

Cytokine polymorphisms in silicosis and other pneumoconioses

Berran Yucesoy,¹ Val Vallyathan,² Douglas P. Landsittel,³
Petia Simeonova⁴ and Michael I. Luster⁴

¹Ankara University, Faculty of Pharmacy, Department of Toxicology, Ankara Turkey; ²Pathology and Physiology Research Branch; ³Biostatistics Branch; ⁴Toxicology and Molecular Biology Branch, Health Effects and Laboratory Division, National Institute for Occupational Safety and Health, Morgantown, WV, USA

Abstract

Silicosis and coal workers' pneumoconiosis are complex multifactorial lung diseases whose etiopathogenesis are not well defined. It is generally accepted that fibrotic lung disorders are mediated by macrophage-derived cytokines and growth factors. There is evidence showing a crucial role for tumor necrosis factor- α (TNF- α) and interleukin-1 (IL-1) in inflammation caused by silica dust and in the transition from simple to progressive massive fibrosis. In this review we discuss genetic polymorphisms responsible for regulating the production of these proinflammatory cytokines and their role in modifying silicosis severity. (*Mol Cell Biochem* **234/235**: 219–224, 2002)

Key words: silicosis, pneumoconiosis, cytokines, polymorphism, TNF- α

Introduction

Among interstitial lung disorders, silicosis and coal workers' pneumoconiosis (CWP) are the most widespread fibrotic lung diseases. Silicosis, very rarely an isolated form of pneumoconiosis in coal workers, is a chronic fibrosing disease of the lungs produced by prolonged and extensive exposure to free crystalline silica. When workers inhale silica, the lung tissue reacts by developing fibrotic nodules and scarring around the trapped silica particles. This pulmonary fibrotic condition is called silicosis and usually occurs against a background of a simple nodular or macular CWP. Workers in mines, foundries, blasting operations, stone, clay and glass manufacturing encounter silica [1, 2]. In the United States, between 1979 and 1996, 2,694 deaths were attributed to silicosis. About 1.6 million workers are believed to have been exposed to silica dust, and almost 60,000 are expected to suffer from some degree of silicosis. CWP, also known as black lung disease, is caused by inhaling coal mine dust. When the disease progresses from simple to complicated pneumoconiosis, the condition is called progressive massive

fibrosis. An estimated 4.5% of coal miners are affected and about 0.2% have scarring on the lungs, the most severe form of the disease. Between 1979 and 1996, 14,156 deaths were attributed to black lung disease [2, 3].

Although their pathophysiology has not been fully understood, several lines of evidence suggest the participation of cytokines produced by alveolar macrophages (AM), at least in the initiation of the alveolitis. The AM is a critically important cell playing a prominent role in lung inflammation via the production of a large panel of mediators including cytokines, reactive oxygen species, enzymes and arachidonic acid metabolites [4, 5].

Inflammatory cytokines as candidate genes for fibrotic lung diseases

Cytokines play key roles in immune responses, inflammation and fibrosis. The cytokines receiving the most attention to date, in relation to pulmonary diseases, include IL-1, TNF-

α , platelet-derived growth factor (PDGF), transforming growth factor- β (TGF- β), insulin-like growth factor I (IGF-I), and interleukin-6 (IL-6) [6–9]. Experimental animal and clinical studies reveal that TNF- α and IL-1 are important in regulating fibrotic mediators in silicosis. In this respect, increased expression of inflammatory cytokines corresponds to pathological changes in lungs of silicotic rodents [10–14]. A major role of TNF- α in pulmonary fibrosis is supported by evidence obtained from TNF- α deficient mice, which are resistant to developing fibrosis from silica [15,16]. In humans, the local release of IL-1 and TNF- α has been shown to coincide with pathogenesis of the disease [17, 18]. Coal mine dust-stimulated release of TNF- α from peripheral blood monocytes (PBM) was also increased in subjects with pneumoconiosis [19–21] while higher levels of spontaneous TNF- α and IL-1 secretion by AMs were observed in patients with CWP [22]. In addition, elevated mRNA levels of TNF- α have been observed in lungs of subjects with pneumoconiosis [23]. These results indicate that AMs are involved in chronic lung inflammatory reactions to mineral dusts, partly by way of cytokine secretion. Moreover, cytokine secretion by AMs was suggested to be an early event in response to mineral dust exposure.

Associations between disease and IL-1 and TNF- α polymorphisms

Multifactorial diseases involve complex interactions among multiple genes and environmental factors. Susceptibility depends on both intrinsic features of the host and the influence of environmental factors [24]. Genetic factors such as polymorphisms are usually not, by themselves, sufficient for most diseases but modify the extent or severity of the disease after it has been initiated. As with other multifactorial diseases, there is a wide inter-individual variability of susceptibility to silicosis. The role of genetic and environmental or physiological factors as disease modifiers may be described as shown in Fig. 1. This pattern is similar to the model of clinical expression of adult periodontitis outlined by Kornman *et al.* [25].

Polymorphisms in cytokine genes have been reported to contribute to the recognized stable inter-individual variation in the level of cytokine production rates [26–28]. Inter-individual differences in spontaneous as well as stimulated production of IL-1 and TNF- α support the possibility that silicosis and pneumoconiosis severity are related to the genetic propensity of the host to produce these proteins. At the IL-1 and TNF loci, some allelic variants have been found to be significantly over-represented in inflammatory diseases. These variations affect the level of TNF- α expression in response to various stimuli. In humans the gene encoding for TNF- α

is located on chromosome 6 between HLA-B and DR, within the class III region of the major histocompatibility complex, and is a candidate gene for autoimmune and inflammatory diseases [29, 30]. Two SNPs, at positions –308 and –238 in the promoter region, [30, 31] are associated with a variety of immune and inflammatory diseases, such as CWP, malaria, leishmaniasis, celiac disease, chronic bronchitis, psoriasis and systemic lupus erythematosus [32–38]. Due to the high degree of linkage disequilibrium across the MHC, TNF- α expression may depend on polymorphisms in the TNF- α promoter region or a linkage association with the HLA genotype [31, 39]. Therefore, it is difficult to determine which genes on a haplotype are important in the etiology of a disease. The –308 variant of TNF- α is reported to be associated with the HLA A1, B8, DR3, DR4 and the DQ2 haplotypes. DR2 positive genotypes have been reported to produce low levels of TNF- α whereas the DR3 and DR4 genotypes produce high levels [30, 40]. Therefore, the increased production of TNF- α could contribute to the increased incidence of autoimmune diseases observed in individuals with an HLA A1, B8 and DR3 haplotypes [41].

Polymorphisms within the human IL-1 gene cluster on chromosome 2 have been associated with several chronic inflammatory diseases [42]. The minor variant of the IL-1RA VNTR in linkage disequilibrium with exon 2 (+2018) has been associated with systemic lupus erythematosus, ulcerative colitis, lichen sclerosis and alopecia areata [43–46]. Two variants in the IL-1 α gene at sites –889 and +4845 are over-represented in juvenile rheumatoid arthritis and chronic polyarthritis [47, 48]. The IL-1 β (+3953) variant has been found to be prevalent in severe periodontitis and psoriasis [25, 49].

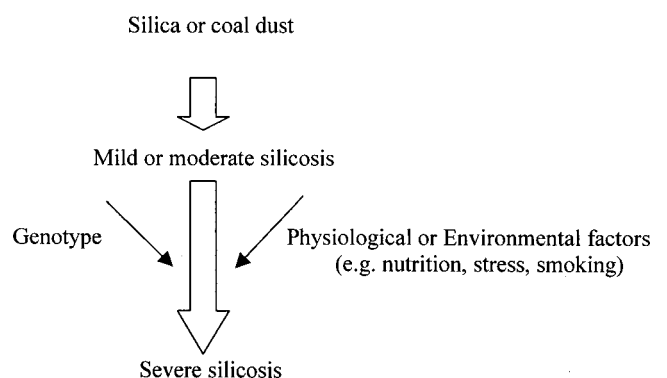


Fig. 1. In the gene by environmental interaction in a biological sense, silica or coal dust are the causal elements producing disease, but specific genotypes and physiological or environmental factors may modify the clinical expression of disease after it has been initiated.

IL-1 and TNF genotypes in silicosis and CWP

In caucasian patients with silicosis, the frequency of HLA-B7 was found to be lower than that in dust-exposed and non-exposed referents and the highest risk of developing severe fibrosis was found to be associated with the HLA-Aw19-B18 haplotype [50, 51]. Immunogenetic analysis revealed that susceptibility to silicosis is associated with HLA-Bw54 in the Japanese population, suggesting that a TNF- α allele, in linkage disequilibrium with this haplotype, might predispose individuals to silicosis. The major gene for silicosis was also reported to be mapped near the HLA-B locus [52]. The frequency of DR8 was elevated in German coal miners with CWP, whereas the frequency of DR1 and DR52 was reduced in miners without CWP [53]. In another study, an increased presence of the -308 variant in the TNF- α promoter was reported in ex-coal miners with mild CWP [38].

In most chronic inflammatory diseases, whatever the role of environmental factors, there are genetic components which cannot be attributed to those linked to the MHC [54]. In view of the genetic findings and the chronic inflammatory nature of silicosis, we investigated whether polymorphisms in the IL-1 and TNF- α genes are associated with the incidence and/or severity of this disease. In this study, all the subjects were selected from a total of 6580 autopsy cases submitted to the National Coal Workers' Autopsy Study from 1972–1996. From these subjects, a random sample of 325 cases was selected and genotyped for at least one of the polymorphisms. Additional 164 autopsy subjects without any evidence of pulmonary disease were defined as controls. Cases with pulmonary silicosis were reviewed and graded according to the criteria and schema developed by a joint committee of the National Institute for Occupational Safety and Health (NIOSH) and College of American Pathologists. Lesions were graded subjectively into three grades of severity; mild, moderate and severe, based on profusion and size of lesions in the sections. All individuals included in the study were Caucasian, males and worked as underground coal miners. Table 1 summarizes the distribution of age, smoking status and years of exposure by disease status.

The polymorphisms that were investigated, distribution of genotypes and allelic frequencies are listed in Table 2. Sub-

jects with severe silicosis were compared to subjects with moderate disease and to subjects with no silicosis. Odds ratios were calculated using a logistic regression model after adjusting for years of occupational exposure. The odds ratio represents the odds of being a case (i.e. proportion of cases divided by proportion of controls) in subjects with the polymorphism divided by the odds of being a case in subjects without the polymorphism. We observed a strong association between silicosis and the TNF- α (-238) variant, as the frequency of this allele were significantly reduced in moderate disease and significantly predictive of severe disease (adjusted odds ratio 0.5 and 4.0, respectively). This implies that individuals with the TNF- α (-238) variant are predisposed to more rapid development of severe silicosis, which would account for the apparently protective effect on moderate outcomes since those individuals are progressing past moderate status with a higher probability. Regardless of disease severity, the TNF- α (-308) variant showed an increased risk for both moderate and severe disease (adjusted odds ratios of 3.6 and 1.6, respectively). The distribution of the minor variant did not show a consistent relationship with disease since the association was confounded by occupational exposure [55].

The proportion of the IL-1RA (+2018) allele 2 genotype was increased in miners with silicosis (0.27) compared to controls (0.16) [56]. This minor variant was significantly increased in miners with both moderate and severe silicosis suggesting that this variant affects susceptibility to silicosis rather than severity. Although there was no association with the IL-1 β variant, an allelic association between IL-1RA and IL-1 α was found ($p = 0.04$) [55]. This may also represent a susceptibility factor for silicosis as the IL-1/IL-1RA ratio is important in the regulation of inflammatory processes [57].

Much more is known about the environmental causes of silicosis than about the genes influencing disease. It is theoretically possible that a gene might have no independent effect itself on silicosis occurrence but in combination with another gene or a specific environmental exposure confer an increased risk. In this respect, examination of two-way gene-gene interactions provides insight into the contribution of these SNPs and silicosis. After adjusting for exposure, while the IL-1RA and TNF- α (-308) interaction showed a strong independent association between each SNP and moderate disease, the presence of both variants led to much higher odds

Table 1. Age, smoking status and years of exposure by disease status

Population	Number of patient	Mean (range; S.D.)		
		Age	Years smoking	Years exposure
Controls	164	63.2 (50–87; 8.0)	20.4 (0–50; 16.4)	21.3 (1–58; 13.3)
Moderate	140	66.9 (27–87; 9.2)	20.5 (0–70; 19.1)	34.4 (10–52; 10.1)
Severe	185	68.7 (39–93; 8.8)	17.9 (0–60; 18.4)	34.2 (1–55; 11.3)
Overall	489	66.3 (27–93; 9.0)	19.5 (0–70; 18.0)	29.9 (1–58; 13.2)

Adapted from ref. [55].

Table 2. Distribution of genotypes and allele frequencies

Disease status	Normal: 1/1 alleles	Carrier: 1/2 or 2/2	Allele 2 frequency	AdjustedOR (CI)**
TNF- α (-308) ^a				
Controls	75	79	0.27	1.00
Moderate	40	97	0.37	3.59 (2.0–6.4)
Severe	83	74	0.24	1.61 (0.9–2.8)
All Silicotic*	123	171	0.30	2.25 (1.4–3.6)
TNF- α (-238) ^a				
Controls	87	73	0.24	1.00
Moderate	91	41	0.16	0.52 (0.3–0.9)
Severe	42	141	0.40	4.00 (2.4–6.8)
All Silicotic	133	182	0.30	1.59 (1.0–2.5)
IL-1RA (+2018) ^a				
Controls	113	44	0.16	1.00
Moderate 54	60	0.35	2.54 (1.4–4.5)	
Severe 95	65	0.22	2.01 (1.2–3.4)	
All Silicotic	149	125	0.27	2.15 (1.3–3.5)
IL-1 α (+4845)				
Controls	125	31	0.10	1.00
Moderate 111	21	0.08	0.47 (0.2–0.9)	
Severe 113	42	0.15	0.90 (0.5–1.6)	
All Silicotic	224	63	0.12	0.76 (0.4–1.3)
IL-1 β (+3953)				
Controls	43	95	0.36	1.00
Moderate 35	75	0.40	0.8 (0.5–1.6)	
Severe	55	88	0.36	0.72 (0.4–1.3)
All Silicotic	90	163	0.38	0.75 (0.4–1.2)

^aSignificantly associated with moderate, severe, and overall disease ($p < 0.05$). *Represents total population studied with silicosis. **Odds ratio (95% confidence limits) adjusted for exposure with logistic regression. Adapted from ref. [55].

for severe disease. Three-way interaction analysis between each gene-gene interaction and exposure led to only marginally significant associations. The general pattern demonstrated in each of these interactions is exemplified by the IL-1 α and TNF- α (-308) association ($p = 0.05$) [55]. The prevalence of silicosis increases with increasing exposure, except in the case where both minor variants are present. For the group in which subjects are an allele 2 carrier in both polymorphisms, there is little or no effect of increasing exposure and this group has the highest proportion of moderate and severe cases for those exposed less than 30 years.

In conclusion, polymorphisms in the genes for IL-1 and TNF- α show both independent and interrelated effects on susceptibility and severity of silicosis in underground miners. These results indicate that the risk of a person acquiring or developing an inflammatory disease is influenced not only by exposure levels, but also by genetic polymorphisms of the cytokine system. Future studies in this area and identification of functional polymorphisms for other candidate genes will allow for a better estimate of determining susceptible populations and will improve human risk assessment.

References

1. Wagner GR: Asbestosis and silicosis. *Lancet* 349: 1311–1315, 1997
2. NIOSH: NIOSH alert: Request for assistance in preventing silicosis and deaths from sandblasting. Cincinnati: US Department of Health and Human Services, Public Health Service, CDC, NIOSH, DHHS publication no. (NIOSH)92-102, 1992
3. Legal and Medical Resource Center for Silicosis Victims; www.silicosis.com; last modified January 23, 2002
4. Lim Y, Kim JH, Kim KA, Chang HS, Park YM, Ahn BY, Phee YG: Silica-induced apoptosis *in vitro* and *in vivo*. *Toxicol Lett* 108: 335–339, 1999
5. Castranova V, Vallyathan V: Silicosis and coal workers' pneumoconiosis. *Environ Health Perspect* 108: 675–684, 2000
6. Kelley J: Cytokines of the lung. *Am Rev Respir Dis* 141: 765–788, 1990
7. Gauldie J, Jordana M, Gerard Cox: Cytokines and pulmonary fibrosis. *Thorax* 48: 931–935, 1993
8. Vaillant P, Menard O, Vignaud JM, Martinet N, Martinet Y: The role of cytokines in human lung fibrosis. *Monaldi Arch Chest Dis* 51: 145–152, 1996
9. Ward PA, Hunninghake GW: Lung inflammation and fibrosis. *Am J Respir Crit Care Med* 157: 123–129, 1998
10. Struhar DJ, Harbeck RJ, Gegen N, Kawada H, Mason RJ: Increased expression of class II antigen of the major histocompatibility complex on alveolar macrophages and alveolar type II cells and interleukin-1

- (IL-1) secretion from alveolar macrophages in an animal model of silicosis. *Clin Exp Immunol* 77: 281–284, 1989
11. Driscoll KE, Lindenschmidt RC, Maurer JK, Higgins JM, Ridder G: Pulmonary response to silica or titanium-dioxide: Inflammatory cells, alveolar macrophage-derived cytokines, and histopathology. *Am J Respir Cell Mol Biol* 2: 381–390, 1990
 12. Mohr C, Gerns D, Graebner C, Hemenway DR, Leslie KO, Absher PM, Davis GS: Systemic macrophage stimulation in rats with silicosis: Enhanced release of tumour necrosis factor- α from alveolar and peritoneal macrophages. *Am J Respir Cell Mol Biol* 5: 395–402, 1991
 13. Davis GS, Pfeiffer LM, Hemenway DR: Persistent overexpression of interleukin-1 β and tumour necrosis factor- α in murine silicosis. *J Environ Pathol Toxicol Oncol* 17: 99–114, 1998
 14. Orfila C, Lepert JC, Gossart S, Frisach MF, Cambon C, Pipy B: Immunocytochemical characterization of lung macrophage surface phenotypes and expression of cytokines in acute experimental silicosis in mice. *Histochem J* 30: 857–67, 1998
 15. Piguet PF, Collart MA, Grau GE, Sappino A, Vassalli P: Requirement of tumor necrosis factor for development of silica-induced pulmonary fibrosis. *Nature* 344: 245–247, 1990
 16. Gossart S, Cambon C, Orfila C, Seguelas MH, Lepert JC: Reactive oxygen intermediates as regulators of TNF- α production in rat lung inflammation induced by silica. *J Immunol* 156: 1540–1548, 1996
 17. Schmidt JA, Oliver CN, Lepe-Zuniga JL, Green I, Gery I: Silica-stimulated monocytes release fibroblast proliferation factors identical to interleukin 1. A potential role for interleukin 1 in the pathogenesis of silicosis. *J Clin Invest* 73: 1462–1472, 1984
 18. Savici D, He B, Geist LJ, Monick MM, Hunninghake GW: Silica increases tumor necrosis factor (TNF) production, in part, by upregulating the TNF promoter. *Exp Lung Res* 20: 613–625, 1994
 19. Borm PJA, Palmen N, Engelen JJM, Buurman WA: Spontaneous and stimulated release of tumor necrosis factor- α (TNF) from blood monocytes of miners with coal workers pneumoconiosis. *Am Rev Respir Dis* 138: 1589–1594, 1988
 20. Schins RP, Borm PJ: Epidemiological evaluation of monocyte TNF- α as an exposure and effect marker in pneumoconiosis: A five year follow up study of coal workers. *Occup Environ Med* 52: 441–450, 1995
 21. Kim KA, Lim Y, Kim JH, Kim EK, Chang HS, Park YM, Ahn BY: Potential biomarker of coal workers' pneumoconiosis. *Eur Respir J* 8: 834–842, 1995
 22. Lasalle P, Gosset P, Aerts C, Fournier E, Lafitte JE, Degreffe JM, Wallaert B, Tonnel AB, Voisin C: Abnormal secretion of interleukin-1 and tumor necrosis factor- α by alveolar macrophages in coal workers' pneumoconiosis: Comparison between simple pneumoconiosis and progressive massive fibrosis. *Exp Lung Res* 16: 73–80, 1990
 23. Vanhee D, Gosset P, Boitelle A, Wallaert B, Tonnel AB: Cytokines and cytokine network in silicosis and coal workers' pneumoconiosis. *Eur Respir J* 8: 834–842, 1995
 24. Katsnelson BA, Polzik EV, Privalova LI: Some aspects of the problem of individual predisposition to silicosis. *Environ Health Perspect* 68: 175–185, 1986
 25. Kornman KS, Crane A, Wang H-Y, di Giovine FS, Newman MG, Pirk FW, Wilson TG, Higginbottom FL, Duff GW: The interleukin-1 genotype as a severity factor in adult periodontal disease. *J Clin Periodontol* 24: 72–77, 1997
 26. Pociot F, Molvig J, Wogensens L, Worsaae H, Nerup J: A TaqI polymorphism in the human interleukin-1 β (IL-1 β gene correlates with IL-1 β secretion *in vitro*. *Eur J Clin Invest* 22: 396–402, 1992
 27. Danis VA, Millington M, Hyland VJ, Grennan D: Cytokine production by normal monocytes: Inter-subject variation and relationship to an IL-1 receptor antagonist (IL-1Ra) gene polymorphism. *Clin Exp Immunol* 99: 303–310, 1995
 28. Perrey C, Pravica V, Sinnott PJ, Hutchinson IV: Genotyping for polymorphisms in interferon- γ , interleukin-10, transforming growth factor- β 1 and tumour necrosis factor- α genes: A technical report. *Trans Immunol* 6: 193–197, 1998
 29. Carrol MC, Katzman P, Alicot EM, Koller BH, Geraghty DE, Orr HT, Strominger JL, Spies T: Linkage map of the human major histocompatibility complex including the tumor necrosis factor genes. *Proc Natl Acad Sci USA* 84: 8535–8539, 1987
 30. Wilson AG, de Vries N, Pociot F, di Giovine FS, van der Potte LB, Duff GW: An allelic polymorphism within the human tumor necrosis factor alpha promoter region is strongly associated with HLA A1, B8, and D3 alleles. *J Exp Med* 177: 557–560, 1993
 31. D'Alfonso S, Richiardi PM: A polymorphic variation in a putative regulation box of the TNF- α promoter region. *Immunogenetics* 3: 150–154, 1994
 32. McGuire W, Hill AVS, Allsopp CEM, Greenwood BM, Kwiatkowski D: Variation in the TNF- α promoter region associated with susceptibility to cerebral malaria. *Nature* 371: 508–511, 1994
 33. Cabrera M, Shaw MA, Sharples C, Williams H, Castes M, Convit J, Blackwell JM: Polymorphism in tumor necrosis factor genes associated with mucocutaneous leishmaniasis. *J Exp Med* 182: 1259–1264, 1995
 34. McManus R, Wilson AG, Mansfield J, Weir DG, Duff GW, Kelleher D: TNF2, a polymorphism of the tumour necrosis- α gene promoter, is a component of the celiac disease major histocompatibility complex haplotype. *Eur J Immunol* 26: 2113–2118, 1996
 35. Huang S-L, Su C-H, Chang S-C: Tumor necrosis factor- α gene polymorphism in chronic bronchitis. *Am J Respir Crit Care Med* 156: 1436–1439, 1997
 36. Arias AI, Giles B, Eiermann TH, Sterry W, Pandey JP: Tumor necrosis factor- α gene polymorphism in psoriasis. *Exp Clin Immunogenet* 14: 118–122, 1997
 37. Sullivan KE, Wooten C, Schmeckpeper Goldman D, Petri MA: A promoter polymorphism of tumor necrosis factor- α associated with systemic lupus erythematosus in African-Americans. *Arthritis Rheum* 40: 2207–2211, 1997
 38. Zhai R, Jetten M, Schins RPF, Franssen H, Borm PJA: Polymorphisms in the promoter of the tumor necrosis factor- α gene in coal miners. *Am J Indust Med* 34: 318–324, 1998
 39. Pociot F, Briant L, Jongeneel CV, Molvig J, Worsaae H, Abbal M, Thomsen M, Nerup J, Cambon-Thomsen A: Association of tumor necrosis factor (TNF) and class II major histocompatibility complex alleles with the secretion of TNF- α and TNF- β by human mononuclear cells: A possible link to insulin-dependent diabetes mellitus. *Eur J Immunol* 23: 224–231, 1993
 40. Jacob CO, Fronck Z, Lewis GD, Koo M, Hansen JA, McDevitt HO: Heritable major histocompatibility complex classII-associated differences in production of tumor necrosis factor- α : Relevance to genetic predisposition to systemic lupus erythematosus. *Proc Natl Acad Sci USA* 87: 1233–1237, 1990
 41. Kroeger KM, Carville KS, Abraham LJ: The -308 tumor necrosis factor- α promoter polymorphism effects transcription. *Mol Immunol* 34: 391–399, 1997
 42. Roux-Lombard P: The interleukin-1 family. *Eur Cytokine Netw* 9: 565–576, 1998
 43. Blakemore AIF, Tarlow JK, Cork MJ, Gordon C, Emery P, Duff GW: Interleukin-1 receptor antagonist gene polymorphism as a disease severity factor in systemic lupus erythematosus. *Arth Rheum* 37: 1380–1385, 1994
 44. Mansfield JC, Holden H, Tarlow JK, di Giovine FS, McDowell TL, Wilson AG, Holdsworth CD, Duff GW: Novel genetic association between ulcerative colitis and the anti-inflammatory cytokine interleukin-1 receptor antagonist. *Gastroenterology* 106: 637–642, 1994

45. Clay FE, Cork MJ, Tarlow JK, Blakemore AIF, Harrington CI, Lewis F, Duff GW: Interleukin 1 receptor antagonist gene polymorphism association with lichen sclerosis. *Hum Genet* 94: 407–410, 1994
46. Tarlow JK, Clay FE, Cork MJ, Blakemore AIF, McDonagh AJG, Messenger AG, Duff GW: Severity of alopecia areata is associated with a polymorphism in the interleukin-1 receptor antagonist gene. *J Invest Dermatol* 103: 387–390, 1994
47. McDowell TL, Symons JA, Ploski R, Forre O, Duff GW: A genetic association between juvenile rheumatoid arthritis and a novel interleukin-1 α polymorphism. *Arthritis Rheum* 38: 221–228, 1995
48. Jouvenne P, Chaudhary A, Buchs N, di Giovine FS, Duff GW, Miossec P: Possible genetic association between interleukin-1 α gene polymorphism and the severity of chronic polyarthritis. *Eur Cytokine Netw* 10: 33–36, 1999
49. Di Giovine FS, Cork MJ, Crane A, Mee JB, Duff GW: Novel genetic association of an IL-1 β gene variation at +3953 with IL-1 β protein production and psoriasis. *Cytokine* 7: 606, 1995
50. Gualde N, De Leobardy J, Serizay B, Malinvaud O: HL-A and silicosis. *Am Rev Respir Dis* 116: 334–336, 1977
51. Koskinen H, Tiilikainen A, Nordman H: Increased prevalence of HLA-Aw19 and the phenogroup Aw19, B18 in advanced silicosis. *Chest* 83: 848–852, 1983
52. Honda K, Kimura A, Dong R-P, Tamai H, Nagato H, Nishimura Y, Sasazuki T: Immunogenetic analysis of silicosis in Japan. *Am J Respir Cell Mol Biol* 8: 106–111, 1993
53. Rihs HP, Lipps P, May-Taube K, Jager D, Schmidt EW, Hegemann JH, Baur X: Immunogenetic studies on HLA-DR in German coal miners with and without coal workers' pneumoconiosis. *Lung* 172: 347–354, 1994
54. Wordsworth P, Bell J: Polygenic susceptibility in rheumatoid arthritis. *Ann Rheum Dis* 50: 343–346, 1991
55. Yucesoy B, Vallyathan V, Landsittel DP, Sharp DS, Weston A, Burleson G, Simeonova PP, McKinstry M, Luster MI: Association of TNF- α and IL-1 gene polymorphisms with silicosis. *Toxicol Appl Pharmacol* 172: 75–82, 2001
56. Yucesoy B, Vallyathan V, Landsittel DP, Sharp DS, Matheson JM, Burleson F, Luster MI: Polymorphisms of the IL-1 gene complex in coal miners with silicosis. *Am J Ind Med* 39: 286–291, 2001
57. Casini-Raggi V, Kam L, Chong YJT, Fiocchi C, Pizarro TT, Cominelli F: Mucosal imbalance of IL-1 and IL-1 receptor antagonist in inflammatory bowel disease. A novel mechanism of chronic intestinal inflammation. *J Immunol* 154: 2434–2440, 1995