**Supplemental Figure 1. Overview of gating strategy of lung cells.** Following treatments, mice were euthanized, lavage fluid removed, pulmonary vasculature perfused, and lung cells dissociated and analyzed by flow cytometry. Initial gating excluded debris (data not shown). Next, CD45+ lung leukocytes were identified (R1 gate). Lymphocytes gated based on characteristic forward and side-scatter properties of CD45+ leukocytes and lymphocytes subsets deter-mined by characteristic CD3, CD4, and CD8 staining (data not shown). Next, lymphocytes were excluded, and neutrophils gated based on Ly-6G staining properties (R3). Next, neutrophils were excluded, and CD11 macrophages gated on CD11c staining and forward scatter properties (R4). Respective populations were utilized in experimental studies.

**Supplemental Figure 2. Cell surface staining of CD80, CD86, and IA-b on CD11c+ macrophages from WT and SRA KO mice following ODE exposures and recovery.** Mice were intra-nasally treated with ODE daily for 3 wk followed by no treatment for 1 wk (recovery ODE) whereupon they were euthanized, lavage fluid removed, pulmonary vasculature perfused, and lung cells dissociated. Alveolar macrophages (CD11chi/CD11blo) and exudative macrophages (CD11chi/CD11bhi) were identified using the gating strategy depicted in Figure 5. Representative histograms from one of four individual mice displaying cell surface expression of macrophage-associated proteins CD80, CD86, and I-Ab. Gray-shaded histogram shows isotype staining and black histogram shows specific staining. Insufficient exudative macrophages from saline-treated animals to conduct adequate flow cytometry analysis.