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The Role of Co-Infections in Mother-to-Child Transmission of HIV[§]

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Abstract

In HIV-infected women, co-infections that target the placenta, fetal membranes, genital tract, and breast tissue, as well as systemic maternal and infant infections, have been shown to increase the risk for mother-to-child transmission of HIV (MTCT). Active co-infection stimulates the release of cytokines and inflammatory agents that enhance HIV replication locally or systemically and increase tissue permeability, which weakens natural defenses to MTCT. Many maternal or infant co-infections can affect MTCT of HIV, and particular ones, such as genital tract infection with herpes simplex virus, or systemic infections such as hepatitis B, can have substantial epidemiologic impact on MTCT. Screening and treatment for co-infections that can make infants susceptible to MTCT *in utero*, peripartum, or postpartum can help reduce the incidence of HIV infection among infants and improve the health of mothers and infants worldwide.

Keywords

Co-infections; HIV; infections; infant; mother-to-child transmission

INTRODUCTION

Despite the success of antenatal HIV screening and prevention of mother-to-child transmission of HIV (PMTCT) programs, in 2009, an estimated 370,000 children became infected during the perinatal or breastfeeding period [1]. Highly effective interventions for PMTCT, such as antiretroviral agents during pregnancy and labor [2-9], caesarean section delivery [10-13], postexposure prophylaxis to the infant [14-18], and avoidance of breastfeeding [19, 20], can reduce risk for mother-to-child transmission of HIV (MTCT) to

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CONFLICT OF INTEREST

The authors confirm that this article content has no conflicts of interest.

PATIENT CONSENT

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HUMAN/ANIMAL RIGHTS

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<2% [21-23]. Unfortunately, these interventions are not uniformly administered, particularly in resource-limited countries with the greatest need [24].

Effective PMTCT interventions address the main risk factors for MTCT, which are maternal HIV load and infant exposure to infected fluids [25, 26]. High maternal HIV load is the strongest risk factor for MTCT, and reduction of HIV load with antiretroviral interventions significantly reduces risk [27-34]. Infant exposure to infected fluids during labor and delivery is associated with duration of ruptured membranes [35], type of delivery [10, 11], and presence of other active sexually transmitted infections (STIs) in the vaginal canal [25, 36, 37]. Postnatal exposure to HIV-contaminated milk is correlated with duration of breastfeeding, which is associated with MTCT [20]. Without PMTCT intervention, the overall risk for HIV transmission to the infant is 25%-40%, with an estimated 10%-25% of infections occurring during pregnancy, 35%-40% during labor and delivery, and 35%-40% during breastfeeding [26, 38].

In the absence of antiretroviral treatment, natural defense mechanisms can protect against MTCT. The human placenta is an efficient barrier to the exchange of maternal and fetal circulations, despite HIV tropism for placental cells and frequent infection of the placenta [39]. Postpartum, the blood-milk barrier keeps the HIV load in breast milk generally 100 times lower than the HIV load in plasma [40], possibly because of the many antimicrobial and immunomodulatory factors in breast milk [41, 42], including cell-free HIV inhibitors that likely inactivate many potentially infectious virions [43-45]. Disruption of natural barriers to infection occurs during chorioamnionitis, labor and delivery, or mastitis, and these barrier breaches can increase risk for MTCT. Co-infections can trigger the release of inflammatory agents that increase tissue permeability and cytokines that stimulate HIV replication [46, 47]. Several co-infections in the HIV-infected mother or exposed infant have been shown to increase MTCT risk, and addressing these co-infections may reduce MTCT and improve overall maternal and child health (Table 1) [48].

MATERNAL REPRODUCTIVE TRACT INFECTIONS

Infections of the Placenta and Fetal Membranes

In HIV-infected, pregnant women who are not taking antiretrovirals, the placenta is a reservoir for HIV replication [49]; however, it can also provide an efficient barrier to HIV transmission for more than 90% of exposed infants [39]. Evolution of the placenta's structure and function during gestation generates a barrier between maternal and fetal circulations through which HIV must cross by endocytosis or by an injured villous surface, such as occurs in chorioamnionitis. The placenta also confers protection by producing soluble factors and receptors, such as cytokines, chemokines, and MHC class I molecules. Co-infections of the placenta may affect its anti-HIV properties by altering local expression of cytokines, chemokines, and their co-receptors, as evidenced by increased expression of the CCR5 HIV co-receptor on placental macrophages infected with malaria in a *Plasmodium falciparum* dominated region [50], or by stimulating HIV replication, as evidenced *in vitro* with an antigen of *P. falciparum* [51].

Chorioamnionitis is inflammation of the membranes surrounding the fetus that is caused primarily by infection ascending from the lower genital tract. It can be caused by several different microorganisms, such as bacterial vaginosis-associated bacteria, *Neisseria gonorrhoea*, *Chlamydia trachomatis*, *Trichomonas vaginalis*, or *Group B Streptococcus* [52]. Chorioamnionitis may disrupt the placental barrier allowing HIV-infected maternal lymphocytes to enter the amniotic fluid. It has been associated with increased intrauterine risk for MTCT [53-57], but it has not necessarily been found to be an independent risk factor [29, 58]. Chorioamnionitis is also associated with preterm labor and premature rupture of the membranes [59]. These conditions have been associated with increased MTCT [60-63], but the causality of the association is unclear because HIV infection predisposes women to preterm delivery [64]. In a Kenyan study of 250 mother-infant pairs, histologic chorioamnionitis was found to be an independent risk factor for peripartum MTCT (adjusted odds ratio [AOR], 3.9; 95% confidence interval [CI], 1.2-12.5) after researchers assessed the effect of prolonged rupture of membranes and controlled for low infant birth weight and maternal plasma and genital tract HIV load [53]. Similarly, in a study conducted at seven sites in the United States (n=175), acute histologic chorioamnionitis was found significantly more often in placental tissue from HIV-infected mothers who transmitted the virus to their infants than from mothers who did not transmit the virus to their infants (37.5% vs 11.3%, p=0.008), and it remained a predictor after adjustment for HIV load and prolonged rupture of membranes [54]. In this study, histologic chorioamnionitis was significantly correlated with clinical chorioamnionitis, but clinical chorioamnionitis was not a predictor of MTCT. This study also assessed whether HIV load influenced the association between histologic chorioamnionitis and transmission and found no interaction. A study in former Zaire (now DRC) found chorioamnionitis to be associated with MTCT in both univariate and multivariate analyses (AOR, 2.5; 95% CI, 1.2-5.2), with a significant interaction between maternal immune status and the effect of chorioamnionitis; only among nonimmunocompromised women (as indicated by neither a low CD4+ T lymphocyte count nor an elevated CD8+ T lymphocyte count) was chorioamnionitis associated with significantly increased risk of HIV transmission (relative risk, 4.2; 95% CI, 1.3-13.7) [55]. An interaction between maternal immune status and the effect of chorioamnionitis on MTCT was also observed in a Ugandan study in which, in the absence of immune suppression, the AOR for MTCT was 2.87 (95% CI, 1.04-7.90) for women with chorioamnionitis when they were compared with women who did not have chorioamnionitis [56]. These data suggest that inflammation of the placental membranes may have a greater effect on risk for transmission among women who might otherwise be at low risk for perinatal transmission. Using the assumption that the association between chorioamnionitis and HIV transmission among nonimmunocompromised women is causal, the authors in the Ugandan study estimated that up to 34% of HIV transmissions could be prevented with treatment of placental inflammation in non-immunocompromised women. However, a randomized controlled trial that evaluated two courses of antibiotic treatment at 20-24 weeks gestation (metronidazole 250 mg and erythromycin 250 mg, three times per day, orally for 7 days) and during labor (metronidazole 250 mg and ampicillin 500 mg every 4 hours) among HIV-infected and uninfected pregnant women in Africa failed to show an effect on histologic chorioamnionitis or risk for MTCT [65, 66]. As a result, the study was terminated early by the data safety and monitoring board overseeing the research. This trial, as well as

an observational cohort study in Africa that systematically screened and treated common STIs, documented no difference in MTCT by the presence of histologic chorioamnionitis [66, 67].

Lower Genital Tract Infections

Genital tract infections increase intrapartum risk for MTCT by increasing infant exposure to HIV in genital secretions from the mother during labor and delivery. Common vaginal infections such as bacterial vaginosis (BV) and vulvovaginal candidiasis, STIs such as trichomoniasis and herpes simplex virus type 2 (HSV-2), and cervicitis are associated with increased HIV shedding in the genital tract that declines upon treatment for these infections [26, 68-72].

HSV-2 is the leading cause of genital ulcers. With a seroprevalence of 60%-90% among HIV-infected populations, it likely has a significant effect on HIV infection [73]. HSV-2 ulcers contain high levels of HIV RNA, possibly from the homing of activated T cells [74, 75], and also provide a breach in the mucosa that increases infant exposure to HIV virions or infected cells beneath the epithelium. Consequently, genital ulcers are an important risk factor for MTCT [25,36,37,76]. Even among clinically asymptomatic, serologically positive women, HSV-2 shedding remains common and is associated with increased HIV shedding in the genital tract of co-infected women [77, 78]. Two studies, one in the United States and one in Kenya, found that clinical HSV-2 was associated with increased MTCT but asymptomatic HSV-2 was not [36, 37]. More recently, two nested case-control studies from Thailand and Zimbabwe found that serologic HSV-2 was associated with increased risk for MTCT, independent of maternal HIV load [76, 79]. In the Thai study, genital HSV-2 shedding was independently associated with a threefold increase in intrapartum HIV transmission [76]. In the Zimbabwe study, an estimated 28.4% of intrapartum HIV transmissions were potentially attributable to serologic HSV-2 infection [79]. Asymptomatic HSV-2 infection may affect MTCT through more generalized effects on the mother's HIV load because of immune activation or direct interactions between the viruses [80]. Plasma HIV load increases upon subclinical HSV-2 reactivation and declines upon successful viral suppression with acyclovir treatment [81-83]. Clinical trials have shown that HSV-2 suppressive therapy significantly reduces plasma and cervical HIV load [46, 83-88]. However, results from clinical trials of acyclovir on sexual transmission of HIV have shown no reduction in acquisition or transmission [86,89,90].

Acyclovir is actively transported to the amniotic fluid and breast milk [91], and both acyclovir and valacyclovir, which is rapidly absorbed and converted to acyclovir, have good safety profiles in pregnant women and have not been associated with congenital malformations or infant toxicity [92-94]. The maternal and infant safety profile for maternally administered valacyclovir suppressive therapy was not altered in the context of antiretroviral prophylaxis for PMTCT in a randomized trial [95]. In another report from this trial, which was conducted among pregnant and postpartum Kenyan women co-infected with HSV-2 and HIV, valacyclovir significantly reduced HIV load in plasma by 0.5 log₁₀ copies/mL during pregnancy and after 6 weeks postpartum, compared with the placebo, and reduced the levels of HIV load in breast milk at 6 and 14 weeks postpartum [96].

Theoretically, suppression of HIV load in plasma and breast milk should translate to reduced MTCT. The Kenyan trial found no difference in HIV transmission at 12 months, but the study was not designed to answer this question. Larger clinical trials are warranted because suppressive therapy for HSV-2 could be easily administered within the existing PMTCT infrastructure and could provide maternal benefits of reduced frequency and severity of HSV-2 reactivations, which may slow HIV disease progression, and reduce the risk for HIV transmission.

Syphilis, caused by infection with the bacterium *Treponema pallidum*, is a less common cause of genital ulcer disease, and its role in MTCT is unclear [79, 97-100]. One large prospective cohort study conducted in Malawi reported that syphilis was associated with a 2.7-fold increase in both intrauterine (HIV-positive at birth) and intrauterine/postpartum (HIV-positive at 6 or 12 weeks postpartum) MTCT after adjustment for maternal HIV load and other confounders [97]. A study in Ukraine in which researchers were unable to adjust for maternal HIV load reported that serologically confirmed syphilis was associated with a fivefold increase in MTCT [98]. However, studies conducted in Zimbabwe and Texas found no association between active syphilis and perinatal HIV transmission [79, 100]. These findings are consistent with the Rakai trial of presumptive treatment of STIs, which reported reductions in maternal rates of bacterial STIs but not in perinatal HIV transmission [101].

Human papilloma virus (HPV) infection is the most common STI, and its prevalence, incidence, and persistence are highly correlated with HIV infection and immune status [102, 103]. Some HPV types may cause genital warts, and two studies have reported a significant, unadjusted association between genital warts and increased risk for MTCT among co-infected women [54, 104]. This finding may be secondary to genital warts acting as a proxy for advanced HIV disease and higher maternal HIV load.

In women who have BV, the healthy vaginal flora is replaced with high levels of anaerobic microorganisms and byproducts that can damage vaginal epithelium and degrade cervical mucus. Evidence suggests that BV increases women's risk for STIs, including gonorrhea, chlamydial infection, trichomoniasis, HPV, herpes simplex virus, and HIV [105-109]. Among HIV-infected women, BV is associated with increased shedding of HIV in the genital tract. In a recent prospective study in Kenya, HIV-infected women diagnosed with BV at 32 weeks gestation had a threefold higher risk for *in utero* HIV transmission than women with normal vaginal flora [110]. However, a multi-site randomized controlled trial of metronidazole versus placebo found no difference in MTCT rates, despite a 16% reduction in BV [66]. Further studies are needed to answer the question of whether restoration of normal vaginal flora can reduce MTCT.

Treatment of concurrent genital infections can reduce genital HIV shedding [72], but whether this translates to reduced MTCT is unknown. Besides the aforementioned treatments for specific infections, randomized trials of virucidal vaginal washes, such as benzalkonium chloride or diluted chlorhexidine, on MTCT have failed to consistently demonstrate an effect [111-114].

MASTITIS

Breast milk contains both antimicrobial and immunomodulatory factors that protect infants against various infectious diseases and support the development of the infant's immune system. Because breast milk contains cell-free HIV inhibitors and because of the milk-blood barrier [42], the concentration of HIV is typically 10-100 times lower in breast milk than in plasma [34]. However, infection or inflammation of breast tissue can increase the HIV load in breast milk, which is a strong predictor of postnatal transmission [34].

An estimated 10%-33% of women experience mastitis, typically during the early breastfeeding period or during the mixed feeding and weaning stages. Clinical mastitis may be characterized by cracked nipples and sores, suppuration, pain, swelling, and erythema. Cracked nipples frequently bleed during infant feeding, thereby increasing the exposure to plasma and cell-associated virus, and several studies have reported associations between cracked or bleeding nipples and increased postnatal transmission of HIV [115, 116]. Subclinical mastitis is characterized by an elevated milk leukocyte count or elevated sodium or sodium/potassium ratio. Like clinical mastitis, it can alter the cellular tight junctions that regulate breast epithelial permeability. Both clinical and subclinical mastitis are associated with increased HIV load in breast milk [117-122] and its correlate, postnatal HIV transmission [25, 116, 117, 119, 121], and researchers have estimated that up to 50% of postnatal transmissions may be attributable to the more common subclinical mastitis [123].

Most mastitis is sterile inflammation with isolation of only normal skin flora, but up to 40%-50% of subclinical and clinical mastitis cases may be attributable to *Staphylococcus aureus* [124, 125]. A study among 75 HIV-infected, postpartum women with subclinical mastitis in Malawi evaluated the effect of oral amoxicillin/clavulanic acid on breast milk leukocyte count and HIV load. One week after treatment, there was a >50% reduction in the proportion of mothers with elevated breast milk leukocyte counts; after 4-12 weeks, study results showed a significant reduction in breast milk HIV load compared with baseline data [126]. However, the HIV load remained higher among women with past clinical mastitis than among those without mastitis at baseline. These findings suggest that the effect of inflammation on the alveolar ducts persists for several weeks after treatment.

Interestingly, mastitis has been associated with increased breast milk concentrations of α -defensins, a group of anti-HIV peptides released by the innate immune system, and researchers have suggested that mastitis may provide a protective effect when HIV load is low in plasma [127]. An analysis of plasma and breast milk samples collected as part of a study in Zimbabwe found that mastitis was associated with postnatal transmission of HIV only when HIV load was high in maternal plasma ($>3.7 \log_{10}$ copies/mL) [121]. For women with HIV loads $<3.7 \log_{10}$ copies/mL, the point estimate for the odds of postnatal transmission was in the direction of a protective effect (0.26; 95% CI, 0.04-1.94). These data suggest that, when HIV loads are low in maternal plasma, the increased immune factors in breast milk associated with mastitis may outweigh the effect of any increase in HIV replication.

GINGIVITIS

In many parts of the world, including sub-Saharan Africa, it is common for mothers to pre-masticate food for their infants during the weaning period [128]. In 2009, there was a case series report of three pediatric HIV infections among infants who were HIV-negative at birth, not breast fed, and had no known causes of infection besides exposure to food pre-masticated by a HIV-infected caregiver [129]. For two of the infants, it was a HIV-infected mother who pre-masticated the food; for the other infant of an HIV-negative mother, it was an HIV-infected great aunt. In two of the cases, there was evidence of bleeding gums. The exposure to blood in pre-masticated food along with the compromised oral mucosa of an infant may facilitate transmission via pre-masticated food. A case-control investigation in 2010 at 6 U.S. HIV clinics found no significant difference between 11 infants with late-diagnosed HIV-infection and 35 HIV-exposed, uninfected infants in the proportion that were fed food pre-masticated by an HIV-infected caregiver (27% vs 20%, respectively) [130]. This report also presented the findings from a cross-sectional investigation of pre-mastication prevalence, which was 31% among 154 primary caregivers of children aged 6 months. Pre-mastication appears to be a rather common practice of caregivers, but it is contraindicated for HIV-infected caregivers [131].

MATERNAL SYSTEMIC CO-INFECTIONS

As mentioned, maternal HIV load is highly correlated with risk for HIV transmission to the infant [29, 30, 132-134], and concurrent, systemic maternal infections can inadvertently stimulate HIV replication and increase HIV load. Any infection that raises maternal plasma HIV load, and its correlates, HIV load in the genital tract and in breast milk, could affect the risk for HIV transmission.

Hepatitis B

Up to 90% of HIV-infected persons worldwide have serologic evidence of hepatitis B virus (HBV) infection, and 10% have chronic HBV [135, 136]. Co-infection with HIV accelerates the rate of disease progression of HBV infection and is associated with an increased carriage rate for the hepatitis B e antigen, chronic HBV infection, and high HBV DNA levels, all of which are associated with increased risk for MTCT of HBV [136-141]. Among women with very high HBV DNA levels during pregnancy ($>10^9$ copies/mL), HBV transmission to the infant is substantial (as high as 32%) despite immunoprophylaxis with vaccine and immunoglobulin [142]. Although HIV/HBV co-infection may be related to increased HBV transmission to the infant, HBV infection, as measured by the presence of hepatitis B surface antigen, does not seem to be independently associated with increased MTCT of HIV [143, 144]. However, women co-infected with HIV/HBV are significantly more immunosuppressed than HIV-monoinfected women, which is an independent risk factor for MTCT [145].

Hepatitis C

Although hepatitis C virus (HCV) infection is less common than HBV, rates of co-infection among HIV-infected persons are about 30% worldwide, and the prevalence of HCV

infection is particularly high among HIV-positive injection drug using women [145, 146]. HIV-infected persons exposed to HCV are less likely to clear the virus spontaneously, and upon infection, co-infected persons have increased HCV RNA levels in plasma and progress to cirrhosis faster than HCV-monoinfected persons [147-149]. Increased maternal HCV RNA levels are associated with perinatal transmission of HCV [150-154], and HIV-co-infected mothers are at increased risk for perinatal transmission of HCV [150, 151, 155, 156]. With respect to the role of HCV infection on MTCT of HIV, HIV disease progression, and death, the data are conflicting. HCV seropositivity is associated with severe immunosuppression among HIV-infected persons ($p < .001$) [145], and several studies have documented increased risk for MTCT of HIV among co-infected women [155-159]. The likely mechanism for increased MTCT associated with HCV seropositivity is increased maternal HIV load.

GB Virus C

GB virus C (GBV-C) is a flavivirus closely related to HCV that was discovered in 1995 [160]. Although no clinical disease has been associated with GBV-C, it can be transmitted vertically and has been associated with reduced MTCT of HIV [161, 162].

Cytomegalovirus

Cytomegalovirus (CMV) seropositivity in HIV-infected adults is almost universal, and the two viruses can infect the same cell types, with the viral products of each virus capable of activating the other virus *in vitro*. Researchers have hypothesized that the two infections are interrelated, with each virus predisposing a person to a higher risk of contracting the other virus. The prevalence of congenital CMV infection among infants born to HIV-infected women is relatively high, with rates ranging from 2% to 7% [163-168]; rates in the general newborn population are 0.7% [169]. A simple bivariate analysis of data from eight studies ($n=6,265$) suggests that congenital CMV infection occurs significantly more often among HIV-infected infants (9%) than those who are not infected (2%) [165], but this comparison was heavily weighted by the largest study of 4,797 infants in the French Perinatal Cohort Study. Findings from that study also suggest a role of maternal immunosuppression in increased CMV transmission based on a lower risk for congenital CMV infection among HIV-infected mothers after highly active antiretroviral therapy became available [166]. A recent study in Thailand of 97 HIV-infected infants and 196 HIV-exposed uninfected infants matched for maternal HIV load reported that congenital CMV infections were more common in HIV-infected (14%) than HIV-uninfected (3%) infants [170]. This study also examined the timing of CMV infection relative to HIV infection and found that *in utero* CMV infection correlated with both *in utero* and intrapartum HIV infection, whereas intrapartum CMV infection correlated with intrapartum, but not *in utero*, HIV infection, suggesting that fetal CMV infection may predispose infants to *in utero* HIV infection. *In vitro* studies have also described the ability of CMV to facilitate HIV infection [171-173]. Infants with CMV and HIV infection may be more prone to rapid progression of HIV disease [167].

Epstein-Barr Virus

Another almost universal HIV co-infection is Epstein-Barr virus (EBV). In a multi-site U.S. study of 279 HIV-infected, pregnant women who were also EBV seropositive, EBV shedding was marginally associated with MTCT of HIV [61]. However, this association did not reach statistical significance in a multivariate analysis that adjusted for maternal immune status and other confounders.

HHV-6

HIV-infected mothers also seem to transmit human herpes virus 6 (HHV-6) to infants more frequently than HIV-uninfected mothers [174]. However, a study of perinatally HIV-infected infants from Thailand found the rate of HHV-6 infection was lower in HIV-infected children, but that HHV-6 co-infection correlated with faster progression of HIV disease [175].

HHV-8

Cross-sectional studies have demonstrated a significant association between the seroprevalences of HIV and HHV-8, also known as Kaposi's Sarcoma-associated herpesvirus [176-179]. In Sub-Saharan Africa, where HHV-8 is highly prevalent, infection is commonly acquired in childhood through exposure to infected saliva from other children or caregivers, with the seroprevalence generally increasing with age [180-182]. Vertical transmission of HHV-8 during birth or through breast milk is rare [177, 180-185]; HHV-8 is rarely detected in breast milk [186]. Two large studies, a prospective study in Zambia [184] and a cross-sectional study in South Africa [187], found that maternal HHV-8 status was not an independent risk factor associated with HHV-8 transmission to children, suggesting that other household or non-familial contacts contribute to horizontal transmission. These studies also reported that maternal HIV status does not affect infant HHV-8 status. Infant HIV status, however, was an important predictor, and both studies reported a significant increased risk for HHV-8 among HIV-infected children [184, 187]. A small study of 15 co-infected, pregnant women in Italy found that HHV-8 load significantly increased late in pregnancy and was associated with a significant increase in HIV-1 shedding in the genital tract, suggesting that HHV-8 co-infection may increase MTCT of HIV [188]. However, results from a study in Zambia failed to show an association between HHV-8 infection and transmission of HIV to the infant [177].

Tuberculosis

Tuberculosis (TB) is a leading cause of disease and death worldwide and is of particular concern among those infected with HIV, who are 20-30 times more likely to develop TB [189]. For women, the greatest burden of TB occurs during the reproductive years (ages 15-49 years) [190]. Active TB infection increases HIV load and is associated with immunosuppression, which may explain the association between TB and MTCT [191-195]. To date, few studies have investigated whether TB increases the risk for MTCT independently of HIV load. A small study of 42 HIV-infected, pregnant women with active TB in South Africa reported a 19% rate of *in utero* transmission, which was higher than the 5%-10% rate reported in resource-limited countries at that time [195]. A recent study in

India found that maternal TB was associated with a 2.5-fold increase in the odds for MTCT after adjusting for maternal and infant factors [196].

Malaria

Populations with a high prevalence of both HIV and parasitic infections overlap geographically and socio-economically. In addition, the level of immune compromise associated with HIV increases a person's susceptibility to parasitic diseases. Concurrent infection causes chronic immune activation, which increases the risk for reactivation of both infections and higher HIV load.

In the most severely affected countries in Sub-Saharan Africa, more than 10% of adults are HIV-infected and more than 90% are exposed to malaria. Pregnant women are at increased risk for malaria which is associated with intrauterine growth retardation, preterm delivery, low birth weight, still birth, early neonatal death, and maternal anemia [197, 198], especially among primigravidae women, among whom malaria tends to be more severe. HIV infection further increases the risk for malaria parasitemia and clinical malaria, and among pregnant women, the risk of placental malaria and adverse birth outcomes [199-202]. Furthermore, new evidence suggests that malaria parasites may be more likely to develop wild-type mutations after exposure to sulfadoxine-pyrimethamine as intermittent preventive treatment during pregnancy among HIV-infected women [203]. Malaria also has been associated with a temporary increase in HIV replication and plasma HIV load [204, 205], and associations with blood parasitemia and MTCT of HIV may be secondary to an increase in maternal HIV load. Placental malaria has been associated with increased placental and plasma HIV load [206]. Several studies have examined the association between placental malaria and MTCT of HIV independently of HIV load or CD4+ T lymphocyte count, with inconsistent results [207-212]. The effect may be limited to cases with high parasitemia (>10,000 parasites/ml), as suggested by a study in Western Kenya, where the risk for MTCT increased when parasitemia was high but decreased when it was low (<10,000), compared with malaria-negative controls [202]. Other studies have shown associations between placental malaria and placental characteristics that may increase susceptibility to *in utero* HIV infection, such as placental inflammation [213], increased CC-chemokine production [214, 215], a shift in cytokine production from Th2 to Th1-type responses [216], and increased expression of the CCR5 HIV co-receptor on placental macrophages [50]. Inconsistencies in study results may be due, in part, to differences in the epidemiology of malaria in different settings, which could affect maternal immunity, or to different methods of detecting placental parasitemia. Although it remains unknown whether malaria increases MTCT, malaria during pregnancy is associated with obstetrical problems and adverse birth outcomes and warrants extensive screening and intervention programs in endemic areas.

Other Parasitic Co-Infections

Toxoplasmosis is a parasitic disease caused by the protozoon *Toxoplasma gondii*, and it is estimated that 25% to 30% of people world wide are infected [217]. Congenital toxoplasma infection results from primary maternal infection or reactivation of a past infection during pregnancy, with the risk of congenital toxoplasma transmission ranging from 10% during the first trimester, when the placental barrier is most efficient, to 60-70% in the third

trimester [218]. Conversely, the severity of fetal damage is greatest when infection occurs earlier in gestation, which often leads to severe abnormalities or abortion [219]. In HIV-infected women, the risk of toxoplasmosis, primarily from reactivation of a past infection, increases substantially when the CD4+ T lymphocyte count falls below 100 cells/ μ l. In the pre-HAART era, there were case reports of transmission of both toxoplasmosis and HIV to infants of co-infected mothers, with many of these infants having rapid HIV disease progression [220, 221]. A recent retrospective study of HIV-infected pregnant women and their infants in Brazil evaluated predictors for MTCT of HIV, which occurred among 3.7% (15/401) of infants. Maternal neurotoxoplasmosis during gestation was associated with a 7 times higher risk for infant HIV infection, and congenital toxoplasmosis, which occurred in 6 infants, was associated with 24 times higher risk of infant HIV infection [222].

The protozoan parasite *Trypanosoma cruzi*, which is endemic to Latin America, causes Chagas disease and can be transmitted transplacentally to the fetus. Human placental histocultures co-infected with *T. cruzi* and HIV have shown lower levels of chemokines that downmodulate *T. cruzi* replication (IL-6, IL-8, IP-10, and MCP-1) [223]. Another study suggests that *T. cruzi* inhibits HIV replication at the placental level [224]. However, simultaneous congenital transmission of *T. cruzi* and HIV has been reported [225].

Helminth infections are very prevalent in areas of Asia and Africa [226-229]; they affect immune homeostasis [230], which may influence risk for HIV acquisition and progression. One study found that women with HIV and helminth infections had a higher risk for MTCT of HIV [231]. Two randomized clinical trials have evaluated treatment of helminthic infections as a strategy to reduce the progression of HIV disease and found improvements in immune responses and CD4+ T lymphocyte counts in persons infected with some helminths [232, 233]. Periodic deworming is now recommended in Sub-Saharan Africa as a component of comprehensive HIV care for women and children.

INFANT CO-INFECTIONS

Although some infants become infected with HIV *in utero*, the majority are infected perinatally or postnatally. During labor and delivery, infants whose mothers have HIV are exposed to infected blood and vaginal secretions that may be inadvertently ingested. Prematurity of the skin, mucous membranes, and gastrointestinal tract is the likely mechanism for increased risk of HIV infection to premature infants. In newborns, the gastrointestinal tract is immature: it has a thin mucosa; lower levels of gastric acidity, enzyme activity, and mucus production; and no secretory IgA. These factors increase gastrointestinal permeability and make it possible for HIV to traverse the epithelium to the lamina propria, where HIV likely infects lymphocytes or is taken up by macrophages [234]. Inflammation of the gastrointestinal tract due to infection could weaken barrier defenses and facilitate passage of HIV. Some infants may develop oral candidiasis upon colonization from exposure to *Candida* during vaginal delivery. Candidiasis results in inflammation of the infant's oral mucosa and gastrointestinal tract and an influx of activated CD4+ T lymphocytes and macrophages; it has also been associated with increased risk for postnatal HIV transmission [116].

Diarrhea is associated with increased intestinal permeability [235], and any infection that causes diarrhea has the potential to increase MTCT to exposed infants. Although studies on the effects of diarrheal diseases on MTCT are lacking, increased intestinal permeability due to contamination of other liquids and foods given to the infant is a proposed biological mechanism for the increased risk for postnatal HIV transmission during the first few months of life among infants in resource-limited countries who are fed a mix of breast milk and other foods compared with infants who are exclusively breastfed [20, 236-238]. Mixed feeding poses the same risks for contamination and diarrhea as artificial feeding, but also increases intestinal permeability and risk for HIV infection because of continued exposure to HIV-infected breast milk.

Co-infections in HIV-infected infants are thought to in part explain the higher risk for death by 2 years of age among untreated, HIV-infected children in Sub-Saharan Africa (45%-59%) compared to those in Europe and the United States (10%-20%) [239-245]. A major cause of stillbirth and neonatal death is untreated maternal syphilis [246]. HIV-infected women have a higher prevalence of untreated or inadequately treated syphilis during pregnancy, which places their newborns at higher risk for congenital syphilis and its outcomes, including blindness, deafness, progressive intellectual deterioration, and death [100]. In Sub-Saharan Africa, up to 80% of children may acquire CMV during their first year of life, regardless of HIV status [247], and MTCT appears to be more frequent among infants with congenital CMV infection [167]. HIV and CMV co-infection in infants is associated with higher peak CMV viral loads, prolonged detection of CMV in plasma, rapid progression of HIV disease, and a greater than twofold increase in risk for death [167, 247, 248]. Other perinatal viral infections, such as HHV-6, have also been associated with faster progression of HIV disease [175].

STRATEGIES FOR PREVENTION AND TREATMENT

Antiretroviral treatment for HIV-infected women during pregnancy, delivery, and breastfeeding decreases the risk for MTCT by reducing the HIV load in plasma and breast milk, and it improves maternal health by reconstituting CD4+ T lymphocytes and lowering her risk for opportunistic infections. Administration of antiretrovirals is the most important intervention to decrease MTCT, and these medications should be made available to pregnant women in all settings.

Existing PMTCT programs could also incorporate available, inexpensive, and safe strategies for the prevention and treatment of co-infections during pregnancy and breastfeeding to further reduce MTCT of HIV. The World Health Organization (WHO) currently recommends prevention, screening, and treatment for certain infections at antenatal consultations [249]. In areas with endemic malaria, WHO recommends that pregnant women sleep under insecticide-treated mosquito bed nets and receive 2-3 treatment courses of sulfadoxine-pyrimethamine. Screening and treatment for syphilis are recommended at the first antenatal visit and ideally repeated at 36 weeks or delivery in areas of high risk. For other curable genital tract infections, WHO suggests using syndromic-based algorithms for diagnosis and treatment [250]. However, syndromic management has poor sensitivity and specificity for many of these infections, which can lead to overtreatment or under treatment

in many cases [251, 252]. Several trials are evaluating the use of azithromycin-based combination therapies for intermittent use during pregnancy. Azithromycin has shown high cure rates (>96.5%) for syphilis, *N gonorrhoeae*, and *C trachomatis*, as well as efficacy in reducing risk for *T vaginalis* [253-256]. A meta-analysis of antibiotic therapy for treatment of BV in pregnancy showed high efficacy (odds ratio, 0.17: 95% CI, 0.15-0.20; 10 trials, 4,357 women) and, when administered before 20 weeks, significantly reduced the risk for preterm birth [257]. Postpartum, WHO recommends prophylaxis against bacterial and protozoan opportunistic infections with cotrimoxazole for HIV-infected mothers and for HIV-infected or exposed infants from age 6 weeks throughout breastfeeding and longer for infants that are HIV-infected [258].

Given the limitation of syndromic management of certain co-infections, presumptive, intermittent treatment of prevalent co-infections for all women may be the preferable approach to improving pregnancy outcomes and reducing MTCT of HIV in resource-limited countries [259]. Alternatively, novel rapid diagnostic tests are becoming increasingly available, and frequent screening throughout the antenatal period and targeted treatment may be a better option in some settings. Testing of HIV-infected women for HBV infection during pregnancy should be part of routine antenatal care in all settings. This practice can guide decisions on selection of appropriate antiretroviral regimens and ensure timely administration of HBV vaccine and immunoglobulin to exposed infants [141]. Finally, ensuring that women are immunized against important pathogens, such as tetanus, influenza, or pertussis, before or during pregnancy can help prevent infections that are associated with concurrent immune activation and transient increases in HIV load during pregnancy. Immunization of the mother can also protect infants from these pathogens. Expanding HBV and HPV vaccine programs to cover young women before pregnancy has the potential to offer far-reaching health benefits at the population level.

CONCLUSION

Screening and treatment for co-infections that predispose infants to *in utero*, peripartum, or postpartum HIV infection can help to further reduce the incidence of HIV among infants while providing health benefits to both the mother and infant.

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REFERENCES

- [1]. UNAIDS Report on the Global AIDS Epidemic. 2010.
- [2]. Connor EM, Sperling RS, Gelber R, et al. Pediatric AIDS Clinical Trials Group Protocol 076 Study Group. Reduction of maternal-infant transmission of human immunodeficiency virus type 1 with zidovudine treatment. *N Engl J Med.* 1994; 331:1173–80. [PubMed: 7935654]
- [3]. Shaffer N, Chuachoowong R, Mock PA, et al. Bangkok Collaborative Perinatal HIV Transmission Study Group. Short-course zidovudine for perinatal HIV-1 transmission in Bangkok, Thailand: a randomised controlled trial. *Lancet.* 1999; 353:773–80. [PubMed: 10459957]
- [4]. Dabis F, Msellati P, Meda N, et al. 6-month efficacy, tolerance, and acceptability of a short regimen of oral zidovudine to reduce vertical transmission of HIV in breastfed children in Cote

- d'Ivoire and Burkina Faso: a double-blind placebo-controlled multicentre trial. DITRAME Study Group. *Diminution de la Transmission Mere-Enfant*. *Lancet*. 1999; 353:786–92. [PubMed: 10459959]
- [5]. Wiktor SZ, Ekpini E, Karon JM, et al. Short-course oral zidovudine for prevention of mother-to-child transmission of HIV-1 in Abidjan, Cote d'Ivoire: a randomised trial. *Lancet*. 1999; 353:781–5. [PubMed: 10459958]
- [6]. Guay LA, Musoke P, Fleming T, et al. Intrapartum and neonatal single-dose nevirapine compared with zidovudine for prevention of mother-to-child transmission of HIV-1 in Kampala, Uganda: HIVNET 012 randomised trial. *Lancet*. 1999; 354:795–802. [PubMed: 10485720]
- [7]. Petra Study T. Efficacy of three short-course regimens of zidovudine and lamivudine in preventing early and late transmission of HIV-1 from mother to child in Tanzania, South Africa, and Uganda (Petra study): a randomised, double-blind, placebo-controlled trial. *Lancet*. 2002; 359:1178–86. [PubMed: 11955535]
- [8]. Lallemand M, Jourdain G, Le Coeur S, et al. Single-dose perinatal nevirapine plus standard zidovudine to prevent mother-to-child transmission of HIV-1 in Thailand. *N Engl J Med*. 2004; 351:217–28. [PubMed: 15247338]
- [9]. Kesho Bora Study G, de Vincenzi I. Triple antiretroviral compared with zidovudine and single-dose nevirapine prophylaxis during pregnancy and breastfeeding for prevention of mother-to-child transmission of HIV-1 (Kesho Bora study): a randomised controlled trial. *Lancet Infect Dis*. 2011; 11:171–80. [PubMed: 21237718]
- [10]. The International Perinatal HIV Group. The mode of delivery and the risk of vertical transmission of human immunodeficiency virus type 1--a meta-analysis of 15 prospective cohort studies. *N Engl J Med*. 1999; 340:977–87. [PubMed: 10099139]
- [11]. European Mode of Delivery C. Elective caesarean-section versus vaginal delivery in prevention of vertical HIV-1 transmission: a randomised clinical trial. *Lancet*. 1999; 353:1035–9. [PubMed: 10199349]
- [12]. Kind C, Rudin C, Siegrist CA, et al. Swiss Neonatal HIV Study Group. Prevention of vertical HIV transmission: additive protective effect of elective Cesarean section and zidovudine prophylaxis. *AIDS*. 1998; 12:205–10. [PubMed: 9468370]
- [13]. Mandelbrot L, Le Chenadec J, Berrebi A, et al. Perinatal HIV-1 transmission: interaction between zidovudine prophylaxis and mode of delivery in the French Perinatal Cohort. *JAMA*. 1998; 280:55–60. [PubMed: 9660364]
- [14]. Taha TE, Kumwenda NI, Gibbons A, et al. Short postexposure prophylaxis in newborn babies to reduce mother-to-child transmission of HIV-1: NVAZ randomised clinical trial. *Lancet*. 2003; 362:1171–7. [PubMed: 14568737]
- [15]. Thior I, Lockman S, Smeaton LM, et al. Breastfeeding plus infant zidovudine prophylaxis for 6 months vs formula feeding plus infant zidovudine for 1 month to reduce mother-to-child HIV transmission in Botswana. A randomized trial: the Mashi Study. *JAMA*. 2006; 296:794–805. [PubMed: 16905785]
- [16]. Kumwenda NI, Hoover DR, Mofenson LM, et al. Extended antiretroviral prophylaxis to reduce breast-milk HIV-1 transmission. *N Engl J Med*. 2008; 359:119–29. [PubMed: 18525035]
- [17]. Chasela CS, Hudgens MG, Jamieson DJ, et al. Maternal or infant antiretroviral drugs to reduce HIV-1 transmission. *N Engl J Med*. 2010; 362:2271–81. [PubMed: 20554982]
- [18]. Omer SB. Six Week Extended Dose Nevirapine Study T. Twelve-month follow-up of Six Week Extended Dose Nevirapine randomized controlled trials: differential impact of extended-dose nevirapine on mother-to-child transmission and infant death by maternal CD4 cell count. *AIDS*. 2011; 25:767–76. [PubMed: 21330912]
- [19]. Thiry L, Sprecher-Goldberger S, Jonckheer T, et al. Isolation of AIDS virus from cell-free breast milk of three healthy virus carriers. *Lancet*. 1985; 2:891–2. [PubMed: 2864603]
- [20]. Becquet R, Bland R, Leroy V, et al. Duration, pattern of breastfeeding and postnatal transmission of HIV: pooled analysis of individual data from West and South African cohorts. *PLoS one*. 2009; 4:e7397. [PubMed: 19834601]

- [21]. Guidelines on HIV and infant feeding: principles and recommendations for infant feeding in the context of HIV and a summary of evidence. Joint United Nations Programme on HIV/AIDS; Geneva: 2010.
- [22]. Antiretroviral drugs for treating pregnant women and preventing HIV infection in infants: Recommendations for a public health approach. World Health Organization; Geneva: 2010.
- [23]. [Accessed Nov. 13, 2012] Recommendations for use of antiretroviral drugs in pregnant HIV-1-infected women for maternal health and interventions to reduce perinatal HIV transmission in the United States. Available at <http://aidsinfo.nih.gov/contentfiles/lvguidelines/PerinatalGL.pdf>
- [24]. World AIDS Day Report 2011. UNAIDS; 2011.
- [25]. John GC, Nduati RW, Mbori-Ngacha DA, et al. Correlates of mother-to-child human immunodeficiency virus type 1 (HIV-1) transmission: association with maternal plasma HIV-1 RNA load, genital HIV-1 DNA shedding, and breast infections. *J Infect Dis.* 2001; 183:206–12. [PubMed: 11120927]
- [26]. Lehman DA, Farquhar C. Biological mechanisms of vertical human immunodeficiency virus (HIV-1) transmission. *Rev Med Virol.* 2007; 17:381–403. [PubMed: 17542053]
- [27]. Dickover RE, Garratty EM, Herman SA, et al. Identification of levels of maternal HIV-1 RNA associated with risk of perinatal transmission. Effect of maternal zidovudine treatment on viral load. *JAMA.* 1996; 275:599–605. [PubMed: 8594240]
- [28]. Sperling RS, Shapiro DE, Coombs RW, et al. Pediatric AIDS Clinical Trials Group Protocol 076 Study Group. Maternal viral load, zidovudine treatment, and the risk of transmission of human immunodeficiency virus type 1 from mother to infant. *N Engl J Med.* 1996; 335:1621–9. [PubMed: 8965861]
- [29]. Mofenson LM, Lambert JS, Stiehm ER, et al. Pediatric AIDS Clinical Trials Group Study 185 Team. Risk factors for perinatal transmission of human immunodeficiency virus type 1 in women treated with zidovudine. *N Engl J Med.* 1999; 341:385–93. [PubMed: 10432323]
- [30]. Garcia PM, Kalish LA, Pitt J, et al. Women and Infants Transmission Study Group. Maternal levels of plasma human immunodeficiency virus type 1 RNA and the risk of perinatal transmission. *N Engl J Med.* 1999; 341:394–402. [PubMed: 10432324]
- [31]. Mock PA, Shaffer N, Bhadrakom C, et al. Bangkok Collaborative Perinatal HIV Transmission Study Group. Maternal viral load and timing of mother-to-child HIV transmission, Bangkok, Thailand. *AIDS.* 1999; 13:407–14. [PubMed: 10199232]
- [32]. Fawzi W, Msamanga G, Renjifo B, et al. Predictors of intrauterine and intrapartum transmission of HIV-1 among Tanzanian women. *AIDS.* 2001; 15:1157–65. [PubMed: 11416718]
- [33]. Neveu D, Viljoen J, Bland RM, et al. Cumulative exposure to cell-free HIV in breast milk, rather than feeding pattern per se, identifies postnatally infected infants. *Clin Infect Dis.* 2011; 52:819–25. [PubMed: 21367736]
- [34]. Rousseau CM, Nduati RW, Richardson BA, et al. Longitudinal analysis of human immunodeficiency virus type 1 RNA in breast milk and of its relationship to infant infection and maternal disease. *J Infect Dis.* 2003; 187:741–7. [PubMed: 12599047]
- [35]. International Perinatal HIVG. Duration of ruptured membranes and vertical transmission of HIV-1: a meta-analysis from 15 prospective cohort studies. *AIDS.* 2001; 15:357–68. [PubMed: 11273216]
- [36]. Chen KT, Segu M, Lumey LH, et al. Genital herpes simplex virus infection and perinatal transmission of human immunodeficiency virus. *Obstet Gynecol.* 2005; 106:1341–8. [PubMed: 16319261]
- [37]. Drake AL, John-Stewart GC, Wald A, et al. Herpes simplex virus type 2 and risk of intrapartum human immunodeficiency virus transmission. *Obstet Gynecol.* 2007; 109:403–9. [PubMed: 17267842]
- [38]. Kourtis AP, Lee FK, Abrams EJ, Jamieson DJ, Bulterys M. Mother-to-child transmission of HIV-1: timing and implications for prevention. *The Lancet Infect Dis.* 2006; 6:726–32.
- [39]. Spector SA. Mother-to-infant transmission of HIV-1: the placenta fights back. *J Clin Invest.* 2001; 107(3):267–9. [PubMed: 11160148]

- [40]. Rousseau CM, Nduati RW, Richardson BA, et al. Association of levels of HIV-1-infected breast milk cells and risk of mother-to-child transmission. *J Infect Dis.* 2004; 190:1880–8. [PubMed: 15499546]
- [41]. Lepage P, Van de Perre P. The immune system of breast milk: antimicrobial and anti-inflammatory properties. *Adv Exp Med Bio.* 2012; 743:121–37. [PubMed: 22454346]
- [42]. Kourtis AP, Butera S, Ibegbu C, Belec L, Duerr A. Breast milk and HIV-1: vector of transmission or vehicle of protection? *Lancet Infect Dis.* 2003; 3:786–93. [PubMed: 14652204]
- [43]. Kazmi SH, Naglik JR, Sweet SP, et al. Comparison of human immunodeficiency virus type 1-specific inhibitory activities in saliva and other human mucosal fluids. *Clin Vaccine Immunol.* 2006; 13:1111–8. [PubMed: 16928883]
- [44]. Lyimo MA, Howell AL, Balandya E, Eszterhas SK, Connor RI. Innate factors in human breast milk inhibit cell-free HIV-1 but not cell-associated HIV-1 infection of CD4+ cells. *J Acquir Immune Defic Syndr.* 2009; 51:117–24. [PubMed: 19346967]
- [45]. Henrick BM, Nag K, Yao XD, Drannik AG, Aldrovandi GM, Rosenthal KL. Milk matters: soluble Toll-like receptor 2 (sTLR2) in breast milk significantly inhibits HIV-1 infection and inflammation. *PLoS One.* 2012; 7:e40138. [PubMed: 22792230]
- [46]. Barnabas RV, Celum C. Infectious co-factors in HIV-1 transmission herpes simplex virus type-2 and HIV-1: new insights and interventions. *Curr HIV Res.* 2012; 10:228–37. [PubMed: 22384842]
- [47]. Pawlowski A, Jansson M, Skold M, Rottenberg ME, Kallenius G. Tuberculosis and HIV co-infection. *PLoS Pathogens.* 2012; 8:e1002464. [PubMed: 22363214]
- [48]. Ellington SR, King CC, Kourtis AP. Host factors that influence mother-to-child transmission of HIV-1: genetics, co-infections, behavior and nutrition. *Future Virol.* 2011; 6:1451–69.
- [49]. Zachar V, Zacharova V, Fink T, et al. Genetic analysis reveals ongoing HIV type 1 evolution in infected human placental trophoblast. *AIDS Res Hum Retroviruses.* 1999; 15:1673–83. [PubMed: 10606090]
- [50]. Tkachuk AN, Moormann AM, Poore JA, et al. Malaria enhances expression of CC chemokine receptor 5 on placental macrophages. *J Infect Dis.* 2001; 183:967–72. [PubMed: 11237815]
- [51]. Ayoub A, Badaut C, Kfutwah A, et al. Specific stimulation of HIV-1 replication in human placental trophoblasts by an antigen of *Plasmodium falciparum*. *AIDS.* 2008; 22:785–7. [PubMed: 18356610]
- [52]. Taha TE, Gray RH. Genital tract infections and perinatal transmission of HIV. *Ann N Y Acad Sci.* 2000; 918:84–98. [PubMed: 11131738]
- [53]. Mwanyumba F, Gaillard P, Inion I, et al. Placental inflammation and perinatal transmission of HIV-1. *J Acquir Immune Defic Syndr.* 2002; 29:262–9. [PubMed: 11873075]
- [54]. Van Dyke RB, Korber BT, Popek E, et al. The Ariel Project: A prospective cohort study of maternal-child transmission of human immunodeficiency virus type 1 in the era of maternal antiretroviral therapy. *J Infect Dis.* 1999; 179:319–28. [PubMed: 9878014]
- [55]. St Louis ME, Kamenga M, Brown C, et al. Risk for perinatal HIV-1 transmission according to maternal immunologic, virologic, and placental factors. *JAMA.* 1993; 269:2853–9. [PubMed: 8098783]
- [56]. Wabwire-Mangen F, Gray RH, Mmiro FA, et al. Placental membrane inflammation and risks of maternal-to-child transmission of HIV-1 in Uganda. *J Acquir Immune Defic Syndr.* 1999; 22:379–85. [PubMed: 10634200]
- [57]. Temmerman M, Nyong'o AO, Bwayo J, Fransen K, Coppens M, Piot P. Risk factors for mother-to-child transmission of human immunodeficiency virus-1 infection. *Am J Obstet Gynecol.* 1995; 172:700–5. [PubMed: 7856710]
- [58]. Schwartz DA, Sungkarat S, Shaffer N, et al. Placental abnormalities associated with human immunodeficiency virus type 1 infection and perinatal transmission in Bangkok, Thailand. *J Infect Dis.* 2000; 182:1652–7. [PubMed: 11069236]
- [59]. Romero R, Espinoza J, Goncalves LF, Kusanovic JP, Friel L, Hassan S. The role of inflammation and infection in preterm birth. *Semin Reprod Med.* 2007; 25:21–39. [PubMed: 17205421]

- [60]. Landesman SH, Kalish LA, Burns DN, et al. Obstetrical factors and the transmission of human immunodeficiency virus type 1 from mother to child. The Women and Infants Transmission Study. *N Engl J Med*. 1996; 334:1617–23. [PubMed: 8628356]
- [61]. Pitt J, Schluchter M, Jenson H, et al. Maternal and perinatal factors related to maternal-infant transmission of HIV-1 in the P2C2 HIV study: the role of EBV shedding. Pediatric Pulmonary and Cardiovascular Complications of Vertically Transmitted HIV-1 Infection (P2C2 HIV) Study Group. *J Acquir Immune Defic Syndr Hum Retrovirol*. 1998; 19:462–70. [PubMed: 9859959]
- [62]. Kuhn L, Steketee RW, Weedon J, et al. Distinct risk factors for intrauterine and intrapartum human immunodeficiency virus transmission and consequences for disease progression in infected children. Perinatal AIDS Collaborative Transmission Study. *J Infect Dis*. 1999; 179:52–8. [PubMed: 9841822]
- [63]. Simonds RJ, Steketee R, Nesheim S, et al. Impact of zidovudine use on risk and risk factors for perinatal transmission of HIV. Perinatal AIDS Collaborative Transmission Studies. *AIDS*. 1998; 12(3):301–8. [PubMed: 9517993]
- [64]. Mirpuri J, Jain L. Issues of prematurity and HIV infection. *Clin Perinatol*. 2010; 37:887–905. [PubMed: 21078457]
- [65]. Goldenberg R, Mwatha A, Read J, et al. The HPTN 024 Study: The efficacy of antibiotics to prevent chorioamnionitis and preterm birth. *Am J Obstet Gynecol*. 2006; 194:650–61. [PubMed: 16522393]
- [66]. Taha TE, Brown ER, Hoffman IF, et al. A phase III clinical trial of antibiotics to reduce chorioamnionitis-related perinatal HIV-1 transmission. *AIDS*. 2006; 20:1313–21. [PubMed: 16816561]
- [67]. Ladner J, Leroy V, Hoffman P, et al. Pregnancy and HIV Study Group. Chorioamnionitis and pregnancy outcome in HIV-infected African women. *J Acquir Immune Defic Syndr Hum Retrovirol*. 1998; 18:293–8. [PubMed: 9665509]
- [68]. Spinillo A, Debiaggi M, Zara F, Maserati R, Polatti F, De Santolo A. Factors associated with nucleic acids related to human immunodeficiency virus type 1 in cervico-vaginal secretions. *BJOG*. 2001; 108:634–41. [PubMed: 11426900]
- [69]. Cu-Uvin S, Hogan JW, Caliendo AM, et al. Association between bacterial vaginosis and expression of human immunodeficiency virus type 1 RNA in the female genital tract. *Clinical Infect Dis*. 2001; 33:894–6. [PubMed: 11512096]
- [70]. Sha BE, Zariffard MR, Wang QJ, et al. Female genital-tract HIV load correlates inversely with *Lactobacillus* species but positively with bacterial vaginosis and *Mycoplasma hominis*. *J Infect Dis*. 2005; 191:25–32. [PubMed: 15592999]
- [71]. Al-Harhi L, Roebuck KA, Olinger GG, et al. Bacterial vaginosis-associated microflora isolated from the female genital tract activates HIV-1 expression. *J Acquir Immune Defic Syndr*. 1999; 21:194–202. [PubMed: 10421242]
- [72]. Wang CC, McClelland RS, Reilly M, et al. The effect of treatment of vaginal infections on shedding of human immunodeficiency virus type 1. *J Infect Dis*. 2001; 183:1017–22. [PubMed: 11237825]
- [73]. Corey L, Wald A, Celum CL, Quinn TC. The effects of herpes simplex virus-2 on HIV-1 acquisition and transmission: a review of two overlapping epidemics. *J Acquir Immune Defic Syndr*. 2004; 35:435–45. [PubMed: 15021308]
- [74]. Schacker T, Hu HL, Koelle DM, et al. Famciclovir for the suppression of symptomatic and asymptomatic herpes simplex virus reactivation in HIV-infected persons. A double-blind, placebo-controlled trial. *Ann Intern Med*. 1998; 128:21–8. [PubMed: 9424977]
- [75]. Koelle DM, Abbo H, Peck A, Ziegweid K, Corey L. Direct recovery of herpes simplex virus (HSV)-specific T lymphocyte clones from recurrent genital HSV-2 lesions. *J Infect Dis*. 1994; 169:956–61. [PubMed: 8169426]
- [76]. Bollen LJ, Whitehead SJ, Mock PA, et al. Maternal herpes simplex virus type 2 co-infection increases the risk of perinatal HIV transmission: possibility to further decrease transmission? *AIDS*. 2008; 22:1169–76. [PubMed: 18525263]

- [77]. Mbopi-Keou FX, Gresenguet G, Mayaud P, et al. Interactions between herpes simplex virus type 2 and human immunodeficiency virus type 1 infection in African women: opportunities for intervention. *J Infect Dis.* 2000; 182:1090–6. [PubMed: 10979904]
- [78]. McClelland RS, Wang CC, Overbaugh J, et al. Association between cervical shedding of herpes simplex virus and HIV-1. *AIDS.* 2002; 16:2425–30. [PubMed: 12461416]
- [79]. Cowan FM, Humphrey JH, Ntozini R, Mutasa K, Morrow R, Iliff P. Maternal Herpes simplex virus type 2 infection, syphilis and risk of intra-partum transmission of HIV-1: results of a case control study. *AIDS.* 2008; 22:193–201. [PubMed: 18097221]
- [80]. Van de Perre P, Segondy M, Foulongne V, et al. Herpes simplex virus and HIV-1: deciphering viral synergy. *Lancet Infect Dis.* 2008; 8:490–7. [PubMed: 18652995]
- [81]. Mole L, Ripich S, Margolis D, Holodniy M. The impact of active herpes simplex virus infection on human immunodeficiency virus load. *J Infect Dis.* 1997; 176:766–70. [PubMed: 9291329]
- [82]. Schacker T, Zeh J, Hu H, Shaughnessy M, Corey L. Changes in plasma human immunodeficiency virus type 1 RNA associated with herpes simplex virus reactivation and suppression. *J Infect Dis.* 2002; 186:1718–25. [PubMed: 12447756]
- [83]. Nagot N, Ouedraogo A, Foulongne V, et al. Reduction of HIV-1 RNA levels with therapy to suppress herpes simplex virus. *N Engl J Med.* 2007; 356:790–9. [PubMed: 17314338]
- [84]. Baeten JM, Strick LB, Lucchetti A, et al. Herpes simplex virus (HSV)-suppressive therapy decreases plasma and genital HIV-1 levels in HSV-2/HIV-1 co-infected women: a randomized, placebo-controlled, cross-over trial. *J Infect Dis.* 2008; 198:1804–8. [PubMed: 18928378]
- [85]. Dunne EF, Whitehead S, Sternberg M, et al. Suppressive acyclovir therapy reduces HIV cervicovaginal shedding in HIV- and HSV-2-infected women, Chiang Rai, Thailand. *J Acquir Immune Defic Syndr.* 2008; 49:77–83. [PubMed: 18667923]
- [86]. Celum C, Wald A, Lingappa JR, et al. Acyclovir and transmission of HIV-1 from persons infected with HIV-1 and HSV-2. *N Engl J Med.* 2010; 362:427–39. [PubMed: 20089951]
- [87]. Zuckerman RA, Lucchetti A, Whittington WL, et al. Herpes simplex virus (HSV) suppression with valacyclovir reduces rectal and blood plasma HIV-1 levels in HIV-1/HSV-2-seropositive men: a randomized, double-blind, placebo-controlled crossover trial. *J Infect Dis.* 2007; 196:1500–8. [PubMed: 18008230]
- [88]. Delany S, Mlaba N, Clayton T, et al. Impact of aciclovir on genital and plasma HIV-1 RNA in HSV-2/HIV-1 co-infected women: a randomized placebo-controlled trial in South Africa. *AIDS.* 2009; 23:461–9. [PubMed: 19155993]
- [89]. Celum C, Wald A, Hughes J, et al. Effect of aciclovir on HIV-1 acquisition in herpes simplex virus 2 seropositive women and men who have sex with men: a randomised, double-blind, placebo-controlled trial. *Lancet.* 2008; 371:2109–19. [PubMed: 18572080]
- [90]. Watson-Jones D, Weiss HA, Rusizoka M, et al. Effect of herpes simplex suppression on incidence of HIV among women in Tanzania. *N Engl J Med.* 2008; 358:1560–71. [PubMed: 18337596]
- [91]. Kimberlin DF, Weller S, Whitley RJ, et al. Pharmacokinetics of oral valacyclovir and acyclovir in late pregnancy. *Am J Obstet Gynecol.* 1998; 179:846–51. [PubMed: 9790357]
- [92]. Pasternak B, Hviid A. Use of acyclovir, valacyclovir, and famciclovir in the first trimester of pregnancy and the risk of birth defects. *JAMA.* 2010; 304:859–66. [PubMed: 20736469]
- [93]. Stone KM, Reiff-Eldridge R, White AD, et al. Pregnancy outcomes following systemic prenatal acyclovir exposure: Conclusions from the international acyclovir pregnancy registry, 1984-1999. *Birth Defects Res A Clin Mol Teratol.* 2004; 70:201–7. [PubMed: 15108247]
- [94]. Kimberlin DW, Whitley RJ, Wan W, et al. Oral acyclovir suppression and neurodevelopment after neonatal herpes. *N Engl J Med.* 2011; 365:1284–92. [PubMed: 21991950]
- [95]. Drake AL, Roxby AC, Kiarie J, et al. Infant safety during and after maternal valacyclovir therapy in conjunction with antiretroviral HIV-1 prophylaxis in a randomized clinical trial. *PLoS One.* 2012; 7:e34635. [PubMed: 22509337]
- [96]. Drake AL, Roxby AC, Ongecha-Owuor F, et al. Valacyclovir suppressive therapy reduces plasma and breast milk HIV-1 RNA levels during pregnancy and postpartum: a randomized trial. *J Infect Dis.* 2012; 205:366–75. [PubMed: 22147786]

- [97]. Mwapasa V, Rogerson SJ, Kwiek JJ, et al. Maternal syphilis infection is associated with increased risk of mother-to-child transmission of HIV in Malawi. *AIDS*. 2006; 20:1869–77. [PubMed: 16954728]
- [98]. Thorne C, Malyuta R, Semenenko I, et al. Mother-to-child transmission risk is increased among HIV-infected pregnant women in Ukraine with serological test results positive for syphilis. *Clin Infect Dis*. 2008; 47:1114–5. [PubMed: 18800938]
- [99]. Lee MJ, Hallmark RJ, Frenkel LM, Del Priore G. Maternal syphilis and vertical perinatal transmission of human immunodeficiency virus type-1 infection. *Int J Gynaecol Obstet*. 1998; 63:247–52. [PubMed: 9989893]
- [100]. Schulte JM, Burkham S, Hamaker D, et al. Syphilis among HIV-infected mothers and their infants in Texas from 1988 to 1994. *Sex Transm Dis*. 2001; 28:315–20. [PubMed: 11403187]
- [101]. Gray RH, Wabwire-Mangen F, Kigozi G, et al. Randomized trial of presumptive sexually transmitted disease therapy during pregnancy in Rakai, Uganda. *Am J Obstet Gynecol*. 2001; 185:1209–17. [PubMed: 11717659]
- [102]. Ahdieh L, Klein RS, Burk R, et al. Prevalence, incidence, and type-specific persistence of human papillomavirus in human immunodeficiency virus (HIV)-positive and HIV-negative women. *J Infect Dis*. 2001; 184:682–90. [PubMed: 11517428]
- [103]. Palefsky JM, Minkoff H, Kalish LA, et al. Cervicovaginal human papillomavirus infection in human immunodeficiency virus-1 (HIV)-positive and high-risk HIV-negative women. *J Natl Cancer Inst*. 1999; 91:226–36. [PubMed: 10037100]
- [104]. Mandelbrot, Mayaux, Bongain, et al. Obstetric factors and mother-to-child transmission of human immunodeficiency virus type 1: The French perinatal cohorts. *Am J Obstet Gynecol*. 1996; 175:661–7. [PubMed: 8828431]
- [105]. Atashili J, Poole C, Ndumbe PM, Adimora AA, Smith JS. Bacterial vaginosis and HIV acquisition: a meta-analysis of published studies. *AIDS*. 2008; 22:1493–501. [PubMed: 18614873]
- [106]. Brotman RM, Klebanoff MA, Nansel TR, et al. Bacterial vaginosis assessed by gram stain and diminished colonization resistance to incident gonococcal, chlamydial, and trichomonal genital infection. *J Infect Dis*. 2010; 202:1907–15. [PubMed: 21067371]
- [107]. Chernes TL, Meyn LA, Krohn MA, Lurie JG, Hillier SL. Association between acquisition of herpes simplex virus type 2 in women and bacterial vaginosis. *Clin Infect Dis*. 2003; 37:319–25. [PubMed: 12884154]
- [108]. Kaul R, Nagelkerke NJ, Kimani J, et al. Prevalent herpes simplex virus type 2 infection is associated with altered vaginal flora and an increased susceptibility to multiple sexually transmitted infections. *J Infect Dis*. 2007; 196:1692–7. [PubMed: 18008255]
- [109]. Wiesenfeld HC, Hillier SL, Krohn MA, Landers DV, Sweet RL. Bacterial vaginosis is a strong predictor of *Neisseria gonorrhoeae* and *Chlamydia trachomatis* infection. *Clin Infect Dis*. 2003; 36:663–8. [PubMed: 12594649]
- [110]. Farquhar C, Mbori-Ngacha D, Overbaugh J, et al. Illness during pregnancy and bacterial vaginosis are associated with in-utero HIV-1 transmission. *AIDS*. 2010; 24:153–5. [PubMed: 19952542]
- [111]. Gaillard P, Mwanjumba F, Verhofstede C, et al. Vaginal lavage with chlorhexidine during labour to reduce mother-to-child HIV transmission: clinical trial in Mombasa, Kenya. *AIDS*. 2001; 15:389–96. [PubMed: 11273219]
- [112]. Biggar RJ, Miotti PG, Taha TE, et al. Perinatal intervention trial in Africa: effect of a birth canal cleansing intervention to prevent HIV transmission. *Lancet*. 1996; 347:1647–50. [PubMed: 8642957]
- [113]. Mandelbrot L, Msellati P, Meda N, et al. 15 Month follow up of African children following vaginal cleansing with benzalkonium chloride of their HIV infected mothers during late pregnancy and delivery. *Sex Transm Infect*. 2002; 78:267–70. [PubMed: 12181464]
- [114]. Msellati P, Meda N, Leroy V, et al. DITRAME Study Group. Safety and acceptability of vaginal disinfection with benzalkonium chloride in HIV infected pregnant women in west Africa: ANRS 049b phase II randomized, double blinded placebo controlled trial. *Sex Transm Infect*. 1999; 75:420–5. [PubMed: 10754950]

- [115]. Tess BH, Rodrigues LC, Newell ML, Dunn DT, Lago TD. Infant feeding and risk of mother-to-child transmission of HIV-1 in Sao Paulo State, Brazil. Sao Paulo Collaborative Study for Vertical Transmission of HIV-1. *J Acquir Immune Defic Syndr Hum Retrovirol.* 1998; 19:189–94. [PubMed: 9768630]
- [116]. Embree JE, Njenga S, Datta P, et al. Risk factors for postnatal mother-child transmission of HIV-1. *AIDS.* 2000; 14:2535–41. [PubMed: 11101065]
- [117]. Semba RD, Kumwenda N, Hoover DR, et al. Human immunodeficiency virus load in breast milk, mastitis, and mother-to-child transmission of human immunodeficiency virus type 1. *J Infect Dis.* 1999; 180(1):93–8. [PubMed: 10353866]
- [118]. Willumsen JF, Filteau SM, Coutoudis A, Uebel KE, Newell ML, Tomkins AM. Subclinical mastitis as a risk factor for mother-infant HIV transmission. *Adv Exp Med Biol.* 2000; 478:211–23. [PubMed: 11065074]
- [119]. Willumsen JF, Filteau SM, Coutoudis A, et al. Breastmilk RNA viral load in HIV-infected South African women: effects of subclinical mastitis and infant feeding. *AIDS.* 2003; 17:407–14. [PubMed: 12556695]
- [120]. Phiri W, Kasonka L, Collin S, et al. Factors influencing breast milk HIV RNA viral load among Zambian women. *AIDS Res Hum Retroviruses.* 2006; 22:607–14. [PubMed: 16831084]
- [121]. Lunney Kevin M, Iliff P, Mutasa K, et al. Associations between breast milk viral load, mastitis, exclusive breast-feeding, and postnatal transmission of HIV. *Clin Infect Dis.* 2010; 50:762–9. [PubMed: 20121424]
- [122]. Gantt S, Shetty AK, Seidel KD, et al. Laboratory indicators of mastitis are not associated with elevated HIV-1 DNA loads or predictive of HIV-1 RNA loads in breast milk. *J Infect Dis.* 2007; 196:570–6. [PubMed: 17624843]
- [123]. Piwoz EG, Ross J, Humphrey J. Human immunodeficiency virus transmission during breastfeeding: knowledge, gaps, and challenges for the future. *Adv Exp Med Biol.* 2004; 554:195–210. [PubMed: 15384577]
- [124]. Nussenblatt V, Lema V, Kumwenda N, et al. Epidemiology and microbiology of subclinical mastitis among HIV-infected women in Malawi. *Int J STD AIDS.* 2005; 16:227–32. [PubMed: 15829023]
- [125]. Matheson I, Aursnes I, Horgen M, Aabo O, Melby K. Bacteriological findings and clinical symptoms in relation to clinical outcome in puerperal mastitis. *Acta Obstet Gynecol Scand.* 1988; 67:723–6. [PubMed: 3250184]
- [126]. Nussenblatt V, Kumwenda N, Lema V, et al. Effect of antibiotic treatment of subclinical mastitis on human immunodeficiency virus type 1 RNA in human milk. *J Trop Pediatr.* 2006; 52:311–5. [PubMed: 16595526]
- [127]. Bosire R, John-Stewart GC, Mabuka JM, et al. Breast milk alpha-defensins are associated with HIV type 1 RNA and CC chemokines in breast milk but not vertical HIV type 1 transmission. *AIDS Res Hum Retroviruses.* 2007; 23:198–203. [PubMed: 17331027]
- [128]. Pelto GH, Zhang Y, Habicht JP. Premastication: the second arm of infant and young child feeding for health and survival? *Matern Child Nutr.* 2010; 6:4–18. [PubMed: 20073131]
- [129]. Gaur AH, Dominguez KL, Kalish ML, et al. Practice of feeding pre-masticated food to infants: a potential risk factor for HIV transmission. *Pediatrics.* 2009; 124:658–66. [PubMed: 19620190]
- [130]. Ivy W 3rd, Dominguez KL, Rakhmanina NY, et al. Premastication as a route of pediatric HIV transmission: case-control and cross-sectional investigations. *J Acquir Immune Defic Syndr.* 2012; 59:207–12. [PubMed: 22027873]
- [131]. Premastication of food by caregivers of HIV-exposed children—nine U.S. sites, 2009–2010. *MMWR Morb Mortal Wkly Rep.* 2011; 60:273–5.
- [132]. Shaffer N, Roongpisuthipong A, Siriwasin W, et al. Bangkok Collaborative Perinatal HIV Transmission Study Group. Maternal virus load and perinatal human immunodeficiency virus type 1 subtype E transmission, Thailand. *J Infect Dis.* 1999; 179:590–9. [PubMed: 9952365]
- [133]. Chuachoowong R, Shaffer N, Siriwasin W, et al. Bangkok Collaborative Perinatal HIV Transmission Study Group. Short-course antenatal zidovudine reduces both cervicovaginal human immunodeficiency virus type 1 RNA levels and risk of perinatal transmission. *J Infect Dis.* 2000; 181:99–106. [PubMed: 10608756]

- [134]. Mayaux MJ, Dussaix E, Isopet J, et al. SEROGEST Cohort Group. Maternal virus load during pregnancy and mother-to-child transmission of human immunodeficiency virus type 1: the French perinatal cohort studies. *J Infect Dis.* 1997; 175:172–5. [PubMed: 8985214]
- [135]. Alter MJ. Epidemiology of viral hepatitis and HIV co-infection. *J Hepatol.* 2006; 44(1 Suppl):S6–9. [PubMed: 16352363]
- [136]. Thio CL, Locarnini S. Treatment of HIV/HBV co-infection: clinical and virologic issues. *AIDS Rev.* 2007; 9:40–53. [PubMed: 17474312]
- [137]. Piroth L, Sene D, Pol S, et al. Epidemiology, diagnosis and treatment of chronic hepatitis B in HIV-infected patients (EPIB 2005 STUDY). *AIDS.* 2007; 21(10):1323–31. [PubMed: 17545709]
- [138]. Soriano V, Puoti M, Bonacini M, et al. Care of patients with chronic hepatitis B and HIV co-infection: recommendations from an HIV-HBV International Panel. *AIDS.* 2005; 19:221–40. [PubMed: 15718833]
- [139]. Lee C, Gong Y, Brok J, Boxall EH, Gluud C. Effect of hepatitis B immunisation in newborn infants of mothers positive for hepatitis B surface antigen: systematic review and meta-analysis. *BMJ.* 2006; 332:328–36. [PubMed: 16443611]
- [140]. Hoffmann CJ, Thio CL. Clinical implications of HIV and hepatitis B co-infection in Asia and Africa. *Lancet Infect Dis.* 2007; 7:402–9. [PubMed: 17521593]
- [141]. Kourtis AP, Bulters M, Hu DJ, Jamieson DJ. HIV-HBV co-infection--a global challenge. *N Engl J Med.* 2012; 366:1749–52. [PubMed: 22571198]
- [142]. Xu WM, Cui YT, Wang L, et al. Lamivudine in late pregnancy to prevent perinatal transmission of hepatitis B virus infection: a multicentre, randomized, double-blind, placebo-controlled study. *J Viral Hepat.* 2009; 16:94–103. [PubMed: 19175878]
- [143]. Menendez C, Sanchez-Tapias JM, Kahigwa E, et al. Prevalence and mother-to-infant transmission of hepatitis viruses B, C, and E in Southern Tanzania. *J Med Virol.* 1999; 58:215–20. [PubMed: 10447415]
- [144]. Tess BH, Rodrigues LC, Newell ML, Dunn DT, Lago TD, Sao Paulo Collaborative Study for Vertical Transmission of HIV-1. Breastfeeding, genetic, obstetric and other risk factors associated with mother-to-child transmission of HIV-1 in Sao Paulo State, Brazil. *AIDS.* 1998; 12:513–20. [PubMed: 9543450]
- [145]. Landes M, Newell ML, Barlow P, et al. Hepatitis B or hepatitis C co-infection in HIV-infected pregnant women in Europe. *HIV Med.* 2008; 9:526–34. [PubMed: 18554310]
- [146]. Sherman KE, Rouster SD, Chung RT, Rajcic N. Hepatitis C Virus prevalence among patients infected with Human Immunodeficiency Virus: a cross-sectional analysis of the US adult AIDS Clinical Trials Group. *Clin Infect Dis.* 2002; 34:831–7. [PubMed: 11833007]
- [147]. Benhamou Y, Bochet M, Di Martino V, et al. The Multivirc Group. Liver fibrosis progression in human immunodeficiency virus and hepatitis C virus co-infected patients. *Hepatology.* 1999; 30:1054–8. [PubMed: 10498659]
- [148]. Puoti M, Bonacini M, Spinetti A, et al. Liver fibrosis progression is related to CD4 cell depletion in patients co-infected with hepatitis C virus and human immunodeficiency virus. *J Infect Dis.* 2001; 183:134–7. [PubMed: 11087200]
- [149]. Soriano V, Barreiro P, Nunez M. Management of chronic hepatitis B and C in HIV-co-infected patients. *J Antimicrob Chemother.* 2006; 57:815–8. [PubMed: 16556638]
- [150]. Thomas DL, Villano SA, Riester KA, et al. Women and Infants Transmission Study. Perinatal transmission of hepatitis C virus from human immunodeficiency virus type 1-infected mothers. *J Infect Dis.* 1998; 177:1480–8. [PubMed: 9607823]
- [151]. Zanetti AR, Tanzi E, Romano L, et al. A prospective study on mother-to-infant transmission of hepatitis C virus. *Intervirology.* 1998; 41:208–12. [PubMed: 10213898]
- [152]. Okamoto M, Nagata I, Murakami J, et al. Prospective reevaluation of risk factors in mother-to-child transmission of hepatitis C virus: high virus load, vaginal delivery, and negative anti-NS4 antibody. *J Infect Dis.* 2000; 182:1511–4. [PubMed: 11023474]
- [153]. Dal Molin G, D'Agaro P, Ansaldi F, et al. Mother-to-infant transmission of hepatitis C virus: rate of infection and assessment of viral load and IgM anti-HCV as risk factors. *J Med Virol.* 2002; 67:137–42. [PubMed: 11992574]

- [154]. Dowd Kimberly A, Hershov Ronald C, Yawetz S, et al. Maternal Neutralizing Antibody and Transmission of Hepatitis C Virus to Infants. *J Infect Dis.* 2008; 198:1651–5. [PubMed: 18928374]
- [155]. Tovo PA, Palomba E, Ferraris G, et al. Italian Study Group for HCV Infection in Children. Increased risk of maternal-infant hepatitis C virus transmission for women co-infected with human immunodeficiency virus type 1. *Clin Infect Dis.* 1997; 25:1121–4. [PubMed: 9402369]
- [156]. Papaevangelou V, Pollack H, Rochford G, et al. Increased transmission of vertical hepatitis C virus (HCV) infection to human immunodeficiency virus (HIV)-infected infants of HIV- and HCV-co-infected women. *J Infect Dis.* 1998; 178:1047–52. [PubMed: 9806033]
- [157]. Giovannini M, Tagger A, Ribero ML, et al. Maternal-infant transmission of hepatitis C virus and HIV infections: a possible interaction. *Lancet.* 1990; 335:1166. [PubMed: 1971901]
- [158]. Hershov RC, Riestler KA, Lew J, et al. Women and Infants Transmission Study. Increased vertical transmission of human immunodeficiency virus from hepatitis C virus-co-infected mothers. *J Infect Dis.* 1997; 176:414–20. [PubMed: 9237706]
- [159]. Paccagnini S, Principi N, Massironi E, et al. Perinatal transmission and manifestation of hepatitis C virus infection in a high risk population. *Pediatr Infect Dis J.* 1995; 14:195–9. [PubMed: 7761184]
- [160]. Karayiannis P, McGarvey MJ. The GB hepatitis viruses. *J Viral Hepat.* 1995; 2:221–6. [PubMed: 8745313]
- [161]. Bhanich Supapol W, Remis RS, Raboud J, et al. Mother-to-Child Transmission of GB Virus C in a Cohort of Women Co-infected with GB Virus C and HIV in Bangkok, Thailand. *J Infect Dis.* 2009; 200:227–35. [PubMed: 19508162]
- [162]. Zanetti AR, Tanzi E, Romano L, et al. The Lombardy Study Group on Vertical/Perinatal Hepatitis Viruses Transmission. Multicenter trial on mother-to-infant transmission of GBV-C virus. *J Med Virol.* 1998; 54:107–12. [PubMed: 9496368]
- [163]. Chandwani S, Kaul A, Bebenroth D, et al. Cytomegalovirus infection in human immunodeficiency virus type 1-infected children. *Pediatr Infect Dis J.* 1996; 15:310–4. [PubMed: 8866799]
- [164]. Doyle M, Atkins JT, Rivera-Matos IR. Congenital cytomegalovirus infection in infants infected with human immunodeficiency virus type 1. *Pediatr Infect Dis J.* 1996; 15:1102–6. [PubMed: 8970220]
- [165]. Duryea EL, Sanchez PJ, Sheffield JS, et al. Maternal human immunodeficiency virus infection and congenital transmission of cytomegalovirus. *Pediatr Infect Dis J.* 2010; 29:915–8. [PubMed: 20431424]
- [166]. Guibert G, Warszawski J, Le Chenadec J, et al. Decreased risk of congenital cytomegalovirus infection in children born to HIV-1-infected mothers in the era of highly active antiretroviral therapy. *Clin Infect Dis.* 2009; 48:1516–25. [PubMed: 19388872]
- [167]. Kovacs A, Schluchter M, Easley K, et al. Pediatric Pulmonary and Cardiovascular Complications of Vertically Transmitted HIV Infection Study Group. Cytomegalovirus infection and HIV-1 disease progression in infants born to HIV-1-infected women. *N Engl J Med.* 1999; 341:77–84. [PubMed: 10395631]
- [168]. Mussi-Pinhata MM, Yamamoto AY, Figueiredo LT, Cervi MC, Duarte G. Congenital and perinatal cytomegalovirus infection in infants born to mothers infected with human immunodeficiency virus. *J Pediatr.* 1998; 132:285–90. [PubMed: 9506642]
- [169]. Kenneson A, Cannon MJ. Review and meta-analysis of the epidemiology of congenital cytomegalovirus (CMV) infection. *Rev Med Virol.* 2007; 17:253–76. [PubMed: 17579921]
- [170]. Khamduang W, Jourdain G, Sirirungsi W, et al. The interrelated transmission of HIV-1 and cytomegalovirus during gestation and delivery in the offspring of HIV-infected mothers. *J Acquir Immune Defic Syndr.* 2011; 58:188–92. [PubMed: 21792064]
- [171]. Skolnik PR, Kosloff BR, Hirsch MS. Bidirectional interactions between human immunodeficiency virus type 1 and cytomegalovirus. *J Infect Dis.* 1988; 157:508–14. [PubMed: 2830343]

- [172]. Ho WZ, Harouse JM, Rando RF, Gonczol E, Srinivasan A, Plotkin SA. Reciprocal enhancement of gene expression and viral replication between human cytomegalovirus and human immunodeficiency virus type 1. *J Gen Virol.* 1990; 71:97–103. [PubMed: 2154540]
- [173]. McKeating JA, Griffiths PD, Weiss RA. HIV susceptibility conferred to human fibroblasts by cytomegalovirus-induced Fc receptor. *Nature.* 1990; 343:659–61. [PubMed: 2154697]
- [174]. D'Agaro P, Burgnich P, Comar M, et al. HHV-6 is frequently detected in dried cord blood spots from babies born to HIV-positive mothers. *Curr HIV Res.* 2008; 6(5):441–6. [PubMed: 18855654]
- [175]. Kositanont U, Wasi C, Wanprapar N, et al. Primary infection of human herpesvirus 6 in children with vertical infection of human immunodeficiency virus type 1. *J Infect Dis.* 1999; 180:50–5. [PubMed: 10353860]
- [176]. Klaskala W, Brayfield BP, Kankasa C, et al. Epidemiological characteristics of human herpesvirus-8 infection in a large population of antenatal women in Zambia. *J Med Virol.* 2005; 75:93–100. [PubMed: 15543582]
- [177]. Brayfield BP, Phiri S, Kankasa C, et al. Postnatal human herpesvirus 8 and human immunodeficiency virus type 1 infection in mothers and infants from Zambia. *J Infect Dis.* 2003; 187:559–68. [PubMed: 12599072]
- [178]. Collenberg E, Ouedraogo T, Ganame J, et al. Seroprevalence of six different viruses among pregnant women and blood donors in rural and urban Burkina Faso: A comparative analysis. *J Med Virol.* 2006; 78:683–92. [PubMed: 16555290]
- [179]. Hladik W, Dollard SC, Downing RG, et al. Kaposi's sarcoma in Uganda: risk factors for human herpesvirus 8 infection among blood donors. *J Acquir Immune Defic Syndr.* 2003; 33:206–10. [PubMed: 12794556]
- [180]. Calabro ML, Gasperini P, Fiore JR, Barbierato M, Angarano G, Chieco-Bianchi L. Intrafamilial transmission of human herpesvirus 8. *J Natl Cancer Inst.* 2001; 93:154–6. [PubMed: 11208891]
- [181]. Plancoulaine S, Abel L, van Beveren M, et al. Human herpesvirus 8 transmission from mother to child and between siblings in an endemic population. *Lancet.* 2000; 356:1062–5. [PubMed: 11009141]
- [182]. Lyall EG, Patton GS, Sheldon J, et al. Evidence for horizontal and not vertical transmission of human herpesvirus 8 in children born to human immunodeficiency virus-infected mothers. *Pediatr Infect Dis J.* 1999; 18:795–9. [PubMed: 10493340]
- [183]. Mayama S, Cuevas LE, Sheldon J, et al. Prevalence and transmission of Kaposi's sarcoma-associated herpesvirus (human herpesvirus 8) in Ugandan children and adolescents. *Int J Cancer.* 1998; 77:817–20. [PubMed: 9714046]
- [184]. Minhas V, Crabtree KL, Chao A, et al. Early childhood infection by human herpesvirus 8 in Zambia and the role of human immunodeficiency virus type 1 co-infection in a highly endemic area. *Am J Epidemiol.* 2008; 168:311–20. [PubMed: 18515794]
- [185]. Mantina H, Kankasa C, Klaskala W, et al. Vertical transmission of Kaposi's sarcoma-associated herpesvirus. *Int J Cancer.* 2001; 94:749–52. [PubMed: 11745472]
- [186]. Brayfield BP, Kankasa C, West JT, et al. Distribution of Kaposi sarcoma-associated herpesvirus/human herpesvirus 8 in maternal saliva and breast milk in Zambia: implications for transmission. *J Infect Dis.* 2004; 189:2260–70. [PubMed: 15181574]
- [187]. Malope BI, Pfeiffer RM, Mbisa G, et al. Transmission of Kaposi sarcoma-associated herpesvirus between mothers and children in a South African population. *J Acquir Immune Defic Syndr.* 2007; 44:351–5. [PubMed: 17195763]
- [188]. Lisco A, Barbierato M, Fiore JR, et al. Pregnancy and human herpesvirus 8 reactivation in human immunodeficiency virus type 1-infected women. *J Clin Microbiol.* 2006; 44:3863–71. [PubMed: 16943357]
- [189]. Global tuberculosis control: key findings from the December 2009 WHO report. *Wkly Epidemiol Rec.* 2010; 85:69–80. [PubMed: 20210259]
- [190]. WHO global tuberculosis control report 2010. Summary. *Cent Eur J Public Health.* 2010; 18:237. [PubMed: 21361110]
- [191]. Day JH, Grant AD, Fielding KL, et al. Does tuberculosis increase HIV load? *J Infect Dis.* 2004; 190:1677–84. [PubMed: 15478075]

- [192]. Goletti D, Weissman D, Jackson RW, et al. Effect of Mycobacterium tuberculosis on HIV replication. Role of immune activation. *J Immunol.* 1996; 157:1271–8. [PubMed: 8757635]
- [193]. Zhang Y, Nakata K, Weiden M, Rom WN. Mycobacterium tuberculosis enhances human immunodeficiency virus-1 replication by transcriptional activation at the long terminal repeat. *J Clin Invest.* 1995; 95:2324–31. [PubMed: 7738195]
- [194]. Toossi Z, Mayanja-Kizza H, Hirsch CS, et al. Impact of tuberculosis (TB) on HIV-1 activity in dually infected patients. *Clin Exp Immunol.* 2001; 123:233–8. [PubMed: 11207653]
- [195]. Pillay T, Adhikari M, Coovadia HM, Moodley J, Khan M, Sullivan JL. *In utero* HIV infection in pregnancies complicated by tuberculosis in Durban, South Africa. *Arch Dis Child Fetal Neonatal Ed.* 2004; 89:F468–9. [PubMed: 15321976]
- [196]. Gupta A, Bhosale R, Kinikar A, et al. Maternal Tuberculosis: A Risk Factor for Mother-to-Child Transmission of Human Immunodeficiency Virus. *J Infect Dis.* 2011; 203:358–62. [PubMed: 21208928]
- [197]. Kalilani L, Mofolo I, Chaponda M, Rogerson SJ, Meshnick SR. The effect of timing and frequency of Plasmodium falciparum infection during pregnancy on the risk of low birth weight and maternal anemia. *Trans R Soc Trop Med Hyg.* 2010; 104:416–22. [PubMed: 20207387]
- [198]. van Geertruyden JP, Thomas F, Erhart A, D'Alessandro U. The contribution of malaria in pregnancy to perinatal mortality. *Am J Trop Med Hyg.* 2004; 71:35–40. [PubMed: 15331817]
- [199]. Verhoeff FH, Brabin BJ, Hart CA, Chimsuku L, Kazembe P, Broadhead RL. Increased prevalence of malaria in HIV-infected pregnant women and its implications for malaria control. *Trop Med Int Health.* 1999; 4:5–12. [PubMed: 10203167]
- [200]. van Eijk AM, Ayisi JG, ter Kuile FO, et al. HIV increases the risk of malaria in women of all gravidities in Kisumu, Kenya. *AIDS.* 2003; 17:595–603. [PubMed: 12598780]
- [201]. Steketee RW, Wirima JJ, Bloland PB, et al. Impairment of a pregnant woman's acquired ability to limit Plasmodium falciparum by infection with human immunodeficiency virus type-1. *Am J Trop Med Hyg.* 1996; 55:42–9. [PubMed: 8702036]
- [202]. Ayisi JG, van Eijk AM, ter Kuile FO, et al. The effect of dual infection with HIV and malaria on pregnancy outcome in western Kenya. *AIDS.* 2003; 17:585–94. [PubMed: 12598779]
- [203]. Menendez C, Serra-Casas E, Scahill MD, et al. HIV and placental infection modulate the appearance of drug-resistant Plasmodium falciparum in pregnant women who receive intermittent preventive treatment. *Clin Infect Dis.* 2011; 52:41–8. [PubMed: 21148518]
- [204]. Hoffman IF, Jere CS, Taylor TE, et al. The effect of Plasmodium falciparum malaria on HIV-1 RNA blood plasma concentration. *AIDS.* 1999; 13:487–94. [PubMed: 10197377]
- [205]. Kublin JG, Patnaik P, Jere CS, et al. Effect of Plasmodium falciparum malaria on concentration of HIV-1-RNA in the blood of adults in rural Malawi: a prospective cohort study. *Lancet.* 2005; 365:233–40. [PubMed: 15652606]
- [206]. Mwapasa V, Rogerson SJ, Molyneux ME, et al. The effect of Plasmodium falciparum malaria on peripheral and placental HIV-1 RNA concentrations in pregnant Malawian women. *AIDS.* 2004; 18:1051–9. [PubMed: 15096809]
- [207]. Brahmabhatt H, Sullivan D, Kigozi G, et al. Association of HIV and malaria with mother-to-child transmission, birth outcomes, and child mortality. *J Acquir Immune Defic Syndr.* 2008; 47:472–6. [PubMed: 18332766]
- [208]. Inion I, Mwanyumba F, Gaillard P, et al. Placental malaria and perinatal transmission of human immunodeficiency virus type 1. *J Infect Dis.* 2003; 188:1675–8. [PubMed: 14639538]
- [209]. Bulterys PL, Chao A, Dalai SC, et al. Placental malaria and mother-to-child transmission of human immunodeficiency virus-1 in rural Rwanda. *Am J Trop Med Hyg.* 2011; 85:202–6. [PubMed: 21813835]
- [210]. Msamanga GI, Taha TE, Young AM, et al. Placental malaria and mother-to-child transmission of human immunodeficiency virus-1. *Am J Trop Med Hyg.* 2009; 80:508–15. [PubMed: 19346367]
- [211]. Ayisi JG, van Eijk AM, Newman RD, et al. Maternal malaria and perinatal HIV transmission, western Kenya. *Emerg Infect Dis.* 2004; 10:643–52. [PubMed: 15200854]
- [212]. Brahmabhatt H, Kigozi G, Wabwire-Mangen F, et al. The effects of placental malaria on mother-to-child HIV transmission in Rakai, Uganda. *AIDS.* 2003; 17:2539–41. [PubMed: 14600529]

- [213]. Moormann AM, Sullivan AD, Rochford RA, et al. Malaria and pregnancy: placental cytokine expression and its relationship to intrauterine growth retardation. *J Infect Dis.* 1999; 180:1987–93. [PubMed: 10558956]
- [214]. Abrams ET, Brown H, Chensue SW, et al. Host response to malaria during pregnancy: placental monocyte recruitment is associated with elevated beta chemokine expression. *J Immunol.* 2003; 170:2759–64. [PubMed: 12594307]
- [215]. Chaisavaneeyakorn S, Moore JM, Mirel L, et al. Levels of macrophage inflammatory protein 1 alpha (MIP-1 alpha) and MIP-1 beta in intervillous blood plasma samples from women with placental malaria and human immunodeficiency virus infection. *Clin Diagn Lab Immunol.* 2003; 10:631–6. [PubMed: 12853396]
- [216]. Fried M, Muga RO, Misore AO, Duffy PE. Malaria elicits type 1 cytokines in the human placenta: IFN-gamma and TNF-alpha associated with pregnancy outcomes. *J Immunol.* 1998; 160:2523–30. [PubMed: 9498798]
- [217]. Robert-Gangneux F, Darde ML. Epidemiology of and diagnostic strategies for toxoplasmosis. *Clin Microbiol Rev.* 2012; 25:264–96. [PubMed: 22491772]
- [218]. Dunn D, Wallon M, Peyron F, Petersen E, Peckham C, Gilbert R. Mother-to-child transmission of toxoplasmosis: risk estimates for clinical counselling. *Lancet.* 1999; 353(9167):1829–33. [PubMed: 10359407]
- [219]. Desmonts G, Couvreur J. A prospective study of 378 pregnancies. Congenital toxoplasmosis. *N Engl J Med.* 1974; 290:1110–6. [PubMed: 4821174]
- [220]. Mitchell CD, Erlich SS, Mastrucci MT, Hutto SC, Parks WP, Scott GB. Congenital toxoplasmosis occurring in infants perinatally infected with human immunodeficiency virus 1. *Pediatr Infect Dis J.* 1990; 9:512–8. [PubMed: 2371084]
- [221]. *Pediatric AIDS: The Challenge of HIV Infection in Infants, Children, and Adolescents.* 2nd ed. Williams and Wilkins; 1994.
- [222]. Delicio AM, Milanez H, Amaral E, et al. Mother-to-child transmission of human immunodeficiency virus in ten years period. *Reprod Health.* 2011; 8:35. [PubMed: 22129112]
- [223]. Teixeira MM, Gazzinelli RT, Silva JS. Chemokines, inflammation and *Trypanosoma cruzi* infection. *Trends Parasitol.* 2002; 18:262–5. [PubMed: 12036740]
- [224]. Dolcini GL, Solana ME, Andreani G, et al. *Trypanosoma cruzi* (Chagas' disease agent) reduces HIV-1 replication in human placenta. *Retrovirology.* 2008; 5:53. [PubMed: 18593480]
- [225]. Freilij H, Altchek J, Muchnik G. Perinatal human immunodeficiency virus infection and congenital Chagas' disease. *Pediatr Infect Dis J.* 1995; 14:161–2. [PubMed: 7746707]
- [226]. Belyhun Y, Medhin G, Amberbir A, et al. Prevalence and risk factors for soil-transmitted helminth infection in mothers and their infants in Butajira, Ethiopia: a population based study. *BMC Public Health.* 2010; 10:21. [PubMed: 20085635]
- [227]. Naish S, McCarthy J, Williams GM. Prevalence, intensity and risk factors for soil-transmitted helminth infection in a South Indian fishing village. *Acta Tropica.* 2004; 91:177–87. [PubMed: 15234667]
- [228]. Nguyen PH, Nguyen KC, Nguyen TD, et al. Intestinal helminth infections among reproductive age women in Vietnam: prevalence, co-infection and risk factors. *Southeast Asian J Trop Med Public Health.* 2006; 37:865–74. [PubMed: 17333727]
- [229]. Yatch NJ, Yi J, Agbenyega T, et al. Malaria and intestinal helminth co-infection among pregnant women in Ghana: prevalence and risk factors. *Am J Trop Med Hyg.* 2009; 80:896–901. [PubMed: 19478245]
- [230]. van Riet E, Hartgers FC, Yazdanbakhsh M. Chronic helminth infections induce immunomodulation: consequences and mechanisms. *Immunobiology.* 2007; 212:475–90. [PubMed: 17544832]
- [231]. Gallagher M, Malhotra I, Mungai PL, et al. The effects of maternal helminth and malaria infections on mother-to-child HIV transmission. *AIDS.* 2005; 19:1849–55. [PubMed: 16227793]
- [232]. Walson JL, Otieno PA, Mbuchi M, et al. Albendazole treatment of HIV-1 and helminth co-infection: a randomized, double-blind, placebo-controlled trial. *AIDS.* 2008; 22:1601–9. [PubMed: 18670219]

- [233]. Kallestrup P, Zinyama R, Gomo E, et al. Schistosomiasis and HIV-1 infection in rural Zimbabwe: effect of treatment of schistosomiasis on CD4 cell count and plasma HIV-1 RNA load. *J Infect Dis.* 2005; 192:1956–61. [PubMed: 16267767]
- [234]. Winter H. Gastrointestinal tract function and malnutrition in HIV-infected children. *J Nutr.* 1996; 126:2620S–2S. [PubMed: 8861924]
- [235]. Goto K, Chew F, Torun B, Peerson JM, Brown KH. Epidemiology of altered intestinal permeability to lactulose and mannitol in Guatemalan infants. *J Pediatr Gastroenterol Nutr.* 1999; 28:282–90. [PubMed: 10067729]
- [236]. Iliff PJ, Piwoz EG, Tavengwa NV, et al. Early exclusive breastfeeding reduces the risk of postnatal HIV-1 transmission and increases HIV-free survival. *AIDS.* 2005; 19:699–708. [PubMed: 15821396]
- [237]. Coutoudis A, Pillay K, Spooner E, Kuhn L, Coovadia HM, South African Vitamin A Study Group. Influence of infant-feeding patterns on early mother-to-child transmission of HIV-1 in Durban, South Africa: a prospective cohort study. *Lancet.* 1999; 354:471–6. [PubMed: 10465172]
- [238]. Kuhn L, Sinkala M, Kankasa C, et al. High uptake of exclusive breastfeeding and reduced early post-natal HIV transmission. *PLoS One.* 2007; 2:e1363. [PubMed: 18159246]
- [239]. The European Collaborative Study. Natural history of vertically acquired human immunodeficiency virus-1 infection. *Pediatrics.* 1994; 94:815–9. [PubMed: 7970995]
- [240]. Abrams EJ, Weedon J, Steketee RW, et al. New York City Perinatal HIV Transmission Collaborative Study Group. Association of human immunodeficiency virus (HIV) load early in life with disease progression among HIV-infected infants. *J Infect Dis.* 1998; 178:101–8. [PubMed: 9652428]
- [241]. Biggar RJ, Janes M, Pilon R, et al. Virus levels in untreated African infants infected with human immunodeficiency virus type 1. *J Infect Dis.* 1999; 180:1838–43. [PubMed: 10558939]
- [242]. Obimbo EM, Mbori-Ngacha DA, Ochieng JO, et al. Predictors of early mortality in a cohort of human immunodeficiency virus type 1-infected african children. *Pediatr Infect Dis J.* 2004; 23:536–43. [PubMed: 15194835]
- [243]. Richardson BA, Mbori-Ngacha D, Lavreys L, et al. Comparison of human immunodeficiency virus type 1 viral loads in Kenyan women, men, and infants during primary and early infection. *J Virol.* 2003; 77:7120–3. [PubMed: 12768032]
- [244]. Prendergast A, Tudor-Williams G, Jeena P, Burchett S, Goulder P. International perspectives, progress, and future challenges of paediatric HIV infection. *Lancet.* 2007; 370:68–80. [PubMed: 17617274]
- [245]. Newell ML, Coovadia H, Cortina-Borja M, et al. Mortality of infected and uninfected infants born to HIV-infected mothers in Africa: a pooled analysis. *Lancet.* 2004; 364:1236–43. [PubMed: 15464184]
- [246]. Peeling RW, Mabey D, Fitzgerald DW, Watson-Jones D. Avoiding HIV and dying of syphilis. *Lancet.* 2004; 364:1561–3. [PubMed: 15519615]
- [247]. Slyker JA, Lohman-Payne BL, John-Stewart GC, et al. Acute cytomegalovirus infection in Kenyan HIV-infected infants. *AIDS.* 2009; 23:2173–81. [PubMed: 19617812]
- [248]. Nigro G, Krzysztofiak A, Gattinara GC, et al. Rapid progression of HIV disease in children with cytomegalovirus DNAemia. *AIDS.* 1996; 10:1127–33. [PubMed: 8874630]
- [249]. Global strategy for the prevention and control of sexually transmitted infections: 2006-2015. World Health Organization; 2007.
- [250]. Guidelines for the management of sexually transmitted infections. World Health Organization; Geneva: 2003.
- [251]. Pepin J, Sobela F, Khonde N, et al. The syndromic management of vaginal discharge using single-dose treatments: a randomized controlled trial in West Africa. *Bull World Health Organ.* 2006; 84:729–38. [PubMed: 17128343]
- [252]. Mlisana K, Naicker N, Werner L, et al. Symptomatic vaginal discharge is a poor predictor of sexually transmitted infections and genital tract inflammation in high-risk women in South Africa. *J Infect Dis.* 2012; 206:6–14. [PubMed: 22517910]

- [253]. Riedner G, Rusizoka M, Todd J, et al. Single-dose azithromycin versus penicillin G benzathine for the treatment of early syphilis. *N Engl J Med.* 2005; 353:1236–44. [PubMed: 16177249]
- [254]. Handsfield HH, Dalu ZA, Martin DH, Douglas JM Jr, McCarty JM, Schlossberg D, Azithromycin Gonorrhea Study Group. Multicenter trial of single-dose azithromycin vs ceftriaxone in the treatment of uncomplicated gonorrhea. *Sex Transm Dis.* 1994; 21:107–11. [PubMed: 9071422]
- [255]. Lau CY, Qureshi AK. Azithromycin versus doxycycline for genital chlamydial infections: a meta-analysis of randomized clinical trials. *Sex Transm Dis.* 2002; 29:497–502. [PubMed: 12218839]
- [256]. Luntamo M, Kulmala T, Mbewe B, Cheung YB, Maleta K, Ashorn P. Effect of repeated treatment of pregnant women with sulfadoxine-pyrimethamine and azithromycin on preterm delivery in Malawi: a randomized controlled trial. *Am J Trop Med Hyg.* 2010; 83:1212–20. [PubMed: 21118924]
- [257]. McDonald HM, Brocklehurst P, Gordon A. Antibiotics for treating bacterial vaginosis in pregnancy. *Cochrane Database Syst Rev.* 2007; 1:CD000262. [PubMed: 17253447]
- [258]. Grimwade K, Swingler GH. Cotrimoxazole prophylaxis for opportunistic infections in children with HIV infection. *Cochrane Database Syst Rev.* 2006; 1:CD003508. [PubMed: 16437457]
- [259]. Chico RM, Mayaud P, Ariti C, Mabey D, Ronsmans C, Chandramohan D. Prevalence of malaria and sexually transmitted and reproductive tract infections in pregnancy in sub-Saharan Africa: a systematic review. *JAMA.* 2012; 307:2079–86. [PubMed: 22665107]
- [260]. Kaplan JE, Benson C, Holmes KH, Brooks JT, Pau A, Masur H. Guidelines for prevention and treatment of opportunistic infections in HIV-infected adults and adolescents: recommendations from CDC, the National Institutes of Health, and the HIV Medicine Association of the Infectious Diseases Society of America. *MMWR Recomm Rep.* 2009; 58(RR-4):1–207.

Table 1
Maternal and Infant Co-Infections that have an Impact on MTCT of HIV

Co-Infection	Mechanisms of Affecting MTCT of HIV	Intervention Strategies
Chorioamnionitis	Potential to increase MTCT <i>in utero</i> via placental inflammation or intrapartum via association with preterm labor and premature rupture of membranes	Treatment with antibiotics during pregnancy has not shown value in preventing MTCT of HIV [65, 66]
Lower genital tract infections (HSV, syphilis, BV, trichomoniasis, etc.)	Potential to increase MTCT <i>in utero</i> and intrapartum by increased peripheral blood HIV load and increased genital tract HIV shedding and local mucosal inflammation	Treatment of HSV with valacyclovir in pregnant and postpartum women reduced HIV RNA in plasma and breast milk [95] but did not reduce MTCT; further studies are needed Trials of vaginal cleansing have not shown benefit in reducing MTCT [111-114]
Mastitis	Increased MTCT during breastfeeding from increased viral load in breast milk	Counseling on proper breastfeeding technique. Exclusive breastfeeding was associated with reduced MTCT of HIV, compared with mixed feeding [20, 236-238] Antibiotic treatment of mastitis may reduce breast inflammation [126], however effects on MTCT are unknown
Gingivitis/Periodontal Disease	Bleeding gums or increased HIV shedding in oral secretions associated with periodontal inflammation when food is pre-masticated for infants by an HIV-infected caregiver	HIV-infected caregivers should not pre-masticate food for infants [131]
Viral Co-infections (HBV, HCV, CMV, others)	Increased MTCT due to increased prevalence of co-infections among more immunosuppressed women and increased maternal HIV load associated with co-infections	Prenatal maternal screening for HBV. If positive, infants should receive hepatitis B vaccine and immune globulin soon after birth [141] Antiviral regimens against HBV may be considered during pregnancy [141]
TB	Associated with maternal immune suppression and higher HIV load.	Appropriate maternal treatment with antiretrovirals and regimens against TB may result in improvement in immune status and decreases in VL
Malaria	Possibly increased MTCT <i>in utero</i> and intrapartum due to increase in maternal viral load, preterm delivery and low birth weight	Appropriate malaria preventive measures during pregnancy (intermittent preventive therapy, insecticide-treated bed nets)
Other parasitic infections (toxoplasmosis, trypanosomiasis, helminths)	Associated with maternal immune suppression and with increased HIV load	Periodic deworming in endemic areas may be associated with improvement in immune status and decreases in HIV load Screening for toxoplasmosis during pregnancy; appropriate prophylaxis or treatment, as necessary [260]
Infant Oral Candidiasis	Increased MTCT during breastfeeding due to breaches in oral mucosa and influx of activated CD4+ T lymphocytes and macrophages as well as HIV into the GI tract	Treatment of oral candidiasis will decrease oral mucosal inflammation
Infant Diarrhea	Increased MTCT during breastfeeding due to increased intestinal permeability	Promotion of breastfeeding in resource-limited settings decreases infant diarrheal disease