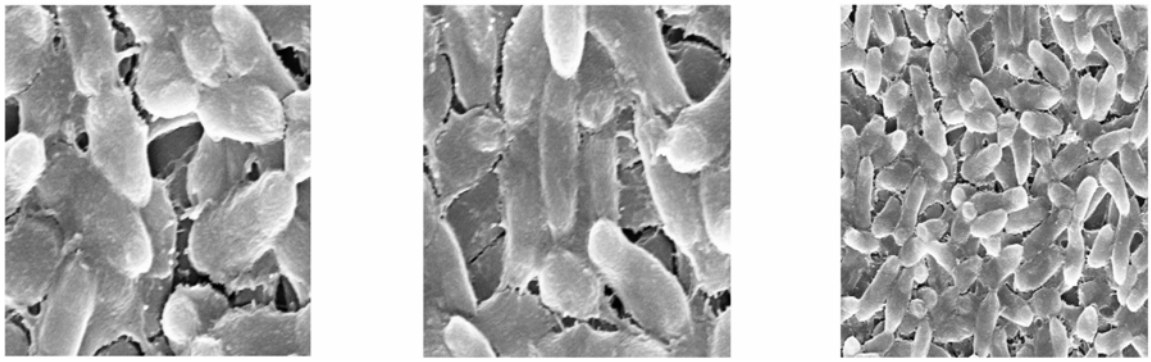

Figure S3. SEM images of wild-type cells treated with sPNAG (400,000x, 200,000x, and 100,000x respectively).

50 ml of fresh LB was inoculated with ~2.5 × 10^{10} wild-type MG1655 cells with a sPNAG content of 0.1U per milliliter of the mixture at 25ºC. A glass slide was provided as biofilm formation surface in a vertical orientation. The biofilm formed on the surface of the glass slide after 12 hours was fixed with 2.5% glutaraldehyde in 200mM sodium cadodylate for 60 minutes. Next, it was gently washed twice with 200mM sodium cadodylate. The sample was post-fixed with 1% osmium tetroxide in sodium cacodylate buffer for 30 minutes. It was washed 4 times, 10 minutes each, with distilled water to remove all the fixative and buffer salts. Next, it was dehydrated sequentially in 35%, 45%, 55%, 65%, 70%, 85%, 95%, and 100% ethanol, 5 minutes each. Then, it was transferred first to 50% ethanol: 50% TMS (tetramethylsilane) followed by 20% ethanol: 80% TMS, 15 minutes each. All the above mentioned steps were carried out at 4ºC. The sample was allowed to air dry at room temperature, and coated with palladium/gold. Finally, the biofilm structures were visualized by a Philips XL30 Field Emission Scanning Electron Microscope.