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Symptomatic Infection and Detection of Vaccine and Vaccine-Reassortant Rotavirus Strains in 5 Children: A Case Series

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Abstract

Vaccine or vaccine-reassortant rotavirus strains were detected in fecal specimens from 5 of 106 (4.7%) immunocompetent children who required treatment for rotavirus gastroenteritis at a large pediatric hospital in Texas in 2009–2010. Four strains were related to pentavalent rotavirus vaccine, whereas one was related to monovalent rotavirus vaccine. The contribution of these strains to each patient's illness was unclear given that 2 patients had prominent respiratory symptoms and 2 were concurrently infected with another pathogen (group F adenovirus and norovirus). Continued monitoring is necessary to assess the role of vaccine strains and vaccine-reassortant strains in pediatric rotavirus infections.

Rotavirus is a nonenveloped, RNA virus belonging to the *Reoviridae* family [1]. Group A rotaviruses, the primary source of human infection, have been genotyped conventionally through characterization of the 2 outer capsid proteins, *VP7* (G-type) and *VP4* (P-type). These capsid proteins contribute to regional and temporal diversity of circulating strains [2, 3].

Two live, attenuated, orally administered vaccines are currently licensed and recommended for use in infants: 3-dose pentavalent vaccine (RotaTeq [RV5]; Merck Vaccines), which contains 5 G1-G4, and P[8] human-bovine reassortant strains, and 2-dose monovalent

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vaccine (Rotarix [RV1]; GlaxoSmith-Kline Biologicals), which contains a single human G1P[8] strain.

Following vaccination, rotavirus vaccine strains replicate in the gastrointestinal tract and may be shed in the stool. In prelicensure trials and 1 postlicensure assessment, viral shedding was observed 7 days after the first dose in 25.6%–26.5% of RV1 recipients and in 8.9%–21.4% of RV5 recipients 1–15 days after any dose [4–6]. Likely transmission of vaccine virus from infants given RV1 to their unvaccinated twins was observed in a randomized clinical trial [7]. Transmission of RV5 virus through fecal shedding has not been formally assessed; however, 1 instance of horizontal transmission of vaccinederived virus to an unvaccinated sibling resulting in acute gastroenteritis (AGE) requiring medical attention has been described [8].

Rotavirus has the ability to undergo reassortment with wild-type or vaccine-derived strains because of its segmented genome [9]. Little is known about the frequency with which reassortment occurs or the resulting clinical outcome. This report describes 5 children with AGE who were found to be shedding rotavirus vaccine strains and/or vaccine-reassortant strains during routine surveillance.

METHODS

Active surveillance for AGE was conducted at Texas Children's Hospital from November 2009 through June 2010. Children aged 15 days–4 years with AGE (3 episodes of diarrhea and/or 1 episode of vomiting) requiring emergency department treatment or hospitalization were offered enrollment. A questionnaire assessing disease symptoms and epidemiologic factors was administered, and fecal specimens were collected within 14 days of illness onset.

Institutional review board approval was obtained from Baylor College of Medicine and Texas Department of State Health Services. Informed consent was obtained from participants' parents/guardians.

Fecal specimens were tested for rotavirus antigen using the Premier Rotaclone kit (Meridian Bioscience, Inc). RNA was extracted from 10% stool suspensions using the MagNA Pure Compact instrument with the MagNA Pure Compact RNA isolation kit (Roche Applied Science). *VP4* and *VP7* genotyping and amplicon sequencing were performed [10], and identification of RV5 and RV1 strains was carried out through comparison with vaccine strain sequences deposited in GenBank [8]. Identification of RV1 was accomplished also by alignment of *NSP2* sequence with that obtained by genomic sequencing of RV1 vaccine itself (unpublished data). Examination of stool samples by electron microscopy (EM) and testing for other agents of gastroenteritis by reverse-transcription polymerase chain reaction (RT-PCR), polymerase chain reaction (PCR), and culture were performed as previously described [8].

RESULTS

A total of 1133 patients with AGE were enrolled. Fecal specimens were obtained from 997 subjects (88.0%); 106 of these (10.6%) were rotavirus-positive by RT-PCR. Subsequent

genomic sequencing of the 106 rotavirus-positive fecal specimens identified 5 (4.7%) patients with vaccine-reassortant and/or vaccine strains. The epidemiologic and clinical characteristics of these 5 patients and their virus characterization findings are described below.

Patient 1

A previously healthy 22-month-old boy presented with a history of 11 episodes of nonbloody, nonbilious emesis occurring over a 2-hour period (Table 1). He was treated with oral ondansetron, orally rehydrated, and discharged. The patient returned 8 hours later with persistent vomiting. Because he was tachycardic and dehydrated, he received intravenous fluids overnight and was discharged the next day. Although the patient had received no doses of rotavirus vaccine, his infant sibling received a second dose of RV5 7 days prior to the patient's illness onset.

The VP7 G1 gene of RV5 component strain WI79-9 (G1P [5]) (99.9% identity in 839 bases sequenced) and the VP4 P[8] (100% identity in 835 bases) and VP6 I2 (100% identity in 338 bases) genes from RV5 strain WI79-4 (G6P[8]) (Table 2) were detected in a fecal specimen obtained 2 days after illness onset. The VP3 M2 gene matched the sequence of strains WI79-4 (G6P[8]), WI78-8 (G3P[5]), and BrB-9 (G4P[5]) (100% identity in 500 bases). The strain appears to be a reassortant derived by insertion of the WI79-9 (G1P[5]) G1 gene onto a WI79-4 (G6P[8]) backbone. Rotaviruses and unidentified particles 30–40 nm in diameter were observed by EM. All other tests were negative.

Patient 2

A 4-month-old boy presented with a 4-day history of rhinorrhea and nonbloody diarrhea, and 3 days of fever. The patient was wheezing and tachypneic with an oxygen saturation of 95%. He was diagnosed with bronchiolitis, treated with levalbuterol and vaponephrine, and observed overnight. The patient received his first dose of RV5 11 days before illness onset.

Genetic analysis of a fecal specimen obtained 9 days after illness onset detected the *VP7* G1 gene of RV5 strain WI79-9 (G1P[5]) (100% identity in 854 bases) and the *VP4* P[8] gene from RV5 strain WI79-4 (G6P[8]) (100% identity in 835 bases) (Table 2). The *VP6* I2 gene detected matched that of RV5 component strains WI79-9 (G1P[5]), SC2-9 (G2P[5]), WI78-8 (G3P[5]), and BrB-9 (G4P[5]) (100% identity in 342 bases). The *VP3* M2 gene matched the sequence of strains WI79-4 (G6P[8]), WI78-8 (G3P[5]), and BrB-9 (G4P[5]) (100% identity in 500 bases). This strain appears to be a double reassortant with either the WI79-9 (G1P[5]) *VP7* and *VP6* genes inserted onto a WI79-4 (G6P[8]) backbone or the VP4 of WI79-4 (G6P[8]) and VP3 of WI79-4 (G6P[8]), BrB-9 (G4P[5]), or WI78-8 (G3P[5]) inserted onto a WI79-9 (G1P[5]) backbone. Rotaviruses and adenoviruses were observed by EM. Real-time PCR was positive for adenovirus group F; all other tests were negative.

Patient 3

A previously healthy 2-month-old girl presented with 4 days of vomiting and 2 days of fever, cough, and conjunctivitis without diarrhea. On physical exam, she was fussy but consolable with mild conjunctival injection and nasal discharge. Laboratory evaluation

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revealed an elevated white blood cell count (19 700/ μ L) with 12% bandemia. Urinalysis and urine culture were negative. The patient was treated with intravenous ceftriaxone and discharged. She received her first dose of RV5 1 day before illness onset.

A fecal specimen was obtained 5 days after illness onset and appeared to contain nonreassortant RV5 component strain BrB-9 (G4P[5]) alone. The *VP7* G4 gene of BrB-9 (G4P[5]) was detected (100% identity in 363 bases) along with *VP4* P [5] (100% identity in 416 bases), *VP6* I2 (100% identity in 342 bases), and *VP3* M2 genes (100% identity in 500 bases) consistent with BrB-9 (G4P[5]). By EM, a few unidentified 30-nm particles were observed; all other tests were negative.

Patient 4

A previously healthy 2-month-old boy presented with 3 days of nonbloody diarrhea and 2 days of fever and irritability. On physical exam, he was fussy but consolable with a slightly distended abdomen and hyperactive bowel sounds. Negative laboratory examinations included complete blood count with differential, urinalysis, and urine culture (catheterized). The patient was orally rehydrated and discharged. He received his first dose of RV5 2 days before illness onset.

Four distinct *VP7* genotypes were detected in a specimen obtained 3 days after illness onset: the G1 gene of RV5 component strain WI79-9 (G1P[5]) (100% identity in 116 bases); the G2 gene of strain SC2-9 (G2P[5]) (100% identity in 202 bases); the G4 gene of strain BrB-9 (G4P[5]) (100% identity in 363 bases); and the G6 gene of strain WI79-4 (G6P[8]) (99.8% identity in 421 bases) (Table 2). The *VP4* P[8] gene (99.9% identity in 835 bases) and *VP6* I2 (100% identity in 342 bases) genes from RV5 strain WI79-4 (G6P[8]) were detected. The *VP3* M2 gene matched the sequence of strains WI79-4 (G6P [8]), WI78-8 (G3P[5]), and BrB-9 (G4P[5]) (100% identity in 500 bases). This specimen appears to contain 4 strains, the WI79-4 (G6P[8]) component strain itself and 3 potential reassortant strains each composed of strain WI79-4 (G6P[8]) but expressing G1, G2, and G4 individually. Rotaviruses were observed by EM, and real-time RT-PCR was positive for norovirus genogroup II. All other tests were negative.

Patient 5

A 6-month-old boy presented with a 1-day history of nonbloody, nonbilious vomiting with diarrhea, lethargy, and fever. His past medical history was notable for failure-to-thrive and gastroesophageal reflux. The patient's 2.5-year-old sister also had been ill with vomiting. On physical exam, he was tachycardic and had generalized pallor. The patient failed oral rehydration and was subsequently treated with intravenous ondansetron and fiuids and was observed overnight. Neither the patient nor his sister had received any doses of either rotavirus vaccine.

Analyses of a fecal specimen obtained 8 days after illness onset identified what appears to be an RV1-derived strain. Sequences of the *VP7* (850 bases), *VP4* (803 bases), and NSP2 (991 bases) genes each shared 99.8% identity with cognate RV1 vaccine strain sequences.

Rotaviruses and unidentified particles 40–45 nm in diameter were observed by EM. All other tests were negative.

DISCUSSION

These patients illustrate a spectrum of potential clinical and virologic outcomes associated with routine use of live, attenuated rotavirus vaccines. The diversity of these occurrences include apparent transmission of an RV5 vaccine-reassortant strain from a vaccinated infant to an unvaccinated sibling (patient 1); detection of an RV5 vaccine-reassortant strain in a recently vaccinated child (patient 2); detection of a nonreassortant RV5 vaccine strain in a recently vaccinated child (patient 3); detection of an RV5 vaccine strain and 3 potential vaccine-reassortant strains in a recently vaccinated child (patient 4); and detection of an RV1 vaccine strain in a nunvaccinated child residing in an area with low RV1 coverage (patient 5).

Although these findings suggest potentially unusual transmission or vaccine replication processes, the contribution of the vaccine or vaccine-reassortant strain to each patient's illness is unclear. Patient 1's illness was clinically consistent with rotavirus AGE and occurred after exposure to a younger sibling who had recently received RV5. This patient's history of exposure and subsequent illness is similar to a prior report of sibling transmission of the same vaccine-derived G1P[8] strain [8], supporting the likely linkage between this vaccine-reassortant strain and patient 1's illness. Patients 2 and 3 had upper respiratory symptoms accompanied by vomiting or diarrhea, and their illnesses were less typical of rotavirus disease alone. Additionally, detection of adenovirus group F in patient 2's specimen provides a plausible alternate etiology. Similarly, although patient 4 had diarrhea without respiratory symptoms, the presence of norovirus genogroup II in his fecal specimen along with the potential vaccine-reassortant strains makes it impossible to discern the cause of his illness. The presence of multiple reassortant RV5 strains in patient 4's stool is very unusual, and these findings will be confirmed using an advanced genetic characterization method such as microarray analysis and/or deep sequencing. Both patient 5 and his 2.5-yearold sister presented with symptoms consistent with rotavirus disease. The RV1 strain detected in this unvaccinated patient's stool is suggestive of an environmental exposure to a vaccine strain that is not widely present in the patient's community. Although RV1 uptake rates in the Houston area during this surveillance period are not known, no enrolled patient received any doses of RV1, thus confirming its rare use. However, because a full epidemiologic investigation was not conducted, it is impossible to determine how this patient's exposure to RV1 occurred or the role of RV1 in the patient's illness. These outcomes highlight the difficulty in establishing clear causative relationships between vaccine-derived strains and clinical illness, despite exhaustive laboratory testing.

The detection of rotavirus vaccine or vaccine-derived reassortant strains in fecal specimens from 5 of 106 (4.7%) immunocompetent, rotavirus-positive children hospitalized or visiting the emergency department with AGE symptoms was surprising. Although the population rate of severe AGE related to vaccine or vaccine-reassortant strains is likely to be low given the large number of vaccinated children and their contacts residing in the catchment area of our surveillance hospital, such events may not be as uncommon as previously suspected.

Continued monitoring is necessary to assess the rate of occurrence and clinical relevance of vaccine strains and vaccine-reassortant strains in symptomatic and asymptomatic pediatric rotavirus infections.

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Table 1

Demographic Characteristics, Clinical Symptoms, and Fecal Testing Results of the 5 Patients Described in This Study

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Characteristic	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5
Age at presentation in months	22.5	4.6	2.9	2.3	6.2
Symptoms					
Diarrhea					
Duration; maximum episodes in 24 h	None	5 d; 4 episodes	3 d; 18 episodes	None	1 d; 5 episodes
Vomiting					
Duration; maximum episodes in 24 h	1 d; 11 episodes	None	None	4 d; 3 episodes	1 d; 12 episodes
Fever					
Duration; maximum	1 d; 38.8°C (rectal)	5 d; 38.6°C (rectal)	1 d; 38.3°C (rectal)	2 d; 39.4°C (axillary)	1 d; 38.6°C (rectal)
Intravenous rehydration	Yes	None	None	None	Yes
Hospitalization for AGE episode	Yes; 23-hour stay	Yes; 23-hour stay	None	None	Yes; 23-hour stay
Race/ethnicity	White, non-Hispanic	White, Hispanic	White, Hispanic	Middle Eastern, non- Hispanic	White, non-Hispanic
Birth history					
Gestation, birth weight	Term, 7 lb 3 oz	Term, 7 lb 13 oz	Term, 9 lb 8 oz	Term, 7 lb 12 oz	Term, 6 lb 9 oz
History of breastfeeding					
Duration	Yes; 7–12 mo	Yes; <1 mo	Yes; 1–3 mo	Yes; <1 mo	Yes; 1–3 mo
Other children in the home	1 sibling aged <6 mo	2 siblings, both aged 5 y	2 siblings, both aged 5 y	None	1 sibling, aged 2.5 y
Daycare attendance	None	None	None	None	None

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Characteristic	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5
Rotavirus vaccination history					
Patient	No doses RV1 or RV5	1st dose RV5 11 d prior to illness onset	1st dose RV5 2 d prior to illness onset	1st dose RV5 11 d prior 1st dose RV5 2 d prior 1st dose RV5 1 d prior to illness onset to illness onset	No doses RV5 or RV1
Other children aged <5 y in the home	2nd dose RV5 7 d before patient's illness onset	νVa	NA ^a	NA ^a	No doses RV5 or RV1
Timing of fecal specimen collection	2 d after illness onset	9 d after illness onset	5 d after illness onset	3 d after illness onset	8 d after illness onset
Rotavirus strain detected	1 RV5 vaccine- reassortant strain	1 RV5 double vaccine- reassortant strain	1 RV5 vaccine strain	1 RV5 vaccine strain & 3 RV5 vaccine- reassortant strains	RV1 vaccine strain
AGE coinfections b	Unidentified particles 30–40 nm in diameter	Adenovirus group F	Unidentified particles 30 nm in diameter	Norovirus genogroup II Unidentified particles 40–45 nm in diameter	Unidentified particles 40–45 nm in diameter

^aOther children in the home were 5 years at the time of the patient's illness, and, therefore, were born before rotavirus vaccine was licensed in the United States.

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b Specimens were tested for norovirus genogroups I and II, astroviruses, sapoviruses, enteroviruses, parechoviruses [11–14], adenovirus, Salmonella, Shigella, Campylobacter, diarrheagenic Escherichia coli, and Vibrios.

		RI	RV5 Component Strains ^b	p_{ns}			Case	Case Strains	
Gene	6-97IW	SC2-9	W178-8	BrB-9	W179-4	Patient 1	Patient 2	Patient 3	Patient 4
VP7	GI (GU565057) ^c	G2 (GU565068)	G3 (GU565079)	G4 (GU565090)	G6 (GU565046)	GI (GU565057)	G1 (GU565057)	G4 (GU565090)	GKGU565057) G2(GU565068) G4(GU565090) G6(GU565046) ^d
VP4	P[5] (GU565055)	P[5] (GU565066)	P[5] (GU565077)	P[5] (GU565088)	P[8] (GU565044)	P[8] (GU565044)	P[8] (GU565044)	P[5] ^e	P[8] (GU565044)
VP6 ^f	<i>VP</i> 6 <i>f</i> 12 (GU565056)	12 (GU565067)	12 (GU565078)	I2 (GU565089)	I2 (GU565045)	I2 (GU565045)	I2 (GU565056 or GU565067 or GU565078 or GU565089)	I2 (GU565056 or GU565067 or GU565078 or GU565089)	I2 (GU565045)
VP3	M1 (GU565054)	M1 (GU565065)	M2 (GU565075)	M2 (GU565087)	M2 (GU565043)	M2 ⁸	M2 ⁸	M2 ⁸	M2 ⁸
A Rotar Within t	ix vaccine (RV1)-de he RV5 component	erived strain was iden strains. the <i>VP1</i> . <i>VP2</i>	^a A Rotarix vaccine (RV1)-derived strain was identified in the specimen obtained from patient 5 and is not included in this table. ^b Within the RV5 component strains. the VP1. VP2. VP6. NSP1. NSP2. NSP3. NSP4. and NSP5/6 of all 5 strains are identical or r	1 obtained from patien NSP3, NSP4, and NSI	nt 5 and is not include <i>P5/6</i> of all 5 strains a	ed in this table. re identical or nearly	identical. The <i>VP3</i> <u>o</u>	but	a^{d} Rotarix vaccine (RV1)-derived strain was identified in the specimen obtained from patient 5 and is not included in this table.
genes of	WI78-8, BrB-9, and	WI79-4 are identical	genes of WI78-8, BrB-9, and WI79-4 are identical or nearly identical. The VP4 genes of WI79-9, SC2-9, WI78-8, and BrB-9 are identical or nearly identical.	The VP4 genes of WI7	79-9, SC2-9, WI78-8,	, and BrB-9 are identi	ical or nearly identica	Γ	
GenBan	k accession number	for reference sequenc	^c GenBank accession number for reference sequence (RV5 component strains) or closest sequence match (case strains).	trains) or closest sequ	sence match (case stra	ains).			
l _{Four dis}	d Four distinct <i>VP7</i> genotypes detected.	s detected.							
One VP.	4 P[5] genotype dete	scted, but 4 sequences	^e One VP4 P[5] genotype detected, but 4 sequences (GU565055, GU565066, GU565077, GU565088) are identical in the region sequenced (416 bases) and could not be distinguished.	066, GU565077, GU	565088) are identical	l in the region sequen	ced (416 bases) and c	could not be distingu	ished.
Within ti	he region of the VP6	$f_{\rm Within}$ the region of the VP6 gene sequenced (342 bases),		GU565067, GU5650	78, and GU565089 at	re identical, and a sin	gle base substitution	distinguishes GU565	GU565056, GU565067, GU565078, and GU565089 are identical, and a single base substitution distinguishes GU565045 from the other 4 strains.
One VP.	3 M2 genotype deteo	cted, but all 3 M2 seq	⁸ One VP3 M2 genotype detected, but all 3 M2 sequences (GU565075, GU565087, GU565043) are identical in the region sequenced (500 bases) and could not be distinguished.	3U565087, GU56504	13) are identical in the	e region sequenced (2	500 bases) and could	not be distinguished.	

Table 2

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