



# Full-Genome Sequence of a Neuroinvasive West Nile Virus Lineage 2 Strain from a Fatal Horse Infection in South Africa

### Juliet L. D. Mentoor,<sup>a\*</sup> Alison B. Lubisi,<sup>b</sup> Truuska Gerdes,<sup>b</sup> Stacey Human,<sup>a\*</sup> June H. Williams,<sup>c</sup> Marietjie Venter<sup>a,d</sup>

Zoonosis Research Unit, Department Medical Virology, University of Pretoria, Pretoria, South Africa<sup>a</sup>; Virology Molecular Epidemiology and Diagnostics Programme, Agricultural Research Council, Onderstepoort Veterinary Institute, Pretoria, South Africa<sup>b</sup>; Department of Paraclinical Sciences, Faculty of Veterinary Sciences, University of Pretoria, South Africa<sup>c</sup>; Centre for Global Disease Detection, South Africa, Division of Global Health Protection, Centers for Disease Control and Prevention, Pretoria, South Africa<sup>d</sup>

\* Present address: Juliet L. D. Mentoor, Department of Genetics, Faculty of Health, University of Pretoria, Pretoria, South Africa; Stacey Human, Division of Pediatric Infectious Diseases, Emory University, Atlanta, Georgia, USA.

We report here the complete genome sequence of a lineage 2 West Nile virus (WNV) strain that resulted in fatal neurological disease in a horse in South Africa. Several recent reports exist of neurological disease associated with lineage 2 WNV in humans and horses in South Africa and Europe; however, there are a lack of sequencing data from recent fatal cases in Southern Africa, where these strains likely originate. A better understanding of the genetic composition of highly neuroinvasive lineage 2 strains may facilitate the identification of putative genetic factors associated with increased virulence.

Received 8 June 2016 Accepted 10 June 2016 Published 28 July 2016

Citation Mentoor JLD, Lubisi AB, Gerdes T, Human S, Williams JH, Venter M. 2016. Full-genome sequence of a neuroinvasive West Nile virus lineage 2 strain from a fatal horse infection in South Africa. Genome Announc 4(4):e00740-16. doi:10.1128/genomeA.00740-16.

Copyright © 2016 Mentoor et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Marietjie Venter, yds8@cdc.gov.

est Nile virus (WNV) is a mosquito-transmitted flavivirus (family *Flaviviridae*) of global concern (1, 2). In humans, ~20% of infections result in WN fever associated with rash, arthralgia, and myalgia, while <1% may develop severe neurological disease and death (3). In horses, 20% of WNV cases exhibit clinical signs; however, ~90% of these cases may develop neurological disease, with a fatality rate approaching 30% (1). WNV lineage 2 (L2) viruses are endemic to South Africa and Madagascar (4, 5) and have emerged in Europe in the past 10 years, causing outbreaks in humans and horses (6, 7). WNV genome sequencing has provided insight into possible genetic factors that may differentiate between highly pathogenic and attenuated strains (8, 9). Previously, we described WNVL2 strains associated with severe neurological diseases in horses and humans in South Africa (4, 10–13). WNVL2 full-genome sequences from Africa are limited, with the only currently available sequence originating from a nonfatal encephalitis, a fatal hepatitis case, and febrile infections in humans from South Africa (8, 14).

As part of a study investigating WNV as the cause of severe neurological disease in horses in South Africa, an isolate (HS101/ 08) was obtained from a brain tissue sample from a horse that had died of severe neurological signs in 2008, including seizures and complete paralysis (4, 12). This study aimed to characterize this neuroinvasive strain by describing its complete genome sequence and comparing it to both highly and less neuroinvasive lineage 1 and 2 strains. HS101/08 was isolated on Vero cell monolayers, and supernatants were harvested after three passages. Viral particles were concentrated through  $0.2-\mu$ m-pore microfilters (Sigma-Aldrich) and treated with DNase and RNase. Nucleic acid extraction was performed with the High Pure viral nucleic acid isolation kit (Roche Diagnostics, Mannheim, Germany) and the RNA minikit (Qiagen, Hilden, Germany). cDNA was synthesized with the Expand reverse transcriptase kit (Roche Diagnostics). Universal flavivirus and WNV-specific primers were used to amplify the complete genome (15). Cycle sequencing was performed with the ABI BigDye Terminator version 3.1 kit (Applied Biosystems, CA). Phylogenetic and nucleotide analyses were performed using BioEdit (version 7.0.5.2) and MEGA version 7 (16, 17).

The HS101/08 genome sequence was 11,042 bp in size and closely related to neuroinvasive strains SPU116/89 and SA93/01 (accession numbers EF429197 and EF429198) from South Africa and a subclade of neurovirulent human and horse strains from Italy and Greece (7, 8). HS101/08 shared 98% nucleotide identity with SPU116/89 and SA93/01. These two strains are highly neuroinvasive in mice (18, 19). An amino acid sequence comparison between HS101/08 and SA93/01 identified six uncharacterized substitution mutations predicted to have no effect on the structural or nonstructural proteins. We verified the presence of a 154N-S156 glycosylation site in the envelope protein sequences of South African L2 strains (including HS101/08), which were previously suggested to be associated with increased mouse neurovirulence (20). Similarities between the highly neurovirulent HS101/08 isolate and human isolates from cases of severe disease confirms that highly pathogenic L2 strains are still circulating in South Africa. WNV should be considered as a diagnosis in human and animal neurological cases.

**Nucleotide sequence accession number.** The accession number of the complete coding sequence of HS101/08 is JN393308.

### ACKNOWLEDGMENTS

We thank the veterinarians that participated in the surveillance program for neurological disease in South Africa and staff members at the Zoonosis Research Unit involved in the diagnosis of cases. We also thank Liz Botha for critical review of the manuscript. The work presented here formed part of the research performed by Juliet L. D. Mentoor during her M.Sc. degree at the University of Pretoria, Department of Medical Virology.

The findings and conclusions of this report are those of the authors and do not necessarily represent the official position of the U.S. Centers for Disease Control and Prevention.

## FUNDING INFORMATION

This work, including the efforts of Marietjie Venter, was funded by National Health Laboratory Service grant (2009-2011). This work, including the efforts of Juliet L. D. Mentoor and Stacey Human, was funded by National Research Foundation (NRF) grant (2007-2011).

The funders had no role in study design, data collection and interpretation, or the decision to submit the work for publication.

#### REFERENCES

- 1. Chancey C, Grinev A, Volkova E, Rios M. 2015. The global ecology and epidemiology of West Nile virus. Biomed Res Int 2015:376230. http://dx.doi.org/10.1155/2015/376230.
- Gray TJ, Webb CE. 2014. A review of the epidemiological and clinical aspects of West Nile virus. Int J Gen Med 7:193–203. http://dx.doi.org/ 10.2147/IJGM.S59902.
- Sejvar JJ. 2014. Clinical manifestations and outcomes of West Nile virus infection. Viruses 6:606–623. http://dx.doi.org/10.3390/v6020606.
- 4. Venter M, Human S, Zaayman D, Gerdes GH, Williams J, Steyl J, Leman PA, Paweska JT, Setzkorn H, Rous G, Murray S, Parker R, Donnellan C, Swanepoel R. 2009. Lineage 2 West Nile virus as cause of fatal neurologic disease in horses, South Africa. Emerg Infect Dis 15: 877–884. http://dx.doi.org/10.3201/eid1506.081515.
- 5. Wesula Olivia L, Mosomtai G, Symekher S. 2015. West Nile virus, a reemerging virus. Precis Med 2:e604.
- 6. Hernández-Triana LM, Jeffries CL, Mansfield KL, Carnell G, Fooks AR, Johnson N. 2014. Emergence of West Nile virus lineage 2 in Europe: a review on the introduction and spread of a mosquito-borne disease. Front Public Health 2:271.
- Bouzalas IG, Diakakis N, Chaintoutis SC, Brellou GD, Papanastassopoulou M, Danis K, Vlemmas I, Seuberlich T, Dovas CI. 7 February 2015. Emergence of equine West Nile encephalitis in Central Macedonia, Greece, 2010. Transbound Emerg Dis [Epub ahead of print.] http:// dx.doi.org/10.1111/tbed.12334.
- 8. Barzon L, Pacenti M, Franchin E, Lavezzo E, Masi G, Squarzon L, Pagni S, Toppo S, Russo F, Cattai M, Cusinato R, Palù G. 2013. Whole genome

sequencing and phylogenetic analysis of West Nile virus lineage 1 and lineage 2 from human cases of infection, Italy, August 2013. Euro Surveill 18:pii=20591. http://www.eurosurveillance.org/ViewArticle.aspx?Article Id=20591.

- McMullen AR, Albayrak H, May FJ, Davis CT, Beasley DW, Barrett AD. 2013. Molecular evolution of lineage 2 West Nile virus. J Gen Virol 94:318–325. http://dx.doi.org/10.1099/vir.0.046888-0.
- Venter M, Burt FJ, Blumberg L, Fickl H, Paweska J, Swanepoel R. 2009. Cytokine induction after laboratory-acquired West Nile virus infection. N Engl J Med 360:1260–1262. http://dx.doi.org/10.1056/NEJMc0808647.
- Venter M, Swanepoel R. 2010. West Nile virus lineage 2 as a cause of zoonotic neurological disease in humans and horses in southern Africa. Vector Borne Zoonotic Dis 10:659–664. http://dx.doi.org/10.1089/ vbz.2009.0230.
- Williams JH, van Niekerk S, Human S, van Wilpe E, Venter M. 2014. Pathology of fatal lineage 1 and 2 West Nile virus infections in horses in South Africa. J S Afr Vet Assoc 85:1105–1118. http://dx.doi.org/10.4102/ jsava.v85i1.1105.
- Zaayman D, Venter M. 2012. West Nile virus neurologic disease in humans, South Africa, September 2008–May 2009. Emerg Infect Dis 18: 2051–2054. http://dx.doi.org/10.3201/eid1812.111208.
- 14. Botha EM, Markotter W, Wolfaardt M, Paweska JT, Swanepoel R, Palacios G, Nel LH, Venter M. 2008. Genetic determinants of virulence in pathogenic lineage 2 West Nile virus strains. Emerg Infect Dis 14:222–230. http://dx.doi.org/10.3201/eid1402.070457.
- 15. **Botha EM**. 2008. Molecular characterization of South African lineage II West Nile virus isolates and development of a diagnostic assay. M.Sc. dissertation. University of Pretoria, Pretoria, South Africa.
- Kumar S, Hedges SB, Tamura K. 2016. MEGA7: Molecular Evolutionary Genetics Analysis version 7.0 for bigger datasets. Mol Biol Evol 33: 863–869. http://dx.doi.org/10.1093/molbev/msw026.
- 17. Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Ibis Biosciences, Carlsbad, CA.
- Venter M, Myers TG, Wilson MA, Kindt TJ, Paweska JT, Burt FJ, Leman PA, Swanepoel R. 2005. Gene expression in mice infected with West Nile virus strains of different neurovirulence. Virology 342:119–140. http://dx.doi.org/10.1016/j.virol.2005.07.013.
- Williams JH, Mentoor JDL, Van Wilpe E, Venter M. 2014. Comparative pathology of neurovirulent lineage 1 (NY99/385) and lineage 2 (SPU93/ 01) West Nile virus infections in BALB/c mice. Vet Pathol 52:140–151.
- Donadieu E, Lowenski S, Servely J-L, Laloy E, Lilin T, Nowotny N, Richardson J, Zientara S, Lecollinet S, Coulpier M. 2013. Comparison of the neuropathology induced by two West Nile virus strains. PLoS One 8:e84473. http://dx.doi.org/10.1371/journal.pone.0084473.