colistin)-resistant O25b:H4-ST131-*H*30R. The worstcase scenario is the global spread of this isolate, which has acquired resistance to the last-line antimicrobial drug, colistin.

# Acknowledgments

We thank Osamu Kuwahara for providing some of the *E. coli* clinical isolates.

This study was partly supported by grants from JSPS KAKENHI (grant nos. 15H06521 and 25861574) and the Yuasa Memorial Foundation.

## References

- Yokota S, Sato T, Okubo T, Ohkoshi Y, Okabayashi T, Kuwahara O, et al. Prevalence of fluoroquinolone-resistant *Escherichia coli* O25:H4-ST131 (CTX-M-15-nonproducing) strains isolated in Japan. Chemotherapy. 2012;58:52–9. http://dx.doi.org/10.1159/000336129
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing (M100-S25). Wayne (PA): The Institute; 2015.
- Colpan A, Johnston B, Porter S, Clabots C, Anway R, Thao L, et al.; VICTORY (Veterans Influence of Clonal Types on Resistance: Year 2011) Investigators. *Escherichia coli* sequence type 131 (ST131) subclone H30 as an emergent multidrug-resistant pathogen among US veterans. Clin Infect Dis. 2013;57:1256–65. http://dx.doi.org/10.1093/cid/cit503
- Price LB, Johnson JR, Aziz M, Clabots C, Johnston B, Tchesnokova V, et al. The epidemic of extended-spectrumβ-lactamase-producing *Escherichia coli* ST131 is driven by a single highly pathogenic subclone, H30-Rx. mBio. 2013;4: e00377–13. http://dx.doi.org/10.1128/mBio.00377-13
- Lau SH, Reddy S, Cheesbrough J, Bolton FJ, Willshaw G, Cheasty T, et al. Major uropathogenic *Escherichia coli* strain isolated in the northwest of England identified by multilocus sequence typing. J Clin Microbiol. 2008;46:1076–80. http://dx.doi.org/10.1128/JCM.02065-07
- Peirano G, Schreckenberger PC, Pitout JD. Characteristics of NDM-1-producing *Escherichia coli* isolates that belong to the successful and virulent clone ST131. Antimicrob Agents Chemother. 2011;55:2986–8. http://dx.doi.org/10.1128/AAC.01763-10
- Morris D, Boyle F, Ludden C, Condon I, Hale J, O'Connell N, et al. Production of KPC-2 carbapenemase by an *Escherichia coli* clinical isolate belonging to the international ST131 clone. Antimicrob Agents Chemother. 2011;55:4935–6. http://dx.doi.org/10.1128/AAC.05127-11
- Liu YY, Wang Y, Walsh TR, Yi LX, Zhang R, Spencer J, et al. Emergence of plasmid-mediated colistin resistance mechanism MCR-1 in animals and human beings in China: a microbiological and molecular biological study. Lancet Infect Dis. 2016;16:161–8. http://dx.doi.org/10.1016/S1473-3099(15)00424-7
- Xavier BB, Lammens C, Ruhal R, Kumar-Singh S, Butaye P, Goossens H, et al. Identification of a novel plasmid-mediated colistin-resistance gene, *mcr-2*, in *Escherichia coli*, Belgium, 2016. Euro Surveill. 2016;21. http://dx.doi.org/10.2807/1560-7917. ES.2016.21.2730280

Address for correspondence: Toyotaka Sato, Department of Microbiology, Sapporo Medical University School of Medicine, S1 W17, Chuo-ku, Sapporo, 060–8556, Japan; email: sato.t@sapmed.ac.jp

# Dual Emergence of Usutu Virus in Common Blackbirds, Eastern France, 2015

# Sylvie Lecollinet, Yannick Blanchard, Christine Manson, Steeve Lowenski, Eve Laloy, Hélène Quenault, Fabrice Touzain, Pierrick Lucas, Cyril Eraud, Céline Bahuon, Stéphan Zientara, Cécile Beck, Anouk Decors

Author affiliations: ANSES Animal Health Laboratory of Maisons-Alfort, Maisons-Alfort, France (S. Lecollinet, S. Lowenski, C. Bahuon, S. Zientara, C. Beck); ANSES Ploufragan, Ploufragan, France (Y. Blanchard, H. Quenault, F. Touzain, P. Lucas); Departmental Veterinary Laboratory of Haut-Rhin (LVD68), Colmar, France (C. Manson); ENVA, Maisons-Alfort (E. Laloy); ONCFS, Paris, France (C. Eraud, A. Decors)

## DOI: http://dx.doi.org/10.3201/eid2212.161272

To the Editor: Usutu virus (USUV) is a mosquitoborne flavivirus amplified in an enzootic cycle involving passeriform and strigiform birds as reservoir hosts and *Culex* mosquitos as vectors (1). Although originating from Africa, USUV has been introduced at least twice into central and western Europe, leading to substantial bird fatalities in central Europe (particularly in Austria, Hungary, Italy, Germany, and Switzerland) since 1996 (2). Its zoonotic potential has been recently highlighted in Italy in immunosuppressed patients who sought treatment for encephalitis (3).

Even though every country bordering France, apart from Luxembourg, has reported USUV in mosquitoes or wild birds recently, USUV outbreaks had not been reported in France, and only indirect evidence indicated circulation of USUV-like viruses in Eurasian magpies (Pica pica) in southeastern France (4). In 2015, the French event-based surveillance network SAGIR (5) reported increased fatalities of common blackbirds (Turdus merula) in 2 departments in eastern France, Haut-Rhin near the German border and Rhône (Figure). Five birds, 2 in Haut-Rhin and 3 in Rhône, were subjected to molecular detection for flaviviruses. During necropsy, their brains, hearts, livers, and kidneys (from 2 birds only) were sampled for RNA extraction and virus isolation. Tissues were homogenized in DMEM with ceramic beads (Qbiogen) and FastPrep ribolyzer (ThermoSavant). Total RNA was extracted with RNeasy kit (Qiagen) and flavivirus genomic RNA was amplified by conventional reverse transcription PCR with all of the tissues from 2 birds in Haut-Rhin that were found dead on August 5–10, 2015, and from 1 bird sampled on September 23 in Rhône (6). USUV was systematically identified in blackbird tissues



**Figure**. Phylogeny of Usutu virus (USUV) Haut-Rhin strains (black circles) and Rhône strain (black triangle), isolated in 2015 in eastern France compared with reference strains. Inset map shows locations where isolates were obtained. The strains from France are genetically distinct, with a homology of 95.7% at the nucleotide level and 98.8% (3,392–3 aa/3,434 aa) at the amino acid level. The evolutionary history was inferred by using the neighbor-joining method. The optimal tree with the sum of branch length 0.60224968 is shown. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (1,000 replicates) are shown at the nodes. Evolutionary analyses were conducted in MEGA6 (http://www.megasoftware.net), and the evolutionary distances were computed by using the Jukes-Cantor method. The resulting tree is drawn to scale, with branch lengths in the units of number of base substitutions per site. The analysis involved 22 strains, including West Nile virus (WNV) as the root; GenBank accession numbers are indicated. All positions containing gaps and missing data were not included; 10,684 positions were included in the final dataset. An outline of the organism from which the virus was isolated (bat, bird, mosquito, or human) is placed next to the strain name. Scale bar indicates substitutions per site.

by Sanger sequencing of the 1085-nt PCR fragment and BLAST analysis (https://blast.ncbi.nlm.nih.gov). Three USUV isolates were obtained after 2–3 passages in Vero cells, and whole-genome sequencing of every isolate was performed as previously described (7). Postmortem examination revealed hepatomegaly and splenomegaly in a USUV-infected blackbird and marked emaciation and kidney hemorrhages in another infected animal. A subset of samples was submitted for histologic analysis, but no microscopic lesions were found in any of the 3 USUV-positive blackbirds, suggesting that infection was hyperacute. Phylogenetic analysis of the whole genome for the 3 USUV isolates demonstrated close genetic relatedness between USUV isolates from Haut-Rhin, France, and Germany (99.8% nucleotide identity with USUV-5684/ Germany/2011, GenBank accession no. KJ438716) and between strains from Rhône, France, and Spain (99.2% identity with USUV-MB11906/Spain/2006, GenBank accession no. KF573410). Results showed that French USUV strains from Haut-Rhin and Rhône departments were clearly distinct from each other (95.7% nucleotide identity) and arose from  $\geq 2$  independent introduction events. In total, 41–42 nonsynonymous mutations were identified along

the 3,434-aa long polyprotein, with capsid, nonstructural protein 2A, and nonstructural protein 4B having the highest nonsynonymous substitution rates of 96.0% (121/126), 97.4% (221/227), and 97.8% (311/318), respectively.

Symptomatic USUV infections were discovered in wild birds in France, indicating the emergence of USUV in counties in eastern France. Unusual and grouped bird fatalities observed in August and September 2015 in common blackbirds in Haut-Rhin and Rhône did not seem to alter blackbird population dynamics (data not shown). The viral strain recovered in Haut-Rhin, which borders Germany, is genetically similar to USUV strains isolated in central Europe, in particular in southwestern Germany in 2011. Such a finding further exemplifies the continuing and gradual diffusion of the Vienna USUV strain since 2001 (Austria in 2001, Hungary in 2005, Italy and Switzerland in 2006, Germany and Czech Republic in 2011, and Belgium in 2012) (1). The USUV strain isolated from the 1 blackbird in Rhône shared the highest genetic homology with USUV strains identified on 2 occasions in Spain: once in 2006 in Catalonia from C. pipiens mosquitoes and once in 2009 in Andalusia from C. perex*iguus* mosquitoes (8).

Our findings indicate that the USUV/Spain strain can be pathogenic in birds. Symptomatic USUV infections in wild avifauna are difficult to quantify (because of low reporting rates and quick removal of dead birds by scavengers), and dynamic modeling of USUV in Austria indicated that a low proportion (0.2%) of USUV-killed birds had been effectively detected by USUV-specific surveillance programs (9). Mutations between USUV-Rhône2705/France/2015 and USUV-MB11906/Spain could also account for differential virulence in birds. These 2 strains differed by 14 nonsynonymous mutations (online Technical Appendix Table, http://wwwnc.cdc. gov/EID/article/22/12/16-1272-Techapp1.pdf). Although little is known about molecular determinants of USUV virulence, one can try to infer the importance of these mutations from data gained from studies on a closely related flavivirus, West Nile virus. In this respect, none of the 14 mutations observed have been found to be critical in flavivirus virulence.

Concomitantly with USUV emergence in France, another *Culex*-borne flavivirus, West Nile virus, has reemerged in southeastern France (10). Climatic and environmental conditions during the summer of 2015 seem to have promoted the spread of *Culex*-borne pathogens. However, risk factors for flavivirus emergence in France in 2015 have not been comprehensively analyzed. bird samples. We are grateful to the technicians of the Hunting Federation and of the National Hunting and Wildlife Agency for their contribution to wildlife surveillance.

This work was partially supported by the European Union Horizon 2020 Framework Program for Research and Innovation under grant agreement no. 643476 (COMPARE).

### References

- Ashraf U, Ye J, Ruan X, Wan S, Zhu B, Cao S. Usutu virus: an emerging flavivirus in Europe. Viruses. 2015;7:219–38. http://dx.doi.org/10.3390/v7010219
- Weissenböck H, Bakonyi T, Rossi G, Mani P, Nowotny N. Usutu virus, Italy, 1996. Emerg Infect Dis. 2013;19:274–7. http://dx.doi.org/10.3201/eid1902.121191
- Vazquez A, Jimenez-Clavero M, Franco L, Donoso-Mantke O, Sambri V, Niedrig M, et al. Usutu virus: potential risk of human disease in Europe. Euro Surveill. 2011;16:19935.
- Vittecoq M, Lecollinet S, Jourdain E, Thomas F, Blanchon T, Arnal A, et al. Recent circulation of West Nile virus and potentially other closely related flaviviruses in southern France. Vector Borne Zoonotic Dis. 2013;13:610–3. http://dx.doi.org/10.1089/ vbz.2012.1166
- Decors A, Hars J, Faure E, Quintaine T, Chollet J, Rossi S. Le réseau Sagir: un outil de vigilance vis-à-vis des agents pathogènes exotiques [in French]. Bulletin épidémiologique santé animale-alimentation. 2014;66:35–9. [cited 2016 Sep 28]. http://bulletinepidemiologique.mag.anses.fr/sites/default/files/BEPmg-BE66-art10\_0.pdf
- Weissenböck H, Kolodziejek J, Url A, Lussy H, Rebel-Bauder B, Nowotny N. Emergence of Usutu virus, an African mosquitoborne flavivirus of the Japanese encephalitis virus group, central Europe. Emerg Infect Dis. 2002;8:652–6. http://dx.doi.org/10.3201/ eid0807.020094
- Grasland B, Bigault L, Bernard C, Quenault H, Toulouse O, Fablet C, et al. Complete genome sequence of a porcine epidemic diarrhea S gene indel strain isolated in France in December 2014. Genome Announc. 2015;3:e00535-15. http://dx.doi.org/10.1128/ genomeA.00535-15
- Bakonyi T, Busquets N, Nowotny N. Comparison of complete genome sequences of Usutu virus strains detected in Spain, central Europe, and Africa. Vector Borne Zoonotic Dis. 2014;14:324–9. http://dx.doi.org/10.1089/vbz.2013.1510
- Rubel F, Brugger K, Hantel M, Chvala-Mannsberger S, Bakonyi T, Weissenböck H, et al. Explaining Usutu virus dynamics in Austria: model development and calibration. Prev Vet Med. 2008;85:166– 86. http://dx.doi.org/10.1016/j.prevetmed.2008.01.006
- Bahuon C, Marcillaud-Pitel C, Bournez L, Leblond A, Beck C, Hars J, et al. West Nile virus epizootics in Camargue, France, in 2015, and reinforcement of West Nile virus surveillance and control networks. Paris: World Organisation for Animal Health; 2016 [cited 2016 Sep 28]. http://www.oie.int/fileadmin/Home/eng/ Publications\_%26\_Documentation/docs/pdf/bulletin/Bull\_2016-1-ENG.pdf

Address for correspondence: Sylvie Lecollinet, ANSES Laboratoire de santé animale de Maisons-Alfort, 14 rue Pierre et Marie Curie, Maisons-Alfort 94701, France; email: sylvie.lecollinet@anses.fr

#### Acknowledgments

We are grateful to Eliette Gretillat and the whole team at the veterinary laboratory of Haut-Rhin department for processing