Genetic Variants in the Major Histocompatibility Complex Class I and Class II Genes Are Associated With Diisocyanate-Induced Asthma

Dr. Berran Yucesoy, PhD, Dr. Victor J. Johnson, PhD, Dr. Zana L. Lummus, PhD, Dr. Michael L. Kashon, PhD, Dr. Marepalli Rao, PhD, Dr. Hansen Bannerman-Thompson, PhD, Ms. Bonnie Frye, BSc, Ms. Wei Wang, MS, Dr. Denyse Gautrin, PhD, Dr. André Cartier, MD, Dr. Louis-Philippe Boulet, MD, Dr. Joaquin Sastre, MD, Dr. Santiago Quirce, MD, Dr. Susan M. Tarlo, MD, Dr. Dori R. Germolec, PhD, Dr. Michael I. Luster, PhD, and Dr. David I. Bernstein, MD

Health Effects Laboratory Division (Drs Yucesoy and Kashon, Ms Frye, and Ms Wang) NIOSH/CDC, Morgantown, WV; BRT-Burleson Research Technologies (Dr Johnson), Morrisville, NC; Division of Immunology, Allergy and Rheumatology, Department of Medicine (Drs Yucesoy, Lummus, and Bernstein) and Department of Environmental Health (Drs Rao and Bannerman-Thompson), University of Cincinnati, Ohio; Hôpital du Sacré-Cœur de Montréal (Drs Gautrin and Cartier), Université de Montréal; Hôpital Laval (Dr Boulet), Université Laval, Sainte-Foy, Québec, Canada; Department of Allergy (Dr Sastre), Fundación Jiménez Díaz and CIBER de Enfermedades Respiratorias CIBERES; Department of Allergy (Dr Quirce), Hospital La Paz-IdiPAZ and CIBER de Enfermedades Respiratorias CIBERES, Madrid, Spain; Department of Medicine (Dr Tarlo), University of Toronto, Ontario, Canada; Toxicology Branch (Dr Germolec), DNTP/NIEHS, Research Triangle Park, NC; and School of Public Health (Dr Luster), West Virginia University, Morgantown

Abstract

Objective—To investigate the association between single nucleotide polymorphisms (SNPs) located across the major histocompatibility complex and susceptibility to diisocyanate-induced asthma (DA).

Methods—The study population consisted of 140 diisocyanate-exposed workers. Genotyping was performed using the Illumina GoldenGate major histocompatibility complex panels.

Address correspondence to Berran Yucesoy, PhD, Division of Immunology, Allergy and Rheumatology, College of Medicine, University of Cincinnati, Cincinnati, OH 45267-0563; Health Effects Laboratory Division, NIOSH/CDC, Morgantown, WV 26505 (yucesobn@ucmail.uc.edu).

The authors declare no conflict of interest.

The findings and conclusions in this report are those of the authors and do not necessarily represent the views of the National Institute for Occupational Safety and Health. This article may be the work product of an employee or a group of employees of the National Institute of Environmental Health Sciences (NIEHS), National Institutes of Health (NIH); however, the statements, opinions, or conclusions contained therein do not necessarily represent the statements, opinions, or conclusions of NIEHS, NIH, or the US government.

Authors Yucesoy, Johnson, Lummus, Kashon, Rao, Bannerman-Thompson, Frye, Wang, Gautrin, Cartier, Boulet, Sastre, Quirce, Tarlo, Germolec, Luster, and Bernstein have no relationships/conditions/circumstances that present potential conflict of interest.

The JOEM editorial board and planners have no financial interest related to this research.
Results—The HLA-E rs1573294 and HLA-DPB1 rs928976 SNPs were associated with an increased risk of DA under dominant (odds ratio [OR], 6.27; 95% confidence interval [CI], 2.37 to 16.6; OR, 2.79, 95% CI, 0.99 to 7.81, respectively) and recessive genetic models (OR, 6.27, 95% CI, 1.63 to 24.13; OR, 10.10, 95% CI, 3.16 to 32.33, respectively). The HLA-B rs1811197, HLA-DOA rs3128935, and HLA-DQA2 rs7773955 SNPs conferred an increased risk of DA in a dominant model (OR, 7.64, 95% CI, 2.25 to 26.00; OR, 19.69, 95% CI, 2.89 to 135.25; OR, 8.43, 95% CI, 3.03 to 23.48, respectively).

Conclusion—These results suggest that genetic variations within HLA genes play a role in DA risk.
diseases, thus far, only a limited number of HLA genes have been examined with respect to DA. This is the first study investigating the association of single nucleotide polymorphisms (SNPs) located across the entire MHC region with DA in a well-characterized worker population using microarray technology.

MATERIALS AND METHODS

Subjects

The study population consisted of 140 workers exposed to diisocyanates (HDI, MDI, and TDI). Of these, 73 were diagnosed with DA (DA+) on the basis of a positive specific inhalation challenge (SIC) test and 67 were asymptomatic workers (AWs) exposed to HDI. Symptomatic subjects were recruited from occupational pulmonary disease clinics located in Canada (Hôpital du Sacré-Coeur Montréal, Montréal, 124 subjects; Laval Hospital, Sainte-Foy, 12 subjects; University Health Network, Toronto, Ontario, 2 subjects) and Spain (Fundación Jiménez Díaz, Madrid, 2 subjects). The subjects underwent SIC testing with the appropriate work-relevant diisocyanate chemicals according to previously described protocols. Patients were classified as DA+ on the basis of their positive response to SIC. A decrease in FEV$_1$ (forced expiratory volume in the first second of expiration) of at least 20% from prechallenge baseline during the early and/or late asthmatic response was defined as a positive SIC test. The AW controls were recruited in Quebec, Canada, from HDI-exposed painters and evaluated by occupational history, spirometry, and skin prick testing. Data regarding age, sex, ethnicity, smoking status, and time of exposure were collected by questionnaire. Atopy was evaluated by skin prick testing to common aeroallergens, defined by a positive reaction of at least 3 mm greater than saline control for at least one allergen. Antibodies were detected by isotype-specific enzyme-linked immunosorbent assay tests, as previously described. Whole blood was collected for genetic testing. All subjects provided written informed consent, and the study protocol was approved by institutional review boards of National Institute for Occupational Safety and Health and each participating institution.

Genotyping

Genomic DNA was extracted from whole blood samples by using the QIAamp blood kit (QIAGEN Inc, Chatsworth, CA). Genotyping was performed according to the standard protocol provided by Illumina using the MHC Panel Set and Golden Gate protocol (Illumina Inc, San Diego, CA). The MHC SNP set consisted of two oligonucleotide pools, MHC Mapping Panel and MHC Exon-Centric Panel for 1228 and 1293 SNP loci, respectively. Both panels cover 2360 independent loci spaced at an average of 2.08 kb (range, 0.005 to 71.05 kb). Genotyping was performed in a 16-well format universal BeadChips. A total of 250 ng to 1 μg DNA was used for each assay, depending on the source. Genotypes were auto called using the BeadStudio software.

The genotype confidence score of the assay was set to 0.5. Data quality was assessed by controlling for discrepancies between 161 overlapping SNPs in the two panels. SNPs with more than two discrepant calls were removed from further analysis. In the remaining overlapping SNPs, consistency was 99.9%. Replicate sample comparisons within and across
DNA genotyping plates also demonstrated high agreement (data not shown). Alleles that were not called in a sample were coded as missing in the analysis. A total of 140 samples and 2202 SNPs passed our quality control criteria and included in the final analysis.

**Statistical Analyses**

All statistical analyses were conducted using either SAS/STAT for Windows or R programming within Bioconductor. Comparisons between groups on demographic variables were analyzed using two-sample t tests and chi-squared tests for continuous and categorical variables, respectively. Initial analyses, including tests for Hardy-Weinberg Equilibrium, allele, and genotype frequencies, were calculated using the Fisher exact test. A Bonferroni adjustment for multiple comparisons from the array was performed on the 2 by 3 genotype and 2 by 2 allele frequency tables. Unadjusted odds ratios (OR) were calculated using contingency tables. Adjusted ORs were calculated using logistic regression while adjusting for age, exposure time (months), smoking (current/ex/never), pack years, height, sex, and atopic status. Since underlying genetic models are unknown a priori, the association between each SNP and DA status was analyzed using three genetic models. These include a dominant model (comparing homozygous wild-type genotype with variant allele-carrying genotypes), recessive model (comparing wild-type allele-carrying genotypes with homozygous variant genotype), and an additive model. Prior to the final analyses, we utilized the MICE algorithm (Multiple Imputations by Chained Equations) to impute missing data. Three independent imputations were generated and age, sex, height, exposure duration, smoking, and atopy were appended to each data set. There were no substantive differences between the 3 imputations and we present the results from the first imputation. SNAP2 tools were used to update annotations of significant SNPs according to dbSNP135 and to find proxy SNPs within 500 kb based on LD and physical distance. RegulomeDB was used to annotate SNPs with known and predicted regulatory elements.

**RESULTS**

**Subject Characteristics**

The demographic characteristics of the study groups included in the statistical analyses are described in Table 1. A total of 92% of the symptomatic workers diagnosed with DA (DA+) and 99% of the asymptomatic exposed workers (AW) were white French Canadians. Mean age was higher in the DA+ group than in AW controls (42.4 vs 30.0 years). While the DA+ group consisted of subjects exposed to HDI, MDI, and TDI (39, 15, and 18, respectively), the AW controls were exposed to HDI in the workplace. The AW controls had less duration of exposure to isocyanates than the DA+ group (63.4 vs 146.3 months). The frequency of atopy was similar in groups (54% in DA+ and 56% in AWs). The number of pack years was higher in the DA+ group than in AW controls (9.4 vs 6.2 years). The allele frequencies in the control population were in Hardy–Weinberg equilibrium (data not shown).

**Genotype Distribution and Genetic Models**

SNPs in *HLA-E, HLA-B, HLA-DOA, HLA-DQA2*, and *HLA-DPB1* genes (rs1573294, rs1811197, rs3128935, rs7773955, and rs928976, respectively) remained significantly associated with DA after the Bonferroni adjustment for multiple testing. Table 2 shows the
distribution of genotypes in the study population. There were no SNPs that were statistically significant under an additive model and thus only the dominant and recessive models are presented (Table 3). The rs1573294 SNP was associated with an increased risk of DA under dominant and recessive genetic models (OR, 6.27, 95% confidence interval [CI], 2.37 to 16.61; OR, 6.27, 95% CI, 1.63 to 24.13, respectively). The HLA-B rs1811197, HLA-DOA rs3128935, and HLA-DQA2 rs7773955 SNPs conferred an increased risk of DA in a dominant model (OR, 7.64, 95% CI, 2.25 to 26.00; OR, 19.69, 95% CI, 2.89 to 135.25; OR, 8.43, 95% CI, 3.03 to 23.48, respectively). The HLA-DPB1 rs928976 SNP was also associated with higher risk of DA under dominant and recessive genetic models (OR, 2.79, 95% CI, 0.99 to 7.81; and OR, 10.10, 95% CI, 3.16 to 32.33). No association was found between non-HLA gene variants and DA.

### Regulatory Information for Significant Associations

Five unique significant SNPs were used as inputs to the SNP Annotation and Proxy Search tool to find highly correlated SNPs within 500 kb (using an $r^2$ of 1). This led to the identification of an additional 34 correlated SNPs using data from the International HapMap Project. The total set of 39 SNPs was then used as inputs to the RegulomeDB web resource, which integrates data from the ENCODE projects and other data sources regarding various types of functional assays including DNaseI-seq, ChIP-seq, RNAseq, and eQTL analyses. RegulomeDB showed that SNP rs1811197 has the potential to affect antigen binding and is linked to expression of a gene target. We were unable to find information pertaining to the possible functional role for the other significant SNPs.

### DISCUSSION

In this study, significant associations were found between SNPs mapped to the MHC class I (HLA-E, HLA-B) and class II (HLA-DOA, HLA-DQA2, and HLA-DPB1) genes and DA in a group of exposed workers. Serum specific antibodies for diisocyanate antigens and the presence of eosinophils and activated T-cells in bronchial biopsies of workers with DA suggest a mechanistic role for antigen-specific immunological mechanisms.

T cells are activated by the interaction of the T-cell receptor with antigenic peptides complexed to MHC molecules. The class I (HLA-A, -B, -C, -E, -F, and -G) and class II (HLA-DR, -DQ, -DM, and -DP) MHC molecules are responsible for the presentation of antigenic peptides to CD8+ and CD4+ T cells, respectively. Since HLA molecules are highly polymorphic, specific peptide epitopes presented to T cells widely vary across the HLA genes and their alleles. The average SNP density varies from 1 to >60 SNPs per kb across the MHC region and these variations are located mainly in the class I and class II HLA molecules. Therefore, it is plausible that genetic variations in the HLA genes markedly influence individual susceptibility.

In our analysis, two SNPs in MHC class I genes, HLA-B and HLA-E, were independently associated with DA. The carriage of the minor allele for HLA-B rs1811197 SNP was associated with an increased risk of DA. Functional annotation of SNPs using RegulomeDB showed that the rs1811197 affects the expression level of the HLA-C gene. HLA-B and HLA-C are classical MHC class I molecules that play a central role in antigen processing/
presentation and immune regulation. Data on the association between HLA-B and HLA-C SNPs/alleles and asthma are restricted to a few studies. Beghe et al.27 examined possible associations of class I (HLA-A, HLA-B, and HLA-C) alleles with DA in 142 subjects with TDI-asthma and a comparator group of 50 asymptomatic exposed subjects. No significant associations were identified between HLA class I alleles and TDI-asthma.27 Nevertheless, associations between different HLA-B alleles (HLA-B8, B12, B16 and Bw61) and asthma phenotypes have been reported.28–30 HLA-E, a nonclassical MHC class I molecule, plays an important role in both natural and acquired immune responses by binding peptides derived from the leader sequence of other HLA class I molecules. We found rs1573294 SNP that mapped to HLA-E to be associated with an increased risk of DA. Although the functional consequence of this SNP is unknown, this association suggested a possible involvement of the HLA-E region in susceptibility to DA. A functional role for MHC class I molecules in the elicitation of DA has not been defined. Hypothetically, it is possible that reactive diisocyanates, known to penetrate cell membranes, could significantly modify endogenous cytosolic proteins, which could then be taken up by proteasomes and digested into antigenic peptides. Peptides could be transported to the endoplasmic reticulum, bound and captured by class I molecules, and transported to the cell surface for antigen presentation.

The HLA class II region is one of the most gene-dense regions in the human genome and is associated with many diseases and the dense LD pattern can complicate the identification of functional variants. We found a significant association between SNPs mapping to HLA-DPB1, HLA-DQA2, and HLA-DOA genes and increased risk of DA. The HLA-DPB1 rs928976 SNP has not been previously associated with any disease risk, and no functional data have been reported to date. Nevertheless, previous studies found a significant contribution of other HLA-DPB1 alleles to asthma risk. A haplotype including DPB1*05 (DRB1*15-DPB1*05) was strongly associated with TDI-induced asthma in a Korean population13 and the allelic frequencies of HLA DQB1*06-DPB1*05 and DRB1*15-DQB1*06-DPB1*05 were significantly higher in TDI asthmatic patients. The same investigators later conducted another study that included subjects with TDI asthma, asymptomatic exposed controls, and unexposed normal controls using high-resolution HLA analysis. The frequency of the HLA DRB1*1501-DQB1*0602-DPB1*0501 haplotype was found to be significantly higher in TDI asthmatic patients than in asymptomatic exposed and normal controls.12 Genetic variants in the HLA-DP locus were also associated with the risk of other asthma phenotypes, including allergic, pediatric, and aspirin-intolerant asthma.31–33 These reports support our finding and suggest involvement of HLA-DPB1 in asthma pathogenesis.

We also identified two SNPs mapping to the HLA-DOA and DQA2 genes that were associated with DA (rs3128935 and rs7773955, respectively). HLA-DO is a nonclassical class II heterodimer consisting of α and β chains, which are encoded by the HLA-DOA and HLA-DOB genes. HLA-DOA has been proposed to have functional implications in autoimmunity and can inhibit the activity of HLA-DM genes in vitro that regulates the antigen loading and presentation of specific peptides.34 A recent study identified an SNP in HLA-DOA gene (rs9276977) significantly associated with rheumatoid arthritis in African
Nevertheless, there are no studies showing association between HLA-DOA SNPs and asthma phenotypes.

HLA-DQ consists of α and β chains that are encoded by HLA-DQA1 and HLA-DQB1, respectively. These two loci are adjacent to each other and in close genetic linkage to HLA-DR. HLA-DQ was the first asthma-susceptibility locus to be identified and plays a major role in peptide loading of MHC II molecules. Our results showed a significant association between HLA-DQA2 rs7773955 SNP and DA risk. Although the functional role of this SNP is not known, previously reported associations between HLA-DQ alleles and asthma phenotypes suggest a role for DQ in the asthmatic process. Bignon et al. found that the HLA DQB1*0503 and the allelic combination DQB1*0201/0301 were associated with susceptibility to DA. On the contrary, the DQBI*0501 allele and the DQA1*0101-DQB1*0501-DR1 haplotype were reported to be protective. Although not replicated in other European populations, Mapp et al. confirmed these results and also reported an association between HLA-DQB1*0503 allele with an aspartic acid at residue 57 and TDI asthma. Previous genome wide association studies have also shown associations between SNPs in the HLA-DQ/DR region and asthma phenotypes. The major strengths of this study include a well-defined phenotype and selection of candidate regions based on their functional role in disease pathogenesis. In addition, genetic associations were tested while adjusting for potential confounding factors and results were corrected for multiple comparisons.

The major limitations include small sample size due to the relative rarity of DA compared with other types of asthma; however, rigorous phenotypic characterization in this population helps maximize the discriminatory potential between study groups. Another limitation is that the controls were younger and had shorter exposure period than cases. This was unintentional due to difficulty in the recruitment of age-matched workplace controls. Nevertheless, it has been reported that nearly 40% and 60% of subjects exposed to isocyanates become symptomatic within 1 year and after 5 years of exposure, respectively.

Taken together, this study showed novel associations between SNPs in MHC class I (HLA-E, HLA-B) and class II (HLA-DOA, HLA-DQA2, and HLA-DBP1) genes and DA. The present results are consistent with the hypothesis that an immunological mechanism is involved in DA and that genetic variations within HLA genes play a major role in DA risk. Identification of significant polymorphisms and their allelic variations within the MHC is potentially important as the structural diversity of the MHC alleles influences peptide binding and controls disease susceptibility. For example, hypersensitivity syndrome induced by antiviral drug abacavir is strongly associated with the HLA-B*5701 allele that excluding those with this particular allele prior treatment is effective in preventing drug reactions. Further studies are needed to validate the results reported herein and identify causative alleles behind these associations using high-resolution mapping.

Acknowledgments

This work was supported in part by an interagency agreement between National Institute of Environmental Health Sciences and National Institute for Occupational Safety and Health (Agreement No. Y1-ES0001), NIOSH/CDC R01 OH 008795 and CDC Seed Funding for Public Health Genomics Research Program.
References


Learning Objectives

- Discuss previous research on genetic factors potentially contributing to asthma caused by diisocyanates and other low-molecular-weight sensitizers.
- Summarize the new findings on single-nucleotide polymorphisms (SNPs) of the major histocompatibility complex (MHC) associated with diisocyanate-induced asthma (DA).
- Discuss the implications for the mechanism of DA and the role of genetic variations of human leukocyte antigen (HLA) genes.
## TABLE 1

The Demographic Characteristics of the Study Groups

<table>
<thead>
<tr>
<th></th>
<th>AWs (n = 67)</th>
<th>DA+ Cases (n = 73)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (mean ± SE)</td>
<td>30.0 ± 0.93</td>
<td>42.4 ± 1.32*</td>
</tr>
<tr>
<td>Sex (female/male)</td>
<td>5/62</td>
<td>10/63</td>
</tr>
<tr>
<td>Exposure (mean months ± SE)</td>
<td>63.4 ± 2.71</td>
<td>146.3 ± 16.36*</td>
</tr>
<tr>
<td>Exposure (HDI; MDI; TDI)</td>
<td>67 HDI</td>
<td>39; 15; 18</td>
</tr>
<tr>
<td>Ethnicity (% French Canadian)</td>
<td>99</td>
<td>92</td>
</tr>
<tr>
<td>Atopy (positive/negative)</td>
<td>36/27</td>
<td>41/32</td>
</tr>
<tr>
<td>Smoking Status (cur/ex/never)</td>
<td>26/13/28</td>
<td>11/26/36*</td>
</tr>
<tr>
<td>Smoking (pack/years ± SE)</td>
<td>6.2 ± 1.11</td>
<td>9.4 ± 1.66</td>
</tr>
</tbody>
</table>

*P < 0.05.

AW, asymptomatic workers; DA+, symptomatic workers diagnosed with DA; HDI, hexamethylene diisocyanate; MDI, 4,4′-diphenylmethane diisocyanate; TDI, toluene diisocyanate.
<table>
<thead>
<tr>
<th>Gene</th>
<th>SNP</th>
<th>Position</th>
<th>Location</th>
<th>Genotype</th>
<th>DA+ (n = 73) N(%)</th>
<th>AWs (n = 96) N(%)</th>
<th>Fisher Exact Test P (DA+ vs AWs)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HLA-B</td>
<td>rs181197</td>
<td>−2725</td>
<td>flanking→5′UTR</td>
<td>GG</td>
<td>40 (54.8)</td>
<td>62 (92.5)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>HLA-B</td>
<td>rs3573294</td>
<td>−70894</td>
<td>&gt;10 kb coding</td>
<td>GA</td>
<td>21 (28.8)</td>
<td>14 (20.9)</td>
<td>0.0003</td>
</tr>
<tr>
<td>HLA-B</td>
<td>rs3128935</td>
<td>−498</td>
<td>flanking→3′UTR</td>
<td>CT</td>
<td>47 (64.4)</td>
<td>65 (97.0)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>HLA-B</td>
<td>rs1770355</td>
<td>−2444</td>
<td>flanking→3′UTR</td>
<td>TT</td>
<td>26 (35.6)</td>
<td>56 (83.6)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td>HLA-B</td>
<td>rs180976</td>
<td>−496</td>
<td>intron</td>
<td>CC</td>
<td>32 (43.8)</td>
<td>11 (16.4)</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

AW, asymptomatic workers; DA+, symptomatic workers diagnosed with DA; HLA, human leukocyte antigen; SNP, single nucleotide polymorphism.
<table>
<thead>
<tr>
<th>Gene</th>
<th>dbSNP ID</th>
<th>Unadjusted OR (95% CI)</th>
<th>P</th>
<th>Adjusted * OR (95% CI)</th>
<th>P</th>
<th>Unadjusted OR (95% CI)</th>
<th>P</th>
<th>Adjusted * OR (95% CI)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>HLA-E</td>
<td>rs1573294</td>
<td>5.55 (2.68, 11.51)</td>
<td>&lt;0.0001</td>
<td>6.27 (2.37, 16.61)</td>
<td>0.0002</td>
<td>9.80 (3.21, 29.89)</td>
<td>&lt;0.0001</td>
<td>6.27 (1.63, 24.13)</td>
<td>0.0076</td>
</tr>
<tr>
<td>HLA-B</td>
<td>rs1811197</td>
<td>10.23 (3.68, 28.40)</td>
<td>&lt;0.0001</td>
<td>7.64 (2.25, 26.00)</td>
<td>0.0001</td>
<td>8.74 (0.46, 165.5)</td>
<td>0.0519</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HLA-DOA</td>
<td>rs3128935</td>
<td>17.98 (4.07, 79.48)</td>
<td>&lt;0.0001</td>
<td>19.69 (2.89, 135.25)</td>
<td>0.0024</td>
<td>15.22 (0.85, 271.9)</td>
<td>0.0093</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HLA-DQA2</td>
<td>rs7773955</td>
<td>9.20 (4.12, 20.57)</td>
<td>&lt;0.0001</td>
<td>8.43 (3.03, 23.48)</td>
<td>&lt;0.001</td>
<td>35.76 (2.09, 610.9)</td>
<td>0.0001</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HLA-DPB1</td>
<td>rs928976</td>
<td>3.63 (1.71, 7.72)</td>
<td>0.0009</td>
<td>2.79 (0.99, 7.81)</td>
<td>0.0511</td>
<td>13.00 (5.22, 32.39)</td>
<td>&lt;0.0001</td>
<td>10.10 (3.16, 32.33)</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

*Logistic regression models were adjusted for age, sex, atopy, height, exposure duration, and smoking.
†Logit estimators used to adjust for cells with 0 count.
‡Models failed to converge due to cells with 0 counts.

AW, asymptomatic workers; CI, confidence interval; DA+, symptomatic workers diagnosed with diisocyanate asthma; HLA, human leucocyte antigen; OR, odds ratio; SNP, single nucleotide polymorphism.