LETTERS

References

- Yip CC, Lau SK, Woo PC, Yuen KY. Human enterovirus 71 epidemics: what's next? Emerg Health Threats J. 2013;6:19780. http://dx.doi.org/10.3402/ehtj.v6i0.19780
- Brown BA, Oberste MS, Alexander JP Jr, Kennett ML, Pallansch MA. Molecular epidemiology and evolution of enterovirus 71 strains isolated from 1970 to 1998. J Virol. 1999;73:9969–75.
- McMinn PC. Recent advances in the molecular epidemiology and control of human enterovirus 71 infection. Curr Opin Virol. 2012;2:199–205. http://dx.doi.org/10.1016/j.coviro.2012.02.009
- Saxena VK, Sane S, Nadkarni SS, Sharma DK, Deshpande JM. Genetic diversity of enterovirus A71, India. Emerg Infect Dis. 2015;21:123–6.
- Bessaud M, Razafindratsimandresy R, Nougairède A, Joffret ML, Deshpande JM, Dubot-Pérès A, et al. Molecular comparison and evolutionary analyses of VP1 nucleotide sequences of new African human enterovirus 71 isolates reveal a wide genetic diversity. PLoS One. 2014;9:e90624. http://dx.doi.org/10.1371/journal. pone.0090624
- Fischer TK, Nielsen AY, Sydenham TV, Andersen PH, Andersen B, Midgley SE. Emergence of enterovirus 71 C4a in Denmark, 2009 to 2013. Euro Surveill. 2014;19:p=20911. http://dx.doi.org/10.2807/1560-7917.ES2014.19.38.20911
- Hassel C, Mirand A, Lukashev A, TerletskaiaLadwig E, Farkas A, Schuffenecker I, et al. Transmission patterns of human enterovirus 71 to, from and among European countries, 2003 to 2013. Euro Surveill. 2015;20:pi=30005. http://dx.doi.org/10.2807/1560-7917. ES.2015.20.34.30005
- Tee KK, Lam TT, Chan YF, Bible JM, Kamarulzaman A, Tong CY, et al. Evolutionary genetics of human enterovirus 71: origin, population dynamics, natural selection, and seasonal periodicity of the VP1 gene. J Virol. 2010;84:3339–50. http://dx.doi.org/10.1128/ JVI.01019-09
- Kroneman A, Vennema H, Deforche K, Avoort Hvd, Peñaranda S, Oberste MS, et al. An automated genotyping tool for enteroviruses and noroviruses. J Clin Virol. 2011;51:121–5. http://dx.doi.org/10.1016/j.jcv.2011.03.006
- Tong CY, Bible JM, Platt C. A fatal case of enterovirus 71 infection with a single nucleotide variation in domain V of the 5' untranslated region. Pediatr Infect Dis J. 2011;30:1013–4. http://dx.doi.org/10.1097/INF.0b013e318228ba24

Address for correspondence: Sabine Diedrich, National Reference Centre for Poliomyelitis and Enteroviruses, Robert Koch Institute, Seestraße 10, 13353 Berlin, Germany; email: DiedrichS@rki.de

Cerebral Syphilitic Gumma within 5 Months of Syphilis in HIV-Infected Patient

Motoyuki Tsuboi, Takeshi Nishijima, Katsuji Teruya, Yoshimi Kikuchi, Hiroyuki Gatanaga, Shinichi Oka

Author affiliations: National Center for Global Health and Medicine, Tokyo, Japan (M. Tsuboi, T. Nishijima, K. Teruya, Y. Kikuchi, H. Gatanaga, S. Oka); Kumamoto University, Kumamoto, Japan (H. Gatanaga, S. Oka)

DOI: http://dx.doi.org/10.3201/eid2210.160600

To the Editor: Tertiary syphilis, including cerebral syphilitic gumma, usually occurs >10 years after contracting syphilis (1) and is a rare manifestation since the introduction of penicillin (2). However, progression of syphilis is reported to be faster in HIV-infected patients than in those without such infections (3). We report a case of cerebral syphilitic gumma in an HIV-1–infected patient for whom serum samples obtained as recently as 5 months earlier showed negative results for syphilis.

A 21-year-old man infected with HIV came to the AIDS Clinical Center, National Center for Global Health and Medicine (Tokyo, Japan), because of a 2-hour loss of consciousness. He reported an uncomfortable feeling at the back of his head and neck and eye fatigue that lasted for 1 week. His HIV-1 infection was well-controlled with an antiretroviral combination of tenofovir, emtricitabine, and dolutegravir. The patient had a CD4 count of 565 cells/mL and a viremia level below detectable limits (<20 copies/mL). He was not using any other medications.

At examination, his vital signs were within reference ranges. Apart from a tongue bite, physical and neurologic examinations showed no abnormal findings. Results for chest radiograph, Holter electrocardiogram, and electroencephalogram were unremarkable. There were no abnormal ophthalmologic findings. Computed tomography of the brain showed a hypodense lesion at the left frontal lobe (Figure, panel A). Subsequent magnetic resonance imaging showed that the lesion (mass) was hypointense by gadolinium-enhanced, axial, T1weighted imaging (Figure, panel B), hyperintense by T2-weighted imaging, and surrounded by extensive cerebral edema (Figure, panel C).

Symptomatic epilepsy caused by the mass was suspected to have caused the loss of consciousness. This conclusion was based on the intracranial mass, long duration of loss of consciousness, increase in creatine kinase level (471 U/L), and tongue bite.

When the HIV-1 infection was diagnosed in the patient 15 months earlier, results of serum rapid plasma reagin (RPR) and *Treponema pallidum* hemagglutination test (TPHA) were negative. However, during this examination, serum RPR and TPHA titers were 1:32 and 1:10,240, respectively. Results of cerebrospinal fluid (CSF) analysis were compatible with neurosyphilis (4,5) and showed a leukocyte count of 35 cells/mL (2 neutrophils/mL, 33 lymphocytes/mL), a total protein level of 30 mg/dL, a glucose level of 59 mg/dL (serum glucose level 92 mg/dL), an RPR titer of 1:<1, a TPHA titer of 1:160, and a fluorescent treponemal antibody-absorption titer of 1:32.

Cerebral syphilitic gumma was suspected on the basis of neurosyphilis and compatible imaging findings (6) and because other conditions, such as meningioma, primary central nervous system lymphoma, toxoplasmosis,

LETTERS

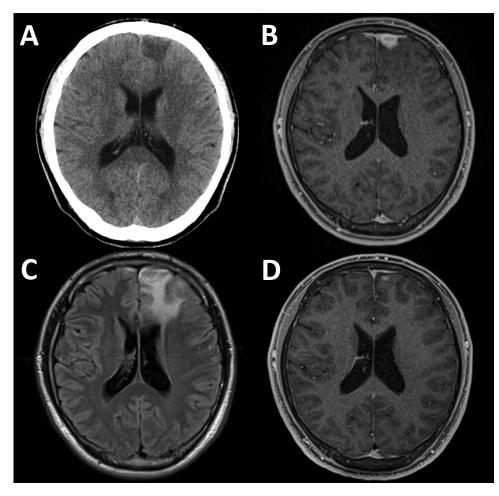


Figure. Diagnostic imaging results for a brain mass in a 21-year-old HIV-positive man with cerebral syphilitic gumma in Tokyo, Japan, for whom serum samples obtained as recently as 5 months earlier showed negative results for syphilis. A) Noncontrast, cranial computed tomography showing a hypodense lesion in the left frontal lobe. B) Gadoliniumenhanced, axial, T1-weighted magnetic resonance imaging (MRI) showing an enhanced lesion (mass) $(14 \times 14 \times 12 \text{ mm})$ adjacent to the enhanced dura in the left frontal lobe, C) Axial. fluid-attenuated inversion recovery MRI showing extensive left frontal edema. D) Two months after treatment for syphilis. gadolinium-enhanced, axial T1weighted MRI showing substantial resolution of the lesion.

cryptococcoma, tuberculoma, or brain abscess, were unlikely. These results were based on radiologic findings; high CD4 cell counts; and negative results for Epstein-Barr virus and malignant cells in CSF; serum cryptococcal antigen; CSF culture for bacteria, mycobacteria, and fungi; and interferon- γ release assay.

We did not perform a biopsy for the patient because of presumed high pretest probability of cerebral syphilitic gumma, the invasiveness of this complication, and the young age of the patient. However, successful therapy confirmed the diagnosis (3).

We empirically treated the patient with intravenous benzylpenicillin (24 million units/d for 14 consecutive days). Clinical symptoms improved shortly after treatment. At 2-month follow-up, all clinical symptoms and signs had resolved, and a 4-fold decrease in RPR titer was observed (RPR titer 1:8, TPHA titer 1:5,120) (7). The brain mass was substantially reduced, which confirmed the diagnosis of cerebral syphilitic gumma (Figure, panel D).

Gumma is a complication of long-term infection with *T. pallidum*, which develops 1–46 years after healing of secondary lesions; most cases develop within 15 years

(1). However, for our patient, cerebral syphilitic gumma developed within 5 months after he contracted syphilis. After written informed consent was obtained, serum samples were obtained from the patient at his first and subsequent clinic visits and stored. Samples obtained at 11 months, 10 months, 9 months, 5 months, 11 weeks, and 5 weeks before detection of the brain mass were then tested retrospectively for RPR and TPHA titers. Results were negative at 11, 10, 9, and 5 months before detection of the brain mass, but TPHA titers became positive (1:80) at 11 weeks before presentation, and RPR titers became positive (1:16) at 5 weeks before presentation. Because RPR and TPHA titers can become positive as late as 6 weeks after infection (8), we believe that the patient contracted syphilis within 5 months before documentation of the cerebral mass.

In conclusion, we report an HIV-1–infected patient in whom cerebral syphilitic gumma developed within 5 months after contracting syphilis. Cerebral syphilitic gumma should be considered in the differential diagnosis of a cerebral lesion in sexually active patients even if they had recently contracted syphilis. Moreover, as guidelines

LETTERS

recommend (9), screening of HIV-infected patients who are sexually active with multiple partners should be conducted every 3–6 months for early detection of syphilis and initiation of proper treatment to prevent transmission and progression to late syphilis.

Acknowledgments

We thank the clinical staff at the AIDS Clinical Center (Tokyo, Japan) for assistance.

This study was supported by Grant-in-Aids for AIDS research from the Japanese Ministry of Health, Labour, and Welfare (H23-AIDS-001 and H24-AIDS-003).

References

- Clark EG, Danbolt N. The Oslo study of the natural history of untreated syphilis; an epidemiologic investigation based on a restudy of the Boeck-Bruusgaard material; a review and appraisal. J Chronic Dis. 1955;2:311–44. http://dx.doi.org/10.1016/0021-9681(55)90139-9
- Noel CB, Moeketsi K, Kies B. Cavernous sinus syndrome, an atypical presentation of tertiary syphilis: case report and review of the literature. Clin Neurol Neurosurg. 2011;113:65–7. http://dx.doi.org/10.1016/j.clineuro.2010.08.007
- Pialoux G, Vimont S, Moulignier A, Buteux M, Abraham B, Bonnard P. Effect of HIV infection on the course of syphilis. AIDS Rev. 2008;10:85–92.
- Ghanem KG, Moore RD, Rompalo AM, Erbelding EJ, Zenilman JM, Gebo KA. Lumbar puncture in HIV-infected patients with syphilis and no neurologic symptoms. Clin Infect Dis. 2009;48:816–21. http://dx.doi.org/10.1086/597096
- Castro R, Prieto ES, da Luz Martins Pereira F. Nontreponemal tests in the diagnosis of neurosyphilis: an evaluation of the Venereal Disease Research Laboratory (VDRL) and the rapid plasma reagin (RPR) tests. J Clin Lab Anal. 2008;22:257–61. http://dx.doi.org/10.1002/jcla.20254
- Fargen KM, Alvernia JE, Lin CS, Melgar M. Cerebral syphilitic gummata: a case presentation and analysis of 156 reported cases. Neurosurgery. 2009;64:568–75. http://dx.doi.org/10.1227/01. NEU.0000337079.12137.89
- Workowski KA, Bolan GA. Sexually transmitted diseases treatment guidelines, 2015. MMWR Recomm Rep. 2015;64:1–137.
- Janier M, Hegyi V, Dupin N, Unemo M, Tiplica GS, Potocnik M, et al. 2014 European guideline on the management of syphilis. J Eur Acad Dermatol Venereol. 2014;28:1581–93. http://dx.doi.org/10.1111/jdv.12734
- Kaplan JE, Benson C, Holmes KK, Brooks JT, Pau A, Masur H, et al. Guidelines for prevention and treatment of opportunistic infections in HIV-infected adults and adolescents: recommendations from CDC, the National Institutes of Health, and the HIV Medicine Association of the Infectious Diseases Society of America. MMWR Recomm Rep. 2009;58:1–207.

Address for correspondence: Takeshi Nishijima, AIDS Clinical Center, National Center for Global Health and Medicine, 1-21-1, Toyama, Shinjuku, Tokyo 162-8655, Japan; email: tnishiji@acc.ncgm.go.jp

African Tick-Bite Fever in Traveler Returning to Slovenia from Uganda

Petra Bogovic, Stanka Lotric-Furlan, Misa Korva, Tatjana Avsic-Zupanc

Author affiliations: University Medical Center Ljubljana, Ljubljana, Slovenia (P. Bogovic, S. Lotric-Furlan); Institute of Microbiology and Immunology, Ljubljana (M. Korva, T. Avsic-Zupanc)

DOI: http://dx.doi.org/10.3201/eid2210.160650

To the Editor: African tick-bite fever (ATBF) is a well known disease in travelers to sub-Saharan Africa (I). The causative agent, *Rickettsia africae*, is transmitted to humans by ticks of the genus *Amblyomma* (I,2). *R. africae* has been isolated or detected in ticks, humans, or both in 22 sub-Saharan countries (3). Most ATBF cases have been described in tourists returning from countries to which it is endemic, most often from South Africa, Zimbabwe, and Botswana (4). We report a case of ATBF in a Slovenian traveler returning from Uganda.

In June 2015, a 29-year-old Slovenian man without underlying illnesses sought care at the Department of Infectious Diseases, University Medical Centre Ljubljana (Ljubljana, Slovenia). He had a 1-day history of fever up to 38°C without chills, 5 days after returning from a 2-week trip to Uganda. He had received vaccines against yellow fever and viral hepatitis A before traveling and did not use antimalarial prophylaxis during his stay in southwestern Uganda. A day before he left Uganda, he noticed a tick bite on his left upper abdomen.

At initial examination, he appeared well. He had a temperature of 37.8°C, pulse rate 75 beats/min, and blood pressure 120/80 mm Hg. Skin examination was remarkable for a solitary papular lesion at the site of tick bite surrounded by a small erythematous halo associated with discrete lymphangitic streaking and painful enlarged left axillar lymph nodes. Results of initially performed routine laboratory tests were normal.

On day 5 of illness, the man was still febrile, with a temperature up to 39°C. Papular skin lesion had developed a dark brown crusted center (compatible with a tache noire), and some new discrete asymptomatic pale papular skin lesions appeared on his left leg and arm. Repeat laboratory testing indicated only mildly increased serum C-reactive protein (16.0 mg/L [reference <5 mg/L]).

The clinical course improved rapidly after treatment began with doxycycline. Fever resolved in 2 days, and enlarged lymph nodes and skin lesions resolved completely within 14 days.

Microbiological procedures to detect for infection with tick-transmitted pathogens were performed to elucidate the