CENTERS FOR DISEASE CONTROL

Current Trends



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Dental personnel may be exposed to a wide variety of microorganisms in the blood and saliva of patients they treat in the dental operatory. These include *Mycobacterium tuberculosis*, hepatitis B virus, staphylococci, streptococci, cytomegalovirus, herpes simplex virus types I and II, human T-lymphotropic virus type III/lymphadenopathy-associated virus (HTLV-III/LAV), and a number of viruses that infect the upper respiratory tract. Infections may be transmitted in dental practice by blood or saliva through direct contact, droplets, or aerosols. Although not documented, indirect contact transmission of infection by contaminated instruments is possible. Patients and dental health-care workers (DHCWs) have the potential of transmitting infections to each other (1).

**Recommended Infection-Control Practices for Dentistry** 

A common set of infection-control strategies should be effective for preventing hepatitis B, acquired immunodeficiency syndrome, and other infectious diseases caused by bloodborne viruses (2-4). The ability of hepatitis B virus to survive in the environment (5) and the high titers of virus in blood (6) make this virus a good model for infection-control practices to prevent transmission of a large number of other infectious agents by blood or saliva. Because all infected patients cannot be identified by history, physical examination, or readily available laboratory tests (3), the following recommendations should be used routinely in the care of all patients in dental practices.

# MEDICAL HISTORY

Always obtain a thorough medical history. Include specific questions about medications, current illnesses, hepatitis, recurrent illnesses, unintentional weight loss, lymphadenopathy, oral soft tissue lesions, or other infections. Medical consultation may be indicated when a history of active infection or systemic disease is elicited.

# **USE OF PROTECTIVE ATTIRE AND BARRIER TECHNIQUES**

1. For protection of personnel and patients, gloves must always be worn when touching blood, saliva, or mucous membranes (7-10). Gloves must be worn by DHCWs when touching blood-soiled items, body fluids, or secretions, as well as surfaces contaminated with them. Gloves must be worn when examining all oral lesions. All work must be completed on one patient, where possible, and the hands must be washed and regloved before performing procedures on another patient. Repeated use of a single pair of gloves is not recommended, since such use is likely to produce defects in the glove material, which will diminish its value as an effective barrier.

2. Surgical masks and protective eyewear or chin-length plastic face shields must be worn when splashing or spattering of blood or other body fluids is likely, as is common in dentistry (11, 12).

## U.S. DEPARTMENT OF HEALTH AND HUMAN SERVICES / PUBLIC HEALTH SERVICE

# Infection Control - Continued

3. Reusable or disposable gowns, laboratory coats, or uniforms must be worn when clothing is likely to be soiled with blood or other body fluids. If reusable gowns are worn, they may be washed, using a normal laundry cycle. Gowns should be changed at least daily or when visibly soiled with blood (13).

4. Impervious-backed paper, aluminum foil, or clear plastic wrap may be used to cover surfaces (e.g., light handles or x-ray unit heads) that may be contaminated by blood or saliva and that are difficult or impossible to disinfect. The coverings should be removed (while DHCWs are gloved), discarded, and then replaced (after ungloving) with clean material between patients.

5. All procedures and manipulations of potentially infective materials should be performed carefully to minimize the formation of droplets, spatters, and aerosols, where possible. Use of rubber dams, where appropriate, high-speed evacuation, and proper patient positioning should facilitate this process.

# HANDWASHING AND CARE OF HANDS

Hands must always be washed between patient treatment contacts (following removal of gloves), after touching inanimate objects likely to be contaminated by blood or saliva from other patients, and before leaving the operatory. The rationale for handwashing after gloves have been worn is that gloves become perforated, knowingly or unknowingly, during use and allow bacteria to enter beneath the glove material and multiply rapidly. For many routine dental procedures, such as examinations and nonsurgical techniques, handwashing with plain soap appears to be adequate, since soap and water will remove transient microorganisms acquired directly or indirectly from patient contact (13). For surgical procedures, an antimicrobial surgical handscrub should be used (14). Extraordinary care must be used to avoid hand injuries during procedures. However, when gloves are torn, cut, or punctured, they must be removed immediately, hands thoroughly washed, and regloving accomplished before completion of the dental procedure. DHCWs who have exudative lesions or weeping dermatitis should refrain from all direct patient care and from handling dental patient-care equipment until the condition resolves (15).

# **USE AND CARE OF SHARP INSTRUMENTS AND NEEDLES**

1. Sharp items (needles, scalpel blades, and other sharp instruments) should be considered as potentially infective and must be handled with extraordinary care to prevent unintentional injuries.

2. Disposable syringes and needles, scalpel blades, and other sharp items must be placed into puncture-resistant containers located as close as practical to the area in which they were used. To prevent needlestick injuries, disposable needles should not be recapped; purposefully bent or broken; removed from disposable syringes; or otherwise manipulated by hand after use.

3. Recapping of a needle increases the risk of unintentional needlestick injury. There is no evidence to suggest that reusable aspirating-type syringes used in dentistry should be handled differently from other syringes. Needles of these devices should not be recapped, bent, or broken before disposal.

4. Because certain dental procedures on an individual patient may require multiple injections of anesthetic or other medications from a single syringe, it would be more prudent to place the unsheathed needle into a "sterile field" between injections rather than to recap the needle between injections. A new (sterile) syringe and a fresh solution should be used for each patient.

# Infection Control - Continued

# INDICATIONS FOR HIGH-LEVEL DISINFECTION OR STERILIZATION OF INSTRUMENTS

Surgical and other instruments that normally penetrate soft tissue and/or bone (e.g., forceps, scalpels, bone chisels, scalers, and surgical burs) should be sterilized after each use. Instruments that are not intended to penetrate oral soft tissues or bone (e.g., amalgam condensers, plastic instruments, and burs) but that may come into contact with oral tissues should also be sterilized after each use, if possible; however, if sterilization is not feasible, the latter instruments should receive high-level disinfection (*3*, *13*, *16*).

# METHODS FOR HIGH-LEVEL DISINFECTION OR STERILIZATION

Before high-level disinfection or sterilization, instruments should be cleaned to remove debris. Cleaning may be accomplished by a thorough scrubbing with soap and water or a detergent, or by using a mechanical device (e.g., an ultrasonic cleaner). Persons involved in cleaning and decontaminating instruments should wear heavy-duty rubber gloves to prevent hand injuries. Metal and heat-stable dental instruments should be routinely sterilized between use by steam under pressure (autoclaving), dry heat, or chemical vapor. The adequacy of sterilization cycles should be verified by the periodic use of spore-testing devices (e.g., weekly for most dental practices) (13). Heat- and steam-sensitive chemical indicators may be used on the outside of each pack to assure it has been exposed to a sterilizing cycle. Heat-sensitive instruments may require up to 10 hours' exposure in a liquid chemical agent registered by the U.S. Environmental Protection Agency (EPA) as a disinfectant/sterilant; this should be followed by rinsing with sterile water. High-level disinfection may be accomplished by immersion in either boiling water for at least 10 minutes or an EPA-registered disinfectant/sterilant chemical for the exposure time recommended by the chemical's manufacturer.

# DECONTAMINATION OF ENVIRONMENTAL SURFACES

At the completion of work activities, countertops and surfaces that may have become contaminated with blood or saliva should be wiped with absorbent toweling to remove extraneous organic material, then disinfected with a suitable chemical germicide. A solution of sodium hypochlorite (household bleach) prepared fresh daily is an inexpensive and very effective germicide. Concentrations ranging from 5,000 ppm (a 1:10 dilution of household bleach) to 500 ppm (a 1:100 dilution) sodium hypochlorite are effective, depending on the amount of organic material (e.g., blood, mucus, etc.) present on the surface to be cleaned and disinfected. Caution should be exercised, since sodium hypochlorite is corrosive to metals, especially aluminum.

# DECONTAMINATION OF LABORATORY SUPPLIES AND MATERIALS

Blood and saliva should be thoroughly and carefully cleaned from laboratory supplies and materials that have been used in the mouth (e.g., impression materials, bite registration), especially before polishing and grinding intra-oral devices. Materials, impressions, and intra-oral appliances should be cleaned and disinfected before being handled, adjusted, or sent to a dental laboratory (17). These items should also be cleaned and disinfected when returned from the dental laboratory and before placement in the patient's mouth. *Because of the ever-increasing variety of dental materials used intra-orally, DHCWs are advised to consult with manufacturers as to the stability of specific materials relative to disinfectant"* and that has a label claim for mycobactericidal (e.g., tuberculocidal) activity is preferred, because mycobacteria represent one of the most resistant groups of microorganisms; therefore, germicides that are effective against mycobacteria are also effective against other bacterial and viral pathogens (15). Communication between a dental office and a dental laboratory with regard to handling and decontamination of supplies and materials is of the utmost importance.

# Infection Control - Continued

## USE AND CARE OF ULTRASONIC SCALERS, HANDPIECES, AND DENTAL UNITS

1. Routine sterilization of handpieces between patients is desirable; however, not all handpieces can be sterilized. The present physical configurations of most handpieces do not readily lend them to high-level disinfection of both external and internal surfaces (see 2 below); therefore, when using handpieces that cannot be sterilized, the following cleaning and disinfection procedures should be completed between each patient: After use, the handpiece should be flushed (see 2 below), then thoroughly scrubbed with a detergent and water to remove adherent material. It should then be thoroughly wiped with absorbent material saturated with a chemical germicide that is registered with the EPA as a "hospital disinfectant" and is mycobactericidal at use-dilution (*15*). The disinfecting solution should remain in contact with the handpiece for a time specified by the disinfectant's manufacturer. Ultrasonic scalers and air/water syringes should be treated in a similar manner between patients. Following disinfection, any chemical residue should be removed by rinsing with sterile water.

2. Because water retraction valves within the dental units may aspirate infective materials back into the handpiece and water line, check valves should be installed to reduce the risk of transfer of infective material (18). While the magnitude of this risk is not known, it is prudent for water-cooled handpieces to be run and to discharge water into a sink or container for 20-30 seconds after completing care on each patient. This is intended to physically flush out patient material that may have been aspirated into the handpiece or water line. Additionally, there is some evidence that overnight bacterial accumulation can be significantly reduced by allowing water-cooled handpieces to run and to discharge water into a sink or container for several minutes at the beginning of the clinic day (19). Sterile saline or sterile water should be used as a coolant/irrigator when performing surgical procedures involving the cutting of soft tissue or bone.

#### HANDLING OF BIOPSY SPECIMENS

In general, each specimen should be put in a sturdy container with a secure lid to prevent leaking during transport. Care should be taken when collecting specimens to avoid contamination of the outside of the container. If the outside of the container is visibly contaminated, it should be cleaned and disinfected, or placed in an impervious bag (20).

## **DISPOSAL OF WASTE MATERIALS**

All sharp items (especially needles), tissues, or blood should be considered potentially infective and should be handled and disposed of with special precautions. Disposable needles, scalpels, or other sharp items should be placed intact into puncture-resistant containers before disposal. Blood, suctioned fluids, or other liquid waste may be carefully poured into a drain connected to a sanitary sewer system. Other solid waste contaminated with blood or other body fluids should be placed in sealed, sturdy impervious bags to prevent leakage of the contained items. Such contained solid wastes can then be disposed of according to requirements established by local or state environmental regulatory agencies and published recommendations (*13,20*).

## Developed by Dental Disease Prevention Activity, Center for Prevention Svcs, Hospital Infections Program, Center for Infectious Diseases, CDC.

Editorial Note: All DHCWs must be made aware of sources and methods of transmission of infectious diseases. The above recommendations for infection control in dental practices incorporate procedures that should be effective in preventing the transmission of infectious agents from dental patients to DHCWs and vice versa. Assessment of quantifiable risks to dental personnel and patients for specific diseases requires further research. There is no current documentation of patient-to-patient blood- or saliva-borne disease transmission from

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procedures performed in dental practice. While few in number, reported outbreaks of dentistto-patient transmission of hepatitis B have resulted in serious and even fatal consequences (9). Herpes simplex virus has been transmitted to over 20 patients from the fingers of a DHCW (10). Serologic markers for hepatitis B in dentists have increased dramatically in the United States over the past several years, which suggests current infection-control practices have been insufficient to prevent the transmission of this infectious agent in the dental operatory. While vaccination for hepatitis B is strongly recommended for dental personnel (21), vaccination alone is not cause for relaxation of strict adherence to accepted methods of asepsis, disinfection, and sterilization.

Various infection-control guidelines exist for hospitals and other clinical settings. Dental facilities located in hospitals and other institutional settings have generally utilized existing guidelines for institutional practice. These recommendations are offered as guidance to DHCWs in noninstitutional settings for enhancing infection-control practices in dentistry; they may be useful in institutional settings also.

#### References

- 1. Ahtone J, Goodman RA. Hepatitis B and dental personnel: transmission to patients and prevention issues. J Am Dent Assoc 1983;106:219-22.
- 2. Crawford JJ. State-of-the-art: practical infection control in dentistry. J Am Dent Assoc 1985;110: 629-33.
- 3. Cottone JA, Mitchell EW, Baker CH, et al. Proceedings of the National Symposium on Hepatitis B and the Dental Profession. J Am Dent Assoc 1985;110:614-49.
- CDC. Acquired immunodeficiency syndrome (AIDS): precautions for health-care workers and allied professionals. MMWR 1983;32:450-1.
- 5. Bond WW, Favero MS, Petersen NJ, Gravelle CR, Ebert JW, Maynard JE. Survival of hepatitis B virus after drying and storage for one week [Letter]. Lancet 1981;1:550-1.
- 6. Shikata T, Karasawa T, Abe K, et al. Hepatitis B e antigen and infectivity of hepatitis B virus. J Infect Dis 1977;136:571-6.
- Hadler SC, Sorley DL, Acree KH, et al. An outbreak of hepatitis B in a dental practice. Ann Intern Med 1981;95:133-8.
- Occupational Safety and Health Administration. Risk of hepatitis B infection for workers in the health care delivery system and suggested methods for risk reduction. U.S. Department of Labor 1983; (CPL 2-2.36).
- 9. CDC. Hepatitis B among dental patients Indiana. MMWR 1985;34:73-5.
- Manzella JP, McConville JH, Valenti W, Menegus MA, Swierkosz EM, Arens M. An outbreak of herpes simplex virus type 1 gingivostomatitis in a dental hygiene practice. JAMA 1984;252: 2019-22.
- 11. Petersen NJ, Bond WW, Favero MS. Air sampling for hepatitis B surface antigen in a dental operatory. J Am Dent Assoc 1979;99:465-7.
- 12. Bond WW, Petersen NJ, Favero MS, Ebert JW, Maynard JE. Transmission of type B viral hepatitis B via eye inoculation of a chimpanzee. J Clin Microbiol 1982;15:533-4.
- 13. Garner JS, Favero MS. Guideline for handwashing and hospital environmental control, 1985. Atlanta, Georgia: Centers for Disease Control, 1985; publication no. 99-1117.
- 14. Garner JS. Guideline for prevention of surgical wound infections, 1985. Atlanta, Georgia: Centers for Disease Control, 1985; publication no. 99-2381.
- 15. CDC. Recommendations for preventing transmission of infection with human T-lymphotropic virus type III/lymphadenopathy-associated virus in the workplace. MMWR 1985;34:682-6, 691-5.
- Favero MS. Sterilization, disinfection, and antisepsis in the hospital. In: Lennette EH, Balows A, Hauslen WJ, Shadomy HJ. Manual of clinical microbiology. Washington, D.C.: American Society of Microbiology, 1985:129-37.
- Council on Dental Therapeutics and Council on Prosthetic Services and Dental Laboratory Relations, American Dental Association. Guidelines for infection control in the dental office and the commercial laboratory. J Am Dent Assoc 1985;110:969-72.

# Infection Control - Continued

- 18. Bagga BSR, Murphy RA, Anderson AW, Punwani I. Contamination of dental unit cooling water with oral microorganisms and its prevention. J Am Dent Assoc 1984;109:712-6.
- 19. Scheid RC, Kim CK, Bright JS, Whitely MS, Rosen S. Reduction of microbes in handpieces by flushing before use. J Am Dent Assoc 1982;105:658-60.
- Garner JS, Simmons BP. CDC guideline for isolation precautions in hospitals. Atlanta, Georgia: Centers for Disease Control, 1983; HHS publication no. (CDC) 83-8314.
- 21. ACIP. Inactivated hepatitis B virus vaccine. MMWR 1982;31:317-22, 327-8.

		15th Week End	ing	Cumulative, 15th Week Ending				
Disease	Apr. 12, 1986	Apr. 13, 1985	Median 1981-1985	Apr. 12, 1986	Apr. 13, 1985	Median 1981-1985		
Acquired Immunodeficiency Syndrome (AIDS)	380	199	N	3,580	1.878	N		
Aseptic meningitis	82	71	58	1,207	1.039	1,144		
Encephalitis: Primary (arthropod-borne				.,	1,000	.,		
& unspec.)	14	21	19	235	271	256		
Post-infectious	2	3	1	25	38	25		
Gonorrhea: Civilian	15.880	15,194	15.194	224,979	223.900	256.543		
Military	221	276	355	4,457	5.335	7.059		
Hepatitis: Type A	405	407	416	6,437	6,114	6,575		
Туре В	498	536	434	7.042	7,139	6.532		
Non A, Non B	54	88	N	932	1,195	N		
Unspecified	95	123	127	1,429	1,482	2.093		
Legionellosis	7	8	N	157	173	N		
Leprosy	7	6	6	78	114	58		
Malaria	12	11	18	201	192	198		
Measles: Total*	167	104	58	1,607	721	721		
Indigenous	165	96	Ň	1,565	597	N		
Imported	2	8	N	42	124	N		
Meningococcal infections: Total	66	48	67	959	894	1.041		
Civilian	66	48	67	957	893	1.040		
Military	-	-	-	2	1	4		
Mumps	137	62	101	970	1,194	1,254		
Pertussis	23	35	35	545	436	436		
Rubella (German measles)	11	11	34	145	115	344		
Syphilis (Primary & Secondary): Civilian	420	433	540	6,821	7,051	8,771		
Military	10	3	7	68	53	109		
Toxic Shock syndrome	9	6	N	95	108	N		
Tuberculosis	368	433	489	5,458	5,361	6,258		
Tularemia	1	-	2	18	24	27		
Typhoid fever	4	8	8	64	74	106		
Typhus fever, tick-borne (RMSF)	2	5	5	18	19	22		
Rabies, animal	110	107	158	1,400	1,302	1,579		

#### TABLE I. Summary-cases specified notifiable diseases, United States

#### TABLE II. Notifiable diseases of low frequency, United States

	Cum 1986		Cum 1986
Anthrax	•	Leptospirosis	13
Botulism: Foodborne	3	Plague	-
Infant (Calif. 1)	15	Poliomyelitis, Paralytic	
Other	-	Psittacosis	16
Brucellosis (Iowa 1)	14	Rabies, human	-
Cholera		Tetanus	1 11
Congenital rubella syndrome	1 1	Trichinosis	7
Congenital syphilis, ages < 1 year Diphtheria	11	Typhus fever, flea-borne (endemic, murine)	5

\*There were no cases of internationally imported measles reported for this week.

	Aseptic Encephalitis Hepatitis (Viral), by type											
	AIDS	Aseptic Menin-	Primary	Post-in-	Gon (Cir	orrhea vilian)	A	epatitis (V B	NA,NB	Unspeci-	Legionel- losis	Leprosy
Reporting Area	Cum.	gitis 1986	Cum.	fectious Cum.	Cum.	Cum.				fied	1986	Cum.
	1986		1986	1986	1986	1985	1986	1986	1986	1986		1986
UNITED STATES	3,580	82	235	25	224,979	223,900	405	498	54	95	7	78
NEW ENGLAND Maine	155 9	2	9	1	5,239 278	7,080 288	18 1	31 2	2	2	-	1
N.H. Vt.	7 2	-	2 2	1	148 83	151 63	1	:	-	:	-	-
Mass R I	92 9	1	2	-	2,240 487	2,621 524	10 1	19 2	1	2	-	1
Conn	36	1	3	-	2,003	3,433	5	8	1	-	-	-
MID ATLANTIC Upstate N.Y.	1,337 118	5 3	38 12	-	41,852 4,519	30,976 4,332	12 5	30 6	2	38	-	7
N.Y. City	908	1	10	-	24,599	14,072	3 4	7	2	32 6	-	7
N.J. Pa	223 88	1	5 11	-	5,721 7,013	6,342 6,230	4	-	-	-	-	-
EN CENTRAL	176	9	47	4	27,554	32,404	19 7	73 28	8 1	3	2 1	4
Ohio Ind	30 24	4	15 5	2	7,715 4,299	8,200 3,061	1	7	-	-	-	-
III. Mich	71 46	1 3	6 20	2	4,074 9,706	9,243 9,380	7 4	16 22	1 6	3	1	3 1
Wis	5	-	1	-	1,760	2,520	-	-	•	-	-	-
W.N. CENTRAL Minn	74 33	4 1	6 4	5	10,395 1,561	11,487 1,701	8 1	19 · 2	2 1	-	1	1
lowa Mo	8 20	2	2	-	1,043 5,159	1,221 5,298	1	3 10	1	-	1	-
N Dak	2	-	-		103	84. 208	• 1	1	-	-	-	-
S. Dak Nebr	3	1	-	- 5	696	1,136 1,839	4	1	-	-	-	-
Kans	7 481	19	39	5 11	1,624 50,318	48,254	32	2 84	7	7	1	1
S ATLANTIC Del	9	-	3	-	965	1,071	-	1	, 1	-	-	÷
Md D C	45 69	6	10	-	7,214 4,480	7,723 4,036	6 3	25 1	i	1	-	-
Va W Va	51 2	2	14 6	-	5,068 718	5,170 659	1	4 2,	-	2	-	1
N C S C	21 14	1	5	1	9,949 5,328	9,203 6,083	4 	11 11 ·	1	1	-	-
Ga Fla	64 206	10	1	10	16,596	14,309	4 14	13 16	1 2	- 3	1	-
E.S. CENTRAL	32	1	18	1	19,564	19,645	5	44 ·	2	1	-	-
Ky Tenn	10 13	1	8 1	1	2,318 7,490	2,196 7,676	1 2	9 28	1	1	-	-
Ala Miss	5	-	9	-	5,605 4,151	6,119 3,654	1 1	6, 1	:	2	-	-
W S CENTRAL	304	13	19	-	28,854	31,341	52	34 •	2 ·	24	2	5
Ark La	9 42	- 1	2	-	2,666 5,015	2,993 6,522	-3	- 8,	-	1	-	-
Okla Tex	16 237	12	4 13	-	3,334 17,839	3,199 18,627	6. 43	• 5 21	2	22	1 1	- 5
MOUNTAIN	82	1	11	1	7,580	7,281	25	20	5	4	-	7
Mont. Idaho	1	-	-	1	184 232	222 244	1	1	-	1	-	-
Wyo	2	-	2	-	172 1,929	199 2.191	5	-	2	1	-	3
Colo. N Mex.	36	-	1	-	780	846	3	i	1	1	-	2
Ariz. Utah	21 6	1	4	-	2,356 308	2,143 297	8 5	10 1	2	-	-	-
Nev	9	-	1	-	1,619	1,139	3	2	-	-	-	2
PACIFIC Wash	939 34	28 2	48 5	2	33,623 2,506	35,432 2,553	234 16	163 7	24	16	1	52 6
Oreg. Calif.	18 869	25	41	2	1,316 28,501	1,847 29,591	40 176	13 138	1 23	15	1	40
Alaska Hawaii	8 10	1	2	2	931 369	871 570	1	1		1	-	6
Guam	-	-	-		28	52	-		-	-	-	1
P.R. V.I.	32	-	2		651 66	1,109 130	4	5	1	1	-	:
Pac. Trust Terr Amer. Samoa	-	-	-	:	42 12	235	1	1	-	-	-	1
					12							

# TABLE III. Cases of specified notifiable diseases, United States, weeks ending April 12, 1986 and April 13, 1985 (15th Week)

N Not notifiable

			~		1300		pril 13, 1	305	1501	TOOR					
Paparting Area	Malaria	Meas Indigenous		_	sles (Rubeola)		Menin- gococcal Infections	Mumps		Pertussis			Rubella		
Reporting Area	Cum. 1986	1986	Cum. 1986	1986	Cum. 1986	Cum. 1985	Cum. 1986	1986	Cum. 1986	1986	Cum. 1986	Cum. 1985	1986	Cum. 1986	Cum. 1985
UNITED STATES	S 201	165	1,565	2	42	721	959	137	970	23	545	436	11	145	115
NEW ENGLAND	11	1	10	-	-	40	69	-	25	3	38	21	-	1	5
Maine N.H.	-	2	-	-	-	:	14 3	:	- 5	-	2 14	2 13	-	1	2
Vt.	1	-	-	-	-		9	-	-	-	1	2	-	:	-
Mass: R.I.	6 1	1	9 1	-		40	14 10	-	1 4	-	9 1	3 1	-	-	3
Conn.	3	-	-	-	-	-	19	-	15	3	11	-	-	-	-
MID ATLANTIC	25	116	616	-	3	53	163	3	55	1	73	54	-	23	29
Upstate N.Y. N.Y. City	3 8	1 16	2 91	-	2 1	27 20	45 36	-	21 5	1	48 3	30 7	-	15 5	777
N.J.	3	99	523	-	-	6	27	3	13	-	4	í	-	3	3
Pa.	11	-	-	-	•	-	55	-	16	-	18	16	-	-	12
E.N. CENTRAL	5	10	158	-	2	243	115	95	484	4	131	66	з	4	9
Ohio Ind.	1	-	-	:		12	54 10	4	53 15	1 2	62 16	13 11	-	-	-
III.	2	10	89	-	-	139	27	87	276	-	15	12	3	3	3
Mich. Wis.	2	-	69		- 2	48 43	24	4	64 76	1	14 24	7 23	-	1	5 1
	_						-	-		-					
W.N. CENTRAL Minn.	5 2	8	78	1	1	4	52 12	3	45 1	2	32 15	38 11	1	5	7
lowa	1	-	-		-	-	6	1	7	1	5	2	-	-	
Mo. N. Dak.	2	:	-	1 §	1	2	18	2	9 2	1	4 2	8	-	1	•
S. Dak.	-	-	-	-	-	-	-		1	-	-	6	-	-	:
Nebr. Kans.	-	8	78	:	-	1	6 10	-	25	-	- 6	1 10	1	4	;
S. ATLANTIC								-		•			•		
Del.	25	21	240	-	4	101	206 1	7	71	3	102	106	-	6	7
Md.	3	3	9	-	2	4	28	-	4	-	21	34	-	-	1
D.C. Va.	6	-		-	-	1 12	2 37	-	- 9	-	- 9	2	-	-	•
W. Va. N.C.	-	2	2	-	-	2	3	2	25	2	4	-		-	
S.C.	3	16	218	-			32 24	3 2	7 9	-	15 2	7	-	-	2
Ga. Fla.	3	-	-	-	1	8	30	-	5	1	43	45	-	-	-
	8	-	11	-	1	74	49	-	12	-	8	18	-	6	4
E.S. CENTRAL Ky.	4	1	1	-	-	-	50	6	13	-	15	4	-	1	1
Tenn.	2	1	1	-	-	-	9 23	6	2	-	1 5	1	-	1	1
Ala. Miss.	2	-	-	-	-	-	13	-	ĩ	-	9	2	-	-	-
	-	-	-	-	-	-	5	-	1	-	-	-	-	-	-
W.S. CENTRAL Ark.	17	2	287 265	-	12	23	71	2	70	-	24	40	3	30	13
La.	4	-	265	-	-	1	8 8	2	6	-	1 3	8	-	-	1
Okla. Tex.	2 11	2	2 20	-	12	22	12	N	N	-	20	31	-		-
		2		•			43	2	64	-	-	-	3	30	12
MOUNTAIN Mont.	5	-	48	:	6 1	175 120	36 4	7	101 2	2	70	21	-	-	3
Idaho	1	-	-	-	-	20	1	-	2	-	15	3	:	-	ī
Wyo. Colo.	1	-	-	•	3	-3	2 7	1	- 5	-	-	-	-	-	-
N. Mex.	-	-	15	:	2	1	4	Ň	Ň	-	16 8	8 3	:	-	i
Ariz. Utah	2	-	33		-	49	12	6	88	2	23	3	-	-	i
Nev.	1	-	-		-	-	3	-	1 3		8	4	:	-	-
PACIFIC	104	6	127	1	14	82	197	14	106	8	60	86		76	
Wash. Oreg.	9	-	23	-	7	1	28	-	4	2	25	12	4	75 1	41
Calif.	8 87	3	- 85	1 §	2 5	2 72	15 148	N 13	N 94	1 5	3 29	16	-	-	1
Alaska Hawaii	-	-	-	-	-	-	5	-	2	-	1	53 2	3	73	32
	-	3	19	•	-	7	1	1	6	-	2	3	1	1	8
Guam P.R.	1	1	3	-	-	10	-	1	2	-	-	-	-	2	1
V.I.	1		-		-	40 9	2	1	15 6	1	3	1	:	-	4
Pac. Trust Terr. Amer. Samoa	-	-	-	-	-	-	-	-	-		-	-	-	-	-
cinel Jamoa	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-

# TABLE III. (Cont'd.) Cases of specified notifiable diseases, United States, weeks ending April 12, 1986 and April 13, 1985 (15th Week)

\*For measles only, imported cases includes both out-of-state and international importations. §Out-of-state

N. Not notifiable U: Unavailable

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		April	12, 1986 a	nd April	13, 1985	(15th We	ek)		
Reporting Area	Syphilis (Civilian) (Primary & Secondary)		Toxic- shock Syndrome	Tuber	culosis	Tula- remia	Typhoid Fever	Typhus Fever (Tick-borne) (RMSF)	Rabiès, Animal
	Cum. 1986	Cum. 1985	1986	Cum. 1986	Cum. 1985	Cum. 1986	Cum. 1986	Cum. 1986	Cum. 1986
UNITED STATES	6,821	7,051	9	5,458	5,361	18	64	18+3	1,400
NEW ENGLAND Maine	140	158	2	167	191	-	3	1	1
N.H.	10 6	5 3	2	18 3	16 6	-	-	-	-
Vt. Mass.	6 67	87	-	7 82	3	-	-	:	-
R.I.	8	5	-	82 11	115 16	-	2	1	- 1
Conn.	43	58	-	46	35	-	1	-	-
MID ATLANTIC	1,003	949	-	1,087	1,046	-	7	1	118
NY. City	51 570	65 606		174 526	157 561	-	1 4	1	20
N.J.	196	206	-	197	92	-	2	-	-
Pa.	186	72	-	190	236	-	-	-	98
E.N. CENTRAL	175	339	2	700	674	-	4	-	27
nd.	34 40	34 29	2	103 86	128 83		:	-	2 7
N.	39	178	-	304	300	-	-	-	8
Mich. Nis.	43 19	82 16	-	167 40	128 35	•	3 1	-	3 7
		75	-		141		3	1+1	, 184
N N CENTRAL	67 8	75 19		156 36	141	6	3	-	184
owa	5	11	-	11	22	1	-	-	40
Λο. IDak	38 2	29	-	85 3	66 2	5	2	-	16 46
5 Dak	1	4	-	5	7		-	-	41
Nebr. Cans	8 5	3 9	-	4 12	7 14	-		<u>1</u> 1	5 16
ATLANTIC	1,767	1,790	_	1,073	1,085	4	6	6+1	372
Del	10	14	-	13	9	-	-	-	-
Md.	142 105	137 96	-	77 42	86 52	1	-	-	231
D C /a	139	90	-	100	52 79	1	2	1	60
N. Va.	3	2	-	40	25	-	-	2	8
	155 201	211 230		137 128	131 140	1	2	2	10
Ga	-	-	-	132	159	1	-	٦١	42
la	1,012	1,003	-	404	404	-	2	-	21
S. CENTRAL	486 25	635 25	2 2	495 135	469 99	3 2	-	5 1	74 24
(y. Tenn.	202	173	-	135	138	1	-	-	24 30
Ala	163	220	-	162	163	-	-	1	20
Aiss	96	217	-	62	69	•	-	3	-
N.S. CENTRAL Ark	1,586 77	1,729 82	1	665 92	556 40	4	2	4 🕂 1	197 42
.a.	247	306	-	125	82	-	-		42
Okla.	47	48 1,293	1	56 392	69	1	1	21	18
ſex.	1,215		-		365	-	1	2	133
AOUNTAIN Aont	190	238	-	108	123	-	2	•	231
daho	3 1	2	-	5 5	19 3		-	-	88
Vyo.	-	5	-	-	1		-	-	102
Colo. I. Mex	61 22	55 27	-	1 25	16 22	:	-	-	2
riz	80	133	-	55	53		1	-	39
Jtah 1 Nev.	3 20	3 12	-	4 13	3 6	-	1	-	-
ACIFIC	1,407	1,138	2	1,007	1,076	1	37		196
Vash	27	43	2	56	53	-	2	-	190
)reg. Calif	28	29	-	36	35	-	-	-	
Calif. Naska	1,337	1,045	2	844 17	892 44	1	33	-	190 6
lawaii	15	21	-	54	52		2	-	-
iuam	1	2	-	-	12	-	-		-
.R.	245	262 1	-	76 1	84 1	-	2	-	14
ac. Trust Terr.	45	15	-	ż	23	-	6	-	-
mer. Samoa	-	-	-	-	-	-	-	-	-

# TABLE III. (Cont'd.) Cases of specified notifiable diseases, United States, weeks ending April 12, 1986 and April 13, 1985 (15th Week)

U: Unavailable

TABLE IV.	Deaths in 121 U.S. cities," week ending	
	April 12, 1986 (15th Week)	

	All Causes, By Age (Years)								All Causes, By Age (Years)						
Reporting Area	All Ages	≥65	45-64	25-44	1-24	<1	P&I** Total	Reporting Area	All Ages	≥65	45-64	25-44	1-24	<1	P&I** Total
NEW ENGLAND	675	486	116	28	18	27	66	S. ATLANTIC	1,359	867	308	101	44	37	75
Boston, Mass.	192	125	34	14	5	14	22	Atlanta, Ga.	157	100	35	18	2 7	2	5 9
Bridgeport, Conn. Cambridge, Mass.	38 29	27 24	7 3	1 2	2	1	4 5	Baltimore, Md. Charlotte, N.C.	237 82	153 50	53 21	19 6	2	5 3	6
Fall River, Mass.	20	16	4		-	-	-	Jacksonville, Fla.	120	73	32	ž	3	5	7
Hartford, Conn.	61	44	10	3	2	2	6	Miami, Fla.	130	77	35	10	7	1	3
Lowell, Mass.	27	21	5	-	1	-	2	Norfolk, Va.	30	15	9	1	3 2	2	3 10
Lynn, Mass. New Bedford, Mas	15 s. 28	12 18	3 9	-	1	-	1 2	Richmond, Va. Savannah, Ga.	92 48	57 29	21 14	5 3	1	6 1	2
New Haven, Conn.	50	39	7	1	i	2	3	St. Petersburg, Fla.		92	14	5	4	i	10
Providence, R.I.	55	39	9	2	-	5	1	Tampa, Fla.	77	57	11	4	2	3	5
Somerville, Mass.	7 52	6 40	1 9	-	-3	-	1	Washington, D.C.	223	129	54	21	10	8	13 2
Springfield, Mass. Waterbury, Conn.	37	25	9	2	2	1	8 4	Wilmington, Del.	47	35	9	2	1	•	2
Worcester, Mass.	64	50	ś	3	1	2	7	E.S. CENTRAL	860	521	200	67	37	35	58
								Birmingham, Ala.	112	66	29	9	6	2	6
	2,864	1,926	580	234	69	54	145	Chattanooga, Tenn		46	25	5	4	1	5
Albany, N.Y. Allentown, Pa.	49 20	38 17	8 2	2 1	-	1	3	Knoxville, Tenn	100	66	20	8	3 4	3 2	10 11
Buffalo, N.Y.	129	83	29	9	3	5	15	Louisville, Ky. Memphis, Tenn.	105 218	70 137	23 45	6 16	4 6	14	10
Camden, N.J.	43	27	11	2	-	š	2	Mobile, Ala.	58	32	15	6	1	4	3
Elizabeth, N.J.	15	11	4	-	-	-	-	Montgomery, Ala	62	36	12	5	8	1	2
Erie, Pa.†	40 45	24 30	11	5	-	-	3	Nashville, Tenn	124	68	31	12	5	8	11
Jersey City, N.J. N.Y. City, N.Y.	45	1,111	4 342	8 148	2 41	1 29	3 68		1,484	881	255	136	49	63	69
Newark, N.J.	66	29	19	11	2	5	4	W.S. CENTRAL Austin, Tex.	78	42	355 17	10	49	8	10
Paterson, N.J.	23	15	6	1	-	1	-	Baton Rouge, La.	22	11	9	2	- 1	-	2
Philadelphia, Pa.	302	203	59	18	13	9	12	Corpus Christi, Tex	27	14	4	6	2	1	1
Pittsburgh, Pa.†	65 29	47 19	14 8	3	1	-	3	Dallas, Tex.	221	107	66	35	5	8	12
Reading, Pa. Rochester, N.Y.	104	78	18	2 6	2	-	3 8	El Paso, Tex	48 105	24 61	13 24	5 7	3 9	3 4	- 7
Schenectady, N.Y.	30	22	3	4	ĩ	-	5	Fort Worth, Tex Houston, Tex §	416	268	87	33	10	18	12
Scranton, Pa.†	31	27	4	-	-	-	1	Little Rock, Ark.	71	43	22	3		3	6
Syracuse, N.Y.	95	70	17	5	2	-	8	New Orleans, La	131	77	29	14	4	7	-
Trenton, N.J. Utica, N.Y.	48 23	32 18	8 5	7	1	-	3	San Antonio, Tex.	201	122	54	12	9	4	
Yonkers, N.Y.	36	25	5 8	2	1	-	1 3	Shreveport, La. Tulsa, Okla.	52 112	35 77	10 20	2 7	6	5 2	1 8
E.N. CENTRAL	2,383	1,575	523	157	53	75	112	MOUNTAIN	710	449	145	54	41	21	30
Akron, Ohio	73	53	13	4	2	1	7	Albuquerque, N.Me	x. 81	54	12	6	8	1	7
Canton, Ohio	35 570	23 368	8	2	2	-	4	Colo. Springs, Colo.		26	16	5	-	1	3
Chicago, III.§ Cincinnati, Ohio	128	368 94	125 22	42 3	12 2	23 7	18 7	Denver, Colo. Las Vegas, Nev.	132 83	84 56	23 16	15 6	7 3	3 2	23
Cleveland, Ohio	154	99	30	14	5	6	6	Ogden, Utah	19	14	3	1	1		1
Columbus, Ohio	129	85	30	8	3	3	5	Phoenix, Ariz.	155	94	31	11	14	5	4
Dayton, Ohio	124	81	33	5	-	5	6	Pueblo, Colo.	22	18	3	-	-	1	2
Detroit, Mich. Evansville, Ind.	282 49	154 36	79 10	37 1	7 1	5 1	8	Salt Lake City, Utah Tucson, Ariz		23 80	11 30	4 6	7	4	
Fort Wayne, Ind.	69	48	13	6	2		3	Tucson, Ariz	121	80	30	0	,	4	
Gary, Ind.	12	7	4	ĩ	-	-	- 1	PACIFIC	2,082	1,404	381	180	61	51	127
Grand Rapids, Mic		38	13	3	2	1	5	Berkeley, Calif	15	11	2	2	-	-	
Indianapolis, Ind. Madison, Wis.	144 38	88 24	41 9	6 2	4 2	5 1	3	Fresno, Calif.	85	60	12	7	2	4	
Milwaukee, Wis.	135	97	20	8	4	6	4	Glendale, Calif. Honolulu, Hawaii	31 80	25 57	3 16	1	1	1 3	5
Peoria, III.	38	29	6	1	ĩ	1	3.	Long Beach, Calif.	92	60	23	5	3	1	
Rockford, III.	44	29	9	3	-	3	4	Los Angeles, Calif.	665	445	117	69	21	8	23
South Bend, Ind.	57	47	7	2	-	1	6	Oakland, Calif.	78	49	14	6	1	8	2
Toledo, Ohio Youngstown, Ohio	172	119 56	39 12	6 3	3 1	5 1	15 3	Pasadena, Calif. Portland, Oreg.	37 133	30 90	4 22	2 10	1 6	- 5	3
Toungstown, Onio	, ,3	50	14	3	•		3	Sacramento, Calif.	133	85	30	16	6	5	14
W.N. CENTRAL	780	550	139	43	29	19	45	San Diego, Calif.	162	105	26	15	6	10	24
Des Moines, Iowa	69	47	16	2	2	2	4	San Francisco, Calif.		77	32	20	-	1	3
Duluth, Minn.	29 22	27 14	1	1	-	-	4	San Jose, Calif.	184	120	38	14	10	2	14
Kansas City, Kans. Kansas City, Mo.	132	14 91	5 30	2 6	1	4	1	Seattle, Wash. Spokane, Wash.	153 56	120 39	22 15	7	2 1	2	5
Lincoln, Nebr.	46	32	6	-	5	3		Tacoma, Wash.	43	39	15	3		4	í
Minneapolis, Minn.	83	60	12	7	3	1	3				-	-			
Omaha, Nebr	93	60	18	.7	6	2	10	TOTAL	13,197	8,659	2,747	1,000	401	382	727
St. Louis, Mo. St. Paul, Minn.	161 77	107 58	28 12	11 5	8 2	7	5								
Wichita, Kans.	68	58	11	2	1	-	7								
teronico, reano.				4		-	'								

 Mortality data in this table are voluntarily reported from 121 cities in the United States, most of which have populations of 100,000 or more. A death is reported by the place of its occurrence and by the week that the death certificate was filed. Fetal deaths are not included
 Pneumonia and influenza.

† Because of changes in reporting methods in these 3 Pennsylvania cities, these numbers are partial counts for the current week. Complete counts will be available in 4 to 6 weeks.

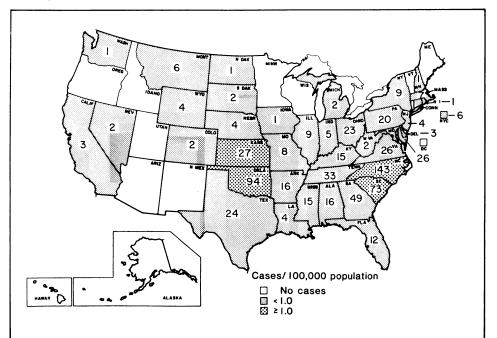
ttTotal includes unknown ages.

§ Data not available. Figures are estimates based on average of past 4 weeks.

# **Rocky Mountain Spotted Fever — United States, 1985**

For 1985, a provisional total of 700 cases of Rocky Mountain spotted fever (RMSF) was reported to *MMWR*, an incidence rate of 0.29 cases per 100,000 population. Oklahoma had the highest incidence rate (94 cases, 2.8/100,000). North Carolina reported the largest number of cases (143 cases, 2.3/100,000). Two other states had incidence rates of 1/100,000 or higher—South Carolina (73 cases, 2.2/100,000) and Kansas (27 cases, 1.1/100,000) (Figure 1).

States submitted case report forms for 587 (84%) of the 700 reported cases. Of these 587 cases, 335 (57%) were laboratory-confirmed by either serologic testing, isolation of spotted fever group rickettsia, or fluorescent antibody staining of biopsy or autopsy material. A case is considered serologically confirmed if testing reveals an indirect fluorescent antibody titer (IFA) of 1:64 or greater, a complement fixation (CF) titer of 1:16 or greater, or a fourfold rise in titer by the CF, IFA, microagglutination (MA), latex agglutination (LA), or indirect hemag-glutination (IHA) assay. An additional 34 (6%) cases were classified as probable cases, as indicated by a fourfold rise in titer or a single titer 1:320 or higher in the Weil-Felix assay or an LA, MA, or IHA single titer of 1:128 or higher. The other 218 (37%) cases were supported by clinical diagnoses alone.



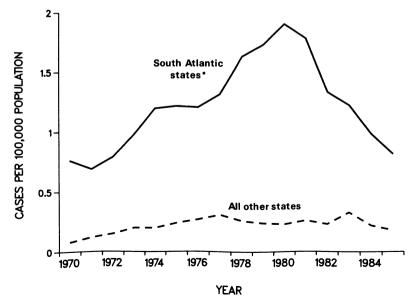
# FIGURE 1. Reported Rocky Mountain spotted fever cases and rates, by state - United States, 1985

# RMSF - Continued

The 1985 surveillance revealed case characteristics similar to those previously reported. Ninety-five percent of patients reported onset of illness between April 1 and September 30, with 66% becoming ill in May, June, or July. Sixty percent of patients were male; 41% were under 20 years of age; and 90% were white. Symptoms reported included fever (94%), head-ache (88%), and myalgia (85%). A rash was reported by 83% and, of these, 73% reported that the rash was noted on the palms or soles. Seventy-three percent of the patients were hospitalized. The overall case-fatality rate was 4%. The case-fatality rate was higher for blacks (16%) than whites (3%) and was higher for individuals 40 years of age or older (9%) than for individuals under age 40 years (2%). Of the patients for whom exposure histories were available, 68% reported a tick bite or attachment, and an additional 24% reported being in a tick-infested area within 14 days (but no tick bite or attachment). Eight percent did not have a known exposure of either type.

Reported by Viral and Rickettsial Zoonoses Br, Div of Viral Diseases, Center for Infectious Diseases, CDC. Editorial Note: The number of reported RMSF cases has waned considerably from the peak of 1,192 cases (0.51/100,000) reported in 1981 (1). The increase in the early 1970s appeared simultaneously in many regions of the United States and was stimulated by the 1970 initiation of a CDC surveillance program (2). The rate of RMSF reported in the South Atlantic states, which increased steadily from 0.76/100,000 in 1970 to a peak of 1.91/100,000 in 1980, has now fallen to 0.82/100,000 (Figure 2). Excluding the South Atlantic states, the rate of RMSF in the other states rose through 1977 and remained fairly constant between 1978 and 1985 (Figure 2). For the third consecutive year, Oklahoma reported the highest incidence of any state.





\*Delaware, Maryland, Virginia, West Virginia, North Carolina, South Carolina, Georgia, Florida, and the District of Columbia.

#### Vol. 35/No. 15

#### RMSF – Continued

Although the number of reported deaths has decreased with the decrease in the number of reported cases, the case-fatality rate has changed little over the last 5 years. Morbidity and mortality may be decreased by knowledge of the epidemiology and early clinical signs of RMSF (3). RMSF should be suspected, and treatment with chloramphenicol or tetracycline strongly considered, among residents of, or visitors to, RMSF-endemic areas who report fever, head-ache, and myalgias, even without a rash, particularly during April through October. Symptoms and signs referable to the pulmonary system (such as cough or rales), the gastrointestinal system (such as nausea, vomiting, or abdominal pain), or the central nervous system (such as stupor, meningismus, or ataxia), are seen with RMSF and should not delay diagnosis or treatment (3). Special attention is warranted for individuals 40 years of age and older, who have a greater likelihood of a fatal outcome, and dark-skinned individuals, in whom a rash may be more difficult to diagnose.

In a recent study of RMSF in a hyperendemic area, a tick bite or exposure was reported for 85% of serologically confirmed cases, compared with 54% of matched controls (4). Rash was reported in 84% of the serologically confirmed cases. However, in 16% of patients, a rash was never noted, and in an additional 10%, a rash did not develop until later than the fifth day after onset of the illness.

Prevention of RMSF is best accomplished by careful inspection of persons who may have been exposed to ticks. Ticks should be removed by grasping them with tweezers as closely as possible to the point of attachment and pulling slowly and steadily (5). If a portion of the mouth part remains, it should be treated like any other small foreign body; it may cause irritation, but it will not increase the risk of contracting RMSF. The fingers, protected with tissue paper, may be used to remove a tick from a person if tweezers are not available, but should always be washed after the removal of a tick. The fingers should not be used to detick dogs. Persons living and working in tick-infested areas should be educated about the prevention, symptoms, and signs of the disease. No vaccine against RMSF is currently available. RMSF cases should be reported to appropriate local and state health departments.

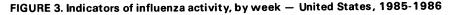
## References

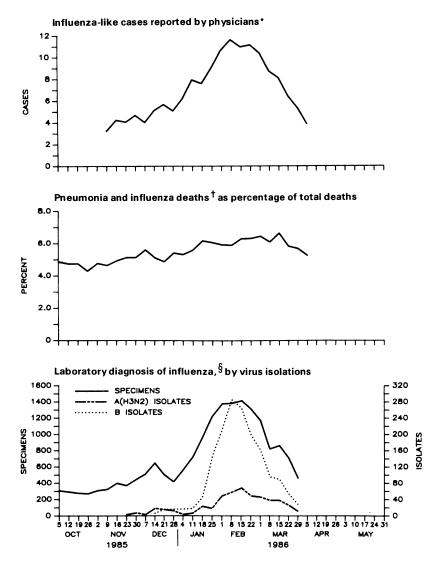
- 1. CDC. Rocky Mountain spotted fever—United States, 1981. MMWR 1982;31:261-3.
- Hattwick MAW, Peters AH, Gregg MB, et al. Surveillance of Rocky Mountain spotted fever. JAMA 1973; 225:1338-43.
- Woodward TE. Rocky Mountain spotted fever: epidemiological and early clinical signs are keys to treatment and reduced mortality. J Infect Dis 1984;150:465-8.
- Wilfert CM, MacCormack JN, Kleeman K, et al. Epidemiology of Rocky Mountain spotted fever as determined by active surveillance. J Infect Dis 1984;150:469-79.
- 5. Needham GR. Evaluation of five popular methods for tick removal. Pediatrics 1985;75:997-1002.

# Current Trends

# Update: Influenza Activity - United States

Influenza activity in the United States continues to decline (Figure 3). Almost all influenza virus isolates this season have been type B (76.1%) or type A(H3N2) (23.8%). However, from February 17 to February 24, type A(H1N1) viruses were isolated from three persons, aged 3, 4, and 31 years, with influenza-like illness who lived in the same neighborhood in west Houston, Texas. Despite continued surveillance, no further type A(H1N1) isolates have been identified in Houston. In Hawaii, one type A(H1N1) virus isolate was reported this season.





\*Reported to CDC by approximately 125 physician members of the American Academy of Family Physicians. A case was defined as a patient with fever 37.8 C (100 F) or greater and at least cough or sore throat.

<sup>†</sup>Reported to CDC from 121 cities in the United States. Pneumonia and influenza deaths include all deaths where pneumonia is listed as a primary or underlying cause or where influenza is listed on the death certificate.

§Reported to CDC by WHO Collaborating Laboratories (including military sources).

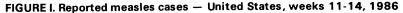
# Influenza — Continued

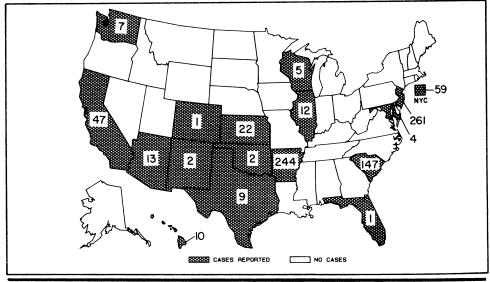
In Birmingham, Alabama, recent laboratory results have shown that an outbreak of influenza among college students reportedly caused by influenza type A(H3N2) virus (1) was primarily associated with type B influenza virus.

Reported by Influenza Research Center, Baylor College of Medicine, Houston, Texas; State and Territorial Epidemiologists; State Laboratory Directors; Statistical Svcs Br, Div of Surveillance and Epidemiologic Studies, Epidemiology Program Office, WHO Collaborating Center for Influenza, Influenza Br, Div of Viral Diseases, Center for Infectious Diseases, CDC.

#### Reference

1. CDC. Update: influenza activity - United States. MMWR 1986;35:135-6, 141.





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The data in this report are provisional, based on weekly reports to CDC by state health departments. The reporting week concludes at close of business on Friday; compiled data on a national basis are officially released to the public on the succeeding Friday.

The editor welcomes accounts of interesting cases, outbreaks, environmental hazards, or other public health problems of current interest to health officials. Such reports and any other matters pertaining to editorial or other textual considerations should be addressed to: ATTN: Editor, *Morbidity and Mortality Weekly Report*, Centers for Disease Control, Atlanta, Georgia 30333.

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