



Published in final edited form as:

Cancer Causes Control. 2016 January ; 27(1): 47–57. doi:10.1007/s10552-015-0681-6.

Energy homeostasis genes and survival after breast cancer diagnosis: The Breast Cancer Health Disparities Study

Andrew J. Pellatt¹, Abbie Lundgreen¹, Roger K. Wolff¹, Lisa Hines², Esther M. John³, and Martha L. Slattery¹

¹University of Utah, Department of Medicine, 383 Colorow, Salt Lake City, Utah 84108

²University of Colorado at Colorado Springs, Department of Biology, 1420 Austin Bluffs Parkway, Colorado Springs, Colorado 80918

³Cancer Prevention Institute of California, Fremont, California, 94538 and Department of Health Research and Policy (Epidemiology) and Stanford Cancer Institute, Stanford University School of Medicine, Stanford, California 94305

Abstract

Purpose—The leptin-signaling pathway and other genes involved with energy homeostasis (EH), have been examined in relation to breast cancer risk as well as to obesity. We test the hypothesis that genetic variation in EH genes influences survival after diagnosis with breast cancer and that body mass index (BMI) will modify that risk.

Methods—We evaluated associations between 10 energy homeostasis genes and survival among 1186 non-Hispanic white (NHW) and 1155 Hispanic/Native American women diagnosed with breast cancer. Percent Native American (NA) ancestry was determined from 104 Ancestry Informative Markers. Adaptive rank truncation product (ARTP) was used to determine gene and pathway significance.

Results—The overall EH pathway was marginally significant for all-cause mortality among women with low NA ancestry ($P_{\text{ARTP}} = 0.057$). Within the pathway, ghrelin (*GHRL*) and leptin receptor (*LEPR*) were significantly associated with all-cause mortality ($P_{\text{ARTP}} = 0.035$ and 0.007 , respectively). The EH pathway was significantly associated with breast cancer-specific mortality among women with low NA ancestry ($P_{\text{ARTP}} = 0.038$). Three genes, cholecystokinin (*CCK*), *GHRL*, and *LEPR* were significantly associated with breast cancer-specific mortality among women with low NA ancestry ($P_{\text{ARTP}} = 0.046$, 0.015 , and 0.046 , respectively) while neuropeptide Y (*NPY*) was significantly associated with breast cancer-specific mortality among women with higher NA ancestry ($P_{\text{ARTP}} = 0.038$). BMI did not modify these associations.

Conclusions—Our data support our hypothesis that certain EH genes influence survival after diagnosis with breast cancer; associations appear to be most important among women with low NA ancestry.

Address correspondence to Dr. Slattery, University of Utah, Department of Medicine, 383 Colorow, Salt Lake City, Utah 84108. The authors have no conflict of interest to report.

Keywords

Breast Cancer; Energy Homeostasis; Leptin Receptor; Ghrelin; Neuropeptide Y; cholecystokinin

The leptin-signaling pathway is positively associated with obesity and has been shown to stimulate the growth of human breast cancer cells. The biological effects of leptin (LEP) are exerted through binding to the leptin receptor (LEPR). This receptor is expressed in a variety of immune cells and has been shown in breast cancer cell lines to have direct communication with estrogen receptor alpha (1). The leptin-signaling pathway, along with other energy homeostasis (EH) genes, have been examined in relation to breast cancer risk as well as to obesity (2). Cocaine and amphetamine regulated transcript protein (*CARTPT*), cholecystokinin (*CCK*), leptin (*LEP*), leptin receptor (*LEPR*), Membrane Bound O-Acyltransferase Domain Containing 4 (*MBOAT4*), melanocortin 4 receptor (*MCR4*), neuropeptide Y (*NPY*), and proopiomelanocortin (POMC) ghrelin/obestatin prepropeptide (*GHRL*), are neuropeptides involved in the regulation of appetite and satiety. Ghrelin/obestatin prepropeptide (*GHRL*) is involved in energy homeostasis and regulation of body weight through its influence on satiety. Polymorphisms in *GHRL* have been linked to breast cancer risk as well as to obesity and insulin levels (3). *GHRL* Membrane Bound O-Acyltransferase Domain Containing 4 (*MBOAT4*) codes the ghrelin O-acyltransferase (GOAT) enzyme that acrylates ghrelin to enable its endocrine actions(4).

While studies have examined the relationship between EH genes with breast cancer risk, there is rationale for their involvement in survival after diagnosis with breast cancer. Variants in *LEP* and *LEPR* have been associated with breast cancer-specific mortality (5). Given the role of EH genes in maintaining body weight, serum levels of adiponectin have been associated with insulin resistance and differences in adipokines such as adiponectin levels have been associated with survival (6). *LEP*, *NPY*, and *GHRL* levels have been shown to regulate growth hormone secretion and promote cell growth (7-11). *LEP* has been shown to have angiogenesis properties and stimulate growth of human breast cancer cells (5, 12, 13). Several neuropeptides have been hypothesized as playing a role in cachexia, or extreme weight loss or wasting after cancer diagnosis (14). Cachexia is associated with decreased survival.

In this study we examine the relationship between ten EH genes and all-cause and breast cancer-specific mortality. These genes were selected because of their association with energy homeostasis and cancer and/or obesity. We evaluate associations by genetic ancestry, given differences in risk associated with these genes by Native American (NA) ancestry (15). Additionally, NA ancestry has been shown to be an important determinate of breast cancer risk among population of mixed Caucasian and NA ancestry, with women with greater NA ancestry having lower incidence of breast cancer than women of European ancestry (16, 17) We evaluate the modifying effects of body mass index (BMI) on survival given the relationship between these genes and BMI and breast cancer (15, 18-20).

Methods

This analysis from the Breast Cancer Health Disparities Study includes participants with information on survival from two population-based case-control studies, the 4-Corners Breast Cancer Study (4-CBCS) and the San Francisco Bay Area Breast Cancer Study (SFBCS) (17). In the 4-CBCS, participants were between 25 and 79 years of age with a histologically confirmed diagnosis of first primary invasive breast cancer (n=1391) between October 1999 and May 2004 (21) and lived in one of the four 4-Corners' states of Arizona, Colorado, New Mexico, or Utah. The SFBCS included women aged 35 to 79 years from the San Francisco Bay Area diagnosed with a first primary histologically confirmed invasive breast cancer (n= 946) between April 1997 and April 2002 (22, 23). All participants provided informed written consent prior to participation. This study was approved by the Institutional Review Boards for Human Subjects at the University of Utah and the Cancer Prevention Institute of California.

Data Harmonization

Data were harmonized across study-specific questionnaires (17). Women were considered post-menopausal if they reported either a natural menopause or if they reported taking hormone therapy (HT) and were still having periods or were at or above the 95th percentile of age for those who reported having a natural menopause (i.e., 12 months since their last period); others were classified as pre-menopausal. Women who reported having a hysterectomy were considered post-menopausal. BMI (kg/m²) was calculated based on self-reported weight during the reference year or weight measured at interview (controls only) if weight during the reference year was not available. Height was based on measured height at interview or self-reported height if the measurement was declined. Categories of BMI were normal BMI (<25.0 kg/m²), overweight (25.0-29.9 kg/m²), or obese (≥ 30 kg/m²). Parity was defined as the number of total pregnancies.

Genetic Data

DNA was extracted from either whole blood or mouthwash samples. Genotyping was completed for 933 women from the 4-CBCS who self-identified as non-Hispanic white (NHW), 412 Hispanic, 8 NA, 14 NHW/Hispanic, 10 NHW/NA, 10 Hispanic/NA, and 4 NHW/Hispanic/NA and for 252 women from the SFBCS who self-reported being NHW and 694 who reported being Hispanic. Women who self-identified as Hispanic and/or NA were considered Hispanic/NA for the analysis. A tagSNP approach was used to characterize variation across candidate genes. TagSNPs were selected using the following parameters: linkage disequilibrium (LD) blocks were defined using a Caucasian LD map and an $r^2=0.8$ based on hapmap data; minor allele frequency (MAF) >0.1; range of -1500 bps from the initiation codon to +1500 bps from the termination codon; and one SNP/LD bin. Coding and non-coding SNPs were included as were both the 5'UTR and 3'UTR areas. We used 104 Ancestry Informative Markers (AIMs) to distinguish European and NA ancestry in the study population (17). All markers were genotyped using a multiplexed bead array assay format based on GoldenGate chemistry (Illumina, San Diego, California). In the current analysis we evaluated tagSNPs for *ADIPOQ* (12 SNPs), *CARTPT* (5 SNPs), *CCK* (4 SNPs), *GHRL* (8 SNPs), *LEP* (9 SNPs), *LEPR* (27 SNPs), *MBOAT4* (1 SNP), *MC4R* (3 SNPs), *NPY* (4

SNPs), and *POMC* (5 SNPs). These genes and SNPs are described in online Supplement Table 1.

Tumor Characteristics and Survival

Data on survival were available from local cancer registries through December of 2013 and included date of death or last follow-up (month and year), underlying cause of death, and SEER summary stage of disease at time of diagnosis. Disease stage was obtained from tumor registries and was coded based on complete pathological reports that included extent of disease, node involvement, and metastasis. Survival (in months) was calculated as the difference between diagnosis date and date of death or last follow-up.

Statistical Methods

The program STRUCTURE was used to compute individual ancestry for each study participant assuming two founding populations (24, 25). A three-founding population model was assessed, but did not fit the population structure with the same level of repeatability and correlation among runs as the two-founding population model. Participants were classified by level of percent NA genetic ancestry. Women who self-reported as being NHW had a low percentage of NA ancestry. Assessment across categories of ancestry was done using cut-points based on the distribution of genetic ancestry in the control population with the goal of creating distinct ancestry groups that had sufficient power to assess associations. Two strata of $\leq 28\%$ and $>28\%$ of NA ancestry were used to evaluate associations.

Associations between SNPs and all-cause and breast cancer-specific mortality were evaluated using Cox proportional hazards models to obtain multivariate hazard ratios (HR) and 95% confidence intervals (CI) for all women and within strata of NA genetic ancestry using SAS version 9.4 (SAS Institute, Cary, NC). Individuals were censored when they were lost to follow-up or if they died of causes other than breast cancer when examining breast cancer-specific mortality. All SNPs were evaluated as a co-dominant model, and if initial analysis suggested too few homozygote variants or the dominant model appeared to fit the data then a dominant model was used. In other instances where a recessive model appeared to fit the data, it was used to evaluate HR estimates. Models were adjusted for age (five-year age categories), study center, BMI (normal, overweight, obese), percent NA ancestry (continuous), parity (categorical), and stage (local, regional, distant).

A major focus of the analysis is the use of the adaptive rank truncated product (ARTP) method that utilizes a highly efficient permutation algorithm to determine the significance of each gene and of the overall pathway with survival (26, 27). This enables us to focus on the significance of the gene and then if genes appear to be significant, we evaluate SNPs that contribute to the gene importance. Using ARTP, we permuted the survival 10,000 times in R version 3.1.3 (R Foundation for Statistical Computing, Vienna, Austria). SNP associations were assessed among the observed and permuted data in R using p values from likelihood-ratio tests comparing fully adjusted Cox proportional hazards models to reduced models excluding the SNP term. The P_{ARTP} is based on assessment of a maximum of five truncation points for each gene and for the pathway. Results included in tables are based on statistically

significant genes from ARTP analysis (p of 0.05 or less) and statistically significant SNPs (p 0.05 or less) that contributed to significant gene p values.

Tests for interaction by ancestry and BMI were calculated using a Wald one degree of freedom (1-df) test; adjustments for multiple comparisons within the gene used the step-down Bonferroni correction, taking into account the correlated nature of the data using the SNP spectral decomposition method proposed by Nyholt (28) and modified by Li and Ji (29).

Results

Approximately the same percentage of women who had low NA ancestry or high NA ancestry died during follow-up (Table 1). Breast cancer was the most common cause of death among women with low NA ancestry (48.9%) and women with high NA ancestry (55.3%) women.

The age, study, menopausal status, and SEER summary stage adjusted HR for all-cause mortality for low NA ancestry versus those with high NA ancestry was 1.13 (95% CI 0.93, 1.38). Further evaluation of all-cause mortality (Table 2) showed that the overall energy homeostasis pathway evaluated was marginally significant among women with low NA ancestry ($P_{\text{ARTP}} = 0.057$). Within the pathway, *GHRL* and *LEPR* were significant ($P_{\text{ARTP}} = 0.035$ and 0.007 respectively). Two SNPs were significantly associated with *GHRL* and 11 SNPs (three in high LD in our data) were associated with *LEPR* in at least one ancestry group. Although no SNP associations were significantly different by NA ancestry after adjustment for multiple comparisons, *GHRL* rs27647 and *LEPR* rs970468, rs10749754, rs1137101, and rs6588147 were significantly different between ancestry groups prior to adjustment for multiple comparisons. Associations did not differ by level of BMI during referent year (data not shown).

Breast cancer-specific mortality HR adjusted for age, study, menopausal status and SEER summary stage for lower NA ancestry relative to higher NA ancestry was 1.15 (95% CI 0.87, 1.52). Although findings associated with energy homeostasis genes were similar for breast cancer-specific mortality as was noted for all-cause mortality, associations were slightly stronger and involved more genes in the pathway (Table 3). The overall pathway was significantly associated with breast cancer-specific mortality among women with low NA ancestry. Three genes, *CCK*, *GHRL*, and *LEPR* were significantly associated with breast cancer-specific mortality among women with low NA ancestry ($P_{\text{ARTP}} = 0.046$, 0.015 , and 0.046 respectively) and *NPY* was significantly associated with breast cancer-specific mortality among women with higher NA ancestry ($P_{\text{ARTP}} = 0.038$). SNPs within these genes that were associated with increased likelihood of dying comparing the rare to more common homozygote variants (*CCK* rs747455, *GHRL* rs35683, rs35682, and rs27647, *LEPR* rs970468, rsrs11585329, rs6588147, and *NPY* rs16129) and better survival with similar comparisons (*LEPR* rs7526141, rs17412175, rs6704167). Associations did not differ by level of BMI during the referent year (data not shown).

Discussion

Our study provides support for an association between EH genes and survival after diagnosis with breast cancer. *GHRL* and *LEPR* appeared to have the greatest influence for both all-cause mortality and breast cancer-specific mortality, with the strongest associations among women with low NA ancestry. *CCK* influenced breast cancer-specific mortality among women with low NA ancestry while *NPY* influenced breast cancer-specific mortality among women with higher NA ancestry. BMI did not appear to modify these associations.

Multiple *GHRL* SNPs showed significant associations with breast cancer-specific mortality. GHRL is a pleiotropic hormone predominately produced in the stomach, and is an endogenous ligand for the Growth Hormone Secretagogue Receptor (GHSR) with two major functions: the stimulation of growth hormone (GH) production and the stimulation of food intake (3). GHRL stimulates the production of GH through the activation of GHSR-1a in the hypothalamus and increases appetite and food intake independent of GHSR. In addition to its orexigenic function, GHRL also functions in cell proliferation; this function, in conjunction with the stimulatory effect on GH secretion from the anterior pituitary renders GHRL a potential factor of tumorigenesis (8). Nonetheless, little evidence has thus far been produced to show an association between *GHRL* polymorphisms and survival in breast cancer patients. However, *GHRL* polymorphisms have been associated with obesity (3) and recent studies have associated obesity with decreased survival in breast cancer patients (30, 31), particularly in Hispanics with morbid obesity (18) but these results are far from conclusive and are contradicted by other studies (32).

GHRL also plays an important role in the maintenance of the GH-IGF1 axis (3). Thus *GHRL* polymorphisms could alter hepatic IGF-1 expression levels and *IGF1* polymorphisms and expression have been associated with breast cancer survival (33, 34). Moreover, in prostate cancer GHRL is highly expressed and has been shown to initiate cross-talk to MAPK signaling cascades, playing an important role in cell proliferation via the activation of the ERK1/2 MAPK pathway, but also through an alternative p38 (MAPK14) pathway (35). While this cross-talk has yet to be shown in breast cancer tissues, both of these MAPK pathways have been associated with breast cancer survival in individuals of lower NA ancestry (36), providing a possible explanation for our observed association between *GHRL* and breast cancer survival. GHRL has also been shown to play a role in the pathophysiology of cachexia, with cachectic patients having higher GHRL concentrations, while cachexia is associated with decreased survival (14). However, other studies have shown that elevated GHRL expression is associated with increased survival in non-cachectic patients (13). Unfortunately we do not have information about how our associated SNPs affect GHRL expression.

LEPR SNPs also showed an association with breast cancer survival. LEPR is a cytokine receptor that is highly expressed in multiple tumors, including breast cancer, and in breast cancer LEPR expression is directly correlated with poor prognosis (37). LEP is predominately secreted by adipose tissue, and functions as an anorexigenic hormone responsible for appetite suppression and maintenance of EH. This control of EH is mediated via LEP induced proteolytic processing of NPY and POMC in the Arcuate nucleus of the

hypothalamus and the subsequent liberation of α -MSH; LEP also negatively regulates the orexigenic hormones NPY and Agouti Related Peptide (AgRP) (38). LEP binding to LEPR initiates multiple signal cascades to mediate its orexigenic effect, including JAK2/STAT3, phosphoinositide 3 kinase (PI3K)/Akt, and ERK MAPK, which can also promote the proliferation and survival of cancer cells (37-39). LEP signaling can also mediate anti-apoptotic effects through the overregulation of bcl-2 and expression of *survivin* and *hey2*, alter microenvironment to favor growth and progression through increases in MMP2 and E-cadherine, and promote angiogenesis through VEGF and VEGFR2 (38).

Therefore LEP can act as a mitogenic, motogenic, prognostic, and angiogenic factor. LEP has also been associated with decreased survival in breast cancer patients (37, 39, 40) and with cachexia (14). While we did not duplicate these findings, we showed an association between multiple *LEPR* SNPs and breast cancer survival. Activation of LEPR leads to downstream signaling via the ERK1/2 MAPK pathway which has been associated with decreased breast cancer survival (36, 41, 42). We have previously reported that this pathway is associated with breast cancer survival in patients of low NA ancestry (36), which correlates with our findings.

CCK was associated with breast cancer-specific mortality in women of low NA ancestry. *CCK* is important in the control of food intake, reducing food intake and promoting satiety (43). However, *CCK* activation of *CCK* Receptor A (*CCKAR*) and *CCK* Receptor B (*CCKBR*) induces the chemotaxis of monocytes (44). Chronic low grade inflammation, as represented by an increased C-reactive protein (CRP), has been negatively associated with breast cancer survival (6). Moreover, *CCK* has been shown to function as an insulin secretagogue and islet derived *CCK* may act locally to prevent β cell apoptosis (45), and hyperinsulinemia is an independent risk factor for poor prognosis in women with breast cancer(6). It has been hypothesized that women with higher NA ancestry develop type 2 diabetes mellitus at a younger age and over time become hypoinsulinemic (21). If this hypothesis is correct it could explain the lower impact of *CCK* SNPs on survival in women of higher NA ancestry.

NPY rs16129 was significantly associated with survival in women of higher NA ancestry. *NPY* is a 36 amino acid peptide released by sympathetic nerves and is a potent trophic factor (46). In the nervous system *NPY* is a neurotransmitter playing a role in cognitive function, feeding behavior and cardiovascular regulation (47). *NPY* has been shown to increase the proliferation and migration of breast cancer cells, angiogenesis, and function in a paracrine manner to stimulate the release of cytokines, such as IL-6, IL-8, TNF- α and VEGF (46, 47). *NPY* Y1R, Y2R and Y5R have been reported in breast cancer lines and breast carcinomas are reported to have a high density of *NPY* receptors; Y5R activation stimulates growth through increased MAPK activity (7, 47, 48). This in turn leads to increased ERK 1/2 phosphorylation. Moreover, chronic stress, which is associated with breast cancer risk, leads to elevated sympathetic neurotransmitter release and sympathetics arising from the lateral and anterior cutaneous branches of the second through the sixth intercostal nerves ensure a constant supply of *NPY* ligands to the breast microenvironment (47). This coupled with the high density of *NPY* receptors may lead to a hyperactivation of the ERK1/2 MAPK system, which is associated with adverse clinical features and poor prognosis (42, 49). Poor clinical

outcome is in part due to the fact that ERK 1/2 MAPK signaling can prime estrogen receptor (ER) signaling, so that overstimulation of the system may drive ER signaling and hence tumor growth independent of an estrogen ligand (42); these hormone refractory breast cancers respond poorly to hormone ablation therapies.

Our findings show that *NPY* SNPs are associated with breast cancer-specific mortality in women of high NA ancestry. One possible explanation for this association is that chronic stress leads to elevated NPY release and that NPY is a potent chemoattractant for monocytes when acting through Y2R and Y5R (47). Moreover, NPY acts in a paracrine fashion to stimulate the release of TNF- α . We have previously shown that women of higher NA ancestry show a greater protective effect with higher intake of dietary antioxidants (50) and that *TNF* SNPs are more strongly associated with breast cancer risk and survival in women of higher NA ancestry (51).

The study has both strengths and limitations. First, we used a tagSNP approach to gather information on the genetic variation across the gene. Our tagSNP approach was implemented on a customized Illumina platform and included SNPs that were validated and considered to have a high probability of yielding results. Our tagSNPs were identified using the Illumina data and were based mainly on Caucasian populations. While this approach allowed us to evaluate genetic variation across the gene, we may have missed important SNPs and therefore important associations. Additionally, we are limited in our knowledge of the functionality of these SNPs, which makes it difficult to determine how SNPs operate in influencing gene expression or protein levels. We were able to examine associations by NA ancestry as well as by BMI, important factors that could modify risk associated with survival. We utilized a two ancestry population since this structure best fit our data. While we had disease stage data we did not have information on treatment. We used both ARTP and Benjamin and Hochberg adjustments for multiple comparisons to identify genes and SNPs of importance for survival. However, findings could still be from chance and need replication in other ethnically diverse populations.

In summary, our results support our hypothesis that EH genes influence survival after diagnosis with breast cancer. *GRHL* and the *LEPR* appear to have the most influence on survival. *CCK* and *NPY* were associated with breast cancer-specific mortality only, while *LEPR* and *GHRL* showed associations with both all-cause and breast cancer specific mortality. The greatest influence of EH genes on survival was found among women with low NA ancestry (i.e. mostly European ancestry), although *NPY* influenced breast cancer-specific survival among women with high NA ancestry only. Body size did not appear to influence these associations with survival. Confirmation of these findings in a similar ethnically diverse population is needed.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

We would also like to acknowledge the contributions of the following individuals to the study: Sandra Edwards and Jennifer Herrick for data harmonization and management; Erica Wolff and Michael Hoffman for laboratory support; Jocelyn Koo for data management for the San Francisco Bay Area Breast Cancer Study; Dr. Tim Byer, Dr. Kathy Baumgartner, and Dr. Anna Giuliano for their contribution to the 4-Corners Breast Cancer Study; and Dr. Josh Galanter for assistance in selection of AIMs markers.

Funding: The Breast Cancer Health Disparities Study was funded by grant CA14002 from the National Cancer Institute to Dr. Slattery. The San Francisco Bay Area Breast Cancer Study was supported by grants CA63446 and CA77305 from the National Cancer Institute, grant DAMD17-96-1-6071 from the U.S. Department of Defense and grant 7PB-0068 from the California Breast Cancer Research Program. The collection of cancer incidence data used in this study was supported by the California Department of Public Health as part of the statewide cancer reporting program mandated by California Health and Safety Code Section 103885; the National Cancer Institute's Surveillance, Epidemiology and End Results Program under contract HHSN261201000036C awarded to the Cancer Prevention Institute of California; and the Centers for Disease Control and Prevention's National Program of Cancer Registries, under agreement #1U58 DP000807-01 awarded to the Public Health Institute. The 4-Corners Breast Cancer Study was funded by grants CA078682, CA078762, CA078552, and CA078802 from the National Cancer Institute. The research also was supported by the Utah Cancer Registry, which is funded by contract N01-PC-67000 from the National Cancer Institute, with additional support from the State of Utah Department of Health, the New Mexico Tumor Registry, and the Arizona and Colorado cancer registries, funded by the Centers for Disease Control and Prevention National Program of Cancer Registries and additional state support. The contents of this manuscript are solely the responsibility of the authors and do not necessarily represent the official view of the National Cancer Institute or endorsement by the State of California Department of Public Health, the National Cancer Institute, and the Centers for Disease Control and Prevention or their Contractors and Subcontractors

References

1. Fusco R, Galgani M, Procaccini C, Franco R, Pirozzi G, Fucci L, et al. Cellular and molecular crosstalk between leptin receptor and estrogen receptor- α in breast cancer: molecular basis for a novel therapeutic setting. *Endocr Relat Cancer*. 2010; 17(2):373–82. [PubMed: 20410173]
2. Llanos AA, Dumitrescu RG, Marian C, Makambi KH, Spear SL, Kallakury BV, et al. Adipokines in plasma and breast tissues: associations with breast cancer risk factors. *Cancer Epidemiol Biomarkers Prev*. 2012; 21(10):1745–55. [PubMed: 22892282]
3. Dossus L, McKay JD, Canzian F, Wilkening S, Rinaldi S, Biessy C, et al. Polymorphisms of genes coding for ghrelin and its receptor in relation to anthropometry, circulating levels of IGF-I and IGFBP-3, and breast cancer risk: a case-control study nested within the European Prospective Investigation into Cancer and Nutrition (EPIC). *Carcinogenesis*. 2008; 29(7):1360–6. [PubMed: 18375957]
4. Lim CT, Kola B, Grossman A, Korbonits M. The expression of ghrelin O-acyltransferase (GOAT) in human tissues. *Endocr J*. 2011; 58(8):707–10. [PubMed: 21646729]
5. Cleveland RJ, Gammon MD, Long CM, Gaudet MM, Eng SM, Teitelbaum SL, et al. Common genetic variations in the LEP and LEPR genes, obesity and breast cancer incidence and survival. *Breast Cancer Res Treat*. 2010; 120(3):745–52. [PubMed: 19697123]
6. Duggan C, Irwin ML, Xiao L, Henderson KD, Smith AW, Baumgartner RN, et al. Associations of insulin resistance and adiponectin with mortality in women with breast cancer. *Journal of clinical oncology: official journal of the American Society of Clinical Oncology*. 2011; 29(1):32–9. [PubMed: 21115858]
7. Sheriff S, Ali M, Yahya A, Haider KH, Balasubramaniam A, Amlal H. Neuropeptide Y Y5 receptor promotes cell growth through extracellular signal-regulated kinase signaling and cyclic AMP inhibition in a human breast cancer cell line. *Mol Cancer Res*. 2010; 8(4):604–14. [PubMed: 20332211]
8. Stefanaki C, Rorris Filippoa-Paschalis, Stamatakos M. The role of ghrelin signals in breast cancer -- A systematic review. *Current Signal Transduction Therapy*. 2012; 7:2470253.
9. Wu JT, Kral JG. Ghrelin: integrative neuroendocrine peptide in health and disease. *Ann Surg*. 2004; 239(4):464–74. [PubMed: 15024307]
10. Dutta D, Ghosh S, Pandit K, Mukhopadhyay P, Chowdhury S. Leptin and cancer: Pathogenesis and modulation. *Indian J Endocrinol Metab*. 2012; 16(Suppl 3):S596–600. [PubMed: 23565495]

11. Hardwick JC, Van Den Brink GR, Offerhaus GJ, Van Deventer SJ, Peppelenbosch MP. Leptin is a growth factor for colonic epithelial cells. *Gastroenterology*. 2001; 121(1):79–90. [PubMed: 11438496]
12. Rose DP, Gilhooly EM, Nixon DW. Adverse effects of obesity on breast cancer prognosis, and the biological actions of leptin (review). *Int J Oncol*. 2002; 21(6):1285–92. [PubMed: 12429979]
13. Gronberg M, Fjallskog ML, Jirstrom K, Janson ET. Expression of ghrelin is correlated to a favorable outcome in invasive breast cancer. *Acta Oncol*. 2012; 51(3):386–93. [PubMed: 22067021]
14. Mondello P, Lacquaniti A, Mondello S, Bolignano D, Pitini V, Aloisi C, et al. Emerging markers of cachexia predict survival in cancer patients. *BMC Cancer*. 2014; 14:828. [PubMed: 25400234]
15. Slattery ML, Lundgreen A, Hines L, Wolff RK, Torres-Mejia G, Baumgartner KN, John EM. Energy homeostasis genes and breast cancer risk: the influence of ancestry, body size, and menopause status, Thre Breast Cancer Health Disparities Study. *Cancer Epidemiology*. 2015
16. Slattery ML, Lundgreen A, Stern MC, Hines L, Wolff RK, Giuliano AR, et al. The influence of genetic ancestry and ethnicity on breast cancer survival associated with genetic variation in the TGF-beta-signaling pathway: The Breast Cancer Health Disparities Study. *Cancer Causes Control*. 2014; 25(3):293–307. [PubMed: 24337772]
17. Slattery ML, John EM, Torres-Mejia G, Lundgreen A, Herrick JS, Baumgartner KB, et al. Genetic variation in genes involved in hormones, inflammation and energetic factors and breast cancer risk in an admixed population. *Carcinogenesis*. 2012; 33(8):1512–21. [PubMed: 22562547]
18. Kwan ML, John EM, Caan BJ, Lee VS, Bernstein L, Cheng I, et al. Obesity and mortality after breast cancer by race/ethnicity: The California Breast Cancer Survivorship Consortium. *Am J Epidemiol*. 2014; 179(1):95–111. [PubMed: 24107615]
19. John EM, Sangaramoorthy M, Hines LM, Stern MC, Baumgartner KB, Giuliano AR, et al. Body size throughout adult life influences postmenopausal breast cancer risk among hispanic women: the breast cancer health disparities study. *Cancer Epidemiol Biomarkers Prev*. 2015; 24(1):128–37. [PubMed: 25352523]
20. John EM, Sangaramoorthy M, Hines LM, Stern MC, Baumgartner KB, Giuliano AR, et al. Overall and Abdominal Adiposity and Premenopausal Breast Cancer Risk among Hispanic Women: The Breast Cancer Health Disparities Study. *Cancer Epidemiol Biomarkers Prev*. 2015; 24(1):138–47. [PubMed: 25352526]
21. Slattery ML, Sweeney C, Edwards S, Herrick J, Baumgartner K, Wolff R, et al. Body size, weight change, fat distribution and breast cancer risk in Hispanic and non-Hispanic white women. *Breast Cancer Res Treat*. 2007; 102(1):85–101. [PubMed: 17080310]
22. John EM, Horn-Ross PL, Koo J. Lifetime physical activity and breast cancer risk in a multiethnic population: the San Francisco Bay area breast cancer study. *Cancer Epidemiol Biomarkers Prev*. 2003; 12(11 Pt 1):1143–52. [PubMed: 14652273]
23. John EM, Phipps AI, Davis A, Koo J. Migration history, acculturation, and breast cancer risk in Hispanic women. *Cancer Epidemiol Biomarkers Prev*. 2005; 14(12):2905–13. [PubMed: 16365008]
24. Falush D, Stephens M, Pritchard JK. Inference of population structure using multilocus genotype data: linked loci and correlated allele frequencies. *Genetics*. 2003; 164(4):1567–87. [PubMed: 12930761]
25. Pritchard JK, Stephens M, Donnelly P. Inference of population structure using multilocus genotype data. *Genetics*. 2000; 155(2):945–59. [PubMed: 10835412]
26. Yu K, Li Q, Bergen AW, Pfeiffer RM, Rosenberg PS, Caporaso N, et al. Pathway analysis by adaptive combination of P-values. *Genetic epidemiology*. 2009; 33(8):700–9. [PubMed: 19333968]
27. Kai Yu, OL.; William, Wheeler. ARTP Gene Pathway p-values computed using the Adaptive Rank Truncated Product. 2.0.0. R package; 2011.
28. Nyholt DR. A simple correction for multiple testing for single-nucleotide polymorphisms in linkage disequilibrium with each other. *American journal of human genetics*. 2004; 74(4):765–9. [PubMed: 14997420]

29. Li J, Ji L. Adjusting multiple testing in multilocus analyses using the eigenvalues of a correlation matrix. *Heredity*. 2005; 95(3):221–7. [PubMed: 16077740]
30. Scholz C, Andergassen U, Hepp P, Schindlbeck C, Friedl TW, Harbeck N, et al. Obesity as an independent risk factor for decreased survival in node-positive high-risk breast cancer. *Breast cancer research and treatment*. 2015; 151(3):569–76. [PubMed: 25962694]
31. Taghizadeh N, Boezen HM, Schouten JP, Schroder CP, Vries EG, Vonk JM. BMI and Lifetime Changes in BMI and Cancer Mortality Risk. *PLoS One*. 2015; 10(4):e0125261. [PubMed: 25881129]
32. Herlevic VC, Mowad R, Miller JK, Darensburg NA, Li BD, Kim RH. Breast cancer outcomes in a population with high prevalence of obesity. *The Journal of surgical research*. 2015
33. Shin A, Ren Z, Shu XO, Cai Q, Gao YT, Zheng W. Expression patterns of insulin-like growth factor 1 (IGF-I) and its receptor in mammary tissues and their associations with breast cancer survival. *Breast cancer research and treatment*. 2007; 105(1):55–61. [PubMed: 17066319]
34. Muendlein A, Lang AH, Geller-Rhomberg S, Winder T, Gasser K, Drexel H, et al. Association of a common genetic variant of the IGF-1 gene with event-free survival in patients with HER2-positive breast cancer. *J Cancer Res Clin Oncol*. 2013; 139(3):491–8. [PubMed: 23180020]
35. Yeh AH, Jeffery PL, Duncan RP, Herington AC, Chopin LK. Ghrelin and a novel preproghrelin isoform are highly expressed in prostate cancer and ghrelin activates mitogen-activated protein kinase in prostate cancer. *Clinical cancer research: an official journal of the American Association for Cancer Research*. 2005; 11(23):8295–303. [PubMed: 16322288]
36. Slattery ML, Hines LH, Lundgreen A, Baumgartner KB, Wolff RK, Stern MC, et al. Diet and lifestyle factors interact with MAPK genes to influence survival: the Breast Cancer Health Disparities Study. *Cancer causes & control: CCC*. 2014; 25(9):1211–25. [PubMed: 24993294]
37. Zheng Q, Banaszak L, Fracci S, Basali D, Dunlap SM, Hursting SD, et al. Leptin receptor maintains cancer stem-like properties in triple negative breast cancer cells. *Endocrine-related cancer*. 2013; 20(6):797–808. [PubMed: 24025407]
38. Garcia-Robles MJ, Segura-Ortega JE, Fafutis-Morris M. The biology of leptin and its implications in breast cancer: a general view. *Journal of interferon & cytokine research: the official journal of the International Society for Interferon and Cytokine Research*. 2013; 33(12):717–27.
39. Terrasi M, Bazan V, Caruso S, Insalaco L, Amodeo V, Fanale D, et al. Effects of PPARgamma agonists on the expression of leptin and vascular endothelial growth factor in breast cancer cells. *Journal of cellular physiology*. 2013; 228(6):1368–74. [PubMed: 23254958]
40. Snoussi K, Strosberg AD, Bouaouina N, Ben Ahmed S, Helal AN, Chouchane L. Leptin and leptin receptor polymorphisms are associated with increased risk and poor prognosis of breast carcinoma. *BMC Cancer*. 2006; 6:38. [PubMed: 16504019]
41. Miller PC, Clarke J, Koru-Sengul T, Brinkman J, El-Ashry D. A novel MAPK-microRNA signature is predictive of hormone-therapy resistance and poor outcome in ER-positive breast cancer. *Clinical cancer research: an official journal of the American Association for Cancer Research*. 2015; 21(2):373–85. [PubMed: 25370469]
42. Gee JM, Robertson JF, Ellis IO, Nicholson RI. Phosphorylation of ERK1/2 mitogen-activated protein kinase is associated with poor response to anti-hormonal therapy and decreased patient survival in clinical breast cancer. *International journal of cancer Journal international du cancer*. 2001; 95(4):247–54. [PubMed: 11400118]
43. Ribeiro JA, Serquiz AC, Silva PF, Barbosa PB, Sampaio TB, Araujo Junior RF, et al. Trypsin inhibitor from tamarindus indica L. seeds reduces weight gain and food consumption and increases plasmatic cholecystokinin levels. *Clinics*. 2015; 70(2):136–43. [PubMed: 25789523]
44. Pivovarova O, Hornemann S, Weimer S, Lu Y, Murahovschi V, Zhuk S, et al. Regulation of nutrition-associated receptors in blood monocytes of normal weight and obese humans. *Peptides*. 2015; 65:12–9. [PubMed: 25620618]
45. Linnemann AK, Neuman JC, Battiola TJ, Wisinski JA, Kimple ME, Davis DB. Glucagon-like peptide-1 Regulates Cholecystokinin Production in beta-cells to Protect from Apoptosis. *Molecular endocrinology*. 2015 me20151030.

46. Medeiros PJ, Jackson DN. Neuropeptide Y Y5-receptor activation on breast cancer cells acts as a paracrine system that stimulates VEGF expression and secretion to promote angiogenesis. *Peptides*. 2013; 48:106–13. [PubMed: 23932937]
47. Medeiros PJ, Al-Khazraji BK, Novielli NM, Postovit LM, Chambers AF, Jackson DN. Neuropeptide Y stimulates proliferation and migration in the 4T1 breast cancer cell line. *International journal of cancer Journal international du cancer*. 2012; 131(2):276–86. [PubMed: 21823118]
48. Korner M, Reubi JC. NPY receptors in human cancer: a review of current knowledge. *Peptides*. 2007; 28(2):419–25. [PubMed: 17223228]
49. Lonne GK, Masoumi KC, Lennartsson J, Larsson C. Protein kinase Cdelta supports survival of MDA-MB-231 breast cancer cells by suppressing the ERK1/2 pathway. *J Biol Chem*. 2009; 284(48):33456–65. [PubMed: 19833733]
50. Slattery ML, John EM, Torres-Mejia G, Lundgreen A, Lewinger JP, Stern MC, et al. Angiogenesis genes, dietary oxidative balance and breast cancer risk and progression: the Breast Cancer Health Disparities Study. *International journal of cancer Journal international du cancer*. 2014; 134(3): 629–44. [PubMed: 23832257]
51. Slattery ML, Lundgreen A, Torres-Mejia G, Wolff RK, Hines L, Baumgartner K, et al. Diet and lifestyle factors modify immune/inflammation response genes to alter breast cancer risk and prognosis: the Breast Cancer Health Disparities Study. *Mutation research*. 2014; 770:19–28. [PubMed: 25332681]

Table 1
Description of Study Population by Native American Ancestry

	0-28% Native American Ancestry		29-100% Native American Ancestry	
	N	%	N	%
Study Site				
4-Corners Breast Cancer Study	999	71.31	391	41.60
San Francisco Bay Area Breast Cancer Study	402	28.69	549	58.40
Age (years)				
24-39	98	7.00	80	8.51
40-49	384	27.41	317	33.72
50-59	395	28.19	258	27.45
60-69	338	24.13	194	20.64
>70	186	13.28	91	9.68
Menopausal Status				
Pre-menopausal	469	34.41	367	41.01
Post-menopausal	894	65.59	528	58.99
Self-Reported Race/Ethnicity				
non-Hispanic White	1177	84.01	9	0.96
Hispanic/Native American	224	15.99	931	99.04
Vital Status				
Deceased	297	21.20	186	19.79
Alive	1104	78.80	754	80.21
Cause of Death				
Breast Cancer	145	48.82	104	55.91
Other	152	51.2	82	44.1
SEER Summary Stage				
Local	946	69.10	532	59.91
Regional	407	29.73	348	39.19
Distant	16	1.17	8	0.90

Table 2
Associations between energy homeostasis genes and deaths from any cause by level of Native American ancestry

Pathway P _{ARTP}	Overall			0 - 28% Native American Ancestry			29 - 100% Native American Ancestry			P _{Interaction} (raw, adjusted)
	Death/Person Years	HR	(95% CI)	Death/Person Years	HRR	(95% CI)	Death/Person Years	HR	(95% CI)	
GHRL P_{ARTP}		0.160		0.057			0.713			
rs27647										0.024, 0.122
	TT	213 / 10766	1.00	94 / 5247	1.00		119 / 5519	1.00		
	TC	205 / 10410	1.03 (0.85, 1.26)	148 / 6850	1.24 (0.95, 1.61)		57 / 3560	0.80 (0.58, 1.10)		
	CC	64 / 2492	1.37 (1.03, 1.83)	55 / 2035	1.60 (1.14, 2.24)		9 / 457	0.92 (0.46, 1.83)		
rs3755777										0.271, 0.542
	GG	232 / 10669	1.00	180 / 7861	1.00		52 / 2808	1.00		
	GC/CC	249 / 13081	0.80 (0.66, 0.96)	117 / 6341	0.73 (0.58, 0.92)		132 / 6741	0.94 (0.67, 1.30)		
LEPR P_{ARTP}		0.120		0.007			0.272			
rs7526141										0.357, 1.000
	CC	180 / 8404	1.00	92 / 3844	1		88 / 4559	1.00		
	CT	232 / 11292	0.96 (0.79, 1.17)	152 / 7438	0.83 (0.64, 1.07)		80 / 3854	1.08 (0.80, 1.47)		
	TT	71 / 4063	0.79 (0.59, 1.04)	53 / 2919	0.72 (0.51, 1.01)		18 / 1144	0.81 (0.49, 1.35)		
rs17412175 ²										0.239, 1.000
	TT	197 / 8819	1.00	107 / 4344	1.00		90 / 4475	1.00		
	TA	224 / 11368	0.89 (0.73, 1.08)	147 / 7338	0.83 (0.64, 1.06)		77 / 4029	0.93 (0.69, 1.27)		
	AA	62 / 3572	0.74 (0.56, 0.99)	43 / 2519	0.64 (0.44, 0.92)		19 / 1053	0.92 (0.56, 1.51)		
rs970468										0.007, 0.119
	TT	159 / 8358	1.00	92 / 5392	1.00		67 / 2966	1.00		
	TG	236 / 11613	1.13 (0.93, 1.39)	155 / 7041	1.40 (1.08, 1.83)		81 / 4572	0.82 (0.59, 1.13)		
	GG	88 / 3787	1.31 (1.01, 1.71)	50 / 1768	1.80 (1.26, 2.55)		38 / 2019	0.89 (0.60, 1.33)		
rs6704167 ²										0.223, 1.000
	AA	211 / 9262	1.00	118 / 4602	1.00		93 / 4660	1.00		
	AT	209 / 11106	0.81 (0.67, 0.98)	134 / 7173	0.73 (0.56, 0.93)		75 / 3933	0.89 (0.65, 1.21)		
	TT	62 / 3377	0.74 (0.55, 0.99)	44 / 2413	0.64 (0.45, 0.91)		18 / 964	0.91 (0.55, 1.51)		

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

	Overall			0 - 28% Native American Ancestry			29 - 100% Native American Ancestry			$P_{Interaction}$ (raw, adjusted)
	Death/Person Years	HR	(95% CI)	Death/Person Years	HRR	(95% CI)	Death/Person Years	HR	(95% CI)	
rs1171271 ³										0.065, 0.803
	TT	235 / 12400	1.00	141 / 7700	1.00	1.00	94 / 4700	1.00		
	TC	207 / 9480	1.21	(1.00, 1.46)	1.31	(1.03, 1.67)	76 / 3798	1.07	(0.79, 1.45)	
	CC	41 / 1878	1.27	(0.91, 1.77)	1.71	(1.12, 2.64)	16 / 1060	0.87	(0.51, 1.48)	
rs1171265										0.173, 1.000
	GG	174 / 9463	1.00	105 / 5854	1.00	1.00	69 / 3609	1.00		
	GA	242 / 11186	1.19	(0.98, 1.44)	1.27	(0.99, 1.62)	89 / 4368	1.07	(0.78, 1.46)	
	AA	67 / 3057	1.34	(1.01, 1.78)	1.60	(1.10, 2.32)	28 / 1579	1.06	(0.68, 1.64)	
rs4370791 ³										0.175, 1.000
	AA	215 / 11463	1.00	133 / 7193	1.00	1.00	82 / 4270	1.00		
	AG	226 / 10080	1.22	(1.01, 1.47)	1.28	(1.01, 1.63)	86 / 4019	1.15	(0.85, 1.56)	
	GG	42 / 2193	1.13	(0.81, 1.58)	1.43	(0.92, 2.22)	18 / 1258	0.86	(0.51, 1.44)	
rs10749754 ⁴										0.007, 0.119
	GG	140 / 7055	1.00	76 / 4368	1.00	1.00	64 / 2687	1.00		
	GA/AA	343 / 16703	1.12	(0.92, 1.37)	1.42	(1.09, 1.85)	122 / 6870	0.80	(0.59, 1.09)	
rs1137101 ⁴										0.004, 0.073
	AA	140 / 6982	1.00	76 / 4357	1.00	1.00	64 / 2625	1.00		
	AG/GG	342 / 16667	1.09	(0.90, 1.34)	1.40	(1.07, 1.82)	121 / 6861	0.77	(0.56, 1.04)	
rs11585329										0.079, 0.891
	GG	335 / 17212	1.00	194 / 10036	1.00	1.00	141 / 7176	1.00		
	GT/TT	148 / 6546	1.14	(0.94, 1.38)	1.30	(1.02, 1.65)	45 / 2381	0.91	(0.65, 1.28)	
rs6588147										0.012, 0.165
	AA	186 / 9769	1.00	116 / 6382	1.00	1.00	70 / 3387	1.00		
	AG	220 / 10672	1.15	(0.94, 1.40)	1.29	(1.01, 1.67)	84 / 4315	0.97	(0.70, 1.33)	
	GG	77 / 3261	1.31	(1.00, 1.72)	1.85	(1.30, 2.62)	32 / 1855	0.88	(0.58, 1.34)	

¹ Hazard Ratios (HR) and 95% Confidence Intervals (CI) are adjusted for adjusted for age, study, BMI during referent year, parity, % NA ancestry, and SEER stage. Table includes of genes with ARTP $p < 0.05$ and SNPs with $p < 0.05$

² High linkage disequilibrium (LD) among women with low NA ancestry ($r^2 = 0.80$)

High LD $r^2=0.84$ among women with low NA ancestry and 0.82 among women with high NA ancestry)
High LD $r^2=0.98$ among women with low NA ancestry and 0.97 among women with high NA ancestry)

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

Table 3
Associations between energy homeostasis genes and breast cancer-specific mortality by level of Native American ancestry

Pathway P _{ARTP}	Death/Person Years	Overall			0 - 28% Native American Ancestry			29 - 100% Native American Ancestry			P _{Interaction} (raw, adjusted)
		HR/	(95% CI)	Death/Person Years	HR	(95% CI)	Death/Person Years	HR	(95% CI)		
CCK P_{ARTP}		0.087		0.038		0.248		0.116		0.949, 1.000	
rs747455		0.005		0.046		0.116		0.116		0.949, 1.000	
	GG	1.00		1.00	1.00	1.00	1.00	1.00	1.00		
	GA/AA	1.55	(1.21, 1.98)	74 / 5676	1.55	(1.12, 2.16)	53 / 4039	1.57	(1.06, 2.32)		
GHL P_{ARTP}		0.144		0.015		0.785		0.785		0.051, 0.212	
rs35683 ²		0.087		0.038		0.248		0.116		0.051, 0.212	
	CC	1.00		1.00	1.00	1.00	1.00	1.00	1.00		
	CA	1.00	(0.76, 1.33)	74 / 7425	1.25	(0.83, 1.89)	44 / 4332	0.87	(0.57, 1.30)		
	AA	1.23	(0.85, 1.78)	37 / 2783	1.65	(1.03, 2.65)	8 / 939	0.73	(0.34, 1.56)		
rs35682 ²		0.087		0.038		0.248		0.116		0.035, 0.212	
	AA	1.00		1.00	1.00	1.00	1.00	1.00	1.00		
	AG	0.96	(0.72, 1.28)	70 / 7271	1.20	(0.79, 1.82)	43 / 4378	0.83	(0.55, 1.26)		
	GG	1.30	(0.90, 1.86)	42 / 3046	1.74	(1.09, 2.75)	9 / 992	0.77	(0.37, 1.60)		
rs27647		0.072		0.046		0.130		0.130		0.170, 0.509	
	TT	1.00		1.00	1.00	1.00	1.00	1.00	1.00		
	TC	1.06	(0.80, 1.40)	66 / 6850	1.15	(0.78, 1.68)	36 / 3560	1.01	(0.66, 1.53)		
	CC	1.80	(1.23, 2.63)	33 / 2035	2.18	(1.38, 3.44)	7 / 457	1.25	(0.56, 2.80)		
LEPR P_{ARTP}		0.072		0.046		0.130		0.130		0.322, 1.000	
rs7526141		0.072		0.046		0.130		0.130		0.322, 1.000	
	CC	1.00		1.00	1.00	1.00	1.00	1.00	1.00		
	CT	0.91	(0.69, 1.19)	71 / 7438	0.74	(0.51, 1.08)	46 / 3854	1.10	(0.73, 1.65)		
	TT	0.68	(0.45, 1.01)	25 / 2919	0.60	(0.37, 0.98)	8 / 1144	0.69	(0.33, 1.47)		
rs17412175 ³		0.072		0.046		0.130		0.130		0.432, 1.000	
	TT	1.00		1.00	1.00	1.00	1.00	1.00	1.00		
	TT	1.00		1.00	1.00	1.00	1.00	1.00	1.00		

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript

	Overall		0 - 28% Native American Ancestry				29 - 100% Native American Ancestry				$P_{Interaction}$ (raw, adjusted)
	Death/Person Years	HR ¹	(95% CI)	Death/Person Years	HR	(95% CI)	Death/Person Years	HR	(95% CI)		
rs970468	TA 113 / 11368	0.84	(0.64, 1.10)	74 / 7338	0.84	(0.59, 1.21)	39 / 4029	0.78	(0.52, 1.18)		
	AA 29 / 3572	0.64	(0.42, 0.97)	19 / 2519	0.52	(0.31, 0.90)	10 / 1053	0.87	(0.44, 1.72)	0.175, 1.000	
	TT 78 / 8358	1.00		42 / 5392	1.00		36 / 2966	1.00			
	TG 123 / 11613	1.23	(0.92, 1.63)	81 / 7041	1.68	(1.14, 2.45)	42 / 4572	0.77	(0.49, 1.21)		
	GG 48 / 3787	1.49	(1.03, 2.15)	22 / 1768	1.77	(1.05, 3.01)	26 / 2019	1.17	(0.70, 1.95)		
rs6704167 ³	AA 115 / 9262	1.00		59 / 4602	1.00		56 / 4660	1.00		0.274, 1.000	
	AT 103 / 11106	0.72	(0.55, 0.94)	66 / 7173	0.67	(0.47, 0.96)	37 / 3933	0.72	(0.47, 1.09)		
	TT 30 / 3377	0.67	(0.45, 1.01)	19 / 2413	0.52	(0.31, 0.88)	11 / 964	1.02	(0.53, 1.95)		
rs1171271	TT 112 / 12400	1.00		65 / 7700	1.00		47 / 4700	1.00		0.418, 1.000	
	TC 113 / 9480	1.32	(1.02, 1.72)	67 / 5682	1.37	(0.97, 1.93)	46 / 3798	1.25	(0.83, 1.89)		
	CC 24 / 1878	1.45	(0.93, 2.26)	13 / 818	1.72	(0.94, 3.16)	11 / 1060	1.17	(0.60, 2.27)		
rs11585329	GG 174 / 17212	1.00		92 / 10036	1.00		82 / 7176	1.00		0.039, 0.681	
	GT/TT 75 / 6546	1.15	(0.88, 1.51)	53 / 4165	1.46	(1.04, 2.05)	22 / 2381	0.79	(0.49, 1.27)		
rs6588147	AA 89 / 9769	1.00		52 / 6382	1.00		37 / 3387	1.00		0.073, 1.000	
	AG 116 / 10672	1.23	(0.93, 1.63)	70 / 6358	1.47	(1.02, 2.12)	46 / 4315	0.95	(0.61, 1.47)		
	GG 44 / 3261	1.54	(1.06, 2.22)	23 / 1406	2.10	(1.27, 3.47)	21 / 1855	1.09	(0.64, 1.87)		
NPY P_{ARTP}		0.546			0.365			0.038			
rs16129	GG 73 / 8259	1.00		37 / 4048	1.00		36 / 4211	1.00		0.015, 0.043	
	GT 132 / 11354	1.26	(0.94, 1.68)	83 / 7017	1.20	(0.81, 1.79)	49 / 4336	1.34	(0.86, 2.09)		
	TT 44 / 4134	1.19	(0.81, 1.74)	25 / 3124	0.82	(0.49, 1.36)	19 / 1010	2.35	(1.32, 4.19)		

¹ Hazard Ratios (HR) and 95% Confidence Intervals (CI) are for primary invasive cases; adjusted for age, study, BMI during referent year, parity, % NA ancestry, and SEER stage. Table includes genes with ARTP $p < 0.05$ and SNPs with $p < 0.05$

² High LD ($r^2 = 0.95$ among women with low NA ancestry and 0.96 among women with high NA ancestry)

High LD among women with low NA ancestry ($r^2=0.80$)

Author Manuscript

Author Manuscript

Author Manuscript

Author Manuscript