

Genome Sequences of Nine *Bordetella holmesii* Strains Isolated in the United States

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An increasing number of pertussis-like cases are attributed to the emergent pathogen *Bordetella holmesii*. The genomes of 9 clinical isolates show that they are clonal, lack the virulence factors encoded by *B. pertussis*, and are more similar to nonpertussis bordetellae. New markers for *B. holmesii* can be developed using these sequences.

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An increasing number of pertussis-like cases in the United States and Europe are attributed to the emergent pathogen *Bordetella holmesii* (1, 2). An analysis of the 2010–2011 outbreak in Ohio showed that *B. holmesii* was detected in nearly 20% of pertussis-like illnesses (1), up from ~1% in the 1990s (3, 4). Only three whole-genome sequences are available for this group of pathogens (5–7). *B. holmesii* genomes do not encode known *Bordetella pertussis* virulence factors, though they share a genomic island that contains the IS481 insertion element (7) commonly used to diagnose *B. pertussis* infections and leading to common misidentifications. More specific diagnostic tests were developed using the *B. holmesii*-specific IS1001 element (4), but it is unclear whether this marker is sufficient due to sparse genomic data.

Here, we report the genome sequences of 9 clinical isolates obtained between 2004 and 2011: six from patients with bacteremia (five from blood and one from synovial fluid) and three respiratory isolates from patients with pertussis-like symptoms. A respiratory isolate was from an infant who had a coinfection with *B. pertussis*. Genomic DNA was prepared (8) and sequenced using a combination of 3- or 5-kb mate-pair (~30× coverage) and 100-bp Illumina paired-end reads (~50× coverage). After quality trimming, all reads were used in assemblies with Celera Assembler

6.1 (9) or Velvet Assembler. Underlying consensus sequences and gaps were improved using custom scripts. All genomes had between 119 and 213 contigs (Table 1). The overall G+C content was ~62.6%, with genome sizes ranging from 3.55 Mb to 3.59 Mb.

B. holmesii isolates belonged to the same multilocus sequencing type (MLST), as seen in *B. pertussis* strains (10), suggesting that the *B. holmesii* isolates were also clonal. The genomes were annotated and predicted to have between 3,118 and 3,285 genes. The genomic content of the *B. holmesii* strains was more similar to those of *Bordetella avium* and *Bordetella petrii* than to those of *B. pertussis* or *B. bronchiseptica*. However, nearly 66% of the genes were shared with *B. pertussis* or *Bordetella bronchiseptica*. Almost 400 genes were shared by all *B. holmesii* isolates but were not present in any other bordetellae, likely due to acquisition via horizontal transfer. Many of these genes were involved in the transport and detoxification of organic compounds and antibiotics. Each strain had between 24 and 114 unique genes, including one strain that had residual members of a degraded type III secretion system, as seen in *Escherichia coli* (11). As expected, the IS481 element was present in all genomes (32 to 65 copies), as was BhlIS1001 (5 to 21 copies). The acellular vaccine targets of pertus-

TABLE 1 Isolate characteristics and accession numbers

<i>B. holmesii</i> isolate ID	State	Yr isolated	Source	Genome length (bp)	Total no. of contigs	GenBank accession no.
H572	Colorado	2010	Synovial fluid	3,585,459	119	JFZY000000000
H585	Minnesota	2010	Blood	3,587,402	150	JFZZ000000000
H629	New York	2010	Blood	3,475,248	190	JGVZ000000000
H635	California	2010	Respiratory fluid	3,569,022	173	JGAA000000000
H643	Pennsylvania	2010	Blood	3,614,976	193	JGWD000000000
H719	Minnesota	2011	Blood	3,578,998	149	JGWA000000000
H785	Oregon	2011	Respiratory fluid	3,565,090	161	JGWB000000000
H809	New York	2011	Blood	3,584,230	153	JMGZ000000000
04P3421	Massachusetts	2004	Respiratory fluid	3,595,240	213	JGWC000000000

sis toxin, pertactin, and fimbriae were not present, while filamentous hemagglutinin was encoded by all *B. holmesii* genomes.

These findings suggest that circulating *B. holmesii* isolates in the United States emerged from a single genetic background more similar to nonpertussis bordetellae. The genomes are a resource for understanding the pathogenicity and evolution of *B. holmesii* and for further developing detection and differentiation strategies.

Nucleotide sequence accession numbers. The *B. holmesii* whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. described in Table 1. The version described in this paper is the first version.

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REFERENCES

- Rodgers L, Martin SW, Cohn A, Budd J, Marcon M, Terranella A, Mandal S, Salamon D, Leber A, Tondella M-L, Tatti K, Spicer K, Emanuel A, Koch E, McGlone L, Pawloski L, LeMaile-Williams M, Tucker N, Iyer R, Clark TA, DiOrio M. 2013. Epidemiologic and laboratory features of a large outbreak of pertussis-like illnesses associated with cocirculating *Bordetella holmesii* and *Bordetella pertussis*—Ohio, 2010–2011. *Clin. Infect. Dis.* 56:322–331. <http://dx.doi.org/10.1093/cid/cis888>.
- Njamkepo E, Bonacorsi S, Debruyne M, Gibaud SA, Guillot S, Guiso N. 2011. Significant finding of *Bordetella holmesii* DNA in nasopharyngeal samples from French patients with suspected pertussis. *J. Clin. Microbiol.* 49:4347–4348. <http://dx.doi.org/10.1128/JCM.01272-11>.
- Yih WK, Silva EA, Ida J, Harrington N, Lett SM, George H. 1999. *Bordetella holmesii*-like organisms isolated from Massachusetts patients with pertussis-like symptoms. *Emerg. Infect. Dis.* 5:441–443. <http://dx.doi.org/10.3201/eid0503.990317>.
- Antila M, He Q, de Jong C, Aarts I, Verbakel H, Bruisten S, Keller S, Haanperä M, Mäkinen J, Eerola E, Viljanen MK, Mertsola J, van der Zee A. 2006. *Bordetella holmesii* DNA is not detected in nasopharyngeal swabs from Finnish and Dutch patients with suspected pertussis. *J. Med. Microbiol.* 55:1043–1051. <http://dx.doi.org/10.1099/jmm.0.46331-0>.
- Planet PJ, Narechania A, Hymes SR, Gagliardo C, Huard RC, Whittier S, Della-Latta P, Ratner AJ. 2013. *Bordetella holmesii*: initial genomic analysis of an emerging opportunist. *Pathog. Dis.* 67:132–135. <http://dx.doi.org/10.1111/2049-632X.12028>.
- Tatti KM, Loparev VN, Ranganathan Ganakammal S, Changayil S, Frace M, Weil MR, Sammons S, MacCannell D, Mayer LW, Tondella ML. 2013. Draft genome sequences of *Bordetella holmesii* strains from blood (F627) and nasopharynx (H558). *Genome Announc.* 1(2):e00056-13. <http://dx.doi.org/10.1128/genomeA.00056-13>.
- Diavatopoulos DA, Cummings CA, van der Heide HG, van Gent M, Liew S, Relman DA, Mooi FR. 2006. Characterization of a highly conserved island in the otherwise divergent *Bordetella holmesii* and *Bordetella pertussis* genomes. *J. Bacteriol.* 188:8385–8394. <http://dx.doi.org/10.1128/JB.01081-06>.
- Chomczynski P, Sacchi N. 1987. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal. Biochem.* 162:156–159. <http://dx.doi.org/10.1006/abio.1987.9999>.
- Miller JR, Delcher AL, Koren S, Venter E, Walenz BP, Brownley A, Johnson J, Li K, Mobarry C, Sutton G. 2008. Aggressive assembly of pyrosequencing reads with mates. *Bioinformatics* 24:2818–2824. <http://dx.doi.org/10.1093/bioinformatics/btn548>.
- Harvill ET, Goodfield LL, Ivanov Y, Meyer JA, Newth C, Cassiday P, Tondella ML, Liao P, Zimmerman J, Meert K, Wessel D, Berger J, Dean JM, Holubkov R, Burr J, Liu T, Brinkac L, Kim M, Losada L. 2013. Genome sequences of 28 *Bordetella pertussis* U.S. outbreak strains dating from 2010 to 2012. *Genome Announc.* 1(6):e01075-13. <http://dx.doi.org/10.1128/genomeA.01075-13>.
- Ren CP, Chaudhuri RR, Fivian A, Bailey CM, Antonio M, Barnes WM, Pallen MJ. 2004. The ETT2 gene cluster, encoding a second type III secretion system from *Escherichia coli*, is present in the majority of strains but has undergone widespread mutational attrition. *J. Bacteriol.* 186:3547–3560. <http://dx.doi.org/10.1128/JB.186.11.3547-3560.2004>.