



Contribution of antimicrobials to the development of allergic disease

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Antimicrobials represent a broad class of chemicals with the intended purpose of eliminating or controlling the growth of harmful microorganisms. Exposure can occur occupationally or through the use or consumption of consumer products. The use of antimicrobial agents has been associated with an increased incidence of allergic diseases, including asthma, atopic dermatitis, and less commonly, anaphylaxis. Very diverse immunological mechanisms and mediators have been identified in the sensitization response to antimicrobial chemicals and the importance of the local microenvironment in the response is increasingly being recognized. A complete understanding of the mechanisms of allergic diseases resulting from antimicrobial exposure will help to ensure safe environments and exposure limits.

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route, the environment) and by factors inherent to the exposed individual (i.e. genetics, immune status, personal habits). Allergic diseases are characterized by a latency period between primary exposure (sensitization) and symptoms (elicitation) that develop upon subsequent exposures, and may involve IgE and/or non IgE-mediated responses. An IgE-mediated allergic reaction (sometimes called immediate-type hypersensitivity (Type I)) involves the production of Th2 cytokines, such as IL-4 and IL-13, which initiate IgE production by B cells; the elicitation phase (symptoms) is caused by IgE-driven mast cell degranulation. These reactions may occur in various organs resulting in diseases such as asthma, allergic rhinitis, urticaria, and anaphylaxis. A non-IgE-mediated or delayed-type hypersensitivity response (Type IV) is T cell-mediated and elicitation is characterized by excessive inflammation, initiated by responding memory T-cells. The most distinctive feature of a non-IgE-mediated hypersensitivity response is the delay observed between allergen exposure and the elicitation immune response. Following sensitization, subsequent exposures result in secretion of pro-inflammatory cytokines that activate and recruit macrophages and other immune cells. Allergic contact dermatitis (ACD) is an example of a non-IgE-mediated hypersensitivity reaction. Interestingly antimicrobials are low molecular weight chemicals that can induce divergent immune mechanisms and can result in multiple types of allergic disease including those described above. This review will focus on the description of specific antimicrobials that have been associated with sensitization along with research advancements that have helped us to better understand the mechanisms of disease.

Introduction

Antimicrobials represent a broad class of chemicals that include sterilizers, disinfectants, sanitizers, and preservatives with the intended purpose of eliminating or controlling the growth of harmful microorganisms such as bacteria, fungi, and viruses. The range of specificity and the effectiveness of antimicrobial agents are very diverse based on the type/types of chemical(s) used. While the importance of these antimicrobial chemicals is understood, exposure to many of these agents is known to directly induce a variety of health effects including allergic disease [1,2].

Allergic disease can be influenced by several factors elicited by the exposure itself (i.e. source, exposure

Occupational and consumer exposure to antimicrobial chemicals

Workers in health-care and clinical facilities have one of the highest incidences of allergic disease compared to other industrial sectors [3,4]. Individuals in these settings are frequently exposed to a variety of high-level cleaners and disinfectants (alcohol, chlorine, iodine-based agents, phenols, hydrogen peroxide, and quaternary ammonium compounds (QACs)) along with antiseptics for the purposes of sterilization of surfaces and medical/surgical instruments, and reducing the incidence of nosocomial infections. It has been estimated that asthma among health care workers accounts for about 16% of all occupational asthma cases [5] and that up to 24% of these cases are due to exposure to cleaning agents [6], including formaldehyde, glutaraldehyde, hypochlorite bleach,

hydrogen peroxide, and enzymatic cleaners [1,7]. Additionally, nurses have been identified to have a higher risk of asthma compared to administrative staff, and tasks with the highest incidence of asthma include those resulting from dilution of disinfection products by manual mixing [8]. In general, there are also increased rates for ACD for health care workers compared to non-health care workers [4] due to exposure to numerous antimicrobial agents including formaldehyde and biocides [9,10]. Although most common in health care workers, there is also an increased incidence for allergic disease in janitors/cleaners, hairdressers, dental assistants, veterinarians, food preparation/service workers, and metalworkers as a result of antimicrobial exposures [9].

Some of the most common allergens in the health care profession include biocides and disinfectants commonly used for the sterilization of medical devices which are sensitive to normal heat or steam sterilization and the disinfection of surfaces [11]. Exposure to the high-level disinfectants, glutaraldehyde and OPA, has been associated with dermatitis and occupational asthma [12–15]. QACs (sprays and wet-wipe products used for disinfecting surfaces and floors) are also common occupational allergens and have been associated with both contact dermatitis and occupational asthma [8,11,16–19]. Additionally, clinical antibacterial hand sanitizers and soaps containing chemicals including chloroxylenol, triclosan, and cocamide diethanolamine have also been associated with allergic disease [11,20,21]. Chlorhexidine gluconate is commonly used as an antiseptic applied directly to the skin to prepare for surgery. Although rare, serious allergic reactions including anaphylaxis have been reported following exposure [22].

In addition to occupational exposures to antimicrobial chemicals, there is also concern for exposures to the general population via consumer and food products including: cleaners and disinfectants for non-critical surfaces, algaecides, fabric softeners, cosmetics, moisturizers, antistatic agents, and food additives and preservatives. The use of antiseptics, disinfectants, detergents, and preservatives has also increased their incorporation into consumer products that are utilized orally (toothpaste and mouthwash) or applied to the skin or eyes (nasal sprays, makeup, lotions, and ophthalmic drops) for the purpose of decreasing microbial contamination and reducing the incidence of pathogen-induced illness. In general, antimicrobials exhibit a wide spectrum of activity; and new ones are constantly being developed/modified for specific purposes.

Novel mediators involved in sensitization to antimicrobial agents

As mentioned earlier, exposure to antimicrobial chemicals can result in multiple hypersensitivity pathways/disease outcomes (i.e. both type I/type IV; ACD and

asthma), which increases the complexity of the immunological mechanism driving the response. Further research is needed to evaluate the hazard-potential and to fully understand the immunological mechanisms that induce and exacerbate allergic diseases. Recently, our laboratory found that a QAC, didecylmethyl ammonium chloride (DDAC), is a strong dermal sensitizer [16]. Although DDAC displays many of the immunological attributes of a T-cell mediated sensitizer, dermal exposure also results in increased production of IgE and Th1 and Th2 cytokines, indicating a mixed-type allergic response [23]. Further investigation into these responses showed that DDAC induced high levels of expression of the Th2-skewing cytokine TSLP, which has been shown to activate Type 2 Innate Lymphoid cells (ILC2s) in the skin. ILC2s, a subset of innate lymphocytes that lack rearranged antigen-specific receptors and produce Type 2 cytokines, have recently emerged as mediators of allergic disease [24]. Following DDAC exposure, ILC2s in the skin were rapidly activated, and their activation coincided with the production of type 2 cytokines in the absence of T cells, providing a potential mechanism for the initiation of the mixed-type allergic response.

As exemplified by the study described above, it is becoming increasingly recognized that sensitization can be influenced by the local microenvironment. Recently, mitochondrial dysfunction has been shown to influence multiple hypersensitivity pathways [25], and antimicrobial chemicals have been demonstrated to negatively affect mitochondrial function in cells present at sites of antimicrobial exposure [26,27]. Triclosan has been identified as a mitochondrial uncoupler (decreases ATP and increases oxygen consumption rate) of mast cells, human primary keratinocytes, and *in vivo* in live zebrafish at non-cytotoxic concentrations [28,29]. Additional mitochondrial effects including altered morphology, calcium levels, and membrane potential were also observed following exposure [30]. Similarly, QAC exposure induced mitochondrial fragmentation and shifts in mitochondrial bioenergetics in epithelial cells [26]. Many of these mitochondrial alterations, including morphology and bioenergetics dysfunction, can elicit proinflammatory upregulation which could influence hypersensitivity response [29]. Understanding how novel mediators (which are often tissue-specific) initiate the development of allergic immune responses at the site of antigen contact will be crucial to predicting the allergic nature of antimicrobials and understanding the type of disease that may manifest due to exposure [31].

Antimicrobials and increased allergic susceptibility

While certain antimicrobials, including those described above, are known to induce sensitization, others such as triclosan have been associated with allergic disease, although not directly sensitizing. In addition to its clinical

use, triclosan is used as a preservative, fungicide, and biocide in household and personal care products [29,32,33]. Research suggests that triclosan exposure may be at least in part responsible for recent increases in the frequency of asthma and allergic disease [20,34,35]. Additional studies have revealed that topical exposure to triclosan augmented the allergic response to an experimental allergen through a thymic stromal lymphopoietin (TSLP)-mediated pathway in a mouse model of asthma [34,36]. The increased Th2 response would likely result in a decreased Th1 response, which could potentially increase an individual's susceptibility to infection and/or disease [37]. Triclosan has been also shown to have direct effects on skin cells [38], which can influence the local microenvironment and immune response [39]. More specifically triclosan has been identified to induce abundant expression of S100A8/A9 in the skin, which acts as an endogenous ligand for the intracellular signaling receptor TLR4, which is responsible for activation of the innate immune system [30]. Similar to triclosan, parabens are preservatives found in a wide range of products encountered daily. Parabens were identified to be useful antibacterial and antifungal agents leading to their use as a preservative. The odorless, colorless, and inexpensive qualities made them a primary choice when trying to extend the shelf life of cosmetics, pharmaceuticals, and other consumer goods. In a review of cosmetic ingredients, 87%–93% of cosmetic products contained at least one paraben [9]. Despite frequent use, the incidence of allergy to parabens is relatively low [4]. Interestingly, disruption of the epidermis including compromised or inflamed skin, which can result from genetic predisposition, is often a prerequisite for ACD resulting from paraben exposure [40,41]. While research does suggest an association between paraben exposure and asthma, existing literature is conflicting [42–44]. Confounding factors include: prenatal versus postnatal exposures; prevalence versus morbidity outcomes, and sex-differences. Additional investigations are warranted.

Antimicrobials and the microbiome

As most antimicrobials used in both the workplace and commercial products are broad spectrum, the potential exists that antimicrobials applied in products may influence an individual's microbiome. The importance of the microbiome in immune responses is increasingly being recognized, including the development of allergic disease. 16S rRNA sequencing results revealed that 13-week triclosan exposure in drinking water induced significant perturbations in mouse gut bacterial assemblages with distinct trajectories compared to controls [45]. Metagenomics sequencing results indicated a remarkable enrichment of gut bacterial genes related to triclosan resistance, stress response, antibiotic resistance, and heavy metal resistance. However, much less research has been conducted on the effects of antimicrobials on the skin microbiome. Many chemical

sensitizers are also antimicrobial agents [12,16,18] and while these agents may be beneficial in protecting against pathogenic bacteria, the influence of exposure on resident bacterial populations warrants further investigations. Recently, skin commensals have been shown to play an important role in Th1 and Th2 balance along with the regulation of anti-inflammatory responses to chemical allergens [46]. Research has also shown that germ-free mice have elevated levels of TSLP, which suggests a role for the microbiota in ameliorating stress signals released by keratinocytes which could potentially influence subsequent immune responses [47]. Additionally, a recent study found that germ-free mice contain largely undifferentiated mast cells and reveal that the skin microbiota plays a significant role in the recruitment and maturation of dermal immune mast cells [48]. SanMiguel *et al.* found that topical treatment with antibiotics led to significant and stable changes in commensal bacterial populations in mice; however, antiseptics only resulted in short-term perturbations [49]. A follow-up study confirmed the rapid, but short-term effect on microbial communities in humans, and also showed that the depleting effect of antiseptics (80% ethanol and povidone-iodine) was dependent on body site and the composition of the microbial community with more dominant taxa being more resistant [50]. Another study showed that the antimicrobial preservative and QAC benzalkonium chloride (BAC) significantly reduced the microbiome (assessed by positive cultures) of the nasal cavity when delivered as a topical ocular lubricant [51]. BAC is present in more than 70% of ophthalmic drugs, highlighting the ubiquitous use of antimicrobials/preservatives in products used by consumers, and their potential impact on allergic disease [52].

Conclusions

While the use of antimicrobials is important for preventing infections, especially in clinical settings, there is also a responsibility to provide a safe and healthy environment for workers, patients, and consumers. The impact of antimicrobial agents is due not only to their sensitizing potency but also to their broad source of exposure and widespread use in daily life. While research demonstrates a role for antimicrobial chemicals in allergic disease, the exact mechanism of action for sensitization for most of these compounds remains to be investigated and explained. In addition, new chemicals are constantly being synthesized for specific antimicrobial purposes or as potentially fewer toxic alternatives and these may also present unique burdens on the immune system. A complete understanding of the mechanisms of allergic diseases resulting from antimicrobial exposure will allow for surveillance, proper treatment and/or prevention, while hazard identification will lead to risk assessment, which will ensure safe environments and exposure limits.

Disclaimer

The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the National Institute for Occupational Safety and Health, Centers for Disease Control and Prevention.

Conflict of interest statement

Nothing declared.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest

- Li RWH, Lipszyc JC, Prasad S, Tarlo SM: **Work-related asthma from cleaning agents versus other agents.** *Occup Med (Lond)* 2018, **68**:589-592.
- Warshaw EM, Maibach HI, Taylor JS, Sasseville D, DeKoven JG, Zirwas MJ *et al.*: **North American contact dermatitis group patch test results: 2011–2012.** *Dermatitis* 2015, **26**:49-59.
- Walters GI, Moore VC, McGrath EE, Burge PS, Henneberger PK: **Agents and trends in health care workers' occupational asthma.** *Occup Med (Lond)* 2013, **63**:513-516.
- Warshaw EM, Schram SE, Maibach HI, Belsito DV, Marks JG Jr, Fowler JF Jr *et al.*: **Occupation-related contact dermatitis in North American health care workers referred for patch testing: cross-sectional data, 1998 to 2004.** *Dermatitis* 2008, **19**:261-274.
- Pechter E, Davis LK, Tumpowsky C, Flattery J, Harrison R, Reinisch F *et al.*: **Work-related asthma among health care workers: surveillance data from California, Massachusetts, Michigan, and New Jersey, 1993–1997.** *Am J Ind Med* 2005, **47**:265-275.
- Pigatto PD, Polenghi MM, Altomare GF: **Occupational dermatitis in bakers: a clue for atopic contact dermatitis.** *Contact Dermatitis* 1987, **16**:263-271.
- Dumas O, Wiley AS, Quinot C, Varraso R, Zock JP, Henneberger PK *et al.*: **Occupational exposure to disinfectants and asthma control in US nurses.** *Eur Respir J* 2017, **50**.
- Gonzalez M, Jegu J, Kopferschmitt MC, Donnay C, Hedelin G, Matzinger F *et al.*: **Asthma among workers in healthcare settings: role of disinfection with quaternary ammonium compounds.** *Clin Exp Allergy* 2014, **44**:393-406.
- Maier LE, Lampel HP, Bhutani T, Jacob SE: **Hand dermatitis: a focus on allergic contact dermatitis to biocides.** *Dermatol Clin* 2009, **27**:251-264 v-vi.
- Prodi A, Rui F, Fortina AB, Corradin MT, Filon FL: **Healthcare workers and skin sensitization: north-eastern Italian database.** *Occup Med (Lond)* 2016, **66**:72-74.
- Suneja T, Belsito DV: **Occupational dermatoses in health care workers evaluated for suspected allergic contact dermatitis.** *Contact Dermatitis* 2008, **58**:285-290.
- Anderson SE, Umbricht C, Sellamuthu R, Fluharty K, Kashon M, Franko J *et al.*: **Irritancy and allergic responses induced by topical application of ortho-phthalaldehyde.** *Toxicol Sci* 2010, **115**:435-443.
- Franchi A, Franco G: **Evidence-based decision making in an endoscopy nurse with respiratory symptoms exposed to the new ortho-phthalaldehyde (OPA) disinfectant.** *Occup Med (Lond)* 2005, **55**:575-578.
- Fujita H, Ogawa M, Endo Y: **A case of occupational bronchial asthma and contact dermatitis caused by ortho-phthalaldehyde exposure in a medical worker.** *J Occup Health* 2006, **48**:413-416.
- Shaffer MP, Belsito DV: **Allergic contact dermatitis from glutaraldehyde in health-care workers.** *Contact Dermatitis* 2000, **43**:150-156.
- Anderson SE, Shane H, Long C, Lukomska E, Meade BJ, Marshall NB: **Evaluation of the irritancy and hypersensitivity potential following topical application of didecyldimethylammonium chloride.** *J Immunotoxicol* 2016, **13**:557-566.
- Bernstein JA, Stauder T, Bernstein DI, Bernstein IL: **A combined respiratory and cutaneous hypersensitivity syndrome induced by work exposure to quaternary amines.** *J Allergy Clin Immunol* 1994, **94**:257-259.
- Shane HL, Lukomska E, Stefaniak AB, Anderson SE: **Divergent hypersensitivity responses following topical application of the quaternary ammonium compound, didecyldimethylammonium bromide.** *J Immunotoxicol* 2017, **14**:204-214.
- Shutty BG, Scheinman PL: **Occupationally induced allergic contact dermatitis to aerosolized quaternary ammonium compounds.** *Dermatitis* 2017, **28**:369.
- Savage JH, Johns CB, Hauser R, Litonjua AA: **Urinary triclosan levels and recent asthma exacerbations.** *Ann Allergy Asthma Immunol* 2014, **112**:179-181.e2.
- Toletone A, Dini G, Massa E, Bragazzi NL, Pignatti P, Voltolini S *et al.*: **Chlorhexidine-induced anaphylaxis occurring in the workplace in a health-care worker: case report and review of the literature.** *La Medicina del lavoro* 2018, **109**:68-76.
- Aschenbrenner DS: **Rare allergic reaction to topical chlorhexidine gluconate.** *Am J Nurs* 2017, **117**:20.
- Shane HL, Lukomska E, Kashon ML, Anderson SE: **Topical application of the quaternary ammonium compound didecyldimethylammonium chloride activates type 2 innate lymphoid cells and initiates a mixed-type allergic response.** *Toxicol Sci* 2019, **168**:505-518.
- Cosmi L, Liotta F, Maggi L, Annunziato F: **Role of type 2 innate lymphoid cells in allergic diseases.** *Curr Allergy Asthma Rep* 2017, **17**:66.
- Iyer D, Mishra N, Agrawal A: **Mitochondrial function in allergic disease.** *Curr Allergy Asthma Rep* 2017, **17**:29.
- Inacio AS, Costa GN, Domingues NS, Santos MS, Moreno AJ, Vaz WL *et al.*: **Mitochondrial dysfunction is the focus of quaternary ammonium surfactant toxicity to mammalian epithelial cells.** *Antimicrob Agents Chemother* 2013, **57**:2631-2639.
- Weatherly LM, Shim J, Hashmi HN, Kennedy RH, Hess ST, Gosse JA: **Antimicrobial agent triclosan is a proton ionophore uncoupler of mitochondria in living rat and human mast cells and in primary human keratinocytes.** *J Appl Toxicol* 2016, **36**:777-789.
- Shim J, Weatherly LM, Luc RH, Dorman MT, Neilson A, Ng R *et al.*: **Triclosan is a mitochondrial uncoupler in live zebrafish.** *J Appl Toxicol* 2016, **36**:1662-1667.
- Weatherly LM, Gosse JA: **Triclosan exposure, transformation, and human health effects.** *J Toxicol Environ Health Part B Crit Rev* 2017, **20**:447-469.
- Marshall NB, Lukomska E, Nayak AP, Long CM, Hettick JM, Anderson SE: **Topical application of the anti-microbial chemical triclosan induces immunomodulatory responses through the S100A8/A9-TLR4 pathway.** *J Immunotoxicol* 2017, **14**:50-59.
- Shane H, Long CM, Anderson S: **Novel cutaneous mediators of chemical allergy.** *J Immunol* 2019:1-15.
- Fang JL, Stingley RL, Beland FA, Harrouk W, Lumpkins DL, Howard P: **Occurrence, efficacy, metabolism, and toxicity of triclosan.** *J Environ Sci Health C Environ Carcinog Ecotoxicol Rev* 2010, **28**:147-171.
- Glaser A: **The ubiquitous triclosan: a common antibacterial agent exposed.** *Pesticides You* 2004, **24**:12-17.

34. Anderson SE, Franko J, Kashon ML, Anderson KL, Hubbs AF, Lukomska E *et al.*: **Exposure to triclosan augments the allergic response to ovalbumin in a mouse model of asthma.** *Toxicol Sci* 2013, **132**:96-106.
35. Savage JH, Matsui EC, Wood RA, Keet CA: **Urinary levels of triclosan and parabens are associated with aeroallergen and food sensitization.** *J Allergy Clin Immunol* 2012, **130**:453-460.e7.
36. Marshall NB, Lukomska E, Long CM, Kashon ML, Sharpnack DD, Nayak AP *et al.*: **Triclosan induces thymic stromal lymphopoietin in skin promoting Th2 allergic responses.** *Toxicol Sci* 2015, **147**:127-139.
37. Mabbott NA: **The influence of parasite infections on host immunity to co-infection with other pathogens.** *Front Immunol* 2018, **9**:2579.
38. Weatherly LM, Nelson AJ, Shim J, Riitano AM, Gerson ED, Hart AJ *et al.*: **Antimicrobial agent triclosan disrupts mitochondrial structure, revealed by super-resolution microscopy, and inhibits mast cell signaling via calcium modulation.** *Toxicol Appl Pharmacol* 2018, **349**:39-54.
39. Steinhoff M, Brzoska T, Luger TA: **Keratinocytes in epidermal immune responses.** *Curr Opin Allergy Clin Immunol* 2001, **1**:469-476.
40. Cashman AL, Warshaw EM: **Parabens: a review of epidemiology, structure, allergenicity, and hormonal properties.** *Dermatitis* 2005, **16**:57-66 quiz 55-6.
41. Joensen UN, Jorgensen N, Thyssen JP, Petersen JH, Szecsi PB, Stender S *et al.*: **Exposure to phenols, parabens and UV filters: associations with loss-of-function mutations in the filaggrin gene in men from the general population.** *Environ Int* 2017, **105**:105-111.
42. Berger K, Eskenazi B, Balmes J, Holland N, Calafat AM, Harley KG: **Associations between prenatal maternal urinary concentrations of personal care product chemical biomarkers and childhood respiratory and allergic outcomes in the CHAMACOS study.** *Environ Int* 2018, **121**:538-549.
43. Lee-Sarwar K, Hauser R, Calafat AM, Ye X, O'Connor GT, Sandel M *et al.*: **Prenatal and early-life triclosan and paraben exposure and allergic outcomes.** *J Allergy Clin Immunol* 2018, **142**:269-278.e15.
44. Quiros-Alcala L, Hansel NN, McCormack MC, Matsui EC: **Paraben exposures and asthma-related outcomes among children from the US general population.** *J Allergy Clin Immunol* 2019, **143**:948-956.
45. Gao B, Tu P, Bian X, Chi L, Ru H, Lu K: **Profound perturbation induced by triclosan exposure in mouse gut microbiome: a less resilient microbial community with elevated antibiotic and metal resistomes.** *BMC Pharmacol Toxicol* 2017, **18**:46.
46. Knaysi G, Smith AR, Wilson JM, Wisniewski JA: **The skin as a route of allergen exposure: part II. Allergens and role of the microbiome and environmental exposures.** *Curr Allergy Asthma Rep* 2017, **17**:7.
47. Yockey LJ, Demehri S, Turkoz M, Turkoz A, Ahern PP, Jassim O *et al.*: **The absence of a microbiota enhances TSLP expression in mice with defective skin barrier but does not affect the severity of their allergic inflammation.** *J Invest Dermatol* 2013, **133**:2714-2721.
48. Wang Z, Mascarenhas N, Eckmann L, Miyamoto Y, Sun X, Kawakami T *et al.*: **Skin microbiome promotes mast cell maturation by triggering stem cell factor production in keratinocytes.** *J Allergy Clin Immunol* 2017, **139**:1205-1216.e6.
49. SanMiguel AJ, Meisel JS, Horwinski J, Zheng Q, Grice EA: **Topical antimicrobial treatments can elicit shifts to resident skin bacterial communities and reduce colonization by staphylococcus aureus competitors.** *Antimicrob Agents Chemother* 2017, **61**.
50. SanMiguel AJ, Meisel JS, Horwinski J, Zheng Q, Bradley CW, Grice EA: **Antiseptic agents elicit short-term, personalized, and body site-specific shifts in resident skin bacterial communities.** *J Invest Dermatol* 2018, **138**:2234-2243.
51. Onerci Celebi O, Celebi ARC: **The effect of ocular lubricants containing benzalkonium chloride on nasal mucosal flora.** *Cutan Ocul Toxicol* 2018, **37**:305-308.
52. Noecker R: **Effects of common ophthalmic preservatives on ocular health.** *Adv Ther* 2001, **18**:205-215.