

Tumor Necrosis Factor α and Toxicology

Michael I. Luster,* Petia P. Simeonova, Randle Gallucci, and Joanna Matheson

Toxicology and Molecular Biology Branch, Health Effects Laboratory Division, National Institute for Occupational Safety and Health, 1095 Willowdale Road, Morgantown, WV 26505

* Corresponding author: Michael I. Luster, Ph.D., P: (304) 285–6060, F: (304) 285–5708, E-mail: myl6@cdc.gov

ABSTRACT: The molecular cloning of a group of proteins, collectively referred to as cytokines, and including interleukins, chemokines, growth factors, colony stimulating factors, and tumor necrosis factors, has allowed for the increased understanding of the mechanisms for many disease processes as well as provided strategies for the development of novel therapies. Conceptually similar to hormones and peptides, this group of phylogenetically related molecules are also involved in various toxicological processes, including apoptosis, cell repair, and in particular inflammation. In this review, we offer a description of what many believe represents the primary regulatory cytokine, tumor necrosis factor (TNF) α and its role in toxicological processes. For over a decade it has been suspected that this molecule helps mediate the shock state induced by bacterial endotoxin and the wasting diathesis that typifies chronic diseases. Advances in molecular biology that have provided tools to modulate TNF α regulation and synthesis have allowed for the identification of additional roles for TNF α in homeostasis, cellular damage, and repair. This review provides a brief summary of our understanding of TNF α biology followed by a discussion of its role in toxicological responses. This is followed by specific examples of organ-specific and tissue-specific responses to chemical damage where TNF α has been implicated. The review concludes with a review of its implication in human risk assessment, particularly as it relates to genetic polymorphisms of TNF α expression and disease susceptibility.

KEY WORDS: tumor necrosis factor and toxicity, proinflammatory cytokines and chemical damage.

I. INTRODUCTION

Tumor necrosis factor (TNF) α , a 17-kDa polypeptide, was first identified as causing a wasting syndrome in tumor-bearing mice, hemorrhage reduction in the size of some tumors, and necrosis in normal tissues.^{1–3} Although the cytokine is initially synthesized in a 31-kDa precursor form, the N-terminal sequence is removed prior to secretion in a 17-kDa form.⁴ The protein sequence is well conserved as human TNF α shows 80% homology with mouse or rabbit. A va-

riety of cell types produce TNF α . In the pathological processes, tissue fixed macrophages, such as Langerhan's cells, Kupffer cells, and astroglia, are believed to be the major sources of TNF α .⁶ However, other cell types, including endothelial cells, epithelial cells, and fibroblasts, secrete significant amounts when treated with appropriate stimuli. Endotoxin is the major stimulator for TNF α but other agents, including certain cytokines themselves, are also effective. Mitochondrial radical production and a variety of enzymes, including phospholipases,

S mases, and protein kinases, can also participate in the cellular activation of TNF α .⁵ As a number of toxic chemicals possess prooxidant activities and modulate cellular phospholipases, it is conceivable that some toxic agents, such as ultraviolet radiation, may act directly to stimulate TNF α activity. At the transcriptional level, TNF α is regulated by a TATA box but promoted through a series of regulatory sequences, including binding sites for nuclear factor-kappa β (NF- κ B), nuclear factor of activated T cells (NFAT), cAMP-responsive element (CRE), activation transcription factor-2 (ATF-2)/Jun, two SV40 promoter-1 (SP-1), activating protein-1 (AP-1) and AP-2.

Like other cytokines, TNF α confers its signals to target cells through binding to specific cell surface membrane receptors. These receptors are found on almost all nucleated cells. Two distinct receptors mediate the biological activities of TNF α , one of a molecular mass of 55 kDa (p55, R1) and the other of 75 kDa (p75, R2). The p55 receptor is constitutively expressed and historically has been considered the main mediator for TNF α responses (Figure 1). The p75 receptor is inducible and provides for ligand passing and induction of soluble TNF receptor molecules as well as limited responses. In addition, two distinct families of proteins, the TNF receptor-associated factors (TRAFs) family of adaptor proteins and the death domain homologs, help coordinate cell- and tissue-specific responses in this complex pathway by interacting with the intracellular unit of the TNF receptors and initiating cell-signaling pathways.⁷ For noncytotoxic processes, TNF α serves primarily to induce the synthesis of biologically active proteins by causing changes in gene expression. This occurs through a complex signaling pathway referred to as mitogen-activated protein (MAP) kinase/extracellular signal-regulated kinase (ERK), collectively referred to as MAPK. Phosphorylation of these kinases results in activation of nuclear transcription

factors, arguably, the most important being NF- κ B.⁸ NF- κ B is a dimer of the Rel family of proteins that is activated following dissociation of an inhibitor protein belonging to the I κ B family. Once dissociated, an NF- κ B dimer can translocate to the nucleus and alter the expression of genes such as IL-6.⁹

TNF α was originally characterized as a protein inducing necrosis of methylcholanthrene-induced sarcomas. Its cytostatic and cytotoxic activities are associated primarily with certain transformed cell lines, tumor cells, and, to a lesser extent, normal cells. Depending on the target cell, TNF α can induce necrotic or apoptotic cell death.¹⁰ Cytotoxicity by TNF α , unlike its other functions, can occur independently of *de novo* gene transcription and translation and involves mitochondrial production of oxygen radicals generated primarily at the semiquinone site. This requires ceramide, a sphingolipid generated in cells following stimulation with TNF α , which generates H₂O₂ from the mitochondrial electron transport chain. The major function attributed to TNF α is not cytotoxicity, but rather synthesis of gene products that influence the cell as in differentiation, growth stimulation, antiviral activity, immunomodulation, and inflammation.^{5,11} For example, TNF α has been shown to be mitogenic for a number of normal cell types such as fibroblasts, hepatocytes, smooth muscle cells, and lymphocytes. It also increases activities of natural killer (NK) cells, endothelial cells, macrophages/monocytes, neutrophils, and lymphocytes. TNF α stimulates the production of several hormones, including ACTH and thyroid-stimulating hormones.¹² In muscle cells, TNF α stimulates glycolysis and glycogenesis,¹³ whereas in synovial fluid it stimulates the synthesis of PGD₂, plasminogen activator, collagenase, and hyaluronic acid.¹⁴ Because of the influence chronic inflammation plays in disease processes, recent attention in toxicology has focused on its role in regulating inflammatory processes.¹⁻³ This occurs through the

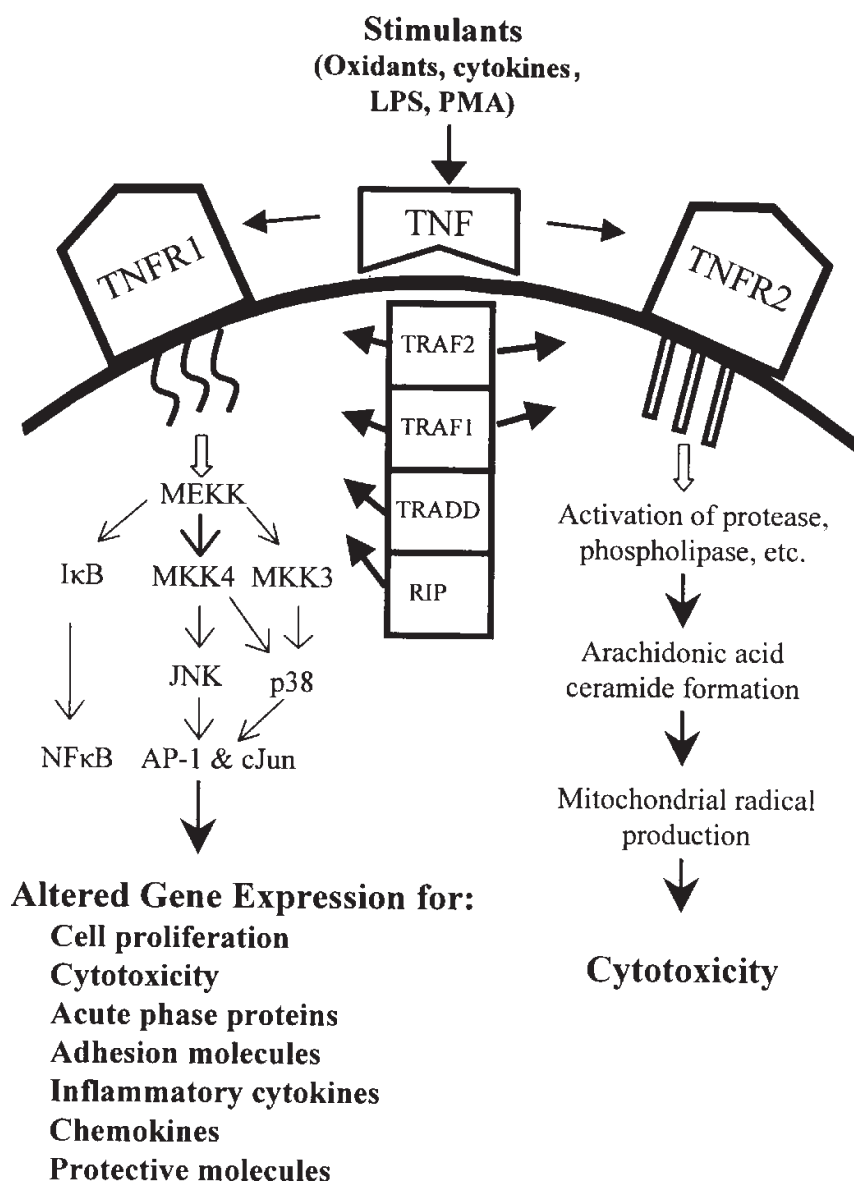


FIGURE 1. Model for TNF α action. TNF α , which is induced by a variety of stimulants, can bind to either the TNFR1 (p55) or TNFR2 (p75) receptor. Although not yet fully established, TNFR1 is the major inducer of biological effects, whereas TNFR2 appears to be confined mainly to cells of the immune system and is responsible for cytotoxic effects. TNF cell signaling, which is highly regulated and complex, occurs via the MEK-MAP kinase kinase (MEKK) pathway, resulting in the activation of the NF- κ B and AP-1 transcription factors. These transcription factors regulate genes involved in cell proliferation, apoptosis, and inflammation. In addition, there is a ceramide-related pathway that leads to cytotoxicity independent of gene transcription. Regulation is also provided through a family of adaptor proteins and death domain homologs (TRAFs) that interact with TNF receptors to produce a cell signal (other abbreviations in text).

ability of TNF α to provide cell signals and regulate genes that code for inflammatory mediators, such as IL-1, IL-6, IL-8, macrophage inflammatory protein (MIP)-2,

granulocyte macrophage-colony stimulating factor (GM-CSF), intracellular adhesion molecule (ICAM)-1, and endothelial leukocyte adhesion molecule (ECAM)-1. In con-

cert, these mediators locally enhance vascular permeability, stimulate the expression of adhesion molecules on endothelial cells, and serve as leukocyte activators and chemoattractants (i.e., chemokines). Most genes that code for inflammatory mediators, including acute phase proteins, contain binding sites for the NF- κ B and, to a lesser extent, AP-1 transcription factors in the cis-acting elements of the promoter region that helps regulate their expression.

II. ROLE IN TOXICOLOGICAL PROCESSES

The role of TNF α in chronic diseases is viewed by many immunologists and toxicologists with considerable interest as more diseases, ranging from arthritis and Alzheimer's to idiopathic pulmonary fibrosis and chronic hepatitis, manifest an inflammatory component that exacerbates, albeit to varying degrees, disease severity.^{5,11} The general hypothesis that links these pathologies is summarized in Figure 2 and is as follows: initial injury, whether initiated by an infectious, chemical, or environmental agent, produces focal tissue necrosis in the target organ through any one of several toxic mechanisms (e.g., lipid peroxidation, mitochondrial damage). As a result of this damage, tissue-fixed macrophages along with adjacent endothelial and epithelial cells become activated and secrete products that cause additional cell damage or induction of inflammatory products. Some of these products are short-lived, such as reactive oxygen species (ROS) and the nitrogen-centered radical, nitric oxide (NO \cdot).¹⁵ Other products, such as arachidonic acid and proinflammatory cytokines, regulate the production of additional inflammatory mediators and, thus, amplify as well as propagate these responses. The recruitment and activation of neutrophils and circulating monocytes into the site are believed to be

important in the toxicological processes, as these cells cause additional damage via degranulation and release of neutral proteinase or the generation of superoxide anions (O₂⁻) via NADPH-oxidase during the respiratory burst. O₂⁻, which itself mediates tissue damage, can be reduced further to other cell-damaging reactive oxygen intermediates (ROIs), including H₂O₂ and hydroxyl radical (OH \cdot).⁶ The ultimate toxicity of O₂⁻ may depend on its ability to bring about the reduction of Fe⁺³ to Fe⁺², providing for the generation of OH \cdot from H₂O₂, and to interact with NO \cdot to generate peroxyxynitrite (ONOO \cdot) and subsequently OH \cdot . The overall effect of this complex series of events is a chronic inflammatory response resulting in tissue damage that may either lead to fibrotic changes or recovery.¹⁶ TNF α is also associated with the activation of repair mechanisms following xenobiotic damage. Although these particular mechanisms have not been well defined, they appear to involve the ability of TNF α to control the replication of various cell types, such as fibroblasts and hepatocytes, by acting as a mitogen as well as stimulating apoptosis.^{5,11}

Support that TNF α can mediate organ-specific toxic or repair responses stems from several observations: (1) elevated levels of TNF α can be found in target organs following exposure to certain occupational and environmental agents; (2) administration of TNF α in experimental animals mimics many of the pathophysiological responses associated with the toxic response; (3) organs in which these types of responses occur contain resident (tissue-fixed) macrophages or macrophage-like cells capable of producing high levels of TNF α ; and (4) inhibition of TNF α prevents many of the pathophysiological or repair responses from occurring. The use of transgenic mice that overexpress TNF α or have a nonfunctional TNF gene (knockouts) and the development of specific pre- and posttranscriptional TNF inhibitors have

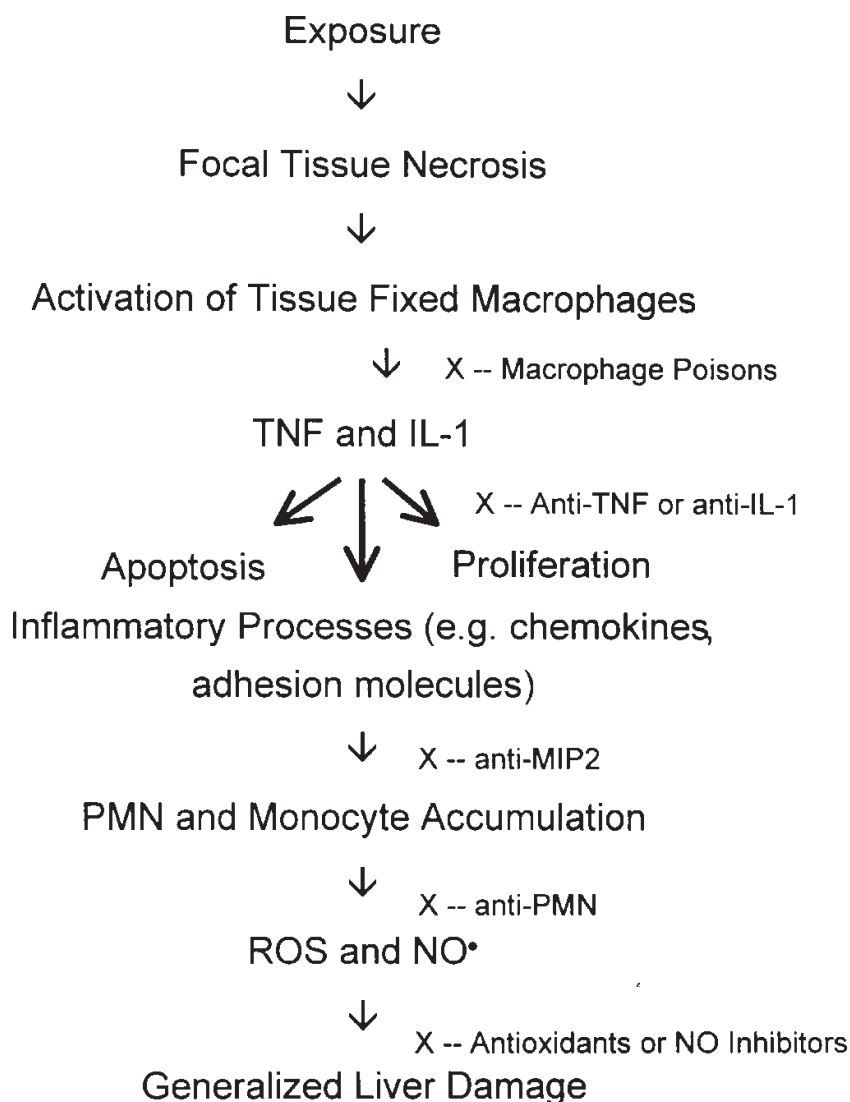


FIGURE 2. Hypothesis of chemical-induced liver injury. Increasing evidence has suggested that inflammation plays a major role in chemical-induced hepatotoxicity and repair. Initial damage produces focal tissue necrosis, resulting in activation of tissue-fixed macrophages. These activated macrophages secrete primary cytokines, including IL-1 and TNF α , which may induce apoptosis, stimulate cell growth, or initiate inflammatory processes. The inflammatory process ultimately results in the recruitment of activated neutrophils and the release of toxic products such as reactive oxygen species (ROS) and nitric oxide. Although exemptions to this scheme exist, the hypothesis is based primarily on the use of different types of inhibitors.

been instrumental in allowing for these observations.

III. TNF α AND ORGAN-SPECIFIC TOXICITY

Although TNF has presumably evolved to provide homeostatic processes by participating in tissue repair and host defense

mechanisms, its overproduction can lead to damage of normal cells and lethality.¹⁸ The pathophysiological sequelae associated with its overproduction or underproduction in chemical toxicities are probably similar to the effects in their idiopathic counterparts such as septic shock, inflammatory lung disease, and autoimmune disease. The following section describes the established asso-

ciations between TNF and organ-specific, chemical-induced toxic responses.

IV. LIVER

Traditionally, the study of xenobiotic-induced hepatic injury has focused on the direct actions of hepatotoxins or their metabolites on cellular targets in the liver. However, emphasis has shifted recently toward indirect mechanisms, and, in particular, the role of nonparenchymal cells and their soluble mediators in liver injury. This was stimulated by the observation that the inactivation or depletion of Kupffer cells decreases toxicity from exposure to hepatotoxins such as galactosamine,¹⁹ phalloidin,²⁰ ethanol,²¹ acetaminophen,²² diethyldithiocarbamate,²³ and carbon tetrachloride.²⁴ Increasing evidence suggests that mediators released from activated Kupffer cells, including TNF α , are important modulators of this injury. In this respect, TNF α proximally mediates acute phase responses, inflammatory cell infiltration, hyperlipidemia, free oxygen radical generation, fibrogenesis, and cholestasis in the liver.^{25–27} Elevated levels of TNF α occur in a variety of acute and chronic liver diseases, including viral or alcoholic hepatitis, biliary obstruction, and ischemia,^{27–29} as well as in experimental animals treated with hepatotoxins such as carbon tetrachloride,³⁰ cadmium chloride,³¹ acetaminophen,³² dimethylnitrosamine,³³ ethanol,³⁴ plant lectins,³⁵ and fumonisin B1.³⁶ A model of xenobiotic-induced inflammatory liver injury has been proposed (see Figure 2) wherein liver damage is thought to occur when a hepatotoxin, such as CCl₄ is metabolized to a highly reactive form (i.e., chloro-trimethyl radical) that is capable of interacting with membrane lipids, resulting in lipid peroxidation and subsequent cellular necrosis.³⁷ Resident macrophages at the site of damage become activated and secrete inflammatory mediators, including TNF α , which causes infiltration of activated neutrophils and monocytes into the damaged organ. These cells produce

additional cytokines, ROS, and reactive nitrogen species that exacerbate the damage. That TNF α is involved in this response can be evidenced by the ability to modulate the pathological sequelae by influencing *in vivo* TNF α synthesis.^{34,38–39} An obligatory role for endotoxin in this response has also been suggested.⁴⁰

Although most studies dealing with TNF α -associated hepatotoxicity have addressed necrotic cell death, hepatic apoptosis has been observed following exposure to hepatotoxic chemicals.^{41–44} The direct role of TNF α in this response has not been examined, although in the presence of transcriptional inhibitors TNF α induces apoptosis in hepatocyte cell cultures.^{45,46} Recently, Leist et al.⁴⁷ reported that the hepatotoxins, amanitin, and actinomycin D, both transcriptional inhibitors, mediate liver failure directly through TNF α activity. Furthermore, mice treated with neutralizing antibodies to TNF α , as well as those deficient in p55 expression, are relatively resistant to the toxicity of either chemical. Under other circumstances, TNF α has been implicated in liver repair following chemical damage through its ability to initiate parenchymal and nonparenchymal cell proliferation.^{39,48–50} In this respect, TNF α is necessary for liver regeneration following partial hepatectomy⁴⁹ and administration of TNF α *in vivo* increases hepatic DNA synthesis within 12 to 24 h.^{50,51} The mechanism by which TNF α stimulates hepatic cell proliferation involves both IL-6-dependent⁵² and -independent processes.¹⁷ Using a CCl₄-induced hepatotoxicity model in rodents, it has been demonstrated that both TNF α induction and liver repair following chemical damage require the induction of early immediate genes, such as *c-jun* and *c-fos*, and activation of the cEBP, AP-1, and NF- κ B transcription factors in hepatocytes.^{39–53} DNA binding sites for these transcription factors are found in the promoter region of the TNF α gene.

A number of chemicals are not toxic but mitogenic for hepatocytes, causing an in-

crease in liver mass *in vivo*. These chemicals have been implicated in liver cancer by acting as tumor promoters. Lead nitrate^{51,54} and ethylene dibromide⁵⁵ both induce TNF α expression in the liver and spleen of experimental animals and stimulate liver hyperplasia without associated necrosis or inflammation. The hyperplasia is temporally similar to that induced by TNF α itself, in that peak DNA synthesis of parenchymal cells is preceded by a wave of nonparenchymal cell proliferation.⁵² However, the induction of growth factors normally associated with liver regeneration, such as hepatocyte growth factor (HGF), transforming growth factor (TGF) α , and TGF β , are not apparent following exposure to these chemicals,⁵⁵ and a definitive role for TNF α in carcinogenesis has yet to be described. Furthermore, these substances may not directly stimulate TNF α , but rather enhance the sensitivity of Kupffer cells to endotoxin.⁵⁴ WY14643 (WY), a prototype peroxisome proliferator and potent rodent liver carcinogen, also increases liver mass and hepatocyte mitogenesis through a TNF α -dependent pathway.⁵⁶ Inhibition of Kupffer cell function by methyl palmitate⁵⁶ or treatment of mice with neutralizing antibodies to TNF α ⁵⁷ prevents hepatic DNA synthesis induced by WY. Although not a peroxisome proliferator, gadolinium chloride (GdCl₃) can induce hepatocyte mitogenesis through modulation of TNF α .⁵⁸ Whereas GdCl₃ has been used to inactivate Kupffer cells, it also induces hepatocyte DNA synthesis⁵⁸ and TNF α expression.⁵⁹ Similar to WY, administration of neutralizing antibodies to TNF α also prevents the mitogenic response to GdCl₃.

V. KIDNEY

Although considerable evidence exists to suggest cytokines play a role in the progression of classic renal diseases, few studies have focused on their role in xenobiotic-

induced nephrotoxicity.⁶⁰⁻⁶³ Regarding the former, the overexpression of cytokines, particularly TNF α and IL-1, are involved in many of the structural and functional changes in inflammatory kidney diseases such as Goodpasture syndrome, crescentic glomerulonephritis, proliferative glomerulonephritis, lupus nephritis, IgA nephropathy, as well as renal allograft rejection (reviewed in rRefs.⁶⁴⁻⁶⁶) This is due, in part, to the ability of TNF α to induce the production of complement components, ROS and prostanoids, in the kidney.⁶⁴ In addition to mediating kidney inflammation, TNF α has been shown to reduce glomerular blood flow, alter glomerular basement membrane permeability, and activate glomerular endothelial and mesangial cells.⁶⁴⁻⁶⁶ The pivotal role TNF α plays in glomerulonephritis has been demonstrated by the ability of neutralizing antibodies to attenuate glomerular injury in experimental nephrotoxic nephritis.⁶⁵ Furthermore, the administration of exogenous TNF α can induce severe proteinuria, glomerular injury, and tubulointerstitial nephritis, thereby mimicking the pathophysiological response in nephrotoxic nephritis.⁶⁵

The importance of TNF α in various classic nephropathies might suggest a similar role in chemical-induced injury. In this respect, suggestive evidence has been presented of a relationship between TNF α , exposure to heavy metals leading to glomerulonephritis and tubulointerstitial lesions, and the development of autoimmune disease.⁶⁷ Direct evidence for an association between nephrotoxicity and TNF α was provided in studies of rodents exposed to CdCl₂, where significant increases in kidney levels of TNF α and IL-6 occurred.⁶⁸ Although a direct role for TNF α in the kidney pathology was not found, IL-6, which is induced by TNF α , was shown to support renal tubular epithelial regeneration, suggesting an involvement in repair processes. A similar role for IL-6 was observed in an experimental model of nephrotoxic nephritis, where IL-6 infusion resulted

in the reduction of TNF and IL-1, along with the inhibition of proteinuria, monocyte infiltration and activation.⁶⁹ Corresponding to the decrements in TNF α and IL-1 levels associated with IL-6 treatment, there were significant increases in the level of IL-1 receptor antagonist and soluble TNF receptor, which serve as endogenous cytokine antagonists.

Adriamycin induces nephrosis characterized by proteinuria, glomerular epithelial damage, and interstitial leukocyte influx.⁷⁰ TNF α expression in this lesion corresponds with cellular infiltration and the presence of the chemotactic protein, interferon-inducible protein (IP)-10 within the glomerulus. The levels of both proteins parallel the development of proteinuria and tubulointerstitial nephritis. Nephrotoxicity of adriamycin is presumably due to a direct interaction of the drug with endogenous glomerular cells resulting in the release of TNF α and platelet activating factor (PAF), which in turn initiates a cascade of events leading to renal damage. The antiviral drug puromycin aminonucleoside represents a model for minimal change nephritis characterized by progressive renal fibrosis.⁷¹ ROS are responsible for the changes in the podocytes, and, similar to adriamycin, high levels of PAF and TNF α in the mesangium are associated with a cellular influx, as well as the synthesis of TGF β , leading to the accumulation of extracellular matrix proteins in the glomerulus and the interstitium. Taken together, these data at least provide suggestive evidence that

TNF α is involved in various aspects of xenobiotic-mediated kidney toxicity.

VI. CENTRAL NERVOUS SYSTEM

The nervous and immune systems show an intimate structural and functional relationship which exhibits extensive bidirectional communication mediated by common regulatory factors and receptors. TNF α is expressed in different brain regions at different stages of development, with the primary cellular sources being astrocytes and glial cells with neurons providing only a minor contribution.⁷²⁻⁷⁴ In addition to locally produced TNF α , peripheral TNF α can penetrate the CNS through a physically or pathologically modified blood-brain barrier.⁷⁴⁻⁷⁶ The expression and occurrence of TNF α in the brain is associated with changes associated with inflammatory processes, examples of which are listed in Table 1. Several studies have demonstrated that TNF α is involved in physiological sleep regulation and increased non-rapid eye movement sleep.⁸⁴ Extensive evidence exists for TNF α involvement in the pathogenesis of inflammatory demyelinating diseases, such as multiple sclerosis and experimental allergic encephalomyelitis (EAE) in mice.^{85,86} Recently, it has been demonstrated that the expression of a TNF α transgene in the CNS of mice triggers the development of a chronic inflammatory demyelinating disease, which includes progressive neurological symptoms such as ataxia and seizures.^{87,88} The pathological features

TABLE 1
Effects of TNF α in the CNS Possibly Associated with Neurodegenerative and Inflammatory Diseases

Response	Ref.
Cytotoxic to oligodendrocytes	77,78
Stimulates neutral sphingomyelinase	79
Up-regulates class I MHC on astrocytes	80
Enhances INF γ -induced class II MHC on astrocytes	81
Mitogenic for primary astrocytes and astrogloma	82
Induces NO synthase causing cerebral vasodilation	83

of this model, which include astrogliosis, reactive microgliosis, aberrant T-cell entry into the CNS, and myelin disruption, are characteristics of human inflammatory and demyelinating diseases. TNF α is also believed to play a key role in the pathogenesis of some CNS parasitic and bacterial infections, such as meningitis and cerebral malaria, as well as AIDS dementia complex, neurodegenerative diseases, and ischemia.⁸⁹⁻⁹¹

The significance of TNF α in CNS pathophysiology has been studied using TNF α neutralization treatments in established animal disease models or transgenic mice that overexpress or underexpress TNF α . For example, TNFR2-deficient mice are resistant to experimental cerebral malaria when compared with TNFR1-deficient or wild-type mice.⁷⁹ This phenomenon has been related to the absence of ICAM expression in brain microvessels of TNFR2-deficient mice. Studies on the role of TNF α in brain ischemic injury are controversial and may be related to differences in the models employed or time-courses studied. TNF α is released during focal ischemia or early reperfusion, and neutralization reduces the injury and improves the neurological outcome.⁹² Using TNFR knockout mice, exacerbation of neuronal damage after focal cerebral ischemia occurred.⁹³

While substantial evidence is provided for TNF α 's role in CNS diseases and trauma, the participation of TNF α in chemically induced neurotoxic responses has been considered only rarely. Overexpression of TNF α in the brain accompanies neurotoxicity in rats and mice associated with trimethyltin exposure, an organotin compound.⁹⁴ Recently, these studies have been confirmed and extended to demonstrate that trimethyltin induces TNF α expression and secretion in glial cell cultures.⁹⁵ Consistent with neurons as the primary target of trimethyltin intoxication following developmental exposure, it has been shown that trimethyltin induces apoptosis in neuronal/glial co-cultures through TNF α originating from glial cells.⁹⁶ In addition to

trimethyltin, a broad group of neurotoxicants, including triethyltin, lead, thallium, and tellurium, selectively affect myelination in the adult, although the role of TNF α in these responses has not been studied.

VII. LUNG

Chronic inflammatory lung diseases, such as idiopathic pulmonary fibrosis, granulomatous diseases, chronic bronchitis, adult respiratory distress syndrome, cystic fibrosis, and asthma, are associated with elevated levels of TNF α in lung fluids.⁹⁷⁻¹⁰¹ Evidence that TNF α is also important in the development of environmental lung diseases stems from both direct and indirect observations. First, a broad spectrum of chemical-induced lung diseases are associated with elevated TNF α level in the lung, including pollutant-induced inflammatory disease,¹⁰²⁻¹⁰⁴ fibrotic disease,¹⁰⁵ hypersensitivity pneumonitis, granulomatous disease,¹⁰⁶⁻¹⁰⁸ occupational asthma, and respiratory hypersensitivity (see Table 2 for specific examples). These elevations can be in excess of 100-fold as occurs during fibrotic reactions in animals as well as humans exposed to bleomycin, silica, or asbestos.^{106,110,111,116} Secondly, TNF α infusion in experimental animals mimics many of the pathological sequelae observed following exposure to pulmonary toxicants such as the growth of fibroblasts, lung collagen deposition, and inflammation.¹²⁵⁻¹²⁸ Finally, mice pretreated with products to inhibit TNF α , such as soluble receptors or neutralizing antibodies, are either resistant or show modified responses to pulmonary toxicity from asbestos, silica, bleomycin, quartz, or ozone.^{117,122,132,133}

The pathophysiologic events associated with environmentally induced inflammatory lung disease are similar to that which occurs from immune complex-induced lung injury. This involves sequentially, the deposition of pulmonary toxicants resulting in cell damage, activation and release of bioactive products, including TNF α , the upregulation of

TABLE 2
Examples of Pulmonary Toxicants Associated with Elevated Levels of TNF α

Agents	Pathology (Ref.)
Amiodarone	Pulmonary toxicity ¹⁰⁹
Beryllium	Granulomas ¹¹⁴
Bleomycin	Fibrosis ¹⁰⁶
Carbon	Monoxide necrosis ¹⁰⁵
Fibers/asbestos, silica	Inflammation/fibrosis/cancer ^{110,111,116,123,124}
Grain dust	Inflammation ^{112,113}
Ozone	Inflammation/irritation ^{120,121,122}
Isocyanates	Respiratory hypersensitivity ¹¹⁵
Lead oxide	Inflammation/cancer ¹¹⁹
Urban/diesel particles	Inflammation ^{117,118}

lung vascular ICAM-1 and E-selectin, the recruitment of blood neutrophils, and ultimately tissue injury and matrix formation by oxidants and proteases from neutrophils and macrophages. As in other organ systems, TNF α , itself does not directly mediate these responses, but rather plays a central role in regulating other mediators. TNF α studies in lung inflammation have focused on its ability to induce chemokines, such as IL-8, MIP-2, and MIP-1 α , which are potent neutrophil and monocyte activators and chemo-attractants.^{122–124} However, TNF α may also produce direct lung effects such as damage to the pulmonary vascular endothelium and subsequent capillary leakage.^{125–128} TNF α is present at low levels in normal lungs, where it may provide a protective role, such as through the expression of superoxide dismutase, a potent antioxidant enzyme.¹²⁹ In this respect, TNF α levels in the lung are reduced in smokers, which may account for their increased susceptibility to infections and decreased incidences to some autoimmune and inflammatory diseases. The role of TNF α in lung fibrogenesis is less clear, but it may have a dual role. During injury, when uncontrolled fibroblast proliferation occurs, TNF α production is increased to slow down the fibroproliferative response, whereas during repair, when fibroblasts participate in structural restitution, it stimulates cell proliferation.¹³⁰ The major source of pulmonary TNF α is thought to be alveolar macro-

phages.¹³¹ However, under most conditions lung endothelial and epithelial cells can also contribute significantly to the TNF α pool.

The signaling events responsible for TNF α induction in the lung are relatively complex, involving both direct and indirect events. TNF α itself as well as certain pollutants, such as vanadium pentoxide, asbestos, silica, and ozone, generate ROS. This in turn can signal the Ras pathway,¹³⁴ ultimately activating AP-1, NF- κ B, and NF-IL-6, members of oxidant-sensitive nuclear transcription factor families, which help regulate genes involved in inflammatory processes, including TNF α .¹³⁵ TNF α can also be induced in an autocrine manner, independent of both Ras and ROS, by the activation of NF- κ B through phosphatase-dependent I κ B activation via members of the TNF receptor-associated factors (see Figure 1).¹³⁶

VIII. SKIN

Keratinocytes, and, to a lesser extent, endothelial cells and resident dendritic cells (Langerhan's cell), are the major sources of dermal cytokines,¹³⁹ including TNF α .^{132,135–139} In the skin, TNF gene expression occurs in the epidermis within minutes after either physical damage or exposure to ultraviolet light (UV-B), contact allergens, or irritants.^{141–147} Irritant contact dermatitis (ICD), caused by agents such as phenol, sodium lauryl sulfate, croton oil, and

picryl chloride, results in the immediate induction of TNF α ,¹⁴⁸ with the levels of TNF α corresponding to the intensity of the inflammatory response.¹⁴⁹ Multiple lines of evidence indicate that TNF α is required for the development of allergic contact dermatitis (ACD) and irritant contact dermatitis (ICD).^{143,145,148,150–152} In this respect, many of the clinical features of ACD or ICD are reduced or prevented in TNF receptor knockout or in wild-type mice administered neutralizing TNF antibodies, whereas intradermal injection of TNF α produces many of their dermal features, including Langerhan's cell migration, epidermal necrosis, leukocyte infiltration, and hemorrhagic necrosis. Furthermore, TNF α can be found at high levels in the skin during acute episodes of irritant or allergic contact dermatitis. TNF α may not directly initiate inflammatory responses by contact irritants. This may occur through the release of preformed IL-1 α from injured keratinocytes that serves to stimulate TNF α production. Mechanical disruption of the skin from tape-stripping induces significant levels of TNF α , IL-1, and IL-6 at the site of the disrupted barrier, suggesting that TNF α is also involved in minor irritation.¹⁴⁷ Similarly, pretreatment of mice with agents that reduce circulating levels of TNF α re-

duce many of the clinical features of minor contact irritant responses such as edema and eosinophil accumulation.^{143–148} When irritants, such as phenol and croton oil, are administered to p55 knockout mice, inflammatory responses are markedly reduced in the knockout mice, compared with the wild type.¹⁴⁵ Furthermore, after intradermal injection of exogenous TNF α , inflammation is significantly lower in the p55 mice, suggesting that, as with most TNF responses, p55 is critical.¹⁴⁵ Surprisingly, ear painting with contact sensitizers results in normal Langerhan's cell migration in p55 knockout mice, whereas p75 knockout mice exhibit a 50% reduction in antigen-positive dendritic cells in the draining lymph nodes, suggesting that the p75 receptor is important in Langerhan's cell migration.¹⁵² In ACD, TNF α is responsible for initiating immune responses by signaling antigen-presenting (Langerhan's) cells of the skin to migrate to regional lymph nodes,¹⁵³ probably via the induction of adhesion molecules, where they present processed antigen to T lymphocytes.

A large number of specific pathologies and clinical manifestations have been ascribed to the overexpression and release of TNF α in the skin and for brevity are summarized in Table 3. As previously indicated,

TABLE 3
Potential Skin and Histological Changes and Diseases Associated with Overexpression of Dermal TNF α

Characteristic	Disease (example)
Histocytosis	Urticaria
Mucocutaneous Leishmaniasis	Psoriasis
Dermatitis Herpetiformis	Eczema
Contact Dermatitis	SLE
Histological occurrences	
Edema/wheal	Scaling
Hemorrhage	Erythema
Necrosis	Thrombi
Ulceration	Vesiculation
Vasoconstriction	
Inflammatory cell influx (PMNs, monocytes, lymphocytes)	

overexpression of TNF α in the skin may not always be associated with pathological events because TNF α has been implicated in epidermal cell proliferation and wound repair by helping to regulate angiogenesis and keratinocyte differentiation.¹⁵⁴ In psoriasis, however, TNF α has been associated with keratinocyte and dermal endothelial cell hyperproliferation through its ability to induce EGF receptors and TGF.

Skin damage from UV-B irradiation, which presents erythema, epidermal proliferation, leukocyte influx, keratinocyte apoptosis, and proinflammatory cytokine production at the site of exposure, is also associated with overexpression of TNF α .¹⁴³ Antigen-specific immunosuppression, which also may occur in response to UV-B, is partially regulated by TNF α .^{155,156} Although the mechanism remains to be clarified, it is believed that UV-induced DNA damage signals for excessive cytokine production and that polymorphisms in the TNF gene may be associated with susceptibility to UVB effects.

IX. TNF α AND GENETIC POLYMORPHISMS

The gene for TNF α is located within the class II region of the MHC, between HLA-B and DR. Its expression is tightly controlled at the transcriptional and post-transcriptional levels. Recently, polymorphisms within the TNF loci in humans have been described.¹⁵⁷⁻¹⁵⁸ In particular, two G vs. A transitions in the promoter region at positions -308 and -238 have been shown to influence the level of TNF α expression in response to various stimuli. The presence of these two alleles have been associated with a variety of immune- and inflammatory-associated diseases, such as autoimmune diseases, psoriasis, and periodontitis.¹⁵⁹⁻¹⁶⁰ These studies are of potential importance in risk assessment, as individuals who possess polymorphisms that express high levels of

inducible TNF α are at considerably higher risk of developing more severe disease. To date, only limited studies have been conducted in humans examining whether such associations exist with xenobiotic-induced diseases. For example, Zhai et al.¹⁶¹ indicated that a strong relationship exists between these TNF α polymorphisms and pneumoconiosis in coal miners, whereas similar associations have been observed in a population with sarcoidosis.¹⁶²

X. CONCLUSIONS

In summary, only recently have toxicologists come to appreciate the role inflammation plays in classic toxicological processes. This relationship can be extremely complex, as inflammation may well be only one facet of a time- and dose-dependent continuum of toxicological and repair processes. Although many mediators are responsible for inflammatory processes, TNF α , and to a lesser extent IL-1 β , have received the most attention because they represent central mediators involved in regulating this process. Not surprisingly, considerable efforts are being undertaken using our newly found understanding of molecular control to develop specific and safe chemical, biological, and molecular inhibitors of TNF α for potential therapeutic use. This effort has also allowed for a better understanding, not only of the pathological sequelae induced by TNF α , but also its influence in normal physiological and repair processes and has allowed an opportunity to better understand the toxicological processes.

REFERENCES

1. Vassali, P., The pathophysiology of tumor necrosis factor, *Ann. Rev. Immunol.*, **1992**; 10: 411-52.
2. Bazzoni, F. and Beutler, B., Seminars in medicine of the Beth Israel Hospital, Boston: the tumor necrosis factor ligand and receptor

- families, *N. Engl. J. Med.*, **1996**; 334: 1717–25.
3. Aggarwal, B. B. and Natarajan, K., Tumor necrosis factors: developments during the last decade, *Eur. Cytokine Network*, **1996**; 7: 93–124.
 4. Kriegler, M., Perez, C., DeFay, K., Albert, I., and Lu, S. D., A novel form of TNF/cahectin is a cell surface cytotoxic transmembrane protein: ramifications for the complex physiology of TNF, *Cell*, **1988**, 53: 45–53.
 5. Beyaert, R. and Fiers, W., Tumor necrosis factor and lymphotoxin, *Cytokines*, **1998**; 24: 335–59.
 6. Schook, L. B. and Laskin, D. L., *Xenobiotics and Inflammation*, Academic Press, San Diego, 1994.
 7. Darnay, B. G. and Aggarwal, B. B., Early events in TNF signaling: a story of associations and dissociations, *J. Leuk. Biol.*, **1997**; 61: 55966.
 8. Miyamoto, S., Maki, M., Schmitt, M. J., Hatanaka, M., and Verma, I. M., Tumor necrosis factor α induced phosphorylation of I κ B α is a signal for its degradation but not association from NF- α B, *Proc. Natl. Acad. Sci.*, **1994**; 91: 12740–4.
 9. Zhang, X. M., Weber, I., and Chen, M. J., Site-directed mutational analysis of human tumor necrosis factor α receptor binding site and structure-functional relationship, *J. Biol. Chem.*, **1992**, 267: 24069–75.
 10. Schmid, D. S., Tite, J. P., and Ruddle, N. H., DNA fragmentation: manifestation of target cell destruction mediated by cytotoxic T-cell clones and cell-free lymphotoxin-containing supernatant, *Proc. Natl. Acad. Sci. USA*, **1986**, 83: 1881–7.
 11. Zhang, M. and Tracey, K. J., Tumor necrosis factor, in *The Cytokine Handbook*, Academic Press, San Diego, **1998**, 19: 517–48.
 12. Milenkovic, L., Rettori, V., Snyder, G. D., Beutler, B., and McCann, S. M., Cachectin alters anterior pituitary hormone release by a direct action *in vitro*, *Proc. Natl. Acad. Sci. USA*, **1989**, 86: 2418–22.
 13. Sherry, B. and Cerami, A., Cachectia/tumor necrosis factor exerts endocrine, paracrine, and autocrine control of inflammatory responses, *J. Cell Biol.*, **1988**, 107: 1269–77.
 14. Brenner, D. A., O'Hara, M., Angel, P., Chojkier, M., and Karin, M., Prolonged activation of jun and collagenase genes by tumor necrosis factor- α , *Nature*, **1989**, 337: 661–3.
 15. Hoffman, M. and Weinberg, J. B., Tumor necrosis factor- α induces increased hydrogen peroxide production and Fc receptor expression, but not increased Ia antigen expression by peritoneal macrophages, *J. Leukocyte Biol.*, **1987**, 42: 704–7.
 16. Jaesche, H., Smith, C. W., Clemens, M. G., Ganey, P. E., and Roth, R. A., Mechanisms of inflammatory liver injury: adhesion molecules and cytotoxicity of neutrophils, *Toxicol. Appl. Pharmacol.*, **1996**; 139: 213–26.
 17. Gallucci, R. M., Simeonova, P. P., Toriumi, W., and Luster, M. I., Tumor necrosis factor- α modulates transforming growth factor-alpha in murine regenerating liver and isolated hepatocytes, **1998**, *submitted*.
 18. Carswell, E. A., Old, L. J., Kassel, R. L., Green, S., Fiore, N., and Williamson, B., An endotoxin-induced serum factor that causes necrosis of tumors, *Proc. Natl. Acad. Sci. USA*, **1975**, 72: 3666–70.
 19. Czaja, M. J., Xu, J., Ju, Y., Alt, E., and Schmiedeberg, P., Lipopolysaccharide-neutralizing antibody reduces hepatocyte injury from acute hepatotoxin administration, *Hepatology*, **1994**; 19: 1282–9.
 20. Barriault, C., Petit, J. L., Gascon-Barre, M., Huet, P. M., Yousef, I. M., and Tuchweber, B., Effect of phalloidin on cholestasis, hemodynamics, and microcirculation in isolated perfused rat liver, *Hepatology*, **1996**; 23: 294–302.
 21. Adachi, Y., Moore, L. E., Bradford, B. U., Gao, W., and Thurman, R. G., Antibiotics prevent liver injury in rats following long-term exposure to ethanol, *Gastroenterology*, **1995**; 108: 218–24.
 22. Laskin, D. L., Gardner, C. R., Price, V. F., and Jollow, D. J., Modulation of macrophage functioning abrogates the acute hepatotoxicity of acetaminophen, *Hepatology*, **1995**; 21: 1045–50.

23. Ishiyama, H., Ogino, K., and Hobara, T., Role of Kupffer cells in rat liver injury induced by diethyldithiocarbamate, *Eur. J. Pharmacol.*, **1995**; 292: 135–41.
24. Edwards, M. J., Keller, B. J., Kauffman, F. C., and Thurman, R. G., The involvement of Kupffer cells in carbon tetrachloride toxicity, *Toxicol. Appl. Pharmacol.*, **1993**; 119: 275–9.
25. Andus, T., Bauer, J., and Gerok, W., Effects of cytokines on the liver, *Hepatology*, **1991**; 13: 364–75.
26. Feingold, K. R., and Grunfeld, C., Tumor necrosis factor- α stimulates hepatic lipogenesis in the rat *in vivo*, *J. Clin. Invest.*, **1987**; 80: 184–90.
27. Tilg, H., The role of cytokines in the pathophysiology of chronic liver diseases, *Int. J. Clin. Lab Res.*, **1993**; 23: 179–85.
28. Bird, G. L., Sheron, N., Goka, A. K., Alexander, G. J., and Williams, R. S., Increased plasma tumor necrosis factor in severe alcoholic hepatitis, *Ann. Intern. Med.*, **1990**; 112: 917–20.
29. Muto, Y., Nouri-Aria, K. T., Meager, A., Alexander, G. J., Eddleston, A. L., and Williams, R., Enhanced tumour necrosis factor and interleukin-1 in fulminant hepatic failure, *Lancet*, **1988**; 2: 72–4.
30. Czaja, M. J., Flanders, K. C., Biempica, L., Klein, C., Zern, M. A., and Weiner, F. R., Expression of tumor necrosis factor- α and transforming growth factor- β 1 in acute liver injury, *Growth Factors*, **1989**; 1: 219–26.
31. Kayama, F., Yoshida, T., Elwell, M. R., and Luster, M. I., Role of tumor necrosis factor- α in cadmium-induced hepatotoxicity, *Toxicol. Appl. Pharmacol.*, **1995**; 131: 224–34.
32. Blazka, M. E., Wilmer, J. L., Holladay, S. D., Wilson, R. E., and Luster, M. I., Role of proinflammatory cytokines in acetaminophen hepatotoxicity, *Toxicol. Appl. Pharmacol.*, **1995**; 133: 43–52.
33. Schook, L. B., Lockwood, J. F., Yang, S. D., and Myers, M. J., Dimethylnitrosamine (DMN)-induced IL-1 β , TNF- α , and IL-6 inflammatory cytokine expression, *Toxicol. Appl. Pharmacol.*, **1992**; 116: 110–6.
34. Iimuro, Y., Gallucci, R. M., Luster, M. I., Kono, H., and Thurman, R. G., Antibodies to tumor necrosis factor attenuate hepatic necrosis and inflammation caused by chronic exposure to ethanol in the rat, *Hepatology*, **1997**; 26: 1530–7.
35. Ksontini, R., Colagiovanni, D. B., Josephs, M. D., Edwards, C. K., Tannahill, C. L., Solorzano, C. C., Norman, J., Denham, W., Clare-Salzler, M., MacKay, S. L., and Moldawer, L. L., Disparate roles for TNF- and Fas ligand in concanavalin A- α induced hepatitis, *J. Immunol.*, **1998**; 160: 4082–9.
36. Dugyala, R. R., Sharma, R. P., Tsunoda, M., and Riley, R. T., Tumor necrosis factor- α as a contributor in fumonisin B1 toxicity, *J. Pharmacol. Exp. Ther.*, **1998**; 285: 317–24.
37. Laskin, D. L. and Pendino, K. J., Macrophages and inflammatory mediators in tissue injury, *Ann. Rev. Pharmacol. Toxicol.*, **1995**; 35: 655–77.
38. Czaja, M. J., Xu, J., and Alt, E., Prevention of carbon tetrachloride-induced rat liver injury by soluble tumor necrosis factor receptor, *Gastroenterology*, **1995**; 108: 1849–54.
39. Bruccoleri, A., Gallucci, R., Germolec, D. R., Blackshear, P., Simeonova, P., Thurman, R. G., and Luster, M. I., Induction of early-immEDIATE genes by tumor necrosis factor contribute to liver repair following chemical-induced hepatotoxicity, *Hepatology*, **1997**; 25: 133–41.
40. Roth, R. A., Harkema, J. R., Pestka, J. P., and Ganey, P. E., Is exposure to bacterial endotoxin a detriment of susceptibility to intoxication from xenobiotic agents?, *Toxicol. Appl. Pharmacol.*, **1997**; 147: 300–11.
41. Ray, S. D., Kamendulis, L. M., Gurule, M. W., Yorkin, R. D., and Corcoran, G. B., Ca²⁺ antagonists inhibit DNA fragmentation and toxic cell death induced by acetaminophen, *FASEB J.*, **1993**; 7: 453–63.
42. Ray, S. D., Sorge, C. L., Kamendulis, L. M., and Corcoran, G. B., Ca(++)-activated DNA fragmentation and dimethylnitrosamine-induced hepatic necrosis: effects of Ca(++)-endonuclease and poly(ADP-ribose) polymerase inhibitors in mice, *J. Pharmacol. Exp. Ther.*, **1992**; 263: 387–94.

43. Goldin, R. D., Hunt, N. C., Clark, J., and Wickramasinghe, S. N., Apoptotic bodies in a murine model of alcoholic liver disease: reversibility of ethanol-induced changes, *J. Pathol.*, **1993**; 171: 73–6.
44. Habeebu, S. S., Liu, J., and Klaassen, C. D., Cadmium-induced apoptosis in mouse liver, *Toxicol. Appl. Pharmacol.*, **1998**; 149: 203–9.
45. Reid, T. R., Torti, F. M., and Ringold, G. M., Evidence for two mechanisms by which tumor necrosis factor kills cells, *J. Biol. Chem.*, **1989**; 264: 4583–9.
46. Leist, M., Gantner, F., Bohlinger, I., Germann, P. G., Tiegs, G., and Wendel, A., Murine hepatocyte apoptosis induced in vitro and in vivo by TNF- α requires transcriptional arrest, *J. Immunol.*, **1994**; 153: 1778–88.
47. Leist, M., Gantner, F., Naumann, H., Bluethmann, H., Vogt, K., Brigelius-Flohe, R., Nicotera, P., Volk, H. D., and Wendel, A., Tumor necrosis factor-induced apoptosis during the poisoning of mice with hepatotoxins, *Gastroenterology*, **1997**; 112: 923–34.
48. Feingold, K. R., Soued, M., and Grunfeld, C., Tumor necrosis factor stimulates DNA synthesis in the liver of intact rats, *Biochem. Biophys. Res. Commun.*, **1988**; 153: 576–82.
49. Akerman, P., Cote, P., Yang, S. Q., McClain, C., Nelson, S., Bagby, G. J., and Diehl, A. M., Antibodies to tumor necrosis factor- α inhibit liver regeneration after partial hepatectomy, *Am. J. Physiol.*, **1992**; 263: G579–85.
50. Satoh, M. and Yamazaki, M., Tumor necrosis factor stimulates DNA synthesis of mouse hepatocytes in primary culture and is suppressed by transforming growth factor- β and interleukin-6, *J. Cell Physiol.*, **1992**; 150: 134–9.
51. Shinozuka, H., Ohmura, T., Katyal, S. L., Zedda, A. I., Ledda-Columbano, G. M., and Columbano, A., Possible roles of non-parenchymal cells in hepatocyte proliferation induced by lead nitrate and by tumor necrosis factor- β , *Hepatology*, **1996**; 23: 1572–7.
52. Cressman, D. E., Greenbaum, L. E., DeAngelis, R. A., Ciliberto, G., Furth, E. E., Poli, V., and Taub, R., Liver failure and defective hepatocyte regeneration in interleukin-6-deficient mice, *Science*, **1996**; 274: 1379–83.
53. Yamada, Y., Kirillova, I., Peschon, J. J., and Fausto, N., Initiation of liver growth by tumor necrosis factor: deficient liver regeneration in mice lacking type I tumor necrosis factor receptor, *Proc. Natl. Acad. Sci. USA*, **1997**; 94: 1441–6.
54. Shinozuka, H., Kubo, Y., Katyal, S. L., Coni, P., Ledda-Columbano, G. M., Columbano, A., and Nakamura, T., Roles of growth factors and of tumor necrosis factor- α on liver cell proliferation induced in rats by lead nitrate, *Lab Invest.*, **1994**; 71: 35–41.
55. Ledda-Columbano, G. M., Columbano, A., Cannas, A., Simbula, G., Okita, K., Kayano, K., Kubo, Y., Katyal, S. L., and Shinozuka, H., Dexamethasone inhibits induction of liver tumor necrosis factor-mRNA and liver growth induced by lead nitrate and ethylene dibromide, *Am. J. Pathol.*, **1994**; 145: 951–8.
56. Rose, M. L., Germolec, D. R., Schoonhoven, R., and Thurman, R. G., Kupffer cells are causally responsible for the mitogenic effect of peroxisome proliferators, *Carcinogenesis*, **1997**; 18: 1453–6.
57. Bojes, H. K., Germolec, D. R., Simeonova, P., Bruccoleri, A., Schoonhoven, R., Luster, M. I., and Thurman, R. G., Antibodies to tumor necrosis factor prevent increases in cell replication in liver due to the potent peroxisome proliferator, WY-14,643, *Carcinogenesis*, **1997**; 18: 669–74.
58. Rai, R. M., Zhang, J. X., Clemens, M. G., and Diehl, A. M., Gadolinium chloride alters the acinar distribution of phagocytosis and balance between pro- and anti-inflammatory cytokines, *Shock*, **1996**; 6: 243–7.
59. Rai, R. M., Loffreda, S., Karp, C. L., Yang, S. Q., Lin, H. Z., and Diehl, A. M., Kupffer cell depletion abolishes induction of interleukin-10 and permits sustained overexpression of tumor necrosis factor- α messenger RNA in the regenerating rat liver, *Hepatology*, **1997**; 25: 889–95.
60. Rotter, B. A., Prelusky, D. B., and Pestka, J. J., Toxicology of deoxynivalenol (vomitoxin), *J. Toxicol. Environ. Hlth.*, **1996**; 48: 1–34.

61. Azcma-Olivera, J. I., Ouyang, Y., Murtha, J., Chu, F. S., and Pestka, J. J., Induction of cytokine mRNAs in mice after oral exposure to the trichothecene v-(deoxynivalenol): relationship to toxin distribution and protein synthesis inhibition, *Toxicol. Appl. Pharmacol.*, **1995**; 133: 109–20.
62. Ideura, T., Yoshimura, A., Shirai, M., Taira, T., and Koshikawa, S., Endotoxin-induced acute tubular necrosis in cirrhotic rats, *Scand. J. Urol. Nephrol.*, **1993**; 27: 433–9.
63. Nagaike, M., Hirao, S., Tajima, H., Noji, S., Taniguchi, S., Matsumoto, K., and Nakamura, T., Renotropic functions of human growth factor in renal regeneration after unilateral nephrectomy, *J. Biol. Chem.*, **1991**; 266: 22781–4.
64. Sedor, J. R., Interleukin-1: a master cytokine in the renal response to injury, in *Molecular Nephrology: Kidney Function in Health and Disease*, Marcek Dekker, New York, **1995**; 631–652.
65. Ostendorf, T., Burg, M., and Floege, J., Cytokines and glomerular injury, *Kid. Blood Pressure Res.*, **1996**; 19: 281–9.
66. Baud, L. and Ardaillou, R., Tumor necrosis factor in renal injury, *Mineral Elect. Metabol.*, **1995**; 21: 336–41.
67. Bigazzi, P. E., Autoimmunity and heavy metals, *Lupus*, **1994**; 3: 449–453.
68. Kayama, F., Yoshida, T., Elwell, M. R., and Luster, M. I., Cadmium-induced renal damage and proinflammatory cytokines: possible role of IL-6 in tubular epithelial cell regeneration, *Toxicol. Appl. Pharmacol.*, **1995**; 134: 26–34.
69. Karkar, A. M., Smith, J., Tam, F. W. K., Pusey, C. D., and Rees, A. J., Abrogation of glomerular injury in nephrotic nephritis by continuous infusion of IL-6, *Kid. Int.*, **1997**; 52: 1313–20.
70. Gomez-Chiarri, M., Ortiz, A., Gonzalez-Cuadrado, S., Seron, D., Emancipator, S., Hamilton, T. A., Barat, A., Plaza, J. J., Gonzalez, E., and Egido, J., Interferon-inducible protein-10 is highly expressed in rats with experimental nephrosis, *Am. J. Pathol.*, **1996**; 148: 301–11.
71. Jones, C. L., Buch, S., Post, M., McCulloch, L., Liu, E., and Eddy, A. A., Pathogenesis of interstitial fibrosis in chronic puromycin aminonucleoside nephrosis, *Kid. Int.*, **1991**; 40: 1020–31.
72. Breder, C. D., Tsujimoto, M., Terano, Y., Scott, D. W., and Saper, C. B., Distribution and characterization of tumor necrosis factor- α -like immunoreactivity in the murine central nervous system, *J. Comp. Neurol.*, **1993**; 337: 543–67.
73. Righi, M., Mori, L., de Libero, G., Sironi, M., Biondi, A., Mantovani, A., Donini, D. S., and Castagnoli, P. R., Monokine production by microglial cell clones, *Eur. J. Immunol.*, **1989**; 19: 1443–8.
74. Breder, C. D., Hazuka, C., Ghayur, T., Klug, C., Huginin, M., Yasuda, K., Teng, M., and Saper, C. B., Regional induction of tumor necrosis factor alpha expression in the mouse brain after systemic lipopolysaccharide administration, *Proc. Natl. Acad. Sci. U.S.A.*, **1994**; 91: 11393–7.
75. Ignatowski, T. A., Noble, B. K., Wright, J. R., Gorfien, J. L., Heffner, R. R., and Spengler, R. N., Neuronal-associated tumor necrosis factor: its role in noradrenergic functioning and modification of its expression following antidepressant drug administration, *J. Neuroimmunol.* **1997**; 79: 84–90.
76. Gahring, L. C., Carlson, N. G., Kulmar, R. A., and Rogers, S. W., Neuronal expression of tumor necrosis factor- α in the murine brain, *Neuroimmunomodulation*, **1996**; 3: 289–303.
77. Selmaj, K. and Raine, S., Tumor necrosis factor mediates myelin and oligodendrocyte damage *in vitro*, *Ann. Neurol.*, **1988**; 23: 339–46.
78. Zajicek, J. P., Wing, M., Scolding, N. J., and Compton, H., Interactions between oligodendrocytes and microglia adherence and killing, *Brain*, **1992**; 115: 1611–31.
79. Chakraborty, G., Ziemba, S., Drivas, A., and Ledeen, R. W., Myelin contains neutral sphingomyelinase activity that is stimulated by tumor necrosis factor- α , *J. Neurosci. Res.*, **1997**; 50: 466–76.
80. Lavi, E., Suzumura, A., Murasko, D. M., Murray, E. M., Silberger, D. H., and Weiss,

- S. R., Tumor necrosis factor induces expression of MHC class I antigens on mouse astrocytes, *J. Neuroimmunol.*, **1988**; 18: 245–53.
81. Vidovic, M., Sparacio, S. M., Elovitz, M., and Benveniste, E. N., Induction and regulation of class II MHC mRNA expression in astrocytes by IFN- γ and TNF- α , *J. Neuroimmunol.*, **1990**; 30: 189–200.
 82. Selmaj, K. W., Farooq, M., Norton, W. T., Raine, C. S., and Brosnan, C. F., Proliferation of astrocytes *in vitro* in response to cytokines: a primary role for tumor necrosis factor, *J. Immunol.* **1990**; 144: 129–35.
 83. Brian, J. E. and Faraci, F. M., Tumor necrosis factor—induced dilation of cerebral arterioles, *Stroke*, **1998**; 29: 509–15.
 84. Fang, J., Wang, Y., and Krueger, J. M., Mice lacking the TNF55 kDa receptor and are involved in physiological sleep regulation fail to sleep more after TNF α treatment, *J. Neurosci.*, **1996**; 17: 5949–55.
 85. Ruddle, N. H., Bergman, C. M., McGrath, K. M., Lingenheld, E. G., Grunnet, M. L., Padula, S. J., and Clark, R. B., An antibody to lymphotoxin and tumor necrosis factor prevents transfer of experimental allergic encephalomyelitis, *J. Exp. Med.*, **1990**; 172: 1193–200.
 86. Selmaj, K., Papierz, W., Glabinski, A., and Kohno, T., Prevention of chronic relapsing experimental autoimmune encephalomyelitis by soluble tumor necrosis factor receptor, *J. Neuroimmunol.*, **1995**; 56: 135–41.
 87. Probert, L., Akassoglou, K., Pasparakis, M., Kontogeorgos, G., and Kollias, G., Spontaneous inflammatory demyelinating disease in transgenic mice showing central nervous system-specific expression of tumor necrosis factor α , *Proc. Natl. Acad. Sci. USA*, **1995**; 92: 11294–8.
 88. Akassoglou, K., Probert, L., Kontogeorgos, G., and Kollias, G., Astrocyte-specific but not neuron-specific transmembrane TNF triggers inflammation and degeneration in the central nervous system of transgenic mice, *J. Immunol.*, **1997**; 158: 438–45.
 89. Lucas, R., Juillard, P., Decoster, E., Redard, M., Burger, D., Donati, Y., Giroud, C., Monso-Hinard, C., De Kesel, T., Buurman, W. A., Moore, M. W., Dayer, J. M., Fiers, W., Bluethmann, H, and Grau, G. E., Crucial role of tumor necrosis factor (TNF) receptor 2 and membrane-bound TNF in experimental cerebral malaria, *Eur. J. Immunol.*, **1997**; 27: 1719–25.
 90. Fillit, H., Ding, W. H., Buee, L., Kalman, J., Alstiel, L., Lawlor, B., and Wolf-Klein, G., Elevated circulating tumor necrosis factor levels in Alzheimer's disease, *Neurosci. Lett.*, **1991**; 129: 318–20.
 91. Ilyin, S. E. and Plata-Salaman, C. R., HIV-1 envelope glycoprotein 120 regulates brain IL-1 system and TNF- α mRNAs *in vivo*, *Brain Res. Bull.*, **1997**; 44: 67–73.
 92. Lavine, S. D., Hofman, F. M., and Zlokovic, B. V., Circulating antibody against tumor necrosis factor-alpha protects rat brain from reperfusion injury, *J. Cerebral Blood Flow Metabol.*, **1998**; 18: 52–8.
 93. Bruce, A. J., Boling, W., Kindy, M. S., Peschon, J., Kraemer, P. J., Carpenter, M. K., Holtsberg, F. W., and Mattson, M. P., Altered neuronal and microglial responses to excitotoxic and ischemic brain injury in mice lacking TNF receptors, *Nature Med.*, **1996**; 2: 788–94.
 94. Maier, W. E., Brown, H. W., Tilson, H. A., Luster, M. I., and Harry, G. J., Trimethyltin increases interleukin (IL)-1 α , IL-6 and tumor necrosis factor- α mRNA levels in rat hippocampus, *J. Neuroimmunol.*, **1995**; 59: 65–75.
 95. Maier, W. E., Bartenbach, M. J., Brown, H. W., Tilson, H. A., and Harry, G. J., Induction of tumor necrosis factor alpha in cultured glial cells by trimethyltin, *Neurochem. Int.*, **1997**; 30: 385–92.
 96. Vivani, B., Corsini, E., Galli, C. L., and Marinovich, M., Glia increase degeneration of hippocampal neurons through release of tumor necrosis factor- α , *Toxicol. Appl. Pharmacol.*, **1998**; 150: 271–6.
 97. Xing, Z., Jordana, M., Kirpalani, H., Driscoll, K. E., Schall, T. J., and Gaudie, J., Cytokine expression by neutrophils and macrophages *in vivo*: endotoxin induces tumor necrosis factor- α , macrophage inflammatory protein-2, interleukin-1 β and interleukin-6, but not RANTES or transforming growth factor- β ₁

- mRNA expression in acute lung inflammation, *Am. J. Respir. Cell Mol. Biol.*, **1994**; 10: 148–53.
98. Bonfield, T. L., Panuska, J. R., Konstan, M. W., Hilliard, K. A., Hilliard, J. B., Ghnaim, H., Berger, M., Inflammatory cytokines in cystic fibrosis lungs, *Am. J. Respir. Crit. Care Med.*, **1995**; 152: 2111–8.
99. Kasahara, K., Kobayashi, K., Shikama, Y., Yoneya, I., Kaga, S., Hashimoto, M., Odagiri, T., Soejima, K., Ide, H., Takahasi, T., and Yoshida, T., The role of monokines in granuloma formation in mice: the ability of interleukin-1 and tumor necrosis factor- α to induce lung granulomas, *Clin. Immunol. Immunopathol.*, **1989**; 51: 419–25.
100. Shah, A., Church, M. K., and Holgate, S. T., Tumor necrosis factor : A potential mediator of asthma, *Clin. Exper. Allergy*, **1995**; 25: 1038–44.
101. Sueoka, N., Sueoka, E., Miyazaki, Y., Okabe, S., Kurosumi, M., Takayama, S., and Fujiki, H., Molecular pathogenesis of interstitial pneumonitis with TNF- α transgenic mice, *Cytokine*, **1998**; 10: 124–31.
102. Kuschner, W. G., d'Allessandro, A., Wong, H., and Blanc, P. D., Early pulmonary cytokine responses to zinc oxide fume inhalation, *Environ. Res.*, **1997**; 75: 7–11.
103. Antonini, J. M., Krishna, M. G. G., and Brain, J. D., Responses to welding fumes: lung injury, inflammation and the release of tumor necrosis factor- α and interleukin-1 β , *Exp. Lung Res.*, **1997**; 23: 20527.
104. Driscoll, K. E., Carter, J. M., Hassenbein, D. G., and Howard, B., Cytokines and particle-induced inflammatory cell recruitment, *Environ. Hlth. Perspect.*, **1997**; 5: 1159–64.
105. Arias-Diaz, J., Villa, N., Hernandez, J., Vara, E., and Balibrea, J. L., Carbon monoxide contributes to the cytokine-induced inhibition of surfactant synthesis by human type II pneumocytes, *Arch. Surg.*, **1997**; 132: 1352–60.
106. Sleijfer, S., Vujaskovic, Z., Limburg, P. C., Koops, H. S., and Mulder, N. H., Induction of tumor necrosis factor as a cause of bleomycin-related toxicity, *Cancer*, **1998**; 82: 970–4.
107. Bost, T. W., Riches, D. W., Schumacher, B., Carre, P. C., Khan, T. Z., Martinez, J. A., and Newman, L. S., Alveolar macrophages from patients with beryllium disease and sarcoidosis express increased levels of mRNA for tumor necrosis factor- and interleukin-6, but not interleukin-1 α , *Am. J. of Respr. Cell Mol. Biol.*, **1994**; 10: 506–13.
108. Tinkle, S. S., Schwitters, P. W., and Newman, L. S., Cytokine production by bronchoalveolar lavage cells in chronic beryllium disease, *Environ. Hlth. Perspect.*, **1996**; 104: 969–71.
109. Reinhart, P. G. and Gairola, C. G., Amiodarone-induced pulmonary toxicity in Fischer rats: release of tumor necrosis factor α and transforming growth factor β by pulmonary alveolar macrophages, *J. Toxicol. Environ. Hlth.*, **1997**; 52: 353–65.
110. Li, X. Y., Lamb, D., and Donaldson, K., The production of TNF α and IL-1-like activity by bronchoalveolar leukocytes after intratracheal instillation of crocidolite asbestos, *Int. J. Exp. Pathol.*, **1993**; 74: 403–11.
111. Simeonova, P. P. and Luster, M. I., Iron and reactive oxygen species in the asbestos-induced tumor necrosis factor- α (TNF- α) response from alveolar macrophages, *Am. J. Respir. Cell Mol. Biol.*, **1995**; 12: 676–83.
112. Vanhee, D., Gosset, P., Marquette, C. H., Wallaert, B., Lafitte, J. J., Gosselin, B., Voisin, C., and Tonnel, A. B., Secretion and mRNA expression of TNF α and IL-6 in the lungs of pneumoconiosis patients, *Am. J. Respir. Crit. Care Med.*, **1995**; 152: 298–306.
113. Jagielo, P. J., Watt, J. L., Quinn, T. J., Knapp, H. R., and Schwartz, D. A., Pentoxifylline does not alter the response to inhaled grain dust, *Chest*, **1997**, 111: 1429–35.
114. Bost, T. W., Riches, D. W., Schumacher, B., Carre, P. C., Khan, T. Z., Martinez, J. A., and Newman, L. S., Alveolar macrophages from patients with beryllium disease and sarcoidosis express increased levels of mRNA for tumor necrosis factor- α and interleukin-6 but not interleukin-1 β , *Am. J. Respir. Cell Mol. Biol.*, **1994**; 10: 506–13.
115. Ban, M., Hettich, D., Goutet, M., and Bonnet, P., TDI inhalation in guinea pigs involves migration of dendritic cells, *Toxicol. Lett.*, **1997**; 93: 185–94.

116. David, G. S., Pfeiffer, L. M., and Hemenway, D. R., Persistent overexpression of interleukin-1 β and tumor necrosis factor- α in murine silicosis, *J. Environ. Pathol. Toxicol. Oncol.*, **1998**; 17: 99–114.
117. Driscoll, K. E., Hassenbein, D. G., Carter, J. M., Kunkel, S. L., Quinlan, T. R., and Mossman, B. T., TNF α and increased chemokine expression in rat lung after particle exposure, *Toxicol. Lett.*, **1995**; 82/83: 483–89.
118. Dong, W., Lewtas, J., and Luster, M. I., Role of endotoxin in tumor necrosis factor expression from alveolar macrophages treated with urban air particles, *Exp. Lung Res.*, **1996**; 22: 577–92.
119. Zelikoff, J. T., Parsons, E., and Schlesinger, R. B., Inhalation of particulate lead oxide disrupts pulmonary macrophage-mediated functions important for host defense and tumor surveillance in the lung, *Environ. Res.*, **1993**; 62: 207–22.
120. Ishii, Y., Yang, H., Sakamoto, T., Nomura, A., Hasegawa, S., Hirata, F., and Bassett, D. J., Rat alveolar macrophage cytokine production and regulation of neutrophil recruitment following acute ozone exposure, *Toxicol. Appl. Pharmacol.*, **1997**; 147: 214–23.
121. Devlin, R. B., McKinnon, K. P., Noah, T., Becker, S., and Koren, H. S., Ozone-induced release of cytokines and fibronectin by alveolar macrophages and airway epithelial cells, *Am. J. Physiol.*, **1994**; 266: L612–9.
122. Pearson, A. C. and Bhalla, D. K., Effects of ozone on macrophage adhesion *in vitro* and epithelial and inflammatory responses *in vivo*: the role of cytokines, *J. Toxicol. Environ. Hlth.*, **1997**; 50: 143–57.
123. Huang, S., Paulauskis, J. D., Godleski, J. J., and Kobzik, L., Expression of macrophage inflammatory protein-2 and KC mRNA in pulmonary inflammation, *Am. J. Pathol.*, **1992**; 141: 981–8.
124. Vanhee, D., Gosset, P., Boitelle, A., Wallaert, B., and Tonnel, A. B., Cytokines and cytokine network in silicosis and coal workers' pneumoconiosis, *Eur. Respir. J.*, **1995**; 8: 834–42.
125. Piguet, P. F., Grau, G. E., and Vassalli, P., Subcutaneous perfusion of tumor necrosis factor induces local proliferation of fibroblasts, capillaries and epidermal cells, or massive tissue necrosis, *Am. J. Pathol.*, **1990**; 136: 103–13.
126. Sime, P. J., Marr, R. A., Gauldie, D., Xing, Z., Hewlett, B. R., Graham, F. L., and Gauldie, J., Transfer of tumor necrosis factor- α to rat lung induces severe pulmonary inflammation and patchy interstitial fibrogenesis with induction of transforming growth factor- β 1 and myofibroblasts, *Am. J. Pathol.*, **1998**; 153: 825–32.
127. White, A. M., Yoshimura, T., Smith, A. W., Westwick, J., and Watson, M. L., Airway inflammation induced by recombinant guinea pig tumor necrosis factor- α , *Am. J. Physiology*, **1997**; 273: 524–30.
128. Sueoka, N., Sueoka, E., Miyazaki, Y., Okabe, S., Kurosumi, M., Takayama, S., and Fujiki, H., Molecular pathogenesis of interstitial pneumonitis with TNF- α transgenic mice, *Cytokine*, **1998**; 10: 124–31.
129. Wong, G. H. W. and Goeddel, D. V., Induction of manganous superoxide dismutase by tumor necrosis factor: possible protective mechanism, *Science*, **1988**; 242: 941–4.
130. Dubaybo, B. A., Role of tumor necrosis factor- in regulating fibrotic lung repair, *Res. Commun. Mol. Pathol. Pharmacol.*, **1998**; 101: 69–79.
131. Ulrich, T., Yin, S., Remick, D., Russell, D., Eisenberg, S., and Kohno, T., Intratracheal administration of endotoxin and cytokines. IV. The soluble tumor necrosis factor receptor type I inhibits acute inflammation, *Am. J. Pathol.*, **1993**; 142: 1335–8.
132. Piguet, P. F., Collart, M. A., Grau, G. E., Sappino, A. P., and Vassalli, P., Requirement of tumor necrosis factor for development of silica-induced pulmonary fibrosis, *Nature*, **1990**; 344: 245–7.
133. Liu, J. Y., Brass, D. M., Hoyle, G. W., and Brody, A. R., TNF-alpha receptor knockout mice are protected from the fibroproliferative effects of inhaled asbestos fibers, *Am. J. Pathol.*, **1998**; 153: 1839–47.

134. Lander, H. M., An essential role for free radicals and derived species in signal transduction, *FASEB J.*, **1997**; 11: 118–24.
135. Baeuerle, P. A. and Baltimore, D., The physiology of the NF- κ B transcription factor, in *Molecular Aspects of Cellular Regulation: Hormonal Control regulation of Gene Transcription*, Cohen, P. and Foulkes J. G., Eds., Elsevier, Amsterdam, **1991**, 409–32.
136. Beutler, B. and van Huffel, C., An evolutionary and functional approach to the TNF receptor/ligand family, *Annal. N.Y. Acad. Sci.*, **1994**; 730: 118–33
137. Nickoloff, B., Karabin, G., Barker, J., Griffiths, C., Sarma, V., Mitra, R., Elder, J., Kunkel, S., and Dixit, V., Cytokine networks: immunobiology surfaces, *J. NIH Res.*, **1991**; 3: 71–3.
138. Kupper, T. S., Production of cytokines by epithelial tissues, *Am. J. Dermatopathol.*, **1989**; 11: 69–73.
139. Schroder, J. M., Cytokine networks in the skin, *J. Invest. Dermatol.*, **1995**; 105: 20–4.
140. Piguet, P. F., Keratinocyte-derived tumor necrosis factor and the physiopathology of the skin, *Springer Semin. Immunopathol.*, **1992**; 13: 345–54.
141. Holliday, M. R., Corsini, E., Smith, S., Basketter, D. A., Dearman, R. J., and Kimber, I., Differential induction of cutaneous TNF α and IL-6 by topically applied chemicals, *Am. J. Contact Derm.*, **1997**; 8: 158–64.
142. Moller, H., Ohlsson, K., Linder, C., Bjorkner, B., and Bruze, M., Cytokines and acute phase reactants during flare-up of contact allergy to gold, *Am. J. Contact Derm.*, **1998**; 9: 15–22.
143. Strickland, I., Rhodes, L. E., Flanagan, B. F., and Friedman, P. S., TNF and IL-8 are upregulated in the epidermis of normal human skin after UVB exposure: correlation with neutrophil accumulation and E-selectin expression, *J. Invest. Dermatol.*, **1997**; 108: 763–8.
144. Corsini, E., Terzoli, A., Bruccoleri, A., Marinovich, M., and Galli, C. L., Induction of TNF *in vivo* by a skin irritant, Tributyltin, through activation of transcription factors: Its pharmacological modulation by anti-inflammatory drugs, *J. Invest. Dermatol.*, **1997**; 108: 892–6.
145. Kondo, S. and Sauder, D. N., Tumor necrosis factor receptor type 1 (p55) is a main mediator for TNF- α -induced skin inflammation, *Eur. J. Immunol.*, **1997**; 27: 1713–8.
146. Luster, M. I., Wilmer, J. L., Germolec, D. R., Spalding, J., Yoshida, T., Gaido, K., Simeonova, P. P., Burleson, F. G., and Bruccoleri, A., Role of keratinocyte-derived cytokines in chemical toxicity, *Toxicol. Lett.*, **1995**; 82/83: 471–6.
147. Wood, L. C., Stadler, A. K., Liou, A., Campbell, I. L., Grunfeld, C., Elias, P. M., and Feingold, K. R., Barrier disruption increases gene expression of cytokines and the 55-kDa TNF receptor in murine skin, *Exp. Dermatol.*, **1997**; 6: 98–104.
148. Piguet, P. F., Grau, G. E., Hauser, C., and Vassalli, P., Tumor necrosis factor is a critical mediator in hapten-induced irritant and contact hypersensitivity reactions, *J. Exp. Med.*, **1991**; 173: 673–9.
149. Lewis, R. W., McCall, J. C., Botham, P. A., and Kimber, I., Investigation of TNF α release as a measure of skin irritancy, *Toxicol. In Vitro*, **1993**; 7: 393–5
150. Wang, B., Fujisawa, H., Zhuang, L., Kondo, S., Shivji, G., Kim, C. S., Mak, T. W., and Sauder, D. N., Depressed Langerhans cell migration and reduced contact hypersensitivity response in mice lacking TNF receptor p75, *J. Immunol.*, **1997**; 159: 6148–55.
151. Hoefakker, S., Caubo, M., Van't Erve, E. H. M., and Roggeveen, M. H., Goersma, W. J. A., Van Joost, T., Notten, W., and Claassen, E., *In vivo* cytokine profiles in allergic and irritant contact dermatitis, *Contact Dermatitis*, **1997**; 33: 258–66.
152. Amar, S., Van Dyke, T. E., Eugster, H.-P., Schultze, N., Koebel, P., and Bleuthmann, H., Tumor necrosis factor-induced cutaneous necrosis is mediated by TNF receptor 1. *J. Inflamm.*, **1995/6**; 47:180–9.
153. Cumberbatch, M. and Kimber, T., TNF α is required for the accumulation of dendritic cells in draining lymph nodes and for optimal contact sensitization, *Immunology*, **1995**; 84: 31–5.

154. Hubner, G., Brauchle, M., Smola, H., Madlener, M., Fassler, R., and Werner, S., Differential regulation of pro-inflammatory cytokines during wound healing in normal and glucocorticoid-treated mice, *Cytokine*, **1996**; 8: 548–56.
155. Hart, P. H., Grimbaldston, M. A., Swift, G. J., Sedgwick, J. D., Korner, H., and Finlay-Jones, J. J., TNF modulates susceptibility to UVB-induced systemic immunomodulation in mice by effects on dermal mast cell prevalence, *Eur. J. Immunol.*, **1998**; 28: 2893–901.
156. Noonan, F. P., Bucana, C., Sauder, D. N., and DeFabo, E. C., Mechanism of systemic immune suppression by UV irradiation *in vivo*, *J. Immunol.*, **1984**; 132: 2408–16.
157. Wilson, A. G., de Vries, N., Poicot, F., di Giovine, F. S., van der Putte, L. B. A., and Duff, G. W., An allelic polymorphism within the human tumor necrosis factor promoter region is strongly associated with HLA A1, B8 and DR3 alleles, *J. Exp. Med.*, **1993**; 177: 557–60.
158. D'Alfonso, S. and Momigliano, R. P., A polymorphic variation in a putative regulation box of the TNF promoter region, *Immunogenetics*, **1994**; 29: 150–4.
159. Arias, A. I., Giles, B., Eiermann, T. H., Sterry, W., and Pandey, J. P., Tumor necrosis factor- α gene polymorphism in psoriasis, *Exp. Clin. Immunogenet*, **1997**; 14: 118–22.
160. Korman, K. S. and diGiovine, F. S., Genetic variations in cytokine expression: A risk factor for severity of adult periodontitis, *Annal. Periodontol.*, **1998**; 3: 327–38.
161. Zhai, R., Jetten, M., Schins, R. P. F., Franssen, H., and Borm, P. J. A., Polymorphisms in the promoter of the tumor necrosis factor- α gene in coal miners, *Am. J. Industr. Med.*, **1998**; 34: 318–24.
162. Seitzer, U., Swider, C., Stuber, F., Suchnicki, K., Lange, A., Richter, E., Zabel, P., Muller-Quernheim, J., Flad, H. D., and Gerdes, J., Tumor necrosis factor promoter gene polymorphism in sarcoidosis, *Cytokine*, **1997**; 9: 787–90.