

Glutathione *S*-transferase μ polymorphism does not explain variation in nitroglycerin responsiveness

Objective: To determine whether the considerable interindividual variability in nitroglycerin-induced venodilation in humans is related to the polymorphic expression of the μ class of glutathione *S*-transferase (GST μ). Recently vascular glutathione *S*-transferase (EC 2.5.1.18) of the μ -class (GST μ), a polymorphic group of enzymes present in only about 60% of the population, have been identified and shown in vitro to possess high metabolic activity toward nitroglycerin. Their clinical relevance is unknown.

Design: Dose-response relationships to nitroglycerin were constructed in vivo measuring changes in compliance of dorsal hand veins in 26 healthy volunteers during local infusion of small amounts of nitroglycerin. Polymerase chain reaction was applied to detect the deoxyribonucleic acid sequence that codes GST μ in whole blood samples.

Results: The GST μ isozyme was present in 15 subjects (58%) and deficient in 11 subjects. Values for mean maximum venodilation ($E_{\max}$) and dose rates producing 50% of $E_{\max}$ (ED_{50}) were not significantly different between the groups with or without GST μ . The respective values were 98% and 103% dilation for $E_{\max}$ and 9 and 16 ng/min for ED_{50} . There was no gender difference in the venodilatory response to nitroglycerin.

Conclusions: Subjects lacking GST μ can clearly respond normally to nitroglycerin, and the large interindividual variability in nitroglycerin potency is not related to the expression of this polymorphic enzyme. Intersubject variability is therefore more likely to be the result of differences in the presence or activity of other vascular enzymes or in steps further distal in the venodilatory cascade. (CLIN PHARMACOL THER 1993;53:463-8.)

Walter E. Haefeli, MD,^a Nidhi Srivastava, Karl T. Kelsey, MD,
John K. Wiencke, PhD, Brian B. Hoffman, MD, and Terrence F. Blaschke, MD
Stanford, San Francisco, and Palo Alto, Calif., and Boston, Mass.

Large interindividual variability in the sensitivity of human blood vessels to the vasodilatory action of nitroglycerin has been reported after systemic adminis-

tration,¹ and substantial variability has also been found during local venous infusions of systemically ineffective doses.²⁻⁴ The vasodilatory effects of nitroglycerin are believed to be mediated through a nitric oxide-like compound liberated during enzymatic denitration in the vascular wall.^{5,6} In rabbits, nitroglycerin metabolism to its dinitrate metabolites is mandatory before vascular action; it is therefore considered to be a pro-drug.^{7,8} In humans, different enzymatic pathways have been reported to metabolize the nitrate,⁹⁻¹² but at the site of action, the vascular wall, the only enzymes with high activity toward nitroglycerin identified thus far belong to the glutathione *S*-transferase family (GST; EC 2.5.1.18).¹³ It is not known whether nitroglycerin metabolism by human glutathione *S*-transferases may produce active intermediates. Indirect evidence obtained in isolated animal vessels suggests that inhibitors of glutathione *S*-trans-

From the Division of Clinical Pharmacology, Department of Medicine, Stanford University Medical Center, Stanford; the Laboratory of Radiobiology, Department of Environmental Health, Harvard School of Public Health, Boston; the Department of Epidemiology and Biostatistics, School of Medicine, University of California, San Francisco; and Geriatric Research, Education, and Clinical Center, Veterans Affairs Medical Center, Palo Alto. Supported by grants AG-05627, ES-00001, and K01-OH00110-01 from the National Institutes of Health, Bethesda, Md.

Received for publication Aug. 6, 1992; accepted Dec. 8, 1992.

Reprint requests: Terrence F. Blaschke, MD, Division of Clinical Pharmacology, Stanford University Medical Center, Room S-169, Stanford, CA 94305-5113.

^aSupported by a grant from the Schweizerische Stiftung für Medizinisch-Biologische Stipendien, Switzerland.

13/1/44771

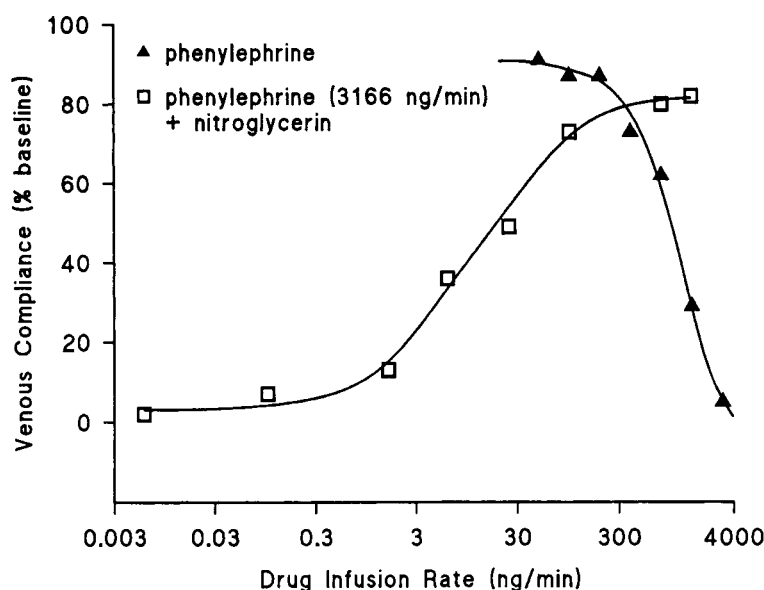


Fig. 1. Vasodilatory action of nitroglycerin in the hand vein of a healthy volunteer expressed as percentage of change from baseline. After constructing a cumulative dose-response curve to phenylephrine (*triangles*), the highest dose of phenylephrine (3166 ng/min) was infused while increasing concentrations of nitroglycerin were added (*squares*). Curves are nonlinear least-squares fits to a four-parameter logistic function²⁴; estimated dose rates exerting 50% of maximum venodilation for phenylephrine and nitroglycerin were 1474 ng/min and 13 ng/min, respectively.

ferase may decrease nitroglycerin-induced vasodilation¹⁴ and that the extent of inhibition is inversely correlated with the amount of dinitrate metabolites formed,¹⁵ which may indicate that glutathione *S*-transferases are involved not only in catabolism of nitroglycerin but also in formation of active metabolites. However, Chung et al.,¹⁶ using microsomes obtained from bovine coronary artery smooth muscle cells, found that nitric oxide production was not primarily mediated by glutathione *S*-transferase. Schröder,¹⁷ using cultured rat lung fibroblasts, observed that cyclic guanosine monophosphate (cGMP) stimulation was not affected by sulfobromophthalein, an inhibitor of glutathione *S*-transferase. In human aorta, two different classes of glutathione *S*-transferase (GST μ and GST α), both of which metabolize nitroglycerin, have been recently isolated; isozymes of the μ class (GST μ) have higher metabolic activity toward nitroglycerin. Because of a gene deletion the expression of some isoforms of GST μ in humans is polymorphic.¹⁸ In about 60% of white persons and about the same fraction of most other races, an intact deoxyribonucleic acid (DNA) sequence encoding this protein is present, and these subjects express GST μ in leukocytes,¹⁹⁻²² liver samples,^{23,24} and other tissues.²⁵ Because of a homozygous deletion of the gene in the re-

maining 40% of the population, no detectable amounts of the isozyme are expressed.¹⁸ Smoking-induced lung cancers²⁴ and chromosomal damage induced by mutagenic drugs²¹ are prone to develop in these GST μ -deficient subjects. The presence or absence of the GST μ gene can be determined by the polymerase chain reaction with primers designed to detect the intact DNA sequence for human GST μ .²⁶ Previous studies have shown that there is complete agreement between the presence or absence of the intact DNA and the metabolic activity of GST μ as measured by the rate of trans-stilbene-oxide metabolism (Wiencke JK, Kelsey KT. Unpublished data).

This study was designed to determine whether part of the considerable interindividual variability in sensitivity of human veins to nitroglycerin is genetically linked to the presence of the GST μ isozyme and whether people lacking the enzyme would have an impaired venodilatory response to nitroglycerin.

MATERIAL AND METHODS

Subjects. Twenty-six healthy volunteers (nine women and 17 men) with a mean age of 27 years (age range, 19 to 53 years) and a mean weight of 73 kg (weight range, 51 to 113 kg) entered the study after giving written informed consent. No subjects were

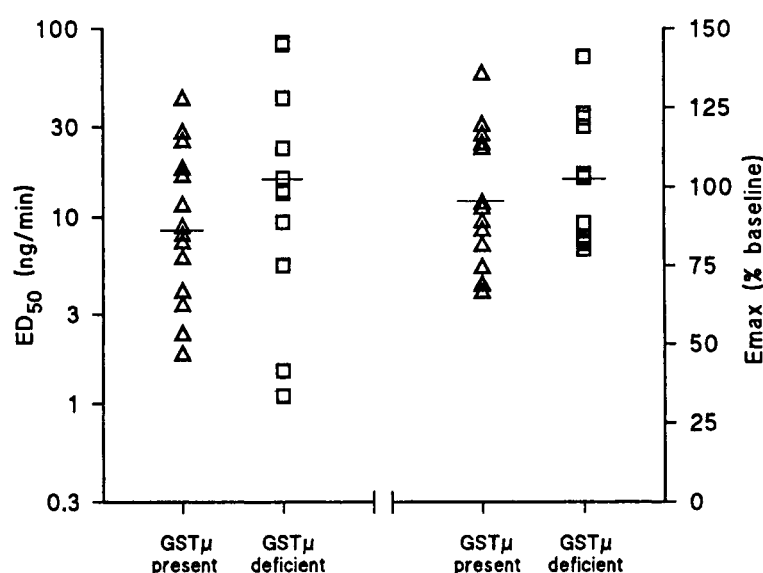


Fig. 2. Dose-response relationship of nitroglycerin in the presence ($n = 15$; triangles) and absence ($n = 11$; squares) of glutathione *S*-transferase (GST μ). Estimates of maximum venodilation ($E_{\max}$) and dose rates exerting 50% of $E_{\max}$ (ED_{50}) were obtained by nonlinear least-squares regression to a four-parameter logistic function.²⁴ Horizontal lines indicate the mean of the respective data set.

Table I. Dose-response relationship of nitroglycerin in healthy volunteers with and without GST μ

	ED_{50} (ng/min)	$\log ED_{50}$ (ng/min)	$E_{\max}$ (% baseline)
GST present ($n = 15$)	8.5	0.93 ± 0.10	95 ± 5
GST deficient ($n = 11$)	16.0	1.20 ± 0.16	103 ± 6
Males ($n = 17$)	11.6	1.06 ± 0.13	98 ± 5
Females ($n = 9$)	7.9	0.90 ± 0.15	88 ± 12

ED_{50} values are expressed as geometric means of individual dose-response curves. Log transformed data of ED_{50} ($\log ED_{50}$) and $E_{\max}$ data are expressed as arithmetic means $\pm$ SEM of individual dose-response curves.

GST, Glutathione *S*-transferase; GST μ , μ class of GST; $E_{\max}$, maximum venodilation; ED_{50} , dose rate exerting 50% of $E_{\max}$.

taking any medications other than oral contraceptives (three subjects), and none had a history of significant medical disease. All had normal physical and electrocardiographic examinations and normal laboratory values (SMA-20, complete blood cell count and urinalysis). The protocol was reviewed and approved by the Medical Committee for the Protection of Human Subjects in Research at Stanford University (Stanford, Calif.).

Volunteers were admitted to the Aging Studies Unit at the Geriatric Research, Education, and Clinical Center at the Veterans Affairs Medical Center, Palo Alto, Calif. They were allowed to eat a light breakfast and were asked to refrain from methylxanthines and alcohol for at least 12 hours before the study. They remained semirecumbent in a quiet room with a constant temperature of $21^\circ \pm 1^\circ$ C throughout the study.

Hand vein compliance technique. Hand vein experiments were performed by use of a general approach and instrumentation similar to that reported by Aellig,²⁷ with modifications as described previously.^{3,4} In brief, through a 23-gauge needle a continuous infusion of physiologic saline solution was delivered into a vein on the dorsal surface of the hand (Harvard Apparatus infusion pump, Harvard Apparatus Inc., South Natick, Mass.). The arm was placed on a padded support above the level of the heart and sloping upward at an angle of 30 degrees from the horizontal to allow for complete emptying of the vein. Changes in the diameter of the vein were recorded with the use of a linear variable differential transformer (Schaevitz type 100 MHR, Schaevitz, Inc., Pennsauken, N.J.) with a freely movable core resting over the summit of the vein under investigation.

Transformer signals were amplified by a Schaevitz signal conditioner (Schaevitz ATA 101) and recorded on a strip-chart recorder (LKB 2210 recorder, LKB Produkter AB, Bromma, Sweden). The difference between the position of the core before and after inflation of a sphygmomanometer cuff on the same arm to 40 mm Hg gives a measure of the diameter changes under a given congestion pressure. For each measurement the blood pressure cuff was inflated and pressure was maintained at 40 mm Hg for approximately 3 minutes, until a stable deflection was observed. The cuff was then deflated to allow continued perfusion of the vein under investigation. After establishing a stable initial baseline with normal saline solution, which was defined as 100% dilation, the vein was constricted with the α_1 -adrenergic agonist phenylephrine to about 20% of its initial diameter; this position of the core was considered to represent 0% dilation. Increasing concentrations of nitroglycerin were then infused locally into the vein together with a constant dose of phenylephrine. Each concentration was administered for at least 6 minutes to permit enough time to reach a stable response.²⁸ Dose rates administered (0.006 ng/min to 1.5 μ g/min) were not sufficiently high to result in any systemic effect as evidenced by the lack of changes in blood pressure and heart rate during frequent measurements with an automated blood pressure monitor (Dinamap, Critikon Inc., Tampa, Fla.) placed on the opposite arm. At the completion of each study a heparinized whole blood sample was frozen for analysis of the GST μ genotype. No untoward effects were observed in any of the subjects.

Polymerase chain reaction for GST μ allele. The GST μ genotype was assessed in white blood cells of the volunteers with polymerase chain reaction as described earlier.²⁶ Primers for actin were used as a control. The determination was done after completion of all hand vein experiments.

Materials. Nitroglycerin (Tridil) was purchased from Du Pont Critical Care (Wilmington, Del.) and phenylephrine (Neo-Synephrine) from Winthrop Pharmaceuticals (New York, N.Y.). To avoid an interaction between the nitrates and the plastic infusion set, all studies were done with use of low absorption non-PVC tubing (Accuset, Imed Corporation, San Diego, Calif.) and syringes (Monoject, Sherwood Medical, St. Louis, Mo.). All intravenous solutions were freshly prepared and used within 3 hours.

Curve fitting and statistical analysis. Data are expressed as mean values $\pm$ SE unless otherwise indicated. Individual dose-response curves, expressed as percentage change from baseline, were fitted to a four-

parameter logistic function, providing estimates of the maximum venodilation ($E_{\max}$) and dose rates exerting 50% of $E_{\max}$ (ED_{50}) (Allfit,²⁹ National Institutes of Health, Bethesda, Md.).

Mean values of the effect parameters for the two groups were compared by use of the unpaired Student *t* test. The relationship between the genetic trait and variability in $E_{\max}$ and log ED_{50} of the two groups was further investigated by linear regression analysis. Confidence intervals of r^2 values were calculated using the Bootstrap method.³⁰ A *p* value of less than 0.05 was considered to indicate statistical significance.

RESULTS

The GST μ enzyme was present in 15 subjects (58%) and deficient in 11 subjects, an enzyme distribution similar to distributions reported earlier.^{20-25,31} As an example, the dose-response relationships of phenylephrine and nitroglycerin in a vein precontracted with phenylephrine in one volunteer are shown in Fig. 1. $E_{\max}$ and log ED_{50} values of nitroglycerin of the two groups are depicted in Fig. 2. As we had seen previously,^{3,4} nitroglycerin potency, expressed as ED_{50} values, varied substantially over a range of almost two orders of magnitude (1.1 to 85 ng/min). There were no significant differences in either pharmacodynamic parameter between the two groups (Table I). Evaluation of the findings by linear regression revealed no correlation between the sensitivity of the subjects to nitroglycerin (ED_{50}) and the GST μ genotype ($r^2 = 0.034$; $p = 0.37$; 95% confidence interval, 0 to 0.26). In accordance with findings in arterial beds³² no gender-related differences in the venodilatory response to nitroglycerin were observed (Table I).

DISCUSSION

Venodilation in response to nitroglycerin is the result of a series of local biochemical events in the vascular wall.^{8,33} After entering the vascular smooth muscle cell, the drug is substrate of several GST isozymes¹³ that metabolize nitroglycerin in the presence of sulfhydryl-containing cofactors³⁴ such as glutathione³⁵ and that mainly belong to the μ class.¹³ The nitrate groups are believed to be the vasoactive components in the nitrate molecule^{8,36} and their enzymatic cleavage from the parent compound is considered a key step in nitroglycerin-induced vasodilation.⁸ After liberation, the nitrate groups exert dilation possibly through formation of an active nitrosothiol intermediate,³³ which activates soluble guanylate cyclase and

subsequently increases in intracellular levels of the second messenger cyclic GMP, a second messenger that is thought to cause vascular relaxation by way of activation of cyclic GMP-dependent protein kinase.^{6,33} Consequently, intersubject variation in any of numerous biochemical steps may lead to interindividual differences in the potency of nitroglycerin in human veins.

This study was designed to evaluate a possible influence of the hereditary GST μ polymorphism on the venous response to nitroglycerin and to relate genetic differences at the very beginning of the vasodilatory cascade to drug action. The findings of this study revealed high potencies of the drug in subjects with and without GST μ deficiency, indicating that the GST μ polymorphism did not determine the subjects' ability to activate nitroglycerin (i.e., subjects without the isozyme maintained their vascular response to nitroglycerin). Consequently, the large interindividual variability in nitroglycerin potency is not caused by the presence or deficiency of this polymorphic enzyme. Moreover, the normal response to nitroglycerin in subjects who totally lack GST μ activity indicates that the polymorphic GST μ plays a minor role in bioactivation of nitroglycerin in human veins and that this polymorphism is not the cornerstone of interindividual variability in nitroglycerin-induced venodilation. The absence of a correlation between GST μ and drug potency could indicate that GST μ is not the rate-limiting step or that other enzymes participate in nitroglycerin bioactivation in humans. Indeed, metabolism of nitroglycerin by enzymes other than GST μ has been reported in human^{37,38} and animal tissues^{16,17} and at least one additional glutathione *S*-transferase isozyme, classified as α -type, with activity against nitroglycerin has been found in human aorta.¹³ Moreover, at least three further pathways metabolizing nitroglycerin have been recently described that involve plasma proteins, hemoglobin, or cytochrome P450 isozymes with nitrate-metabolizing capacity,^{9-12,17} but their involvement in vascular action of nitroglycerin has not been shown thus far. In the case of cytochrome P450 enzymes, animal data suggest that they do not participate significantly in the bioactivation of organic nitrates.^{33,39}

In summary, this study shows that the polymorphic GST μ isozyme is not the cornerstone of interindividual variability in nitroglycerin-induced venodilation and that enzymes other than GST μ must be involved in vascular bioactivation of nitroglycerin in humans. Intersubject variations in nitroglycerin sensitivity are therefore more likely to be the result of differences in

the activity or presence of other enzymes, the availability of cofactors such as sulfhydryl groups, or variation in distal steps in the venodilatory cascade.

We thank Dr. C. Mark Smith for technical assistance and Dr. Ronald Thomas for his statistical advice.

References

1. Armstrong PW, Armstrong JA, Marks GS. Pharmacokinetic-hemodynamic studies of intravenous nitroglycerin in congestive cardiac failure. *Circulation* 1980;62:160-6.
2. Collier JG, Lorge RE, Robinson BF. Comparison of effects of tolmesoxide (RX71107), diazoxide, hydralazine, prazosin, glyceryl trinitrate and sodium nitroprusside on forearm arteries and dorsal hand veins of man. *Br J Clin Pharmacol* 1978;5:35-44.
3. Eichler HG, Hiremath A, Katzir D, Blaschke TF, Hoffman BB. Absence of age-related changes in venous responsiveness to nitroglycerin in vivo in humans. *CLIN PHARMACOL THER* 1987;42:521-4.
4. Haefeli WE, Gumbleton M, Benet LZ, Hoffman BB, Blaschke TF. Comparison of vasodilatory responses to nitroglycerin and its dinitrate metabolites in human veins. *CLIN PHARMACOL THER* 1992;52:590-6.
5. Feelisch M, Noack EA. Correlation between nitric oxide formation during degradation of organic nitrates and activation of guanylate cyclase. *Eur J Pharmacol* 1987;139:19-30.
6. Feelisch M, Kelm M. Biotransformation of organic nitrates to nitric oxide by vascular smooth muscle and endothelial cells. *Biochem Biophys Res Commun* 1991;180:286-93.
7. Brien JF, McLaughlin BE, Breedon TH, Bennett BM, Nakatsu K, Marks GS. Biotransformation of glyceryl trinitrate occurs concurrently with relaxation of rabbit aorta. *J Pharmacol Exp Ther* 1986;237:608-14.
8. Brien JF, McLaughlin BE, Kobus SM, Kawamoto JH, Nakatsu K, Marks GS. Mechanism of glyceryl trinitrate-induced vasodilation. I. Relationship between drug biotransformation, tissue cyclic GMP elevation and relaxation of rabbit aorta. *J Pharmacol Exp Ther* 1988;244:322-7.
9. McDonald BJ, Bennett BM. Cytochrome P-450 mediated biotransformation of organic nitrates. *Can J Physiol Pharmacol* 1990;68:1552-7.
10. Servent D, Delaforge M, Ducrocq C, Mansuy D, Lenfant M. Nitric oxide formation during microsomal hepatic denitration of glyceryl trinitrate: involvement of cytochrome P-450. *Biochem Biophys Res Commun* 1989;163:1210-6.
11. Bennett BM, Kobus SM, Brien JF, Nakatsu K, Marks GS. Requirement for reduced, unliganded hemoprotein for the hemoglobin- and myoglobin-mediated biotransformation of glyceryl trinitrate. *J Pharmacol Exp Ther* 1986;237:629-35.

12. Chong S, Fung HL. Thiol-mediated catalysis of nitroglycerin degradation by serum proteins. Increase in metabolism was not accompanied by S-nitrosothiol production. *Drug Metab Dispos* 1990;18:61-7.
13. Tsuchida S, Maki T, Sato K. Purification and characterization of glutathione transferases with an activity toward nitroglycerin from human aorta and heart. *J Biol Chem* 1990;265:7150-7.
14. Yeates RA, Schmid M, Leitold M. Antagonism of glycerol trinitrate activity by an inhibitor of glutathione S-transferase. *Biochem Pharmacol* 1989;38:1749-53.
15. Lau DTW, Benet LZ. Effects of sulfobromophthalein and ethacrynic acid on glycyl trinitrate relaxation. *Biochem Pharmacol* 1992;43:2247-54.
16. Chung SJ, Chong S, Seth P, Jung CY, Fung HL. Conversion of nitroglycerin to nitric oxide in microsomes of the bovine coronary artery smooth muscle is not primarily mediated by glutathione S-transferases. *J Pharmacol Exp Ther* 1992;260:652-9.
17. Schröder H. Cytochrome P-450 mediates bioactivation of organic nitrates. *J Pharmacol Exp Ther* 1992;262:298-302.
18. Seidegård J, Voracek WR, Pero RW, Pearson WR. Hereditary differences in the expression of the human glutathione transferase active on trans-stilbene oxide are due to a gene deletion. *Proc Natl Acad Sci USA* 1988;85:7293-7.
19. Seidegård J, Pero RW, Miller DG, Beattie EJ. A glutathione transferase in human leukocytes as a marker for the susceptibility to lung cancer. *Carcinogenesis* 1986;7:751-3.
20. Seidegård J, Pero RW. The hereditary transmission of high glutathione transferase activity towards trans-stilbene oxide in human mononuclear leukocytes. *Hum Genet* 1985;69:66-8.
21. Wiencke JK, Kelsey KT, Lamela RA, Toscano WA. Human glutathione S-transferase deficiency as a marker of susceptibility to epoxide-induced cytogenetic damage. *Cancer Res* 1990;50:1585-90.
22. Zhong S, Howie AF, Ketterer B, et al. Glutathione S-transferase mu locus: use of genotyping and phenotyping assays to assess association with lung cancer susceptibility. *Carcinogenesis* 1991;12:1533-7.
23. Harada S, Abei M, Tanaka N, Agarwal DP, Goedde HW. Liver glutathione S-transferase polymorphism in Japanese and its pharmacogenetic importance. *Hum Genet* 1987;75:322-5.
24. Board P, Coggan M, Johnston P, Ross V, Suzuki T, Webb G. Genetic heterogeneity of the human glutathione transferases: a complex of gene families. *Pharmacol Ther* 1990;48:357-69.
25. Strange RC, Faulder CG, Davis BA, et al. The human glutathione S-transferases: studies on the tissue distribution and genetic variation of the GST1, GST2 and GST3 isozymes. *Ann Hum Genet* 1984;48:11-20.
26. Comstock KE, Sanderson BJS, Claflin G, Henner WD. GST1 gene deletion determined by polymerase chain reaction. *Nucleic Acid Res* 1990;18:3670.
27. Aellig WH. A new technique for recording compliance of human hand veins. *Br J Clin Pharmacol* 1981;11:237-43.
28. Luthra A, Borkowski KR, Carruthers SG. Genetic aspects of variability in superficial vein responsiveness to norepinephrine. *CLIN PHARMACOL THER* 1991;49:355-61.
29. De Lean A, Munson PJ, Rodbard D. Simultaneous analysis of families of sigmoidal curves: application to bioassay, radioligand assay, and physiological dose-response curves. *Am J Physiol* 1978;235:E97-102.
30. Efron B. Nonparametric standard errors and confidence intervals. *Can J Statistics* 1981;9:139-72.
31. Suzuki T, Coggan M, Shaw DC, Board PG. Electrophoretic and immunological analysis of human glutathione S-transferase isozymes. *Ann Hum Genet* 1987;51:95-106.
32. Freedman RR, Sabharwal SC, Desai N. Sex differences in peripheral vascular adrenergic receptors. *Circulation Res* 1987;61:581-5.
33. Ahlner J, Andersson RGG, Torfgård K, Axelsson KL. Organic nitrate esters: clinical use and mechanisms of actions. *Pharmacol Rev* 1991;43:351-423.
34. Needleman P, Jakschik B, Johnson EM. Sulfhydryl requirement for relaxation of vascular smooth muscle. *J Pharmacol Exp Ther* 1973;187:324-31.
35. Habig WH, Keen JH, Jakoby WB. Glutathione S-transferase in the formation of cyanide from organic thiocyanates and as an organic nitrate reductase. *Biochem Biophys Res Commun* 1976;64:501-6.
36. Ignarro LJ, Lippman H, Edwards JC, et al. Mechanism of vascular smooth muscle relaxation by organic nitrates, nitrites, nitroprusside and nitric oxide: evidence for the involvement of S-nitrosothiols as active intermediates. *J Pharmacol Exp Ther* 1981;218:739-49.
37. Kamisaka K, Habig WH, Ketley JN, Arias IM, Jakoby WB. Multiple forms of human glutathione S-transferase and their affinity for bilirubin. *Eur J Biochem* 1975;60:153-61.
38. Keen JH, Habig WH, Jakoby WB. Mechanism for the several activities of the glutathione S-transferases. *J Biol Chem* 1976;251:6183-8.
39. Bornfeldt KE, Axelsson KL. Studies on the effect of different inhibitors of arachidonic acid metabolism on glycyl trinitrate-induced relaxation and cGMP elevation in bovine vascular tissue. *Pharmacol Toxicol* 1987;60:110-6.