted into each animal, between the scapulae, via subcutaneous injection. Bats were maintained on mealworms (*Tenebrio molitor*) supplemented with vitamins and an omega fatty acids mixture, and water was available *ad libitum*. Light cycles were set to 12 h of light per day inverted from the natural cycle to allow monitoring of bat activities during facility working hours.

2.3. Viruses and cells

The RCN-*luc* strain used in this study was previously described [32]. The MVA-GFP strain used to create the MVA-*luc* constructs was generously provided by Inviragen (Madison, WI), while RCN-G [34] was kindly provided by the Centers for Disease Control (Atlanta, GA). Recombinant viruses were generated and amplified on cell monolayers of rat embryonic fibroblasts (Rat-2, ATCC #CRL-1764), baby hamster kidney cells (BHK-21, ATCC #CRL-12072), African Green monkey kidney epithelial cells (Vero, ATCC #CCL-18), or primary chicken embryo fibroblasts (CEF, Charles River Laboratories, INC, Wilmington, WA, USA)). Cell cultures were maintained at 37 °C and 5% CO₂ in Dulbecco's Modified Eagle Medium (DMEM) or Opti-MEM® (Life technologies, Madison, WI 53719), supplemented with 2–5% fetal bovine serum (FBS). Viruses were titrated prior to use with plaque dilution assays in 6-well plates.

2.4. Construction of recombinant MVA-*luc*

Recombinant MVA-*luc* viruses were constructed as described elsewhere [39]. Briefly, CEF cells were infected with MVA-GFP at a multiplicity of 0.05 PFU per cell; one hour later the cells were transfected using the FuGENE® reagent protocol (Promega, Fitchburg, WI) with a pI2 transfer plasmid containing (1) DNA flanking segments adjacent to deletion III within the *Hind*III A fragment of MVA, (2) the Red Fluorescent Protein (RFP) under the control of a p11 promoter, and (3) the firefly luciferase gene (*luc*) under the control of a strong

synthetic early/late (SE/L) vaccinia virus promoter upstream Supplementary Fig. S1). At 48–72 h post transfection, the cell cultures were put through three freeze-thaw cycles, harvested, sonicated, and centrifuged at 500g for 5 min at 4 °C. The sonicated cell extracts were plated onto fresh BHK-21 cells and overlaid with 0.8% agarose. After 48–72 h, through the use of specific microscope filters, the recombinant viruses were detected by the presence of the RFP gene, which replaced the GFP gene during homologous recombination. Selected cell/virus samples were sonicated and plated again as described above. After four consecutive rounds of plaque isolation, recombinant MVA-*luc* virus was confirmed by PCR analysis using OneTaq® Quick-Load® 2X Master Mix with Standard Buffer (New England BioLabs Inc., Ipswich, MA 01938, USA) amplifying the insertion at the Del III flanks using ATGCGGCACCTCTCT-TAA as a forward primer and CCAAAGCTTGCACATACATAAGTA as the reverse primer. The virus subsequently amplified in freshly prepared CEF cells.

2.5. Bioluminescent imaging

Biophotonic luminescent imaging (BLI) has been successfully used to assess the infectivity, infection course, and tissue tropism of viruses and candidate vaccine vectors [40]. Unlike traditional pathogenicity studies that require euthanasia of animals at different time points, BLI allows study of the course of infection over time in a single host, increasing the information gained while reducing the number of animals required.

Groups of 8 wild-caught *T. brasiliensis* were separated into two screened flight cages (33" W × 66" D × 84" H) approximately 5 m apart. Four bats from each group were given 10⁹ plaque forming units (PFU) in 100 µl of either RCN-*luc* or MVA-*luc* by intramuscular (IM) injection, split into two 50 µl volumes injected into each thigh muscle. The remaining four bats in each group were given the same amount of virus in 70 µl sterile saline; using a micropipette with sterile tips, the volume split among the nostrils (10 µl given each nare) and mouth (50 µl) for oronasal (ON) exposure. Bats were monitored for 3 h post inoculation for signs of adverse effects. Animals were scanned using an IVIS 200 Biophotonic imager (PerkinElmer, Hopkinton, MA, USA) at one-day post infection (dpi) and every-other day thereafter until 2 consecutive images with less than 100 radiance units (comparable to background) were observed. Bats were scanned prior to infection and confirmed to have no auto-luminescence beyond background levels. Imaging was conducted at roughly the same time period each day, midway through the bats' active period. On imaging days the bats were separated individually into paper lunch bags, weighed, and injected intraperitoneally (i.p) with d-luciferin (Potassium-Luciferin, Gold Biotechnology, St. Louis, MO 63132, dose: 150mg/kg) at 14min prior to imaging the ON group and 26 min prior to imaging the IM group, which were empirically determined to be the time of peak luminescence post substrate exposure on the first day of imaging. All bats were examined for signs of disease or discomfort at the time of substrate injection. For imaging, animals were anesthetized by chamber-delivered isoflurane and positioned in the imager in dorsal recumbency with wings extended laterally, where they were maintained on mask-delivered isoflurane. After imaging the anesthesia was ceased, bats were monitored and given thermal support during recovery. After 11 dpi the bats with remaining detectable luminescence were imaged every 3 days until luminescence was below detectable levels. Images were collected and analyzed using

Living Image software (Caliper Life Sciences, Alameda, California, USA). A region of interest (ROI) was created which covered the entire body of the bat when analyzing the luminescence data.

At 87 dpi, a random group of six bats, all initially exposed to RCN-*luc* by either the ON (n = 4) or IM (n = 2) route, was given a booster exposure to the RCN-*luc* vaccine via the ON route at the same dose. Bats from this group were imaged at 1, 3, and 5 dpi using the same protocol as described above.

2.6. Assessment of viral shedding

During anesthesia for each imaging time-point, oral swabs were collected (CLASSIQSwabs, Copan Flock Technologies, 25125 Brescia, Italy) and placed in a 1.5 ml tube containing 200 μ l DMEM media. Fecal samples were also retrieved when available from the bags in which the individuals were contained during imaging, placed in a 1.5 ml tube without media. All samples were quickly stored at -80 °C until testing. Prior to testing, 100 μ l DMEM media was added to fecal samples, vortexed thoroughly and sonicated four times in a bath sonicator for 15 s. DNA was extracted from 40 μ l of the samples using the Zymo Quick-gDNA™ MiniPrep kit (Zymo Research, Irvine, CA 92614, U.S.A.), and a PCR assay was run to assess for presence of the inoculated virus. PCR was performed with OneTaq® Quick-Load® 2× Master Mix with Standard Buffer (New England BioLabs Inc., Ipswich, MA 01938, USA) and primers targeting the *luc* insert flanks [Supplemental info]. The limit of detection for this PCR protocol was determined to be 0.125 picograms of DNA, or 6.8×10^4 copies, as determined by serial dilution of a known quantity of viral DNA. Any samples positive by PCR were then assessed for levels of live virus by determining the median tissue culture infective dose (TCID₅₀). For RCN-*luc* samples, positive wells were assessed by plaque observation, and the TCID₅₀ was calculated and used to approximate plaque forming units (PFU)/ml by the Spearman & Kärber algorithm [41]. For MVA-*luc* samples, positive wells were assessed for viral titer by the luciferase marker using steadylite plus™ (PerkinElmer Inc, Waltham, MA 02451). Serial dilutions from 10^{-2} to 10^{-7} were made, and 100 μ l from each was used to infect a 96 well plate with BHK-21 cells at ~80% confluency. Due to minimal sample volume left, no smaller dilutions were possible. After 3 days of infection, 100 μ l of the steadylite reagent was added to each well, mixed by pipetting, and after 15 min the plates were scanned in a luminometer (Veritas™ Microplate Luminometer, Turner BioSystems, Inc, Sunnyvale, CA 94085).

2.7. RCN-G immunization

An additional group of five *T. brasiliensis* were housed separately in the same type of caging and given RCN-G via the ON route to assess the immunogenicity of RCN-delivered rabies CVS strain glycoprotein. For this exposure the bats were anesthetized with isoflurane prior to exposure. A dose of 10^8 PFU of viral vaccine was given in 70 μ l sterile saline, split between 50 μ l orally and 10 μ l in each nare. Bats were monitored through their anesthesia recovery for 3 h for potential adverse events. Serum samples were obtained prior to vaccination and at 30 and 60 dpi and tested for the presence of anti-rabies neutralizing antibodies by the rapid fluorescence focus inhibition test (RFFIT). Serum samples from bats given RCN-*luc* (used in the luminescence study) were also collected at 0 and 60 dpi and

used for controls. Serum was collected by making a small lance in the interfemoral vein and collecting up to 200 μ l of blood in a capillary tube (Microvette[®] CB 300 Blood Collection System, Sarstedt AG & Co., Nümbrecht, Germany), which was subsequently centrifuged at 10,000g for 10 min. A micropipette was used to collect the serum from the top of the blood container and transfer it to a separate tube for storage at -80° .

Testing was conducted at the CDC Poxvirus and Rabies Branch using standard RFFIT protocols [42], augmented for smaller volumes of serum as previously described [43]. The assay was run in triplicate for each sample, and the results reported represent average titers. Prior to the 60 dpi sample collection, two bats from the RCN-G group were lost from the study due to non-vaccine related mortalities.

2.8. Statistical analysis

Analysis of the data was performed using the R-commander software package [44]. Weight change was analyzed with repeated measures ANOVA, where 'weight' is a function of group, route, and time, plus all interactions, using individual bats as the repeated measures. Differences in luminescence were analyzed using a linear mixed-effects model fit by the restricted maximum likelihood approach (REML).

3. Results

3.1. In vivo imaging studies

To assess the infectivity, tissue tropism, and course of infection of RCN and MVA in *T. brasiliensis*, bats were infected with recombinant virus expressing the firefly luciferase gene (*luc*). Two routes were assessed; the IM route and the ON route, which is most biologically relevant for wildlife vaccination. Throughout the study, no clinical signs of disease, lesions, or significant weight loss were observed after administration of viral vectors (Supplementary Fig. S2, Supplementary Table S3). All bats infected with *luc*-expressing poxvirus vectors had detectable expression of luminescence by 1 dpi, and peak levels were observed at 1 and 3 dpi for the IM and ON routes, respectively (Figs. 1 and 2). Viral infection via IM exposure was cleared within 7 days for MVA and 9 days for RCN, while infection after ON exposure was cleared within 9 days for MVA and 21 days for RCN (final images for RCN given ON not shown). Statistical analysis revealed that luminescence was significantly higher ($P = 0.028$) in bats that received RCN compared to MVA and significantly higher ($P = 0.032$) for those administered virus by the IM route compared to the ON route. In the IM injected groups, significant viral spread to other areas was not evident. In contrast, an initial site of viral replication was evident in the oral cavity after ON exposure, often followed by a secondary site of expression further down the gastrointestinal tract. All luminescence measurements are listed in Supplementary Table S4).

In the group of bats re-exposed to RCN-*luc* to assess whether prior exposure would affect the infectivity of the viral vectors, no significant difference ($P = 0.33$) was detected in luminescence when compared to initial exposure through 5 dpi (Fig. 3).

3.2. Assessment of viral shedding

PCR analysis of oral swabs revealed the presence of MVA-*luc* DNA in 5 out of 8 bats up to 7 dpi (3 infected IM, 2 infected ON), however no live virus was recovered. RCN-*luc* DNA was present in 4 out of 8 bats up to 9 dpi (2 infected IM, 2 infected ON), with low levels of live virus detected (6.33×10^3 PFU/ml average, with a median of 9.20 PFU/ml in those with detectable virus). Two bats from the RCN-*luc* group that had live virus detected by oral swabs also had PCR positive fecal samples at 7 and 9 dpi, however no live virus was recoverable from these samples in titration. Viral shedding appears to occur at very low levels independent of route of exposure, and with no evidence of shedding viable MVA.

3.3. Antibody responses to RCN-G

To assess the immunogenicity of RCN, an additional group of bats ($n = 5$) was infected via the ON route with RCN expressing rabies virus surface glycoprotein (RCN-G). Again, there was no evidence of clinical disease in any of the vaccinated bats in this experiment. The rabies virus G is very well characterized and known to induce protective humoral immunity to rabies virus [30,45,46]. All bats assessed by RFFIT had negligible Ab titers prior to vaccination (Supplementary Table S5). As a control, bats vaccinated with *luc*-expressing virus for the imaging study ($N = 7$) were bled at 0 and 60 dpi and assessed by RFFIT as well. By 30 dpi all rabies vaccinated bats (5/5) developed Ab titers greater than 0.2 IU/ml (0.20–11.46, with a mean of 5.14), while 7/7 bats that received RCN-*luc* had titers 60.09 IU/ml (Supplementary Table S5). While there is no “protective” level of rabies virus neutralizing antibodies (RVNA), there is a positive correlation between RVNA titers and the level of protection after virus challenge [22,47–50]. Titers between 0.1 and 3.0 IU have been protective for other mammalian species [22,47–50].

4. Discussion

Despite the association between bats and zoonotic diseases, there is currently no significant effort to decrease the incidence of important infectious diseases in these animals. Instead, efforts are more focused on culling of populations (e.g. vampire bats in Latin America) or controlling disease after spill-over into other animal hosts. The continued spillover of rabies virus strains to humans and domestic animals from terrestrial carnivores has led to the development of successful oral rabies vaccination (ORV) programs in Europe and North America. These campaigns often utilize recombinant viral vectors that stimulate immunity to the surface glycoprotein of rabies when ingested orally. When distributed in baits targeted toward certain rabies-carrying species, these vaccines lead to protection from the virus and reduction in local incidence of disease, and even local eradication of some strains of rabies [20,51]. ORV programs are continually evolving and making use of the best available vaccine technology and disease modeling studies. In this study we assessed whether two attenuated pox-viruses might be viable vaccine vectors in a bat species.

Through in vivo imaging studies we have demonstrated that attenuated MVA and RCN are able to infect *T. brasiliensis* via the ON route for a limited time, without causing disease. Our studies demonstrate the tissue tropism and course of infection after exposure of a new animal model, *T. brasiliensis*, to attenuated orthopox-viruses. We show evidence of limited

autologous spread of the viruses after ON exposure, although there is no evidence that the virus spreads outside of GI-associated tissues. MVA was cleared faster and resulted in less detectable luminescence compared to RCN, which may be expected due to the highly attenuated nature of MVA. The lack of significant difference in the levels of viral encoded protein (luciferase) production upon re-infection with RCN, as shown in the booster study (Fig. 3), may be important if bats had been previously exposed to RCN, or if boost inoculations of RCN-vectored vaccine are necessary. While neither vector caused clinical illness, the fact that RCN produced more viral encoded protein (luciferase) over a longer period suggests it is a more immunogenic vector when expressing heterologous antigens. Due to limitations of this study, we were not able to compare the immunogenicity of the vectors directly.

The development of significant levels of rabies neutralizing antibodies after ON exposure to RCN-G demonstrates that the vaccine is highly immunogenic in *T. brasiliensis*. While previous studies have demonstrated the effectiveness of injectable vaccines in this species [52], our study is the first to demonstrate effective mucosal vaccination. An average anti-rabies G titer of 5.14 IU at 60 dpi was detected in bats orally administered RCN-G, which is higher than the levels obtained after vaccination with the VR-G construct in any previous studies, including oral, IM, and scarification routes of exposure (28–30). While these differences may be due to the animal models used, this is additional evidence of the superiority of the RCN vector over vaccinia in bat species. Additionally, the previous studies failed to address the infectivity of vaccinia in the bat host, relying on the lack of clinical disease and development of protective immunity to assess the virus-host interaction. We were unable to assess the duration of immunity past 60 days, but this would be useful information to collect in future studies.

Limited oral shedding of the RCN virus was detected in bats by both PCR and culture of live virus up to 9 dpi, but at very low levels. While there was no evidence of shedding of live MVA virus, there was PCR evidence of vector DNA in oral swabs through 7 dpi; detection of live virus may have been limited due to the method used and the necessary lack of lower dilutions resulting from low sample volumes. One of the advantages of *in vivo* imaging studies is that it is not necessary to sacrifice animals to obtain data, and the bats used in this study went on to be used in a different investigation. Because of this, we were unable to assess organ tissue for evidence of pathology during the infection trials. However, the lack of any apparent morbidity in RCN-treated bats, along with RCN's natural history, record of success in various domestic and non-domestic species, and ability to induce immunity via the oral route, make it a very attractive candidate for use in free-ranging bats.

The results of our experiments provide proof-of-principle that oral vaccination is possible in free-ranging bats. In future work, topical vehicles will be developed that could be used to deliver oral vaccines to the fur coat of bats as they roost or are otherwise congregated. Bats are fastidious animals and spend a large proportion of their time self-grooming [53], which could lead to significant oral exposure to topically applied vaccines. A vaccine that would broadly protect free-ranging bats from rabies virus infection may reduce local incidence of rabies in certain bat populations, limiting the amount of spill-over into humans and other species. *T. brasiliensis* represents a species that roosts in dense colonies and is exposed to

significant levels of circulating rabies virus in their population that may result in human exposures [54]. Therefore, a topically delivered rabies vaccine may be directly applicable to this species, but only if a method for mass application (such as a spray) could be developed. Further research is required to assess the viral vector in other species, such as vampire bats (*Desmodus* spp.), for which topically distributed poison protocols are well developed for culling of colonies [15,55]. While culling has been successful in reducing overall numbers of *Desmodus* at a local level, it has not reduced the incidence of rabies in bat populations and may indeed be counterproductive [17]. Development of topical poxvirus vectored vaccines could potentially lead to effective, applicable, and practical means for reducing disease burden in some free ranging bat populations.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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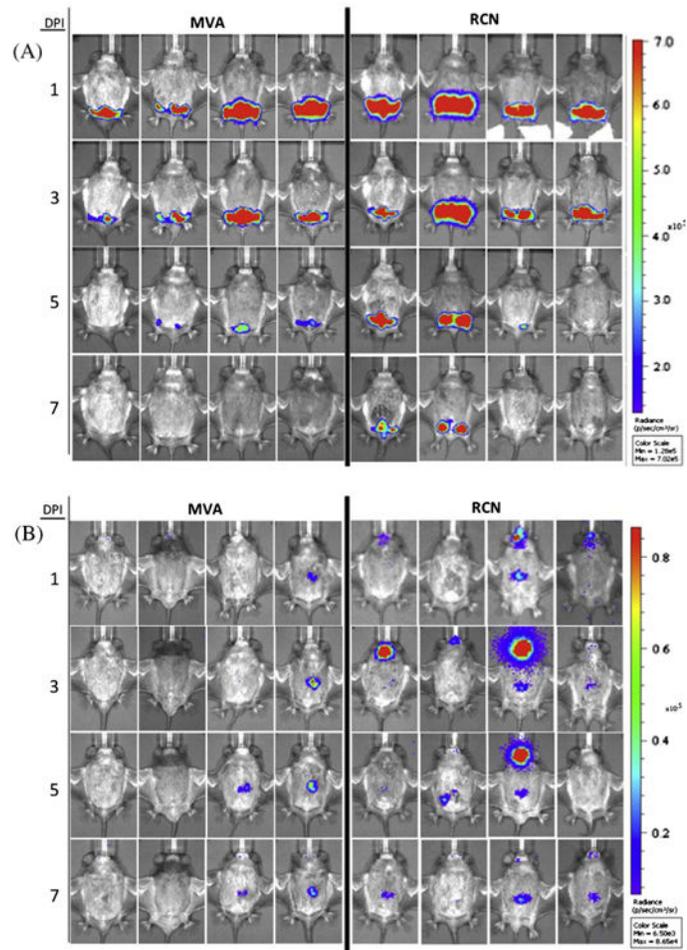


Fig. 1. Luminescent images for bats given attenuated poxviral vectors, raccoon poxvirus (RCN) or modified vaccinia Ankara (MVA) via intramuscular (IM) (A) or oronasal (ON) routes (B). Images were taken with the IVIS 200 Biophotonic imager and analyzed using the Living Image software. For each vector group, the scale of luminescence is given in photons/second/cm²/steradian (p/s/cm²/sr) which has been standardized to compare individuals over time in days post infection (DPI).

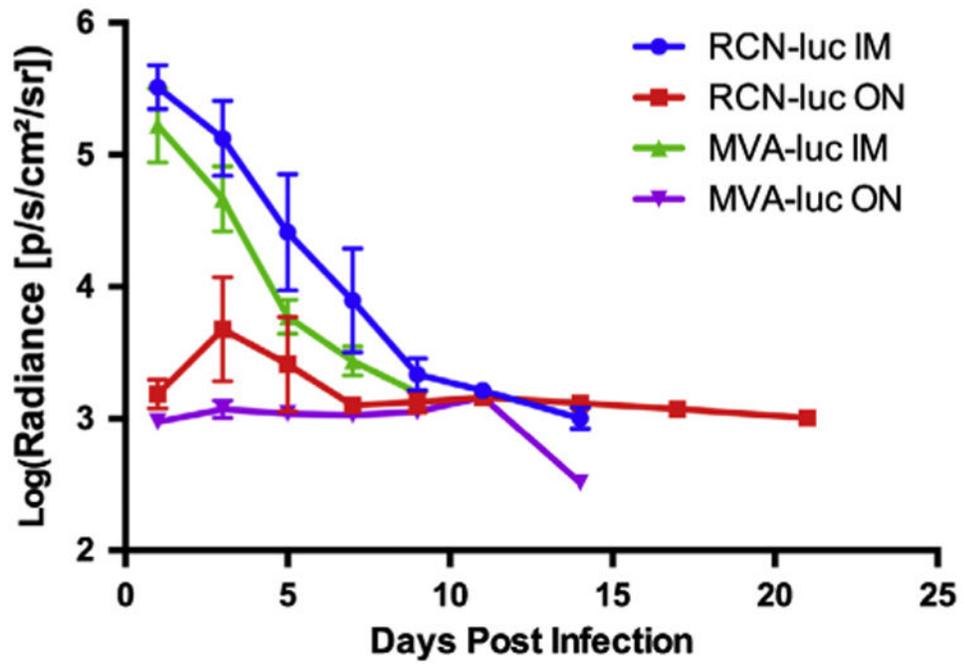


Fig. 2.

Average luciferase expression per vector by route over time for the four groups of bats given luciferase-expressing constructs. Luminescence is given in photons/second/cm²/steradian (p/s/cm²/sr). Luminescence was significantly higher ($P = 0.028$) for those receiving raccoon poxvirus (RCN) than modified vaccinia Ankara (MVA) and also higher ($P = 0.032$) for those administered virus by the intramuscular (IM) route compared to the oronasal (ON) route.

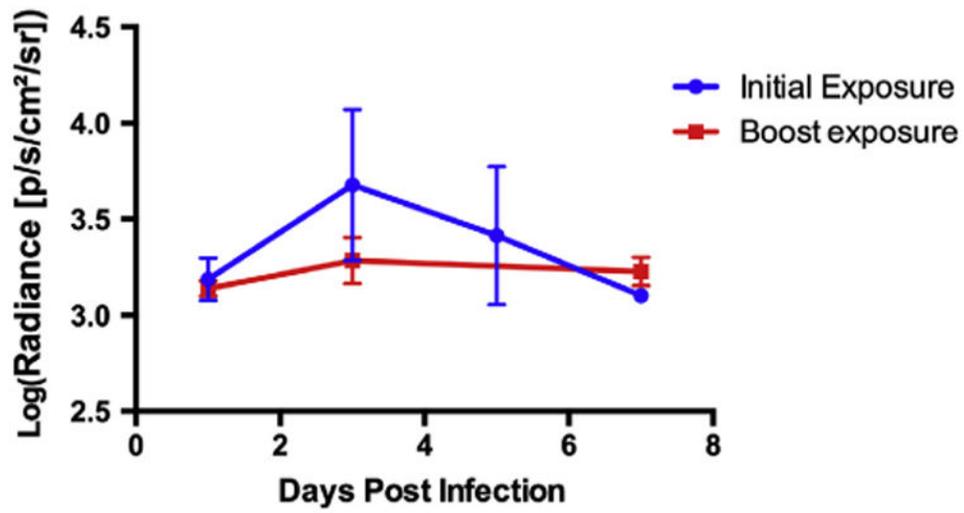


Fig. 3. Average luciferase expression after oronasal (ON) exposure to raccoon poxvirus (RCN) initially and after re-exposure. Luminescence is given in photons/ second/cm²/steradian (p/s/cm²/sr). No significant difference ($P = 0.33$) was detected in luminescence when compared to initial exposure through 5 days post infection.