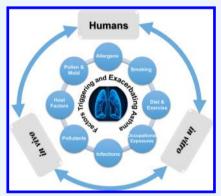


Ins and Outs in Environmental and Occupational Safety Studies of Asthma and Engineered **Nanomaterials**

Marina A. Dobrovolskaia, Michael R. Shurin, Valerian E. Kagan, and Anna A. Shvedova*, L,#

ABSTRACT: According to the Centers for Disease Control and Prevention, approximately 25 million Americans suffer from asthma. The disease total annual cost is about \$56 billion and includes both the direct and indirect costs of medications, hospital stays, missed work, and decreased productivity. Air pollution with xenobiotics, bacterial agents, and industrial nanomaterials, such as carbon nanotubes, contribute to the exacerbation of this condition and are a point of particular attention in environmental toxicology as well as in occupational health and safety research. Mast cell degranulation and activation of Th2 cells triggered either by allergen-specific immunoglobulin E (IgE) or by alternative mechanisms, such as locally produced neurotransmitters, underlie the pathophysiological process of airway constriction during an asthma attack. Other immune and nonimmune cell types, including basophils, eosinophils, Th₁, Th₂, Th₂, macrophages, dendritic cells, and smooth muscle cells, are involved in the inflammatory and allergic responses during asthma, which, under chronic conditions, may progress



without mast cells, the key trigger of the acute asthma attack. To decipher complex molecular, cellular, and genetic mechanisms, many researchers have attempted to develop in vitro and in vivo models to study asthma. Herein, we summarize the advantages and disadvantages of various models and their applicability to nanoparticle evaluation in asthma research. We further suggest that a framework for both in vitro and in vivo methods should be used to study the impact of engineered nanomaterials on asthma etiology, pathophysiology, and treatment.

sthma is an inflammatory disorder occurring in the lungs as a result of repeated immediate-type hypersensitivity and late-phase allergic reactions to the inhaled allergen(s). It is characterized by repeated and reversible airway obstruction, chronic bronchial inflammation, hyperreactivity to bronchoconstrictors, and smooth muscle cell hypertrophy of bronchi. Asthma often coexists with chronic obstructive pulmonary disorder (COPD). The combination of these conditions frequently leads to irreversible airway obstruction, morbidity, and even death. According to the Centers for Disease Control and Prevention (CDC), asthma affects ~8% of Americans (~25 million people). Other reports suggest that 15.7% of asthma cases among adults in the United States (~1.9 million cases) are work related. Overall, about 300 million people worldwide suffer from this disorder,³ and 11% of these cases are associated with workplace conditions, such as exposure to fumes, gases, or dust. The asthma prevalence rate is similar across industrialized countries but is lower in nations with a less developed industrial base. According to the Global Initiative for Asthma (GINA), the prevalence rate has increased by 50% every decade and is accountable for 250,000 deaths annually.³ Another organization whose research efforts focus on this disorder, the Asthma and Allergy Foundation of America, conveyed that direct (hospital stays and medications) and indirect (missed work hours and decreased worker productivity) annual costs of asthma in the United States total over \$56 billion. These data emphasize the impact of the disorder on modern society and justify extensive research in this area.

Published: July 24, 2017

[†]Nanotechnology Characterization Laboratory, Cancer Research Technology Program, Leidos Biomedical Research Inc., Frederick National Laboratory for Cancer Research, NCI at Frederick, Frederick, Maryland 21702, United States

[‡]Department of Pathology, University of Pittsburgh Medical Center, Pittsburgh, Pennsylvania 15213, United States

[§]Department of Immunology, University of Pittsburgh Medical Center, Pittsburgh, Pennsylvania 15213, United States

Departments of Environmental and Occupational Health, Pharmacology and Chemical Biology, Chemistry and Radiation Oncology and Center for Free and Antioxidant Health, University of Pittsburgh, Pittsburgh, Pennsylvania 15260, United States

¹Health Effects Laboratory Division, National Institute of Occupational Safety and Health, Centers for Disease Control and Prevention, Morgantown, West Virginia 26505, United States

Department of Physiology and Pharmacology, West Virginia University, Morgantown, West Virginia 26506, United States

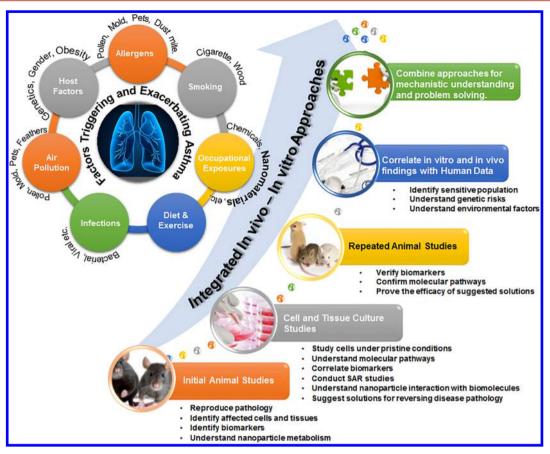


Figure 1. Systemic approaches required for evaluating the asthmatic effects of nanomaterials exposure.

Factors triggering and exacerbating asthma include both immune (i.e., allergen-specific immunoglobulins E, IgE) and non-immune (e.g., cold, certain drugs, exercise) stimuli (Figure 1). Allergen-specific IgE binding to mast cells and basophils and activation of Th2 cells reacting to allergens trigger the pathophysiologic chain of events culminating in an allergic asthma attack. Mast cell degranulation and release of cytokines by Th₂ cells result in recruitment of progressively increasing numbers of T-cells, eosinophils, and basophils. Although the T-cell population in asthma is dominated by Th2 cells, other T-cell subtypes are also involved, including Th₁, Th₁₇, and Th₉ cells. The infiltrating airway lymphocytes continue to stimulate the inflammatory response by secreting cytokines and chemokines, which also support the cellular infiltration into the airway. The inflammatory response finally results in hypertrophy of smooth muscles, thickening of basement membranes, hyperplasia of mucous glands, and continuing cellular infiltration. These alterations of the airway, or airway remodeling, may lead to fibrosis and irreversible airway obstruction in some patients. Collectively, cytokines and mediators secreted by lymphocytes trigger hypertrophy and hyperreactivity of bronchial smooth muscle cells, whereas those released by eosinophils, basophils, and mast cells result in the constriction of airway smooth muscles.

In contrast to the engagement of different cell populations, the molecular mechanisms underlying allergic and non-allergic asthma are insufficiently understood and thus represent major gaps in deciphering the pathogenesis of the disease. One such gap is related to the role of environmental factors and conditions. Specifically, it is well-established that air pollution with xenobiotics, bacterial ligands (e.g., lipopolysaccharide), and

some industrial nanomaterials (carbon nanotubes, CNTs; carbon black; and some diesel exhaust particles) have a significant influence on the development and progression of asthma. Moreover, the effects of these environmental factors also depend on the age of exposed individuals. One hypothesis, termed the hygiene hypothesis, proposes that more frequent bacterial infections, lower air pollution with xenobiotics and diesel exhaust, and higher exposure to commensal flora in a farm setting lead to regulated maturation of the immune system and promote less frequent occurrence of asthma and other Th2-driven responses to non-environmental antigens in less industrialized areas. The same hypothesis states that a cleaner bacterial environment accompanied by greater air pollution in industrialized areas are responsible for the higher prevalence of atopic diseases in these regions. However, it is also known that bacterial and viral infections of the respiratory tract predispose a person to the development of asthma and exacerbate preexisting asthma. The hygiene hypothesis does not explain how such infections promote Th2 and mast cell responses or clarify how non-immune mediators (e.g., cold, physical and mental stress, chemicals, medications) trigger this disease. Genetic differences may also affect susceptibility to asthma, as well as responsiveness to asthma medications. Therefore, more research is needed to understand both cellular and molecular mechanisms underlying this complex disorder.

The respiratory system represents the main route of exposure to aerosolized nanomaterials, by accident or by occupational factors. Although many studies have explored the respiratory effects of engineered nanomaterials, including CNTs, in different animal models, few publications have documented the effects of respiratory nanomaterials in experimental models

ns in uc

A variety of *in vitro* and *in vivo* models have been employed to improve our understanding of asthma and the contributions of environmental nanomaterials to its pathogenesis.

of pulmonary disease. Environmental exposure in sensitized individuals is a critical inducer of an airway inflammatory response. which is a hallmark of the asthmatic lung. Although a variety of factors can initiate inflammation via different cellular and molecular pathways, their cumulative effects lead to increased bronchial reactivity. In the area of environmental toxicology and occupational health, engineered nanomaterials, and particularly carbon-based particles, have recently attracted significant attention as potential triggers of and contributors to asthma.⁶ Due to their high aspect ratio and biopersistance, several studies have emphasized the role of CNTs in asthma (e.g., see refs 7-10). A recent report demonstrated the presence of CNTs in the bronchoalveolar fluid of children with asthma.¹ Another study, comparing gene expression profiles in the blood of individuals exposed to CNTs to those of naïve workers in the same facility, revealed significant alterations in the expression of the genes implicated in diverse cellular processes such as apoptosis, cell-cycle regulation, and proliferation. 12 These findings further fueled the public alertness and triggered more thorough investigations of CNTs' involvement in asthma pathogenesis, including detailed and mechanistic in vivo studies.^{7–12} A variety of *in vitro* and *in vivo* models have been employed to improve both the understanding of asthma and the contributions of environmental nanomaterials to its pathogenesis. The advantages and limitations of these models are discussed below.

The benefits of in vivo studies include the ability to account for particle biodistribution, metabolism, and repeated exposure and to mimic the chronic inhalation conditions that exist in the environment and contribute to lung inflammation and asthma (Table 1). For instance, an in vivo study in a model of asthma demonstrated that pulmonary exposure to carbon nanoparticles (CNPs) induces a significant release of proinflammatory cytokines, modulates proliferation of splenic T-cells, and alters the pattern of inflammatory cells in the lung.¹³ Growing evidence demonstrates that the adverse effects of inhaled CNTs are increased under conditions of pre-existing inflammation such as in allergic asthma. Recent data revealed that exposure to CNTs exacerbates ovalbumin (OVA)-induced allergic airway inflammation in mice by causing airway fibrosis, increasing OVA-induced T-cell proliferation, and amplifying lung Th, cytokines and chemokines. Interestingly, induction of allergic responses exaggerated by CNTs involved activation of B-cells and was accompanied by the production of IgE in the absence of pre-exposure to allergen. 14-16 This effect is not unique to CNTs, as the exposure to another carbon-based nanomaterial, graphene oxide (GO), resulted in potentiation of OVA-triggered asthma potentially through the mechanism involving an increase in chitinase production by macrophages. 13 These and other in vivo studies in animal models suggest that humans with allergic asthma may potentially be more susceptible to the enhanced immune responses and airway remodeling induced by carbon-based nanomaterials than animals. However, due to genetic diversity between different species and even different strains of animals of the same species, as well as differences in the immune responses elicited

Table 1. Advantages and Limitations of In Vitro and In Vivo Methods Used To Study the Role of Engineered Nanomaterials in the Development of Asthma

in vivo	es	bution often does not consider the real nanoparticle concentrations in occupational setting	degradation interspecies immune system differences between rodents and humans make extrapolation of data difficult	difficult to model irregular nature of repeated exposure, especially in the context of other environmental factors	nic inhalation does not account for genetic polymorphisms	en various cell types more time-consuming	onary diseases more cost-intensive	enables working with transgenic and knockout animals to study low-throughput specific cell types or molecular pathways	ı humanized mouse models high possibility of lung overload during intratracheal instillation	difference in biodistribution to secondary organs between inhalation and intratracheal instillation
	advantages	able to account for particle biodistribution	able to account for metabolism and degradation	enables repeated exposure	enables more close mimicking chronic inhalation	enables complex interactions between various cell types	enables modeling of different pulmonary diseases	enables working with transgenic and knock specific cell types or molecular pathways	enables evaluation of human cells in humanized mouse models	enables testing different routes of administration
in vitro	limitations	does not account for particle biodistribution and homing	does not account for particle metabolism and degradation	hard to account for mechanisms involving enables repeated exposure multiple cellular players	cannot account for relevant repeated exposure	low predictive value				
	advantages	able to single out particular cell type	enables deciphering specific molecular pathway(s)	less time-consuming	less cost-intensive	high-throughput	ability to test human cells and biological fluids relevant to disease	possibility of testing exact doses and concentrations	opportunity of long-term cultures and cell lines	cell-specific testing of specific inhibitors, antagonists and antibodies

by mice and humans, extrapolation of *in vivo* data to the human population is often limited, and it is still unknown whether carbon nanotubes can cause or exacerbate asthma in humans.

Additional technical nuances have to be considered when analyzing and interpreting complex in vivo results. Specifically, redistribution of airborne particles from the lungs to other organs and tissues depends on the physicochemical properties of nanomaterials, including composition, size, shape, surface properties, and aspect ratio. 17-19 Routes of nanoparticle entry into the body, their biodistribution, and their metabolism may also contribute significantly to the test results. Administration routes (e.g., inhalation versus intratracheal instillation) may affect the outcomes even in the same animal strain and for the same nanomaterial. For example, inhaled iridium nanoparticles were found in the lung and other organs (liver, spleen, brain, and heart) of experimental rats.^{20,21} In contrast, these nanomaterials administered *via* intratracheal instillation were not found in secondary organs.²² The potential explanation for the difference was associated with their partial ingestion after the inhalation; as such, some of the inhaled particles may be ingested and distributed to the liver and spleen via gastrointestinal absorption.²¹ Thus, the study design is critical for data interpretation.²¹ Intratracheal instillation, however, may lead to lung overload, thus causing the exaggerated picture of pulmonary toxicity of nanomaterials administered via this route. The importance of the administration route and its relevance to the effects of nanoparticles on humans has recently been emphasized.²³

Other factors of special relevance to nanoparticle exposure include changes in nanoparticle composition and physicochemical properties following particle interactions with proteins and other biomolecules and the dose. The changes in nanoparticle composition and physicochemical properties are triggered by adsorption of different biomolecules on the nanoparticle surface, biodegradation of nanomaterials via enzymatic machinery of inflammatory cells, and direct pro-oxidant effects of nanoparticles. Adsorption of biomolecules on the nanoparticle surface leads to the formation of so-called protein and lipid "coronas" whose composition is dynamic and changes during the redistribution of nanoparticles between different compartments in the body. 24,25 For example, it was demonstrated that different protein and lipid components of the lung surfactant, initially integrated on to the nanoparticle surface after inhalation, were replaced by plasma proteins and lipids after the nanoparticles transferred to the systemic circulation.²⁶⁻²⁸ Such "coronas" of nanoparticles may contribute to the particle recognition as pro-asthmatic signals by different types of immune cells.^{29–31} The biodegradation of nanoparticles is also essential for their role in the immune responses and pathogenesis of asthma. The major oxidative enzymatic pathways of inflammatory cells, including myeloperoxidase/ NADPH oxidase of neutrophils and iNOS/NADPH oxidase of macrophages, were reported to degrade carbon-based nanomaterials catalytically.³² In addition to destroying nanoparticles and lowering their concentrations in tissues, these oxidative effects may also lead to the appearance of oxidized proteins, lipids, and carbohydrates, thus affecting their immunogenic potential.³³ Although the exact role and significance of these metabolic transformations of nanoparticles in the pathogenesis of asthma remain to be elucidated, the ability of nanomaterials to affect other enzymatic pathways in an experimental mouse asthma model has been already reported. For example, studies of the mechanisms of up-regulated airway responsiveness

following pulmonary GO exposure in an animal model of Ovalbumin (OVA) sensitization uncovered an intriguing role of pulmonary macrophage-derived chitinase and chitinase-like molecules. 13 The chitinases are hydrolytic enzymes that break down glycosidic bonds in chitin. These enzymes were found to improve allergic responses in murine models of allergy via a mechanism involving the increase in pulmonary macrophages, reduction in alveolar eosinophils, and decrease in serum IgE levels. Interestingly, although the GO induced up-regulation of chitinase expression in the lung, the enzymatic activity of this enzyme was decreased. 13 Based on the molecular modeling studies, the authors suggested that GO binds to the entrance of the catalytic site of chitinase, mimicking interactions with chitinase inhibitors. Thus, by interfering with chitinase, nanomaterials may restrict its role in allergic inflammation and asthma.1

The ability of nanoparticles to alter the activity of different enzymatic systems is not limited to chitinase and has also been reported for other enzymes, for example, α -chymotrypsin.³⁴ The dose of the tested nanomaterials should also be considered. It has been found that dendritic cells' (DCs) exposure to a low or medium dose of fullerenes preceding an antigen (OVA) challenge promoted the ability of DCs to stimulate OVA-specific T-cells. In contrast, a higher dose of fullerenes down-regulated antigen presentation by DCs, whereas carbon nanoparticles did not affect the ability of DCs to stimulate antigen-specific T-cells.³⁵ These differential effects of carbonaceous nanoparticles on DCs may be associated with their capacity to alter expression of the main proteins involved in antigen processing machinery, such as immunoproteasomes.³⁵ These results support the notion that nanoparticles may modify the ability of antigen-presenting cells to process and to present antigens (allergens) and, thus, to control Th₁/Th₂ balance in exposed individuals.

Finally, nanoparticles containing transition metals such as biologically common Fe, Cu, Mn, Cr, Ni, and Co as well as those normally not present in the body (Ce) can all affect the redox environment and cause oxidative stress and injury. In the context of asthma and Th₂ inflammation, pro-oxidant conditions are essential factors that influence exacerbations of asthma episodes and worsen its general course and outcomes. These effects may be related to the lowered effectiveness and levels of reduced glutathione (GSH) and GSH-driven enzymes such as GSH-peroxidases. All of the conditions discussed above emphasize the complexity of biological responses to nanoparticles and nanoparticle interaction with various cells, proteins, and other macromolecules. The inherent complexity of *in vivo* responses to nanoparticle exposure limits the value and the adequacy of the *in vitro* studies.

One common criticism of animal studies is their potentially limited relevance to the human population. ²³ Among the other limitations of *in vivo* models are their high cost and relatively low-throughput. *In vitro* models help to overcome some of these constraints. They offer higher throughput, are less cost intensive, and enable better insights into the contributions of individual cell types and the isolation of specific molecular pathways. Their major drawbacks include the inability to account for particle distribution and metabolism; inability to mimic repeated exposure to multiple, often fluctuating, environmental factors accurately; and do not represent relevant regulatory interactions between various cells, organs, and tissues. Altogether, these limitations lower the predictive value of *in vitro* models. To overcome the limitations of *in vitro*

models pertinent to their inability to mimic chronic inflammation, Chortarea et al. suggest the use of an air-liquid interface cell exposure (ALICE) system, as reported in this issue of ACS Nano.³⁸ This system is efficient in aerosolization of both spherical and fibrous nanomaterials and permits repeated aerosol exposures of cultured bronchial epithelial cells to CNTs. Cells harvested from healthy individuals and asthmatic patients were cultured at the air-liquid interface and treated in vitro with an aerosolized form of CNTs at concentrations corresponding to human occupational lifetime exposures.³⁸ The results showed that chronic CNT exposure elicited a duration-dependent pro-inflammatory and oxidative stress response as well as a significant alteration of the mucociliary clearance mechanism in both healthy and asthmatic cultures. The latter displayed stronger and more durable long-term effects compared to healthy cells, indicating that individuals with asthma may be more prone to adverse effects from CNT exposure compared to non-asthmatic populations. 17 The results also highlight the importance of occupationally relevant subchronic exposures when in vitro models are used in the nanotoxicity hazard assessments. Another recently proposed in vitro system reconstitutes organ-level lung functions on a chip.³⁹ This biomimetic microsystem retains the critical functional alveolar-capillary interface of the human lung. The microdevice reproduces complex integrated organ-level responses to bacteria and inflammatory cytokines introduced into the alveolar space. The lung-mimicking model has been applied to nanotoxicology studies and has demonstrated adequate responses to silica nanoparticles regarding toxic and inflammatory reactions as well as enhanced epithelial and endothelial uptake of nanoparticles and their transport into the underlying microvascular channel.

To overcome the limitations of *in vitro* models pertinent to their inability to mimic chronic inflammation, Chortarea *et al.* suggest the use of an air—liquid interface cell exposure (ALICE) system, as reported in this issue of *ACS Nano*.

For better mechanistic insights into pathophysiological events during asthma development, analysis of *in vitro—in vivo* correlation can be performed (Figure 1). The dominant clearance mechanism of inhaled nanoparticles is particle uptake by the lung resident macrophages. As such, oxidative stress, cytokine secretion, and consequent inflammation were commonly reported in studies with inhaled nanomaterials.

Particle aggregation to larger, micron-sized materials and the presence of other environmental factors, such as endotoxin, were shown to contribute to inflammatory reactions to inhaled nanomaterials. For example, certain airborne particulates are not inflammatory alone but enhance the inflammation caused by endotoxin. Therefore, mitigation of the inhalation toxicity of environmental nanoparticles should include control over contributing contaminants such as endotoxin. Oberdorster *et al.* suggested inclusion of several end-points (reactive oxygen species, lactate dehydrogenase leakage, protein oxidation induction, the number of polymorphonuclear neutrophils in bronchoalveolar fluid) in the study design to correlate *in vitro* and *in vivo* toxicology data. Other researchers have suggested that hemolysis correlates with inflammation *in vivo*.

In vitro—in vivo correlation of the hemolysis tests have been demonstrated by several studies (see review by Brown *et al.*⁴⁹). From a mechanistic standpoint, stroma-free hemoglobin can act as a peroxidase, provided a source of oxidizing equivalents is available. ^{50,51} Under conditions of enhanced inflammation, superoxide radicals generated by macrophages and, to a lesser extent, neutrophils ⁵² can dismutate to yield H₂O₂ that will feed the peroxidase activity of free hemoglobin. Once formed, this poorly controlled pro-oxidant catalytic center can cause severe oxidative stress and oxidative damage to many cells, including red blood cells, thus perpetuating release of hemoglobin and creating a vicious cycle of pro-oxidant reactions. ⁵³

Collectively, the available data indicate that current models do not accurately mimic the complex network of cellular, biochemical, and molecular pathways involved in the development of asthma, especially in different environmental conditions. The advantages and disadvantages of each of the mentioned approaches are summarized in Table 1. Recognizing the limitations and non-ideal nature of each of the approaches is important to arrive at accurate conclusions. Future studies would benefit from better understanding of the nanomaterial effects on various types of immune cells beyond alveolar macrophages. More profound insights into nanoparticle effects on mast cells, neutrophils, eosinophils, basophils, and multiple subsets of T-cells (Th₁, Th₂, Th₁₇, Th₉), as well as their communications with and bidirectorial regulation by macrophages and other antigen-presenting cells, is needed. Systematic approaches based on both in vitro and in vivo studies would fulfill this need (Figure 1).

AUTHOR INFORMATION

Corresponding Author

*E-mail: ats1@cdc.gov.

ORCID

Valerian E. Kagan: 0000-0002-7245-1885

Notes

The content and conclusions of this publication are those of the authors and do not necessarily reflect the views or policies of the Department of Health and Human Services, National Institute for Occupational Safety and Health, Center for Disease Control, nor does mention of trade names, commercial products, or organizations imply endorsement by the U.S. government.

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

The work has been funded with federal funds from the National Cancer Institute, National Institutes of Health, contract HHSN261200800001E and NORA 939051G.

REFERENCES

- (1) C.f.D. Control. Asthma: Recent Statistics, 2017.
- (2) C.f.D.C.a. Prevention. Work-Related Asthma in 22 States. Morbidity and Mortality Weekly Report, 2015; p 343.
- (3) Braman, S. S. The Global Burden of Asthma. *Chest* **2006**, *130*, 4S–12S
- (4) World Health Organization. Global Surveillance, Prevention and Control of Chronic Respiratory Diseases: A Comprehensive Approach, 2017
- (5) A.a.A.F.o. America, Costs of Asthma to Society, 2017.
- (6) Li, N.; Georas, S.; Alexis, N.; Fritz, P.; Xia, T.; Williams, M. A.; Horner, E.; Nel, A. A Work Group Report on Ultrafine Particles (American Academy of Allergy, Asthma & Immunology): Why

Ambient Ultrafine and Engineered Nanoparticles Should Receive Special Attention for Possible Adverse Health Outcomes in Human Subjects. *J. Allergy Clin. Immunol.* **2016**, *138*, 386–396.

- (7) Ronzani, C.; Casset, A.; Pons, F. Exposure to Multi-Walled Carbon Nanotubes Results in Aggravation of Airway Inflammation and Remodeling and in Increased Production of Epithelium-Derived Innate Cytokines in a Mouse Model of Asthma. *Arch. Toxicol.* **2014**, *88*, 489–499.
- (8) Rydman, E. M.; Ilves, M.; Koivisto, A. J.; Kinaret, P. A.; Fortino, V.; Savinko, T. S.; Lehto, M. T.; Pulkkinen, V.; Vippola, M.; Hameri, K. J.; Matikainen, S.; Wolff, H.; Savolainen, K. M.; Greco, D.; Alenius, H. Inhalation of Rod-Like Carbon Nanotubes Causes Unconventional Allergic Airway Inflammation. *Part. Fibre Toxicol.* **2014**, *11*, 48.
- (9) Staal, Y. C.; van Triel, J. J.; Maarschalkerweerd, T. V.; Arts, J. H.; Duistermaat, E.; Muijser, H.; van de Sandt, J. J.; Kuper, C. F. Inhaled Multiwalled Carbon Nanotubes Modulate the Immune Response of Trimellitic Anhydride-Induced Chemical Respiratory Allergy in Brown Norway Rats. *Toxicol. Pathol.* **2014**, *42*, 1130–1142.
- (10) Kayat, J.; Gajbhiye, V.; Tekade, R. K.; Jain, N. K. Pulmonary Toxicity of Carbon Nanotubes: A Systematic Report. *Nanomedicine* **2011**, *7*, 40–49.
- (11) Kolosnjaj-Tabi, J.; Just, J.; Hartman, K. B.; Laoudi, Y.; Boudjemaa, S.; Alloyeau, D.; Szwarc, H.; Wilson, L. J.; Moussa, F. Anthropogenic Carbon Nanotubes Found in the Airways of Parisian Children. *EBioMedicine* **2015**, *2*, 1697–1704.
- (12) Shvedova, A. A.; Yanamala, N.; Kisin, E. R.; Khailullin, T. O.; Birch, M. E.; Fatkhutdinova, L. M. Integrated Analysis of Dysregulated ncRNA and mRNA Expression Profiles in Humans Exposed to Carbon Nanotubes. *PLoS One* **2016**, *11*, e0150628.
- (13) Shurin, M. R.; Yanamala, N.; Kisin, E. R.; Tkach, A. V.; Shurin, G. V.; Murray, A. R.; Leonard, H. D.; Reynolds, J. S.; Gutkin, D. W.; Star, A.; Fadeel, B.; Savolainen, K.; Kagan, V. E.; Shvedova, A. A. Graphene Oxide Attenuates Th₂-Type Immune Responses, but Augments Airway Remodeling and Hyperresponsiveness in a Murine Model of Asthma. *ACS Nano* **2014**, *8*, 5585–5599.
- (14) Inoue, K.; Koike, E.; Yanagisawa, R.; Hirano, S.; Nishikawa, M.; Takano, H. Effects of Multi-Walled Carbon Nanotubes on a Murine Allergic Airway Inflammation Model. *Toxicol. Appl. Pharmacol.* **2009**, 237, 306–316.
- (15) Park, E. J.; Cho, W. S.; Jeong, J.; Yi, J.; Choi, K.; Park, K. Pro-Inflammatory and Potential Allergic Responses Resulting from B Cell Activation in Mice Treated with Multi-Walled Carbon Nanotubes by Intratracheal Instillation. *Toxicology* **2009**, 259, 113–121.
- (16) Ryman-Rasmussen, J. P.; Tewksbury, E. W.; Moss, O. R.; Cesta, M. F.; Wong, B. A.; Bonner, J. C. Inhaled Multiwalled Carbon Nanotubes Potentiate Airway Fibrosis in Murine Allergic Asthma. *Am. J. Respir. Cell Mol. Biol.* **2009**, *40*, 349–358.
- (17) Gregoratto, D.; Bailey, M. R.; Marsh, J. W. Particle Clearance in the Alveolar-Interstitial Region of the Human Lungs: Model Validation. *Radiat. Prot. Dosim.* **2011**, *144*, 353–356.
- (18) Bailey, M. R.; Ansoborlo, E.; Guilmette, R. A.; Paquet, F. Practical Application of the ICRP Human Respiratory Tract Model. *Radiat. Prot. Dosim.* **2003**, *105*, 71–76.
- (19) Bailey, M. R.; Ansoborlo, E.; Guilmette, R. A.; Paquet, F. Updating the ICRP Human Respiratory Tract Model. *Radiat. Prot. Dosim.* **2007**, 127, 31–34.
- (20) Kreyling, W. G.; Semmler, M.; Erbe, F.; Mayer, P.; Takenaka, S.; Schulz, H.; Oberdorster, G.; Ziesenis, A. Translocation of Ultrafine Insoluble Iridium Particles from Lung Epithelium to Extrapulmonary Organs Is Size Dependent but Very Low. *J. Toxicol. Environ. Health, Part A* **2002**, *65*, 1513–1530.
- (21) Oberdorster, G.; Sharp, Z.; Atudorei, V.; Elder, A.; Gelein, R.; Lunts, A.; Kreyling, W.; Cox, C. Extrapulmonary Translocation of Ultrafine Carbon Particles Following Whole-Body Inhalation Exposure of Rats. *J. Toxicol. Environ. Health, Part A* **2002**, *65*, 1531–1543.
- (22) Brown, J. S.; Zeman, K. L.; Bennett, W. D. Ultrafine Particle Deposition and Clearance in the Healthy and Obstructed Lung. *Am. J. Respir. Crit. Care Med.* **2002**, *166*, 1240–1247.

(23) Stern, S. T.; McNeil, S. E. Nanotechnology Safety Concerns Revisited. *Toxicol. Sci.* **2008**, *101*, 4–21.

- (24) Monopoli, M. P.; Aberg, C.; Salvati, A.; Dawson, K. A. Biomolecular Coronas Provide the Biological Identity of Nanosized Materials. *Nat. Nanotechnol.* **2012**, *7*, 779–786.
- (25) Carrillo-Carrion, C.; Carril, M.; Parak, W. J. Techniques for the Experimental Investigation of the Protein Corona. *Curr. Opin. Biotechnol.* **2017**, *46*, 106–113.
- (26) Pozzi, D.; Caracciolo, G.; Digiacomo, G. L.; Colapicchioni, V.; Palchetti, S.; Capriotti, A. L.; Cavaliere, C.; Zenezini Chiozzi, R.; Puglisi, A.; Laganà, A. The Biomolecular Corona of Nanoparticles in Circulating Biological Media. *Nanoscale* **2015**, *7*, 13958–13966.
- (27) Shvedova, A. A.; Pietroiusti, A.; Fadeel, B.; Kagan, V. E. Mechanisms of Carbon Nanotube-Induced Toxicity: Focus on Oxidative Stress. *Toxicol. Appl. Pharmacol.* **2012**, *261*, 121–133.
- (28) Hadjidemetriou, M.; Al-Ahmady, Z.; Mazza, M.; Collins, R. F.; Dawson, K.; Kostarelos, K. *In Vivo* Biomolecule Corona around Blood-Circulating, Clinically Used and Antibody-Targeted Lipid Bilayer Nanoscale Vesicles. *ACS Nano* **2015**, *9*, 8142–8156.
- (29) Mazzolini, J.; Weber, R. J.; Chen, H. S.; Khan, A.; Guggenheim, E.; Shaw, R. K.; Chipman, J. K.; Viant, M. R.; Rappoport, J. Z. Protein Corona Modulates Uptake and Toxicity of Nanoceria *via* Clathrin-Mediated Endocytosis. *Biol. Bull.* **2016**, 231, 40–60.
- (30) Radauer-Preiml, I.; Andosch, A.; Hawranek, T.; Luetz-Meindl, U.; Wiederstein, M.; Horejs-Hoeck, J.; Himly, M.; Boyles, M.; Duschl, A. Nanoparticle—Allergen Interactions Mediate Human Allergic Responses: Protein Corona Characterization and Cellular Responses. *Part. Fibre Toxicol.* 2015, 13, 3.
- (31) Caracciolo, G.; Palchetti, S.; Colapicchioni, V.; Digiacomo, L.; Pozzi, D.; Capriotti, A. L.; La Barbera, G.; Laganà, A. Stealth Effect of Biomolecular Corona on Nanoparticle Uptake by Immune Cells. *Langmuir* **2015**, *31*, 10764–10773.
- (32) Vlasova, I. I.; Kapralov, A. A.; Michael, Z. P.; Burkert, S. C.; Shurin, M. R.; Star, A.; Shvedova, A. A.; Kagan, V. E. Enzymatic Oxidative Biodegradation of Nanoparticles: Mechanisms, Significance and Applications. *Toxicol. Appl. Pharmacol.* **2016**, 299, 58–69.
- (33) Miller, Y. I.; Tsimikas, S. Oxidation-Specific Epitopes as Targets for Biotheranostic Applications in Humans: Biomarkers, Molecular Imaging and Therapeutics. *Curr. Opin. Lipidol.* **2013**, *24*, 426–437.
- (34) De, M.; Chou, S. S.; Dravid, V. P. Graphene Oxide as an Enzyme Inhibitor: Modulation of Activity of Alpha-Chymotrypsin. *J. Am. Chem. Soc.* **2011**, *133*, 17524–17527.
- (35) Tkach, A. V.; Yanamala, N.; Stanley, S.; Shurin, M. R.; Shurin, G. V.; Kisin, E. R.; Murray, A. R.; Pareso, S.; Khaliullin, T.; Kotchey, G. P.; Castranova, V.; Mathur, S.; Fadeel, B.; Star, A.; Kagan, V. E.; Shvedova, A. A. Graphene Oxide, But Not Fullerenes, Targets Immunoproteasomes and Suppresses Antigen Presentation by Dendritic Cells. *Small* **2013**, *9*, 1686–1690.
- (36) Levine, S. J.; Wenzel, S. E. Narrative Review: The Role of Th₂ Immune Pathway Modulation in the Treatment of Severe Asthma and its Phenotypes. *Ann. Intern. Med.* **2010**, *152*, 232–237.
- (37) Koike, Y.; Hisada, T.; Utsugi, M.; Ishizuka, T.; Shimizu, Y.; Ono, A.; Murata, Y.; Hamuro, J.; Mori, M.; Dobashi, K. Glutathione Redox Regulates Airway Hyperresponsiveness and Airway Inflammation in Mice. *Am. J. Respir. Cell Mol. Biol.* **2007**, *37*, 322–329.
- (38) Chortarea, S.; Barosova, H.; Clift, M. J. D.; Wick, P.; Petri-Fink, A.; Rothen-Rutishauser, B. Human Asthmatic Bronchial Cells Are More Susceptible to Subchronic Repeated Exposures of Aerosolized Carbon Nanotubes At Occupationally Relevant Doses Than Healthy Cells. ACS Nano 2017, DOI: 10.1021/acsnano.7b01992.
- (39) Huh, D.; Matthews, B. D.; Mammoto, A.; Montoya-Zavala, M.; Hsin, H. Y.; Ingber, D. E. Reconstituting Organ-Level Lung Functions on a Chip. *Science* **2010**, 328, 1662–1668.
- (40) Brown, D. M.; Kanase, N.; Gaiser, B.; Johnston, H.; Stone, V. Inflammation and Gene Expression in the Rat Lung After Instillation of Silica Nanoparticles: Effect of Size, Dispersion Medium and Particle Surface Charge. *Toxicol. Lett.* **2014**, 224, 147–156.
- (41) Park, J. W.; Lee, I. C.; Shin, N. R.; Jeon, C. M.; Kwon, O. K.; Ko, J. W.; Kim, J. C.; Oh, S. R.; Shin, I. S.; Ahn, K. S. Copper Oxide

Nanoparticles Aggravate Airway Inflammation and Mucus Production in Asthmatic Mice *via* MAPK Signaling. *Nanotoxicology* **2016**, *10*, 445–452.

- (42) Thompson, E. A.; Sayers, B. C.; Glista-Baker, E. E.; Shipkowski, K. A.; Taylor, A. J.; Bonner, J. C. Innate Immune Responses to Nanoparticle Exposure in the Lung. *J. Environ. Immunol. Toxicol.* **2014**, 2, 150–156.
- (43) Inoue, K.; Takano, H. Aggravating Impact of Nanoparticles on Immune-Mediated Pulmonary Inflammation. *Sci. World J.* **2011**, *11*, 382–390.
- (44) Inoue, K.; Takano, H.; Yanagisawa, R.; Hirano, S.; Sakurai, M.; Shimada, A.; Yoshikawa, T. Effects of Airway Exposure to Nanoparticles on Lung Inflammation Induced by Bacterial Endotoxin in Mice. *Environ. Health Perspect.* **2006**, *114*, 1325–1330.
- (45) Inoue, K.; Takano, H.; Yanagisawa, R.; Hirano, S.; Kobayashi, T.; Fujitani, Y.; Shimada, A.; Yoshikawa, T. Effects of Inhaled Nanoparticles on Acute Lung Injury Induced by Lipopolysaccharide in Mice. *Toxicology* **2007**, 238, 99–110.
- (46) Inoue, K. Promoting Effects of Nanoparticles/Materials on Sensitive Lung Inflammatory Diseases. *Environ. Health Prev. Med.* **2011**, *16*, 139–143.
- (47) Han, X.; Corson, N.; Wade-Mercer, P.; Gelein, R.; Jiang, J.; Sahu, M.; Biswas, P.; Finkelstein, J. N.; Elder, A.; Oberdorster, G. Assessing the Relevance of *In Vitro* Studies in Nanotoxicology By Examining Correlations between *In Vitro* and *In Vivo* Data. *Toxicology* 2012, 297, 1–9.
- (48) Clouter, A.; Brown, D.; Hohr, D.; Borm, P.; Donaldson, K. Inflammatory Effects of Respirable Quartz Collected in Workplaces versus Standard DQ12 Quartz: Particle Surface Correlates. *Toxicol. Sci.* **2001**, *63*, 90–98.
- (49) Wildt, B.; Malinauskas, R. A.; Brown, R. P. The Effects of Engineered Nanomaterials on Erythrocytes. In *Handbook of Immunological Properties of Nanomaterials*; Dobrovolskaia, M. A., McNeil, S. E., Eds.; World Scientific Publishing: Singapore, 2013.
- (50) Koch, T.; Duncker, H. P.; Heller, A.; Schaible, R.; van Ackern, K.; Neuhof, H. Effects of Stroma-Free Hemoglobin Solutions on Pulmonary Vascular Resistance and Mediator Release in the Isolated Perfused Rabbit Lung. *Shock* **1994**, *1*, 146–152.
- (51) Kapralov, A.; Vlasova, I. I.; Feng, W.; Maeda, A.; Walson, K.; Tyurin, V. A.; Huang, Z.; Aneja, R. K.; Carcillo, J.; Bayir, H.; Kagan, V. E. Peroxidase Activity of Hemoglobin—Haptoglobin Complexes: Covalent Aggregation and Oxidative Stress in Plasma and Macrophages. J. Biol. Chem. 2009, 284, 30395—30407.
- (52) Shvedova, A. A.; Kisin, E. R.; Murray, A. R.; Gorelik, O.; Arepalli, S.; Castranova, V.; Young, S. H.; Gao, F.; Tyurina, Y. Y.; Oury, T. D.; Kagan, V. E. Vitamin E Deficiency Enhances Pulmonary Inflammatory Response and Oxidative Stress Induced by Single-Walled Carbon Nanotubes in C57BL/6 Mice. *Toxicol. Appl. Pharmacol.* 2007, 221, 339–348.
- (53) Zhu, H.; Du, Q.; Chen, C.; Chang, T. M. The Immunological Properties of Stroma-Free Polyhemolysate Containing Catalase and Superoxide Dismutase Activities Prepared by Polymerized Bovine Stroma-Free Hemolysate. *Artif. Cells Blood Substit. Immobil. Biotechnol.* **2010**, 38, 57–63.