

Published in final edited form as:

Am J Ind Med. 2014 July; 57(7): 748-756. doi:10.1002/ajim.22331.

Arterial Stiffness, Oxidative Stress, and Smoke Exposure in Wildland Firefighters

Denise M. Gaughan, ScD^{1,2,*}, Paul D. Siegel, PhD³, Michael D. Hughes, PhD⁴, Chiung-Yu Chang, ScD², Brandon F. Law, MS³, Corey R. Campbell, MS⁵, Jennifer C. Richards, MS⁶, Stefanos F. Kales, MD², Marcia Chertok, BS², Lester Kobzik, PhD⁷, Phuongson Nguyen, PhD⁷, Carl R. O'Donnell, PhD⁸, Max Kiefer, MS⁵, Gregory R. Wagner, MD², and David C. Christiani, MD²

¹Department of Preventive Medicine and the Institute for Translational Epidemiology, Icahn School of Medicine at Mount Sinai, New York, New York

²Department of Environmental Health (Environmental and Occupational Medicine and Epidemiology Program), Harvard School of Public Health, Boston, Massachusetts

³Health Effects Laboratory Division, National Institute for Occupational Safety and Health, Morgantown, West Virginia

⁴Departmentof Biostatistics, Harvard School of Public Health, Boston, Massachusetts

⁵Western States Office, National Institute for Occupational Safety and Health, Denver, Colorado

⁶Department of Health and Exercise Science, Colorado State University, Fort Collins, Colorado

⁷Department of Environmental Health (Molecular and Integrative Physiological Sciences), Harvard School of Public Health, Boston, Massachusetts

⁸Beth Israel Deaconess Medical Center, Division of Pulmonary and Critical Care Medicine, Boston, Massachusetts

Abstract

Objectives—To assess the association between exposure, oxidative stress, symptoms, and cardiorespiratory function in wildland firefighters.

Methods—We studied two Interagency Hotshot Crews with questionnaires, pulse wave analysis for arterial stiffness, spirometry, urinary 8-iso-prostaglandin F2 α (8-isoprostane) and 8-hydroxy-2'-deoxyguanosine (8-OHdG), and the smoke exposure marker (urinary levoglucosan). Arterial stiffness was assessed by examining levels of the aortic augmentation index, expressed as a percentage. An oxidative stress score comprising the average of z-scores created for 8-OHdG and 8-isoprostane was calculated.

Results—Mean augmentation index % was higher for participants with higher oxidative stress scores after adjusting for smoking status. Specifically for every one unit increase in oxidative

^{*}Correspondence to: Denise M. Gaughan, ScD, Icahn School of Medicine at Mount Sinai, 17 East 102nd Street, D3-132, New York, NY10029-6574. denise.gaughan@mssm.edu.

Disclosure Statement: The authors report no conflicts of interests.

stress score the augmentation index % increased 10.5% (95% CI: 2.5, 18.5%). Higher mean lower respiratory symptom score was associated with lower percent predicted forced expiratory volume in one second/forced vital capacity.

Conclusions—Biomarkers of oxidative stress may serve as indicators of arterial stiffness in wildland firefighters.

Keywords

vascular stiffness; 8-iso-prostaglandin $F2\alpha$; 8-hydroxy-2'-deoxyguanosine; spirometry; levoglucosan

BACKGROUND

Firefighters, both structural and wildland, are known to have cardiovascular and respiratory problems [Musk et al., 1979; Sardinas et al., 1986; Chia et al., 1990; Rothman et al., 1991; Guidotti and Clough, 1992; Liu et al., 1992; Materna et al., 1992; Scannell and Balmes, 1995; Betchley et al., 1997; Austin et al., 2001; Burgess et al., 2001; Kales et al., 2003; Slaughter et al., 2004; CDC, 2006; Gaughan et al., 2008; Yoo and Franke, 2009]. Cardiovascular disease (CVD) events are the leading cause of on-duty and lifetime mortality among structural (career and volunteer) firefighters [Sardinas et al., 1986; Kales et al., 2003; Yoo and Franke, 2009]. The deleterious effects of smoke exposure to structural firefighters have been extensively researched [Musk et al., 1979; Sardinas et al., 1986; Chia et al., 1990; Guidotti and Clough, 1992; Liu et al., 1992; Scannell and Balmes, 1995; Burgess et al., 2001; Kales et al., 2003; CDC, 2006; Yoo and Franke, 2009]. Exposure to particulates and other contaminants, heavy physical exertion and cardiovascular strain have been found to be among the chief health hazards associated with structural firefighting [Gledhill and Jamnik, 1992; Takeyama et al., 2005; Delfino et al., 2009]. Those findings, however, may not be generalizable to wildland firefighters for a number of reasons, including the difference in smoke composition, comparative younger age of wildland firefighters, often shorter career tenure, and the longer duration of respiratory exposures for wildland firefighters. Additionally, structural firefighters routinely wear respiratory protection when responding to fires while wildland firefighters do not.

Fine particulate exposure has been associated with acute changes in cardiovascular and pulmonary function [Vallyathan et al., 1995; Mott et al., 2005; Dominici et al., 2006; Cavallari et al., 2008; Fang et al., 2008, 2009]. Free radical mechanisms have been implicated as a contributing factor in general toxicity, inflammation, asthma, fibrogenesis, bronchopulmonary carcinogenesis, and atherosclerotic plaque formation [Jarjour and Calhoun, 1994; Leonard et al., 2007; LeBlanc et al., 2010].

Wood fires produce smoke with abundant particles in the inhalable range ($<100 \, \mu m$) and contain both carbon radicals and precursors. The latter are able to react with H_2O_2 after exposure to cells and generate the highly reactive hydroxyl radical (OH) from a Fenton-like reaction [Leonard et al., 2007]. The authors additionally observed that carbon radicals were stronger (per unit mass) in larger (coarse) sized particles while OH and other ROS were stronger (per unit mass) in the smaller (ultrafine) sized particles. Finally, the authors noted

that fine and ultrafine woodsmoke particles also significantly increased H₂O₂, DNA strand breaks and lipid peroxidation in exposed RAW 264.7 cells.

Pyrolysis of organic components may increase the potency and or toxicity of particulate by producing ROS. Measurement of levoglucosan, a sugar anhydride by-product of incomplete combustion of cellulose, may be used to indicate relative exposure to products of pyrolysis from burning biomass [Simoneit et al., 1999]. A recent study examined personable exposure to airborne respirable levoglucosan using a Dorr-Oliver cyclone and air sampling pump, and cross-shift changes in lung function in a population of 17 wildland firefighters for 4 days at a large wildland fire [Gaughan et al., 2014]. The authors reported that levoglucosan was found mainly in the respirable fraction, defined as under 2.5 µm, with higher concentrations during fireline construction than in mop-up operations. Furthermore, larger cross-shift declines in forced expiratory volume in one second (FEV₁) were associated with exposure to higher concentrations of respirable levoglucosan (*P*-value <0.05).

Urinary levoglucosan has also been investigated as a biomarker for smoke exposure. Bergauff et al. examined cross-shift changes in urinary levoglucosan in nine firefighters exposed to wood smoke in a controlled setting. They observed elevated urinary levoglucosan following smoke exposure in some but not all firefighters [Bergauff et al., 2010]. Moreover, the authors noted the contribution of dietary intake to urinary levoglucosan levels.

Urinary 8-iso-prostaglandin F2\alpha (8-isoprostane), a biomarker of oxidative stress generated by lipid peroxidation, may serve as a biomarker for atherosclerosis [Cipollone et al., 2000]. Oxidative deoxyribonucleic acid (DNA) damage and repair has also been linked to atherosclerosis. Martinet et al. examined five human carotid endarterectomy specimens and five mammary artery specimens for 8-hydroxy-2'-deoxyguanosine (8-OHdG), an oxidized nucleoside of DNA. The authors observed an increased amount of 8-OHdG in plaques compared to the underlying media or non-atherosclerotic mammary arteries [Martinet et al., 2002]. Urinary 8-OHdG is excreted upon DNA repair and may serve as a non-invasive biomarker of global oxidative DNA damage. For instance, acute changes in urinary 8-OHdG concentrations have been reported in occupational studies examining welders [Kim et al., 2004; Nuernberg et al., 2008].

Elevated arterial stiffness is a characteristic of large artery pathology, a major contributor to CVD, and may serve as an indicator of pre-clinical atherosclerosis and/or hypertension [Blankenhorn and Kramsch, 1989; Duprez and Cohn, 2007]. Assessment of arterial stiffness is done through ultrasound or measurement of pulse wave velocity. The aortic augmentation index, is an indirect measure of systemic arterial stiffness based on pulse wave velocity and is calculated as a percentage. A recent meta-analysis demonstrated that a 10% increase in augmentation index % was associated with a 31.8% increased risk of cardiovascular events and a 34.8% increased risk of total mortality [Vlachopoulos et al., 2010]. The augmentation index % has been successfully implemented in occupational research settings [Nürnberger et al., 2002; Fang et al., 2008].

There are four types of wildland firefighter suppression crews: engine crew, hand crew, helicopter crew, and smoke jumpers. Type 1 Interagency Hotshot Crews are an elite type of hand crew, comprising up to 20 firefighters who construct fire lines using hand tools during the most dangerous phases of fire suppression.

The question addressed by the present study was whether wildland firefighting exposures are associated with oxidative stress concentrations and with pulmonary and vascular function. To answer these questions, we assessed spirometry, vascular function, symptoms, and systemic biomarkers of exposure, inflammation and oxidative stress in members of two Type 1 wildland firefighter Interagency Hotshot Crews.

METHODS

The National Interagency Fire Center (NIFC) Risk Management Committee arranged for two crews to participate in this study: the Alpine Interagency Hotshot Crew, Rocky Mountain National Park, Estes Park, CO and the Pike Interagency Hotshot Crews, Pike and San Isabel National Forests, Monument, CO. The Interagency Hotshot Crews participated in our study by completing questionnaires, pulse wave analysis and spirometry in May 2011. Serum cholesterol, and biomarkers of systemic inflammation (high sensitivity c-reactive protein (hsCRP) and fibrinogen), oxidative stress (8-isoprostane), oxidative DNA damage (8-OHdG), and smoke exposure (urinary levoglucosan) were measured. The study protocol was approved by the Harvard School of Public Health (HSPH) Institutional Review Board and the National Institute for Occupational Safety and Health (NIOSH) Human Subjects Review Board. Informed consent was obtained from each research participant.

Medical Testing Methods

Pulse wave analysis—Vascular function was measured using a pulse wave analysis system according to the manufacturer's instructions [SphygmoCor CP, Atcor Medical Pty Ltd., Sydney, Australia]. Briefly, participants were seated with the dominant arm extended onto a flat surface so that the antecubital fossa was at heart level. Following 5 min of rest, a high-fidelity micro-nanometer was used to flatten the radial artery with gentle pressure. Ten seconds of sequential pulse pressure waveforms were recorded at each reading. The waveforms were then transformed into a corresponding central aortic waveform via a validated transfer function where the systolic part of the central aortic waveform is characterized by a first peak caused by left ventricular ejection and a second peak caused by wave reflection. The difference between the two peaks reflects the degree to which the central aortic pressure is augmented by wave reflection. We calculated each participant's aortic augmentation index %, defined as the ratio of augmented pressure to pulse pressure (i.e., augmentation index % = augmented pressure/pulse pressure ×100) [Nürnberger et al., 2002] and heart rate corrected to 75 beats per minute. Larger augmentation index % values denote increased wave reflection. A minimum of three within-session recordings were obtained from each participant.

Spirometry—Pulmonary function was determined on each participant using an ultrasonic flow spirometer [EasyOneTM Diagnostic Spirometry System 2001, ndd Medical Technologies, Zurich, Switzerland]. Technicians completed a NIOSH-approved spirometry

course followed American Thoracic Society (ATS) guidelines [Miller et al., 2005]. Test results were interpreted using reference values generated from the Third National Health and Nutrition Examination Survey (NHANES III) [Hankinson et al., 1999]. Airways obstruction was defined as a forced expiratory volume in the first second (FEV₁) to forced vital capacity (FVC) ratio below the lower limit of normal according to published reference equations [Pellegrino et al., 2005]. We examined percent predicted FEV₁ (FEV₁%-predicted), percent predicted FVC (FVC %-predicted), and percent predicted FEV₁/FVC (FEV₁/FVC %-predicted). We followed ATS procedure by inquiring about current medications but did not ask participants to abstain from using their medications prior to participating in this study for safety purposes. Reports were reviewed for quality by a respiratory physiologist (C.R.O) experienced in clinical pulmonary function laboratory administration.

Blood—Whole non-fasting serum samples (30 ml/sample) were collected by venous phlebotomy in EDTA tubes, and buffy coat was extracted and stored in cell lyses solution at -20° C for analyses of typical cardiovascular-related biomarkers, specifically, hsCRP and fibrinogen. We additionally examined total cholesterol, high-density lipoprotein cholesterol (HDL-cholesterol), low-density lipoprotein cholesterol (LDL-cholesterol), and triglycerides. These analyses were conducted by Quest Diagnostics Inc., Denver, CO.

Urine—Urine samples were analyzed for 8-OHDG and 8-isoprostane using competitive enzyme-linked immunoassays (EKS-350, Assay Designs, Inc., Ann Arbor, MI; 8-Isoprostane EIA Kit, Cayman Chemical Company, Ann Arbor, MI) as well as for creatinine (picric acid colorimetric assay; Oxford Biomedical Research, Oxford, MI). We also examined urine for levoglucosan concentration. Two hundred microliter of urine or levoglucosan standards (Blank, 6.25–100 µg/ml in saline) were added to 1.5 ml low retention microcentrifuge tubes. To this 30U of urease (Sigma, St. Louis, MO) was added and incubated for 1.0 hr at 37°C. After incubation, 600 µl of cold (4°C) ethanol was added to precipitate the protein. Approximately 400 mg of sodium sulfate was added volumetrically and allowed to sit for 2.0 min. Samples and standards were then centrifuged for 2.0 min at 14,000g. After centrifugation, 600 µl of each were transferred to clean low retention microcentrifuge tubes and evaporated to dryness using a vacuum centrifugation and gentle heat. Two hundred microliter of n-methyl-n-(trimethylsilyl) trifluoroacet (MSTFA) (Sigma Chemical Co.,) was then added to each tube, vortexed and incubated at 72°C for 1.0 hr. After derivatization samples and standards were analyzed on an Agilent 6890 gas chromatograph coupled to an Agilent 5975C mass spectrometer using a 30-m HP5-MS column (Agilent Technologies, Santa Clara, CA). Samples were injected (1 µl) in splitless mode into a 250°C inlet with a 6.0-min solvent delay. Analytes were eluted from the column using 1.0 ml/min helium and an oven temperature program as follows: 70°C for 3.0 min and then ramped at 25.0°C/min to a final temperature of 275°C. The MS source temperature was maintained at 230°C, and the quadrupole temperature was maintained at 150°C. Ions were scanned between 50 and 400 m/z. Levoglucosan from each sample was identified by the MS spectra and retention time (against the known standards) and quantified using the 204 m/z ion. The standard plot from which the samples were extrapolated used a polynomial curve fit of 204 m/z area under the curve ion count versus standard concentration. Specimens were

analyzed by NIOSH's Health Effects Laboratory Division, Allergy and Clinical Immunology Branch, Morgantown, WV.

Questionnaires—The questionnaire was based on two standardized questionnaires, the American Thoracic Society-Division of Lung Disease-78 (ATS-DLD-78) supplemented with questions from NHANES III [Ferris, 1978; Wasserfallen et al., 1997]. This modified questionnaire was designed to acquire information concerning chronic cardiovascular and respiratory conditions; lifetime diagnoses; tobacco history; symptom history; dietary intake; sleep patterns; medication use; and occupational history. A validated symptom scale, with Likert scoring where 0 =none, 1 =trivial, 2 =mild, 3 =moderate, and 4 =severe for upper and lower airways symptoms, was used to derive overall symptom scores by summing the responses to questions about 19 symptoms. Symptoms ascertained included cough, wheeze, sputum production, shortness of breath or chest tightness, and shortness of breath while walking, as well as various eye, nose, and throat symptoms. Subjects additionally completed a semi-quantitative food frequency questionnaire, adult version, 2007. Subjects were asked to report the average daily consumption of various foods in the preceding year. Responses ranged from "never" to "six or more servings per day." The food frequency questionnaire also assessed the frequency of multivitamin and mineral supplement usage. Frequency factors of related foods items were then summed to calculate the daily servings for each food group. The nutrient value of the food item was multiplied by the frequency of consumption in order to obtain macro and micro nutrient intake. Scoring was done by the Nutrition Department, Harvard School of Public Health (https://regepi.bwh.harvard.edu/health/ nutrition.html) [Willett et al., 1985].

Statistical Methods—Descriptive statistics were calculated for demographic and clinical variables. Mean values among subgroups were compared using Student's t-test techniques. Ordinary least squares regression techniques were used to examine associations between augmentation index %, FEV₁%-predicted, FVC %-predicted, FEV₁/FVC %-predicted, 8-OHdG concentration, and 8-isoprostane concentration using the following predictor variables: levoglucosan concentration, serum lipid levels, hsCRP concentration, fibrinogen concentration, LDL-cholesterol, HDL-cholesterol, Interagency Hotshot Crew, volunteer firefighter (yes or no), cumulative time spent fighting fires (seasons), medical diagnoses, allergies, upper and lower respiratory symptom scores, and history of tobacco use. Levoglucosan, 8-isoprostane, 8-OHdG, and hsCRP values were not normally distributed and thus the data were log₁₀-transformed to achieve an approximate normal distribution. We additionally calculated an "oxidative stress score" comprising z-score's of 8-isoprostane and 8-OHdG, where z-value's for each variable were calculated and then averaged to yield a score for each participant. The motivation for calculating the z-score was prompted by the possibility that while 8-isoprostane is considered a marker of lipid peroxidation and 8-OHdG has been linked to DNA damage and repair, they may both be measuring the overall level of oxidative stress in the urine. Thus, we standardized the two biomarkers of oxidative stress on one scale. These values were also examined in multivariable analysis. The z-value was calculated by subtracting the mean value from each subject's value and then dividing by the standard deviation of the values. Multivariable models were chosen based on an initial evaluation using stepwise selection techniques, followed by examining univariate

associations and sequentially adjusting for other predictors. Associations were considered statistically significant if P-values were <0.05 and borderline significant at 0.05 and <0.10. All analyses were conducted using SAS statistical software (version 9.3).

RESULTS

Thirty-eight of the 39 current members of both crews participated in the surveys in May 2011 during their training sessions at their respective home base parks (97%). One member was training off-site the day of our survey and as a result, could not participate. Members of the Pike Interagency Hotshot Crew had been exposed to smoke for 2 days at the Sand Gulch fire 4 days before the testing. The Sand Gulch fire in Wetmore, CO, was a 495-acre wildland fire at 7,000 feet elevation in high difficulty terrain. Fire behavior activity was described by crew members as high/extreme on the first day and low/smoldering on the second. It was reported that the average shift duration each day was 16 hr and it was estimated the crew members were on the fire line performing firefighting activities for 12.5 hr each day.

Participant demographic and clinical characteristics, overall and stratified by crew are shown in Tables I and II. The two crews were very similar. They were all male with a median age of 28 years and had spent a median of 3 years (seasons) working as an Interagency Hotshot Crew member. Approximately 5% of the participants were current smokers and nearly half (42%) reported current chewing tobacco use. Approximately one-third (29%) of the participants were classified as "permanent" employees, the remainder were "seasonal" hires. Additionally, six of the participants (16%) reported working as a volunteer structural firefighter off-season. Values for clinical characteristics were also comparable when examined by crew and within the normal range.

Seven participants reported a history of physician-diagnosed asthma (18%). The median age at asthma diagnosis was 7 years and the median time spent as a firefighter for these seven participants was 2 years (seasons). Among these individuals, the median FEV $_1$ %-predicted was 105%, the median FVC %-predicted was 109%, and the median FEV $_1$ /FVC %-predicted was 96%. Among the four individuals with current asthma, three were currently taking medication for their asthma. One of the seven participants with a history of asthma also reported a smoking history.

Four values for levoglucosan were below the limit of detection and were not included in the analysis. Detectable levels were observed for all values of 8-OHdG, 8-isoprostane, hsCRP, and fibrinogen.

In multivariable analysis, mean augmentation index % was higher for participants with higher oxidative stress scores. Specifically, for every one unit increase in oxidative stress score, mean augmentation index % increased 10.2% (10.2%, 95% CI: 1.35, 19.0%) (Table III). This association remained significant after adjusting for smoking status (P =0.01). No other variables were associated with augmentation index %, including our exposure variable, levoglucosan. Table III details these results.

However, higher levoglucosan concentration was positively associated with oxidative stress scores (Table IV). Specifically, for every twofold increase in log₁₀ levoglucosan

concentration, mean 8-OHdG increased by $\log_{10} 0.14 \,\mu\text{g/ml}$ (95% CI: 0.02, 0.25) (regression estimate 0.41 (95% CI: 0.04, 0.79)) and 8-isoprostane increased by $\log_{10} 0.16$ ng/ml (95% CI: 0.02, 0.29) (regression estimate 0.52 (95% CI: 0.06, 0.97)). Additionally, 8-OHdG values for participants who had recently been exposed to smoke particulate as measured by Interagency Hotshot Crew were higher than those with no recent exposure (P = 0.01). This association became borderline significant after adjusting for levoglucosan (P = 0.07) (Table IV).

Finally, higher lower respiratory symptom score (LRSS) was associated with lower FEV $_1$ /FVC %-predicted after adjusting for smoking status and history of asthma. Specifically, for every twofold increase in mean LRSS, the mean FEV $_1$ /FVC %-predicted dropped on average by 1.66% (95% CI: 0.10%, 3.22%) (regression estimate:–0.83 (95% CI: –1.61, –0.05)). No other variables were associated with FEV $_1$ /FVC %-predicted, most notably, neither oxidative stress score nor levoglucosan.

DISCUSSION

Particulate exposure has been associated with increased arterial stiffness in occupational cohorts. Fang et al. examined changes in augmentation index % in 26 welders over 24 hr on a welding day and non-welding day. Following welding fume exposure, the authors observed an increase in afternoon augmentation index % and a decrease in next morning augmentation index % [Fang et al., 2008]. The results suggested that exposure to welding fume particulate is associated with acute adverse vascular responses. In our homogenous group of healthy workers, we observed higher augmentation index % values for participants with higher oxidative stress values which were also associated with larger levoglucosan values. A possible explanation for this finding could be that our population was younger by comparison (28 vs. 41 years) and/or our assessment occurred 4 days post-exposure to smoke.

Similar to other studies, we observed an association between cross-shift differences in oxidative DNA damage and recent occupational exposure to particulate matter. Kim et al. obtained 5 days of cross-shift urinary 8-OHdG measurements from 20 welders exposed to metal fumes. The authors reported that urinary 8-OHdG levels were significantly elevated in post-shift samples compared to those collected pre-shift [Kim et al., 2004]. Nuernberg et al. [2008] observed similar 8-OHdG elevation post-shift in 63 welders. Moreover, the authors found an unexpected inverse relationship between post-shift and 8 hr post-shift values of 8-isoprostane and $PM_{2.5}$ (P < 0.05). We additionally observed a difference in 8-isoprostane by levoglucosan concentration. We did not observe a difference in mean values of 8-OHdG or 8-isoprostane between participants who identified themselves as having a history of smoking and those who did not (P = 0.51 and 0.64, respectively).

Levoglucosan was positively associated with both biomarkers of oxidative stress, 8-OHdG and 8-isoprostane. Urinary levoglucosan has been shown to increase in mice after exposure to wood smoke [Migliaccio et al., 2009]. However, when examining urinary levoglucosan concentration, it is important to control for dietary confounders such as smoked or fried foods and caramel. Based on responses to the Food Frequency Questionnaire, 32 (84%)

reported consuming two slices of bacon or less once per week; 34 (89%) reported eating one candy bar or less per week. All participants reported eating fried foods at home or away four to six times per week or less, although 22 (58%) reported consuming fried foods less than once per week. Inhaled levoglucosan is probably completely eliminated from the body within 24 hr post-exposure. Thus, it is possible that the elevated levels of levoglucosan we observed were caused by diet, rather than exposure.

There is evidence that particulate matter [PM] produced by woodsmoke fires may be more toxic than PM from ambient air due to the chemical components found in the smoke [Wegesser et al., 2009]. Leonard et al. [2007] examined aerodynamically size-selected aerosol samples at a large wildland fire. The authors found smoke particles in all size fractions [from smaller than $0.056\,\mu m$ to greater than $10\,\mu m$] and a bell-shaped distribution with highest overall mass fraction on filters in the fine range. The authors additionally observed that radical signals were size dependent and resulted in reactive oxygen species (ROS) generation.

Recent animal studies suggest that exposure to ultrafine particles may additionally augment cardiac dysfunction through an ROS mechanism [LeBlanc et al., 2010]. LeBlanc et al. recently demonstrated that local ROS generation can influence vascular reactivity in coronary arterioles. It follows that excessive coronary ROS generation following pulmonary woodsmoke exposure could impair endothelium-dependent arteriolar reactivity.

Seven participants reported a history of asthma; four with current asthma. Similar asthma prevalences were previously observed in this population [Gaughan et al., 2008]. We did not see a difference in %-predicted lung function values in these subjects from the rest nor did we observe an association between oxidative stress score or levoglucosan and a history of asthma. However, we did observe that higher LRSS were associated with lower FEV $_1$ /FVC %-predicted after adjusting for asthma history and smoking status (P < 0.05).

We recognize several limitations to our study. Our sample size was small and the resultant lack of statistical power may have hindered our ability to observe associations if they did exist. While we did observe a significant association between higher oxidative stress scores with recent firefighting activities, the generally qualitative and self-reported nature of exposure characterization in our study may have limited our ability to have identified statistically significant cardiopulmonary effects related to firefighting exposures. Additionally, this was a cross-sectional study with essentially no control group. This design limited examination of acute changes. Our study also lacked cross-shift data during periods in which the crews were working but not fighting a fire. Future studies should examine these individuals cross-shift at a wildland fire and when they are not being exposed to smoke aerosol to ascertain whether any observed changes are part of their normal variation. However, a recent study did not find significant changes in cross-shift lung function on burn days compared to non-burn days [Adetona et al., 2011]. Also, urinary levoglucosan may be more representative of diet than as an indicator of wood-smoke exposure. Finally, all spirometric values were obtained pre-bronchodilator. Post-bronchodilator values would have allowed us to examine reversibility and may have been particularly informative for participants reporting a history of asthma.

This is the first study to examine the association between systemic oxidative stress and arterial stiffness in wildland firefighters. Future studies should additionally examine endothelial function which is governed by smooth muscle tone, collage/elastin, calcification, and other factors that affect the entire arterial wall. The two together could provide different but complementary information on vascular health.

Acknowledgments

We thank the Alpine and Pike Interagency Hotshot Crews for their participation as well as the National Interagency Fire Center, Risk Management Committee, specifically, Chad Fisher, U.S. Department of Interior, National Park Service and Larry Sutton, U.S.D.A. Forest Service for arranging for the Interagency Hotshot Crews to participate. The authors additionally thank Dr. Shona Fang, Li Su, and Lauren Cassidy for assistance preparing for the field surveys and Dr. Kristin Cummings for editorial comments.

References

- Adetona O, Hall DB, Naeher LP. Lung function changes in wildland firefighters working at prescribed burns. Inhal Toxicol. 2011; 23:835–841. [PubMed: 22035123]
- Austin C, Wang D, Ecobichon DJ, Dussault G. Characterization of volatile organic compounds in smoke at municipal structural fires. J Toxicol Environ Health. 2001; 63:437–458.
- Bergauff MA, Ward TJ, Noonan CW, Migliaccio CT, Simpson CD, Evanoski AR, Palmer CP. Urinary levoglucosan as a biomarker of wood smoke: Results of human exposure studies. J Expo Sci Environ Epidemiol. 2010; 20:385–392. [PubMed: 19707249]
- Betchley C, Koenig JQ, vanBelle G, Checkoway H, Reinhardt T. Pulmonary function and respiratory symptoms in forest firefighters. Am J Ind Med. 1997; 31:503–509. [PubMed: 9099351]
- Blankenhorn DH, Kramsch DM. Reversal of atherosis and sclerosis. The two components of atherosclerosis. Circulation. 1989; 79:1–7. [PubMed: 2642753]
- Burgess JL, Nanson CJ, Bolstad-Johnson DM, Gerkin R, Hysong TA, Lantz RC, Sherrill DL, Crutchfield CD, Quan SF, Bernard AM, et al. Adverse respiratory effects following overhaul in firefighters. J Occup Environ Med. 2001; 43:467–473. [PubMed: 11382182]
- Cavallari JM, Eisen EA, Fang SC, Schwartz J, Hauser R, Herrick RF, Christiani DC. PM 2.5 metal exposures and nocturnal heart rate variability: A panel study of boilermaker construction workers. Environ Health. 2008; 7:36. [PubMed: 18613971]
- Centers for Disease Control and Prevention [CDC]. Fatalities among volunteer and career firefighters—United States, 1994–2004. CDC MMWR Morb Mortal Wkly Rep. 2006; 55(16):453–455. [PubMed: 16645570]
- Chia KS, Jeyaratnam J, Chan TB, Lim TK. Airway responsiveness of firefighters after smoke exposure. Br J Ind Med. 1990; 47:524–527. [PubMed: 2393631]
- Cipollone F, Ciabattoni G, Patrignani P, Pasquale M, Di Gregorio D, Bucciarelli T, Davì G, Cuccurullo F, Patrono C. Oxidant stress and aspirin-insensitive thromboxane biosynthesis in severe unstable angina. Circulation. 2000; 102:1007–1013. [PubMed: 10961965]
- Delfino RJ, Brummel S, Wu J, Stern H, Ostro B, Lipsett M, Winer A, Street DH, Zhang L, Tjoa T, et al. The relationship of respiratory and cardiovascular hospital admissions to the southern California wildfires of 2003. Occup Environ Med. 2009; 66:189–197. [PubMed: 19017694]
- Dominici F, Peng RD, Bell ML, Pham L, McDermott A, Zeger SL, Samet JM. Fine particulate air pollution and hospital admission for cardiovascular and respiratory diseases. JAMA. 2006; 295:1127–1134. [PubMed: 16522832]
- Duprez DA, Cohn JN. Arterial stiffness as a risk factor for coronary atherosclerosis. Curr Atheroscler Rep. 2007; 9:139–144. [PubMed: 17877923]
- Fang SC, Eisen EA, Cavallari JM, Mittleman MA, Christiani DC. Acute changes in vascular function among welders exposed to metal-rich particulate matter. Epidemiology. 2008; 19:217–225. [PubMed: 18300696]

Fang SC, Cavallari JM, Eisen EA, Chen JC, Mittleman MA, Christiani DC. Vascular function, inflammation, and variations in cardiac autonomic responses to particulate matter among welders. Am J Epidemiol. 2009; 169:848–856. [PubMed: 19153215]

- Ferris BG. Epidemiology standardization project. Am Rev Respir Dis. 1978; 108:1–113. [PubMed: 742764]
- Gaughan DM, Cox-Ganser JM, Enright PL, Castellan RM, Wagner GR, Hobbs GR, Bledsoe TA, Siegel PD, Kreiss K, Weissman DN. Acute upper and lower respiratory effects in wildland firefighters. J Occup Environ Med. 2008; 50:1019–1028. [PubMed: 18784550]
- Gaughan DM, Piacitelli CA, Chen BT, Law BF, Virji MA, Edwards NT, Enright PL, Schwegler-Berry DE, Leonard SS, Wagner GR, et al. Exposures and cross-shift lung function declines in wildland firefighters. J Occup Environ Hyg. 2014 Feb 25. Epub ahead of print. 10.1080/15459624.2014.895372
- Gledhill N, Jamnik VK. Characterization of the physical demands of firefighting. Can J Sport Sci. 1992; 17:207–213. [PubMed: 1325260]
- Guidotti TL, Clough VM. Occupational health concerns of firefighting. Annu Rev Public Health. 1992; 13:151–171. [PubMed: 1599583]
- Hankinson JL, Odencrantz JR, Fedan KB. Spirometric reference values from a sample of the general U.S. population. Am J Respir Crit Care Med. 1999; 159:179–187. [PubMed: 9872837]
- Jarjour NN, Calhoun WJ. Enhanced production of oxygen radicals in asthma. J Lab Clin Med. 1994; 123:131–136. [PubMed: 8288953]
- Kales SN, Soteriades ES, Christoudias SG, Christiani DC. Firefighters and on-duty deaths from coronary heart disease: A case control study. Environ Health. 2003; 2:14. [PubMed: 14613487]
- Kim JY, Mukherjee S, Ngo LC, Christiani DC. Urinary 8-hydroxy-2'-deoxyguanosine as a biomarker of oxidative DNA damage in workers exposed to fine particulates. Environ Health Perspect. 2004; 112:666–671. [PubMed: 15121508]
- LeBlanc AJ, Moseley AM, Chen BT, Frazer D, Castranova V, Nurkiewicz TR. Nanoparticle inhalation impairs coronary microvascular reactivity via a local reactive oxygen species-dependent mechanism. Cardiovasc Toxicol. 2010; 10:27–36. [PubMed: 20033351]
- Leonard SS, Castranova V, Chen BT, Schwegler-Berry D, Hoover M, Piacitelli C, Gaughan DM. Particle size-dependent radical generation from wildland fire smoke. Toxicology. 2007; 236:103–113. [PubMed: 17482744]
- Liu D, Tager IB, Balmes JR, Harrison RJ. The effect of smoke inhalation on lung function and airway responsiveness in wildland firefighters. Am Rev Respir Dis. 1992; 146:1469–1473. [PubMed: 1456562]
- Martinet W, Knaapen MW, De Meyer GR, Herman AG, Kockx MM. Elevated levels of oxidative DNA damage and DNA repair enzymes in human atherosclerotic plaques. Circulation. 2002; 106:927–932. [PubMed: 12186795]
- Materna BL, Jones JR, Sutton PM, Rothman N, Harrison RJ. Occupational exposures in California wildland firefighting. Am Ind Hyg Assoc J. 1992; 53:69–76. [PubMed: 1317093]
- Migliaccio CT, Bergauff MA, Palmer CP, Simpson CD, Evanoski AR, Palmer CP. Urinary levoglucosan as a biomarker of wood smoke exposure: Observations in a mouse model and in children. Environ Health Perspect. 2009; 117:74–79. [PubMed: 19165390]
- Miller MR, Hankinson J, Brusasco V, Burgos F, Casaburi R, Coates A, Crapo R, Enright P, van der Grinten CP, Gustafsson P, et al. Standardisation of spirometry. Eur Respir J. 2005; 26:319–338. [PubMed: 16055882]
- Mott JA, Mannino DM, Alverson CJ, Kiyu A, Hashim J, Lee T, Falter K, Redd SC. Cardiorespiratory hospitalizations associated with smoke exposure during the 1997Southeast Asian forest fires. Int J Hyg Environ Health. 2005; 208:75–85. [PubMed: 15881981]
- Musk AW, Smith TJ, Peters JM, McLaughlin E. Pulmonary function in firefighters: Acute changes in ventilatory capacity and their correlates. Br J Ind Med. 1979; 36:29–34. [PubMed: 444439]
- Public Use Data File Documentation Number 76200. Hyattsville, MD: Centers for Disease Control and Prevention; 1996. NHANES III Adult and Examination Data Files [CD-ROM].

Nuernberg AM, Boyce PD, Cavallari JM, Fang SC, Eisen EA, Christiani DC. Urinary 8- isoprostane and 8-OHdG concentrations in boilermakers with welding exposure. J Occup Environ Med. 2008; 50:182–189. [PubMed: 18301175]

- Nürnberger J, Keflioglu-Scheiber A, OpazoSaez AM, Wenzel RR, Philipp T, Schäfers RF. Augmentation index is associated with cardiovascular risk. J Hypertens. 2002; 20:2407–2414. [PubMed: 12473865]
- Pellegrino R, Viegi G, Brusasco V, Crapo RO, Burgos F, Casaburi R, Coates A, van der Grinten CP, Gustafsson P, Hankinson J, et al. Interpretative strategies for lung function tests. Eur Respir J. 2005; 26:948–968. [PubMed: 16264058]
- Rothman N, Ford DP, Baser ME, Hansen JA, O'Toole T, Tockman MS, Strickland PT. Pulmonary function and respiratory symptoms in wildland firefighters. J Occup Med. 1991; 33:1163–1167. [PubMed: 1765858]
- Sardinas A, Miller JW, Hansen H. Ischemic heart disease mortality of firemen and policemen. Am J Public Health. 1986; 76:1140–1141. [PubMed: 3740340]
- Scannell CH, Balmes JR. Pulmonary effects of firefighting. Occup Med. 1995; 10:789–801. [PubMed: 8903749]
- Simoneit BRT, Schauer JJ, Nolte CG, Oros DR, Elias VO, Fraser MP, Rogge WF, Cass GR. Levoglucosan, a tracer for cellulose in biomass burning and atmospheric particles. Atmos Environ. 1999; 33:173–182.
- Slaughter JC, Koenig JQ, Reinhardt TE. Association between lung function and exposure to smoke among firefighters at prescribed burns. J Occup Environ Hyg. 2004; 1:45–49. [PubMed: 15202156]
- Takeyama H, Itani T, Tachi N, Sakamura O, Murata K, Inoue T, Takanishi T, Suzumura H, Niwa S. Effects of shift schedules on fatigue and physiological functions among firefighters during night duty. Ergonomics. 2005; 48:1–11. [PubMed: 15764302]
- Vallyathan V, Castranova V, Pack D, Leonard S, Shumaker J, Hubbs AF, Shoemaker DA, Ramsey DM, Pretty JR, McLaurin JL. Freshly fractured quartz inhalation leads to enhanced lung injury and inflammation: Potential role of free radicals. Am J Respir Crit Care Med. 1995; 152:1003–1009. [PubMed: 7663775]
- Vlachopoulos C, Aznaouridis K, O'Rourke MF, Safar ME, Baou K, Stefanadis C. Prediction of cardiovascular events and all-cause mortality with central haemodynamics: A systematic review and meta-analysis. Eur Heart J. 2010; 31:1865–1871. [PubMed: 20197424]
- Wasserfallen JB, Gold K, Schulman KA, Baraniuk JN. Development and validation of a rhinoconjunctivitis and asthma symptom score for use as an outcome measure in clinical trials. J Allergy Clin Immunol. 1997; 100:16–22. [PubMed: 9257782]
- Wegesser TC, Pinkerton KE, Last JA. California wildfires of 2008: coarse and fine particulate matter toxicity. Environ Health Perspect. 2009; 117:893–897. [PubMed: 19590679]
- Willett WC, Sampson L, Stampfer MJ, Rosner B, Bain C, Witschi J, Hennekens CH, Speizer FE. Reproducibility and validity of a semiquantitative food frequency questionnaire. Am J Epidemiol. 1985; 122:51–65. [PubMed: 4014201]
- Yoo HL, Franke WD. Prevalence of cardiovascular disease risk factors in volunteer firefighters. J Occup Environ Med. 2009; 51:958–962. [PubMed: 19620889]

Page 13

TABLE IDemographic Characteristics of Interagency Hotshot Crew Members

Variable	All (N =38)	Alpine interagency hotshot crew (n =18)	Pike interagency hotshot crew (n =20)
Age (years) ^a	29 (4.34)	29.9 (5.44)	28.2 (2.97)
Time spent as a firefighter (years) a	3.65 (3.38)	4.12 (3.89)	3.25 (2.92)
Male %	100	100	100
White, non-Hispanic n, %	34 (90)	15 (83)	19 (95)
Current smoker n, %	2 (5)	1 (6)	1(5)
Former smoker n, %	10 (27)	2 (11)	9 (45)
Current chewing tobacco (yes vs. no) n, %	16 (42)	7 (39)	9 (45)
Volunteer firefighter n, %	6 (16)	5 (28)	1 (5)
Permanent employee n, %	11 (29)	5 (28)	6 (30)

 $^{^{}a}\mathrm{Mean}$ value (standard deviation).

Gaughan et al.

Gaughan et al.

TABLE II

Clinical Characteristics of Interagency Hotshot Crew Members

Variable	All (N =38)	Alpine interagency hotshot crew (n =18)	Pike interagency hotshot crew (n =20)
Augmentation index % (adjusted for heart rate to 75 BPM) a	5 (13)	10 (11)	1.0 (13) ^b
Hypertension (ever) n, %	3 (8)	2 (11)	1 (5)
Elevated cholesterol (ever) n, %	4 (10)	3 (17)	1 (5)
Total cholesterol $(mg/dl)^a$	170 (35.8)	175 (40.1)	164 (28.4)
HDL-cholesterol $(mg/dl)^a$	55.3 (10.6)	54.6 (11.4)	56.25 (9.81)
LDL-cholesterol (mg/dl) a	93.8 (28.1)	101 (32.1)	82.9 (16.5)
Triglycerides (mg/dl) ^a	109 (61.4)	97.6 (39.1)	125.8 (84.0)
HsCRP (mg/L) ^a	1.25 (1.95)	1.23 (2.11)	1.28 (1.78)
Fibrinogen (mg/dl) ^a	270.2 (60.2)	295 (41.2)	221 (64.3)
$\log_{10} 8$ -isoprostane $(ng/ml)^a$	-0.14 (0.42)	-0.20 (0.41)	-0.08 (0.42)
\log_{10} 8-OHdG (ng/ml) a,c	0.75 (0.36)	0.61 (0.39)	0.88 (0.29) ^b
Oxidative stress score ^{a,d}	0.41 (0.45)	0.48 (0.45)	0.34 (0.45)
\log_{10} levoglucosan (µg/ml) a	1.17 (0.31)	1.14 (0.25)	1.20 (0.37)
Asthma (ever) n, %	7 (18)	6 (33)	1 (5)
Allergies (ever) n, %	17 (45)	11 (61)	6 (30)
Upper respiratory symptom score a,e	7.63 (7.1)	7.28 (7.11)	7.95 (7.21)
Lower respiratory symptom score af	2.34 (3.87)	1.56 (2.52)	3.05 (4.73)
Pulmonary function ^a			
FEV ₁ %-predicted	103 (10.2)	101 (10.1)	104 (10.5)
FVC%-predicted	107 (13.0)	104 (11.4)	111 (13.7)
FEV ₁ /FVC	95.6 (9.14)	97.1 (9.80)	94.3 (8.53)

Page 14

 $^{^{}a}$ Mean value and standard deviation.

b Significantly different from Alpine Interagency Hotshot Crew at $P\text{-}\mathrm{value}$ $<\!0.05.$

^c8-hydroxy-2'-deoxyguanosine (8-OHdG).

 $[\]begin{tabular}{l} d Derived from the average of z-scores for 8-isoprostane and 8-hydroxy-2'-deoxyguanosine (8-OHdG). \end{tabular}$

 $^{^{}e}$ Range 0–24; higher score denotes more frequent symptoms.

 $f_{\text{Range 0-19}}$; higher score denotes more frequent symptoms.

TABLE III

Predictors of Arterial Stiffness (Augmentation Index %), Linear Regression Estimates and 95% CIs

Variable ^a	Unadjusted	${\bf Adjusted}^{\pmb{b}}$
Smoking history (ever)	11.2 (2.81, 19.7)	11.5 (3.73, 19.3)
Oxidative stress score ^C	10.2 (1.35, 19.0)	10.5 (2.51, 18.5)

^aAge, chewing tobacco status, HDL-cholesterol, LDL-cholesterol, triglycerides, high sensitivity CRP, fibrinogen, allergy history, lower respiratory symptom score, upper respiratory symptom score, Interagency Hotshot Crew, FEV₁ %-predicted, FVC %-predicted, and FEV₁/FVC %-predicted were not significantly associated with mean augmentation index % values.

 $[^]b{\rm Estimates}$ controlling for all other specified variables in the Table III.

 $[^]c\mathrm{Average}$ z-scores of \log_{10} values of 8-hydroxy-2′-deoxyguanosine (8-OHdG) and 8-isoprostane.

TABLE IV

Predictors of Oxidative DNA Damage and Repair (8-OHdG^a), Linear Regression Estimates and 95% CIs

$\operatorname{Log_{10}}$ 8-OHdG a,b	Unadjusted	Adjusted ^c
Log_{10} levoglucosan d concentration	0.45 (0.07, 0.84)	0.41 (0.04, 0.79)
Interagency Hotshot Crew	0.27 (0.05, 0.50)	0.21 (-0.02, 0.45)

 $[^]a$ 8-hydroxy-2'-deoxyguanosine (8-OHdG).

^bSmoking/chewing tobacco status, hsCRP, HDL-cholesterol, LDL-C, triglycerides, fibrinogen, asthma history, allergy history, lower respiratory symptom score, upper respiratory symptom score, FEV1%-predicted FVC %-predicted, and FEV1/FVC %-predicted, and age were not significantly associated with mean log10 8-hydroxy-2'-deoxyguanosine (8-OHdG) values.

^cEstimates controlling for all other specified variables in the table.

 $^{^{}d}{\rm Levoglucosan\ (LG)}.$