**Supplemental Methods**

Laboratory processing of dialyzers

Arterial blood lines (Seroflow, city state) were plumbed through a Masterflex® peristaltic pump (Cole Parmer, Vernon Hills, IL) attached to an Easy-Load II® pump head (Cole Parmer, Vernon Hills, IL), and attached to arterial blood path inlet of the dialyzer. A 1 liter sterile bottle was used to catch flow through from the venous outlet. The pump ran at 300 mL/min. Peracetic acid levels were measured using LaMotte Insta-Test mid-range peracetic acid strips (LaMotte, Chesterfield, MD) dipped in a 5 mL aliquot of solution. Both the undiluted eluent from the dialyzer (~150 mL) as well as the saline rinse (a 1:6 dilution) were measured.

Next, 10 mL of the dialyzer flush was filtered through 0.22 micron polycarbonate filters, and placed on blood agar (BAP; Becton Dickinson, Franklin Lakes, NJ) and *Burkholderia cepacia* selective agar (BCSA; Remel, Lenexa, KS) plates as previously described (citation). 100 mL of the dialyzer flush was filtered through 0.45 micron gridded nitrocellulose filters and placed on BAP and BCSA. The remainder (~700 mL) was split in half and filtered on to either BAP or BCSA (~350 mL each). All membrane filters were rinsed with 2X volumes of Butterfield Buffer (BB); 20 mL for 10 mL of filtered dialyzer flush, 200 mL for 100 mL of filtered dialyzer flush, and 700 mL for 350 mL of filtered dialyzer flush. All plates were incubated at 30°C and screened for growth at 48 hours.

Laboratory processing of surface samples and water samples

Each 3M™ Sponge-Stick, (St. Paul, NM) was separated from its handle and homogenized at 200 rpm in a blender (Stomacher 400C, Seward UK) in phosphate-buffered saline containing 0.02% Tween® 80 (PBST). The sponge portion was then removed and the homogenate concentrated by centrifugation at 2700 ×g for 20 min. The supernatant was discarded, and the pellet resuspended. A 1:10 dilution was made by adding 0.1 mL of the homogenate to 0.9 mL Butterfield Buffer (BB) and 0.1 mL of both the undiluted and 1:10 dilution were plated onto Tryptic Soy Agar with 5% Sheep’s Blood (TSAII; Becton Dickinson, Sparks, MD) plates, in duplicate. Two aliquots of 0.1 mL and 0.5 mL of the undiluted homogenate were spread on MacConkey II agar (MAC). Trypticase soy broth (TSB) was added to the remaining homogenate and incubated overnight, before 0.1 mL was placed on an MAC plate and struck for isolation. All cultures were incubated for up to 2 days at 35ºC.

Swabs were rolled onto the first quadrant of a MAC plate, and then a loop was used to streak for isolation. The swab was then added to TSB and vortexed in 10 sec intervals for 2 minutes to release any microorganisms. Broth was incubated overnight at 35ºC, vortexed, and then 0.1 mL of broth enrichment was directly struck for isolation onto BAP and MAC. Plates were incubated at 35ºC and screened for suspect colony growth.

Exteriors of containers were sanitized with 70% ethanol, and 1:10 and 1:100 dilutions made in BB. 0.1 mL of the undiluted, 1:10, and 1:100 dilutions were plated onto Reasoner’s 2 Agar (R2A, prepared in house) and tryptic soy agar (TSA, Becton Dickinson, Sparks, MD). 1 mL was passed through a 0.45 micron pore sized membrane filter and placed onto either R2A or TSA. 0.5 mL of the sample was spread in duplicate on MAC, and the remainder filtered onto MAC. The MAC plates were incubated at 35ºC and the R2A and TSA incubated at 30ºC.