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The Epidemiology of Carbapenem-Resistant *Klebsiella*pneumoniae Colonization and Infection among Long-Term Acute Care Hospital Residents

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Abstract

Objective—An improved understanding of carbapenem-resistant *Klebsiella pneumoniae* (CRKP) in long-term acute care hospitals (LTACHs) is needed. The objective of this study was to assess risk factors for colonization or infection with CRKP in LTACH residents.

Methods—A case-control study was performed at a university-affiliated LTACH from 2008 to 2013. Cases were defined as all patients with clinical cultures positive for CRKP and controls were those with clinical cultures positive for carbapenem-susceptible *K. pneumoniae* (CSKP). A multivariable model was developed to identify risk factors for CRKP infection or colonization.

Results—222 patients were identified with *K. pneumoniae* clinical cultures during the study period; 99 (45%) were case patients and 123 (55%) were control patients. On multivariable analysis, factors associated with a significant risk for CRKP colonization or infection were solid organ or stem cell transplantation (OR, 5.05; 95% CI, 1.23 – 20.8, P=0.03), mechanical ventilation (OR, 2.56; 95% CI, 1.24 – 5.28, P=0.01), fecal incontinence (OR, 5.78; 95% CI, 1.52 – 22.0, P=0.01), and exposure in the prior 30 days to meropenem (OR, 3.55; 95% CI, 1.04 – 12.1, P=0.04), vancomycin (OR, 2.94; 95% CI, 1.18 – 7.32, P=0.02), and metronidazole (OR, 4.22; 95% CI, 1.28 – 14.0, P=0.02).

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Conclusions—Rates of colonization and infection with CRKP were high in the LTACH setting, with nearly half of *K. pneumoniae* cultures demonstrating carbapenem resistance. Further studies are needed on interventions to limit the emergence of CRKP in LTACHs, including targeted surveillance screening of high-risk patients and effective antibiotic stewardship measures.

Keywords

Carbapenem-resistant *Klebsiella pneumoniae* (CRKP); risk factors; long-term acute care hospital (LTACH)

INTRODUCTION

Infections due to carbapenem-resistant *Enterobacteriaceae* (CRE) are associated with high mortality rates and limited antibiotic treatment options ^{1–6}. Since the initial identification of a *Klebsiella pneumoniae* carbapenemase (KPC)-producing *K. pneumoniae* isolate in 1996, CRE have rapidly disseminated worldwide^{5,7}. Although multiple genera of Enterobacteriaceae have been found to carry carbapenemase enzymes, carbapenem-resistant *Klebsiella pneumoniae* (CRKP) remains the most epidemiologically important in the United States. Recent studies have emphasized the increasing importance of long-term acute care hospitals (LTACHs) as reservoirs and regional amplifiers of CRKP, with up to 9-fold higher prevalence rates of colonization compared to those in acute care hospitals^{8–11}.

Residents of LTACHs are at increased risk for colonization and infection with CRKP for a number of reasons. LTACHs care for a chronically, critically ill patient population that is characterized by significant debility, prolonged durations of stay, and high rates of invasive device utilization and antibiotic exposure¹². Understanding the epidemiology of CRKP in LTACHs is critical for the development of infection prevention and antibiotic stewardship measures to reduce the emergence of these organisms in the long-term care setting. However, prior studies assessing risk factors for CRE acquisition in LTACHs have been performed during ongoing outbreaks^{13,14}. High levels of colonization pressure during acute outbreaks with multidrug-resistant organisms increases the risk for horizontal transmission, and may not reflect transmission dynamics in other settings¹⁵.

Therefore, given the limited data on the epidemiology of CRKP in LTACHs in the nonoutbreak setting, we performed a case-control study to identify risk factors for infection or colonization with CRKP in LTACH residents.

METHODS

Study design

A retrospective case-control study was performed at a 38-bed, freestanding LTACH within the University of Pennsylvania Health System to identify risk factors for CRKP infection or colonization. Per institutional policy, all patients at the facility have surveillance blood, urine, and sputum or tracheal aspirate cultures performed routinely on admission. Clinical cultures obtained following admission are ordered at the discretion of treating physicians. Contact precautions are implemented for patients with positive CRKP cultures for the duration of their stay. The facility did not institute universal preemptive contact precautions

prior to surveillance culture results during the study period. All patients in the facility are housed in single rooms. The study was reviewed and approved by the Institutional Review Boards of the University of Pennsylvania and Good Shepherd Penn Partners.

All microbiologic isolates were processed at the Hospital of the University of Pennsylvania (HUP) Clinical Microbiology Laboratory. Standard identification and susceptibility testing of *K. pneumoniae* isolates was performed using the semiautomated Vitek 2 identification and susceptibility system (bioMérieux, Durham, NC) and interpreted according to Clinical and Laboratory Standards Institute (CLSI) criteria ¹⁶. Confirmatory KPC polymerase chain reaction (PCR) testing was performed for isolates with a meropenem MIC of 2 μ g/mL and ertapenem MIC < 1 μ g/mL, or a meropenem MIC 2 μ g/mL and ertapenem MIC 1 μ g/mL.

CRKP isolates were tested for susceptibility to tigecycline and colistin or polymyxin B using Etest (bioMérieux, Durham, NC), with isolates with an MIC of $2 \mu g/mL$ determined to be susceptible. Fosfomycin susceptibility was performed using the disk diffusion method as per CLSI guidelines.

Study population

Case patients were defined as all patients with a clinical culture (surveillance culture or clinical culture during LTACH stay) positive for CRKP from July 1 2008 to July 1 2013. Control patients were defined as all patients with a clinical culture positive for carbapenem-susceptible *K. pneumoniae* (CSKP) over the same time period. Patients with positive cultures for both CRKP and CSKP during the study period were included as cases. Only the first positive *K. pneumoniae* culture was included for each unique patient.

Data collection

Data were abstracted from Penn Data Store, a comprehensive electronic database that includes demographic, laboratory, billing, and pharmacy information for all patients treated within the University of Pennsylvania Health System. The following variables were collected: demographics, comorbidities, length of stay (LOS) prior to *K. pneumoniae* culture positivity, location from which the patient was transferred (e.g., acute care hospital, rehabilitation facility), and study year. Comorbid conditions included diabetes mellitus, chronic kidney disease (CKD), chronic pulmonary disease, and malignancy, among others.

Use of antibiotics and immunosuppressive medications (e.g., tacrolimus, cyclosporine) within 30 days of the *K. pneumoniae* culture was also assessed. For purposes of analyses, antibiotics were classified as follows: aminoglycosides; fluoroquinolones; third-generation cephalosporins (i.e. ceftriaxone, ceftazidime); first-generation cephalosporins (i.e. cefadroxil, cefazolin, cephalexin); cefepime; ampicillin/sulbactam; piperacillin-tazobactam; meropenem; metronidazole; vancomycin; linezolid; clindamycin; trimethoprim-sulfamethoxazole; tigecycline; and colistin. Finally, data on the use of mechanical ventilation and presence of a central venous or urinary catheter were collected via medical record review.

Statistical analysis

Cases and controls were characterized by potential risk factors, including demographics, comorbidities, mechanical ventilation, indwelling device use, and antibiotic exposure. Descriptive statistics were conducted to characterize the entire study population. Bivariable analysis of all potential risk factors for CRKP infection or colonization was performed using Fisher's exact test for categorical variables and the Wilcoxon rank sum test for continuous variables. Odds ratios (OR) and 95% confidence intervals (CIs) were calculated to evaluate the strength of each association. Multivariable analyses were then performed using multiple logistic regression. All variables from bivariable analyses with P values <0.05 were evaluated and considered for inclusion in the final multivariable model. Backward stepwise selection was performed for selection of variables in the final explanatory model, with results confirmed using likelihood ratio testing 17 . For all calculations, a two-tailed P value of <0.05 was considered significant.

All analyses were performed using STATA v.13.0 (StataCorp, College Station, Texas).

RESULTS

Study population

A total of 222 unique patients had clinical cultures positive for K. pneumoniae during the study period. The mean age of patients was 69.8 years and 52% were female. The median length of stay prior to K. pneumoniae culture positivity was 6 days (interquartile range [IQR] 1–19) for cases and 4 days (IQR 1–15) for controls, respectively (Table 1). Rates of central venous catheter and urinary catheter utilization were nearly 50% in both groups, with no statistical difference between cases and controls.

Ninety-nine (45%) of 222 *K. pneumoniae* isolates were identified as CRKP. Forty-three of these were isolated within 72 hours of admission, with 29 (67%) transferred from regional academic medical centers and 14 (33%) transferred from community medical centers.

The incidence rate of CRKP acquired after 72 hours of LTACH admission was 1.03 cases per 1,000 patient-days. No temporal trends were noted in CRKP incidence over the duration of the study period (data not shown).

Risk factors for colonization or infection with CRKP

On bivariable analysis (Table 1), there were no significant differences in age (P= 0.53), gender (P= 0.12), or length of LTACH stay prior to K. pneumoniae culture positivity (P= 0.35) between cases and controls.

Colonization or infection with CRKP was significantly associated with non-white race and with the following comorbidities: solid organ or hematopoietic stem cell transplant, mechanical ventilation, cognitive impairment, tracheostomy, and fecal incontinence. There was also an association between CRKP and use of corticosteroids, immunosuppressants, vancomycin, cefepime, meropenem, and metronidazole within the prior 30 days.

On subsequent multivariable analyses (Table 2), the following variables were significant risk factors for infection or colonization with CRKP: solid organ or hematopoietic stem cell transplant (OR, 5.05; 95% CI, 1.23-20.8, P=0.03), mechanical ventilation (OR, 2.56; 95% CI, 1.24-5.28, P=0.01), and fecal incontinence (OR, 5.78; 95% CI, 1.52-22.0, P=0.01). Administration of the following antibiotics within the prior 30 days was also significantly associated with CRKP isolation: meropenem (OR, 3.55; 95% CI, 1.04-12.1, P=0.04), vancomycin (OR, 2.94; 95% CI, 1.18-7.32, P=0.02), and metronidazole (OR, 4.22; 95% CI, 1.28-14.0, P=0.02).

Antibiotic susceptibility profiles of CRKP isolates

Sixteen (17%) of 93 CRKP isolates had a meropenem MIC of 1 μ g/mL and 17 (18%) isolates had a meropenem MIC of 2 μ g/mL. All of these isolates tested positive for KPC by PCR. The remaining 60 CRKP isolates had an MIC >2 μ g/mL. Table 3 demonstrates susceptibility profiles of CRKP isolates. Overall, there was a relatively low rate of resistance to amikacin (31%). Fifty-two of the CRKP isolates underwent tigecycline susceptibility testing, with the majority (83%) identified as susceptible. Forty-nine (94%) of 52 tested CRKP isolates were susceptible to colistin or polymyxin B.

DISCUSSION

In this case-control study, nearly half (45%) of *K. pneumoniae* isolates identified by clinical cultures in a university-affiliated LTACH over a five-year period were resistant to carbapenems. Risk factors for CRKP infection or colonization included solid organ or hematopoietic stem cell transplantation, mechanical ventilation, fecal incontinence, and use of vancomycin, meropenem, or metronidazole in the prior 30 days

The results of our study are notable for a high rate of colonization or infection with CRKP in the study population, with nearly half of K. pneumoniae isolates demonstrating resistance to carbapenems. The high incidence rate of CRKP found in our study in a non-outbreak setting is of major concern, and is nearly fivefold the prevalence reported from acute care hospitals ¹⁸. Notably, 43% of CRKP isolates were detected within 72 hours of arrival to the LTACH, suggesting presence on admission versus acquisition in the LTACH. CRKP isolates identified later during hospitalization may have been acquired through horizontal transmission or through detection of CRKP colonization that preceded admission. While detection of CRKP was likely facilitated through the use of admission clinical culture screening, this finding underscores the importance of LTACHs in the regional epidemiology of CRKP emergence and dissemination. Knowledge of local prevalence rates of CRKP are essential to coordinating a regional approach to CRE control, including enhanced interfacility communication between LTACHs, acute care hospitals, and long-term care facilities, and engagement of local health departments in regional surveillance efforts¹⁹. Furthermore, such high carbapenem resistance rates have important implications for infection prevention and antimicrobial stewardship measures specifically in the LTACH setting, including selection of an empiric antibiotic regimen for infections due to *K. pneumoniae*.

We found that solid organ and hematopoietic stem cell transplant recipients were at increased risk for CRKP infection or colonization. Prior transplantation was previously

identified as a risk factor for CRKP in an acute-care hospital in New York²⁰, but to our knowledge, our study is the first to confirm this association in the LTACH setting. Transplant patients are at significant risk for colonization and/or infection with CRKP due to immunosuppression and high rates of antibiotic use. Given frequent hospitalizations and inter-facility transfers, these patients may also play an important role in the regional dissemination of CRKP²¹. Targeted active surveillance and/or pre-emptive isolation precautions on admission to an LTACH in this patient population may be of particular benefit and should be explored in future studies.

We also found that requirement for mechanical ventilation was a significant risk factor for colonization or infection with CRKP. Mechanical ventilation is a well-described risk factor for CRKP acquisition in acute-care hospitals^{20,22}. The vast majority of ventilated LTACH patients have antecedent intensive care unit (ICU) stays and higher rates of invasive device utilization, increasing their risk for colonization or infection with multidrug-resistant organisms. As ventilation-dependent respiratory failure is the leading admission diagnosis to LTACHs¹², this represents a substantial at-risk population contributing to increased CRKP colonization pressure, and could be targeted with enhanced surveillance measures as part of a coordinated regional CRKP control plan.

Fecal incontinence was also associated with an increased risk of CRKP in LTACH residents. While a study performed in a single LTACH²³ demonstrated that only 0.5% of environmental isolates were positive for CRKP, recent studies have implicated environmental sources as contributing factors to ongoing CRKP outbreaks^{4,24}. While the role of the environment in the transmission of CRE is still unclear, fecal incontinence likely leads to increased environmental contamination and subsequent risk for CRKP transmission. Fecal incontinence is also likely a surrogate for higher dependence on healthcare staff for hygiene and care, with increased opportunities for healthcare worker and patient contact and horizontal transmission of multidrug-resistant organisms. The potential benefit of enhanced precautions (i.e., gown and glove use in LTACH residents with fecal incontinence) should be explored in future studies.

Our analysis of recent antibiotic use found that meropenem, vancomycin, and metronidazole were risk factors for CRKP infection or colonization. Carbapenem use and its association with CRKP acquisition has been well-described in the acute-care hospital setting^{20,25}. The use of broad-spectrum antibiotics, such as those identified in our study, likely exert broad colonization pressure and may select for colonization and/or infection with more resistant strains. Antibiotic utilization is widespread in LTACHs, with rates of vancomycin and carbapenem consumption between the 50th and 75th percentiles of medical ICUs¹⁵. Effective and feasible antibiotic stewardship measures are urgently needed in this setting.

There are several limitations of this study. Given that rectal culture for detection of asymptomatic gastrointestinal colonization was not routinely performed, our results likely underestimated the true burden of CRKP colonization. While our study did not include species of Enterobacteriaceae other than *K. pneumoniae*, thereby potentially underestimating the degree of overall CRE infection or colonization, data from a prior CRE screening program found that 90% of CRE isolates at our institution are CRKP. Finally, the present

study was conducted in a university-affiliated LTACH, and these results may not be generalizable to other LTACHs with differing characteristics.

In conclusion, solid organ or hematopoietic stem cell transplantation, mechanical ventilation, and fecal incontinence were found to be independent risk factors for CRKP infection or colonization in LTACH residents. Patients with these risk factors represent a population who may benefit from enhanced CRKP surveillance measures, including on a regional basis. Recent exposure to meropenem, vancomycin, and metronidazole were also associated with increased risk of CRKP colonization or infection. Future studies are needed on optimal antimicrobial stewardship interventions specifically in the LTACH setting with the goal of reducing the emergence of CRKP.

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References

- 1. Bogan C, Kaye KS, Chopra T, et al. Outcomes of carbapenem-resistant Enterobacteriaceae isolation: matched analysis. Am J Infect Control. 2014; 42:612–620. [PubMed: 24837111]
- Borer A, Saidel-Odes L, Riesenberg K, et al. Attributable mortality rate for carbapenem-resistant Klebsiella pneumoniae bacteremia. Infect Control Hosp Epidemiol. 2009; 30:972–976. [PubMed: 19712030]
- Falagas ME, Tansarli GS, Karageorgopoulos DE, Vardakas KZ. Deaths attributable to carbapenemresistant Enterobacteriaceae infections. Emerg Infect Dis. 2014; 20:1170–1175. [PubMed: 24959688]
- 4. Snitkin ES, Zelazny AM, Thomas PJ, et al. Tracking a hospital outbreak of carbapenem-resistant Klebsiella pneumoniae with whole-genome sequencing. Sci Transl Med. 2012; 4:148ra116.
- Tzouvelekis LS, Markogiannakis A, Psichogiou M, Tassios PT, Daikos GL. Carbapenemases in Klebsiella pneumoniae and other Enterobacteriaceae: an evolving crisis of global dimensions. Clin Microbiol Rev. 2012; 25:682–707. [PubMed: 23034326]
- Gasink LB, Edelstein PH, Lautenbach E, Synnestvedt M, Fishman NO. Risk factors and clinical impact of Klebsiella pneumoniae carbapenemase-producing K. pneumoniae. Infect Control Hosp Epidemiol. 2009; 30:1180–1185. [PubMed: 19860564]
- 7. Yigit H, Queenan AM, Anderson GJ, et al. Novel carbapenem-hydrolyzing beta-lactamase, KPC-1, from a carbapenem-resistant strain of Klebsiella pneumoniae. Antimicrob Agents Chemother. 2001; 45:1151–1161. [PubMed: 11257029]
- 8. Lin MY, Lyles-Banks RD, Lolans K, et al. The importance of long-term acute care hospitals in the regional epidemiology of Klebsiella pneumoniae carbapenemase-producing Enterobacteriaceae. Clin Infect Dis. 2013; 57:1246–1252. [PubMed: 23946222]
- 9. Won SY, Munoz-Price LS, Lolans K, Hota B, Weinstein RA, Hayden MK. Emergence and rapid regional spread of Klebsiella pneumoniae carbapenemase-producing Enterobacteriaceae. Clin Infect Dis. 2011; 53:532–540. [PubMed: 21865189]
- Marchaim D, Chopra T, Bogan C, et al. The burden of multidrug-resistant organisms on tertiary hospitals posed by patients with recent stays in long-term acute care facilities. Am J Infect Control. 2012; 40:760–765. [PubMed: 22285709]

 Marquez P, Terashita D, Dassey D, Mascola L. Population-based incidence of carbapenem-resistant Klebsiella pneumoniae along the continuum of care, Los Angeles County. Infect Control Hosp Epidemiol. 2013; 34:144–150. [PubMed: 23295560]

- 12. Munoz-Price LS. Long-term acute care hospitals. Clin Infect Dis. 2009; 49:438–443. [PubMed: 19548836]
- 13. Chitnis AS, Caruthers PS, Rao AK, et al. Outbreak of carbapenem-resistant enterobacteriaceae at a long-term acute care hospital: sustained reductions in transmission through active surveillance and targeted interventions. Infect Control Hosp Epidemiol. 2012; 33:984–992. [PubMed: 22961017]
- Endimiani A, Depasquale JM, Forero S, et al. Emergence of blaKPC-containing Klebsiella pneumoniae in a long-term acute care hospital: a new challenge to our healthcare system. J Antimicrob Chemother. 2009; 64:1102–1110. [PubMed: 19740911]
- 15. Gould CV, Rothenberg R, Steinberg JP. Antibiotic resistance in long-term acute care hospitals: the perfect storm. Infect Control Hosp Epidemiol. 2006; 27:920–925. [PubMed: 16941316]
- Performance Standards for Antimicrobial Susceptibility Testing; Twentieth Information Supplement. Clinical and Laboratory Standards Institute. Jun. 2010 Vol. 30 M100-S20-U.
- 17. Mickey RM, Greenland S. The impact of confounder selection criteria on effect estimation. Am J Epidemiol. 1989; 129:125–137. [PubMed: 2910056]
- Vital signs: carbapenem-resistant Enterobacteriaceae. MMWR Morb Mortal Wkly Rep. 2013;
 62:165–170. [PubMed: 23466435]
- [Last accessed July 1, 2015] Guidance for Control of Carbapenem-resistant Enterobacteriaceae (CRE): 2012 CRE Toolkit. Centers for Disease Control and Prevention. http://www.cdc.gov/hai/pdfs/cre/CRE-guidance-508.pdf
- Patel G, Huprikar S, Factor SH, Jenkins SG, Calfee DP. Outcomes of carbapenem-resistant Klebsiella pneumoniae infection and the impact of antimicrobial and adjunctive therapies. Infect Control Hosp Epidemiol. 2008; 29:1099–1106. [PubMed: 18973455]
- 21. Mathers AJ, Cox HL, Bonatti H, et al. Fatal cross infection by carbapenem-resistant Klebsiella in two liver transplant recipients. Transpl Infect Dis. 2009; 11:257–265. [PubMed: 19254325]
- Leavitt A, Navon-Venezia S, Chmelnitsky I, Schwaber MJ, Carmeli Y. Emergence of KPC-2 and KPC-3 in carbapenem-resistant Klebsiella pneumoniae strains in an Israeli hospital. Antimicrob Agents Chemother. 2007; 51:3026–3029. [PubMed: 17562800]
- 23. Thurlow CJ, Prabaker K, Lin MY, Lolans K, Weinstein RA, Hayden MK. Anatomic sites of patient colonization and environmental contamination with Klebsiella pneumoniae carbapenemase-producing Enterobacteriaceae at long-term acute care hospitals. Infect Control Hosp Epidemiol. 2013; 34:56–61. [PubMed: 23221193]
- 24. Kotsanas D, Wijesooriya WR, Korman TM, et al. "Down the drain": carbapenem-resistant bacteria in intensive care unit patients and handwashing sinks. Med J Aust. 2013; 198:267–269. [PubMed: 23496403]
- 25. Hussein K, Sprecher H, Mashiach T, Oren I, Kassis I, Finkelstein R. Carbapenem resistance among Klebsiella pneumoniae isolates: risk factors, molecular characteristics, and susceptibility patterns. Infect Control Hosp Epidemiol. 2009; 30:666–671. [PubMed: 19496647]

Table 1

Bivariable analysis of risk factors for infection or colonization with carbapenem-resistant *K. pneumoniae* in long-term acute care hospital residents.

Variable	Cases (n=99) ^a	Controls (n=123) ^a	OR (95% CI)	P value
Age, median years (IQR)	72 (63–79)	72 (62–78)		0.53
Female sex	46 (46)	70 (57)	0.66 (0.37–1.16)	0.12
Non-white race	67 (68)	65 (53)	1.87 (1.00–3.37)	0.03
Comorbidities				
CHF	11 (11)	15 (12)	0.90 (0.35–2.22)	0.80
CKD	28 (28)	23 (19)	1.71 (0.87–3.39)	0.09
Cirrhosis	4 (4)	2 (2)	2.55 (0.36–28.6)	0.27
Diabetes	9 (9)	12 (10)	0.93 (0.33–2.52)	0.87
Obesity b	2 (2)	7 (6)	0.34 (0.03–1.86)	0.17
Chronic lung disease	10 (10)	11 (9)	1.14 (0.41–3.12)	0.78
Malignancy	10 (10)	5 (4)	2.65 (0.79–10.2)	0.08
HIV infection	2 (2)	1 (1)	2.51 (0.13–150)	0.44
Cognitive impairment	18 (18)	7 (6)	3.68 (1.38–10.9)	0.003
Solid organ or hematopoietic stem cell transplant	12 (12)	3 (2)	5.52 (1.42–31.2)	0.004
Clostridium difficile colitis	9 (9)	6 (5)	1.95 (0.59-6.89)	0.21
Sacral decubitus ulcer	8 (8)	4 (3)	2.62 (0.67–12.2)	0.11
Fecal incontinence	13 (13)	3 (2)	6.05 (1.58–33.8)	0.002
Total parental nutrition	9 (9)	17 (14)	0.62 (0.23–1.57)	0.28
Mechanical ventilation	81 (82)	80 (67)	2.19 (1.11–4.42)	0.02
Tracheostomy	87 (90)	91 (77)	2.58 (1.12–6.32)	0.02
Gastrostomy	80 (82)	83 (70)	1.92 (0.97–3.91)	0.05
Central venous catheter	50 (53)	66 (56)	0.88 (0.49–1.56)	0.63
Urinary catheter	50 (51)	49 (41)	1.50 (0.84–2.64)	0.15
$Medications^{C}$				
Corticosteroids	36 (36)	28 (23)	1.94 (1.03–3.65)	0.03
Immunosuppressants	17 (17)	6 (5)	4.04 (1.44–13.0)	0.003
Vancomycin	22 (22)	9 (7)	3.62 (1.50–9.38)	0.001
Cefepime	13 (13)	5 (4)	3.57 (1.13–13.2)	0.01
Meropenem	13 (13)	4 (3)	4.50 (1.32–19.5)	0.006
Metronidazole	15 (15)	6 (5)	3.48 (1.21–11.4)	0.009

Abbreviations: OR, odds ratio; CI, confidence interval; IQR, interquartile range; LOS, length of stay; CHF, congestive heart failure; CKD, chronic kidney disease.

a n (%) unless noted otherwise.

*b*ВМІ 25.

^cReceived within 30 days of the positive culture. Only variables with P < 0.05 are shown.

Table 2

Multivariable model of risk factors for infection or colonization with carbapenem-resistant *K. pneumoniae* in long-term acute care hospital residents.

Variable	OR (95% CI)	P value
Solid organ or hematopoeitic stem cell transplant	5.05 (1.23–20.8)	0.03
Mechanical ventilation	2.56 (1.24–5.28)	0.01
Fecal incontinence	5.78 (1.52–22.0)	0.01
Meropenem ^a	3.55 (1.04–12.1)	0.04
Vancomycin ^a	2.94 (1.18–7.32)	0.02
Metronidazole ^a	4.22 (1.28–14.0)	0.02

Abbreviations: OR, odds ratio; CI, confidence interval.

^aReceipt within prior 30 days.

 Table 3

 Antibiotic susceptibility profiles of carbapenem-resistant *K. pneumoniae* isolates.

Antibiotic	Susceptible n (%)	Intermediate n (%)	Resistant n (%)
Amikacin	66 (69)		30 (31)
Tigecycline ^a	43 (83)	8 (15)	1 (2)
Colistin and/or Polymyxin B ^a	49 (94)		3 (6)
Fosfomycin ^b	27 (75)	5 (14)	4 (11)

^a52 isolates tested.

b₃₆ isolates tested.