

CAD MAPK site reversed the response of effectors resulting in a decreased *de novo*

DNA damage induced G1 cell cycle

Zhiyuan Shen. Molecular Genetics and New Mexico School of Medicine, 915

identified as a BRCA2 and Cip1/p21 to a highly conserved domain of BRCA2 domain of the CDK-inhibitor p21/Cip1. The BCCIP α enhances the inhibitory H1 kinase activity of CDK2, and that of certain tumor cells. An alternatively so expressed in various human tissues.

that overexpression of BCCIP inhibits cell cycle progression, suggesting a potential role of BCCIP in cell cycle. To further test this, RNA interference shutdown the expression of BCCIP α and parental HT1080 cell line underwent the radiation. However, cells with BCCIP were able to arrest in G₁ phase after the same induction of p21/Cip1 by DNA damage was not interfered with RNAi. These results suggest BCCIP in DNA damage-induced G₁-S

Regulation by dibutyryl cyclic AMP in SH-epithelial cell line

Yun-Il Lee, So-Young Kim, Yong-Sung Kim. Pathology and Molecular Biology, College of Medicine, Yonsei University, Yongon-Dong 28, Seoul, Korea, Republic of

cAMP signaling pathway can determine cell proliferation, differentiation, senescence and regulation of regulating cell fate by cAMP are not clear or induction of different cellular response by cAMP is poorly understood. Therefore, to assess the effect of cAMP on cellular cycle changes in expression and activity of cyclin D1 in progression in human neuroblastoma SH-epithelial

treated with cAMP analogue, dibutyryl cAMP. The synthesis and distribution of cells in S phase and phosphorylation of p34 were increased without changes in CDK2 expression of CDK2 related cyclins, cyclin A and B1. The expression of p27Kip1 was decreased and p27Kip1 was stimulated by treatment with dibutyryl cAMP. When the cells were treated with p34 inhibitor, the degradation of p27kip1 was inhibited. It suggested that degradation of p27kip1 is regulated via proteasome.

These results suggest that cAMP promotes cell cycle progression in neuroblastoma SH-SY5Y cells. It increases CDK2 expression and p34 protein, expression of cyclin A and B1 protein expression by stimulating the p34 proteasome.

621.9

Loss of CBF-mediated transcription by deletion of the Cbf-b/Nfy-a gene results in early embryonic lethality, inhibition of cell growth and apoptosis

Anuradha Bhattacharya, Benoit de Crombrugge, Richard Behringer, Sankar Maity. Molecular Genetics, UT MDACC, 1515 Holcombe Blvd., Houston, TX 77030

To understand the physiological function of the mammalian heterotrimeric CCAAT binding factor CBF, in vivo, we have generated a conditional Cbf-b mouse mutant by introducing loxP sites in the murine Cbf-b gene. Controlled expression of cre recombinase deletes the gene, which leads to loss of CBF-mediated transcription, since CBF has no activity in the absence of the CBF-B subunit. Homozygous deletion of Cbf-b in fertilized oocytes causes early mouse embryonic lethality. Similarly, homozygous deletion of Cbf-b in primary cultures of mouse embryonic fibroblast (MEFs) results in inhibition of cell proliferation and later to apoptosis. Cell cycle analysis of the MEFs indicated that Cbf-b null cells show an almost complete block in entry into S-phase and also an arrest in mitosis. An increase in caspase-2 mRNA precedes apoptosis, suggesting that the induction of apoptosis may be caused by activation of caspase-2. Altogether our results demonstrate that: (1) CBF is required for cell cycle progression; (2) CBF is a survival factor in mammalian cells and (3) apoptosis is induced in the absence of CBF-mediated transcription. Ongoing work is testing whether loss of CBF activity inhibits growth of tumors in mice.

621.10

The Role of Endothelial Cells As An Intracellular Mechanism Involved In Antiphospholipid Antibody Upregulation

Monica Charlene Lindsey¹, Sylvia Pierangeli². ¹Biochemistry, Clark Atlanta University, P.O. Box 92637, Atlanta, GA 30314, ²Microbiology/Biochemistry/Immunology, Morehouse School of Medicine, Atlanta, GA

The association of antiphospholipid antibodies with thrombosis in patients with Antiphospholipid Syndrome (APS) is well documented in humans and in animal studies. However, the mechanism(s) by which these antibodies induce thrombosis is the subject of much current study. Previous studies have shown that aPL antibodies upregulate several endothelial cells (ECs) cellular adhesion molecules (CAMs). What is uncertain is the molecular and intracellular events that are induced by aPL interaction with ECs leading to expression of CAM. Although there is a wealth of data suggesting that aPL antibodies activate ECs in vitro and in vivo and they enhance thrombosis, it is unclear what the relative roles are of the ECs adhesion molecules: intracellular cell adhesion molecules-1 (ICAM-1) and vascular cell adhesion monocytes in these processes. We hypothesized that APL antibodies upregulate CAM's on ECs, causing activation. To examine these questions, this investigates the molecular and intracellular events of EC activation mediated by aPL. In this study, we examine the mechanism of aPL by assessing the following areas:

whether aPL-mediated upregulation of CAMs on ECs involves de novo protein synthesis, cytoskeleton movements, and mitogen-activated protein kinase C activation. To achieve these aims, experiments will be conducted through such methods as cell isolation and culture, ELISA, CAM expression, western blot, northern blot, and other molecular biology techniques.

621.11

GADD45 is an Oxidative Stress Response Protein

Jacquelyn Jo Bower, Stephen S. Leonard, Yong Qian, Fei Chen, Xianglin Shi. Pathology and Physiology Research Branch, National Institute for Occupational Safety and Health, 1095 Willowdale Road, Morgantown, West Virginia 26505

Cell cycle arrest is a major mechanism for the repair of DNA damage caused by a variety of insults including UV-induced damage, radiation, and exposure to some transition metals. GADD45 (growth arrest and DNA damage-inducible protein 45) is one of many proteins involved in the regulation of cell cycle arrest. This protein has been shown to interfere with the interaction between the cyclin dependent kinase, cdc2, and Cyclin B1, causing cell cycle arrest at the G₂/M phase. In this report we investigated the involvement of reactive oxygen species

(ROS) in Cr(VI)- and As(III)-induced GADD45 expression in the human airway epithelial cell line, BEAS2B. Cr(VI) induces ROS generation as demonstrated by ESR and confocal microscopy using specific fluorescent dyes for superoxide ($O_2^{\cdot-}$) and H_2O_2 , the precursor of hydroxyl radical ($\cdot OH$). Western blots indicated an induction of GADD45 by Cr(VI) and As(III), which is correlated with the generation of ROS. Here we report that GADD45 is induced in response to ROS generation, particularly $O_2^{\cdot-}$ and $\cdot OH$. Furthermore, the induction of GADD45 by Cr(VI) was suppressed by the over-expression of superoxide dismutase (SOD) and glutathione peroxidase (GPx). Since the overexpression of SOD and GPx substantially decreased GADD45 induction by Cr(VI), we concluded that GADD45 is an oxidative stress response protein.

621.12

Retinoblastoma dependent and independent functions of epidermal keratin K10 in transgenic mice

Mirentxu Santos¹, Sergio Ruiz¹, Carmen Segrelles¹, Marc Vooijs², Hugo Leis¹, Jose Luis Jorcano¹, Jesus M. Paramio¹. ¹Epithelial Damage, Repair and Tissue Engineering, C.I.E.M.A.T., Avda. Complutense 22, MADRID, MADRID 28040 Spain, ²Molecular Biology and Pharmacology, Washington University School of Medicine, Saint Louis, MI

Our previous work in vitro and in transgenic mice, shows that overexpression of a differentiation-associated, keratin K10, inhibits keratinocyte proliferation by sequestering signalling molecules (Akt, and an atypical PKC), thereby altering pRb phosphorylation (Paramio et al., Moll.Cell. Biol 21:7449-7459, 2001; Santos et al., J.Biol. Chem. 277:19122-19130, 2002).

To analyze the specific functions and the relationships between K10 and Retinoblastoma (Rb) in epidermis in vivo, we have crossed our transgenic mice in which human K10 is ectopically expressed in the basal layer of epidermis (bK5hK10 mice) with the epidermal-specific Rb knock out mice (Rb^{f/f} K14 cre mice).

Epidermis in K5K10 mice is hypoplastic, highly confluent and BrdU incorporation is decreased while the skin of Rb cre shows moderate hyperplasia and hyperkeratosis, with increased levels of BrdU incorporation.

The dramatic decrease in epidermal proliferation observed in K5K10 mice, is restored in the Rb^{f/f} K14 cre bK5hK10 mice where a great hyperplasia, hyperkeratosis and elevated levels of BrdU incorporation are observed.

These results suggest that the control of epithelial proliferation in the skin in vivo, elicited by K10, needs Rb functional.

621.13

Ethanol causes p53-dependent cell cycle arrest and apoptosis of SK-N-SH neuroblastoma cells.

Yongil Kwon, Myungae Bae, Wonho Kim, Byoungjoon Song, LMBB/NIAAA/NIH, Park 5 Bldg, room 425, 12420 Parklawn drive, Rockville, Maryland 20892-8110

Chronic alcohol drinking can damage many organs including liver, pancreas, and brain. However, the mechanism of cell or organ damage is still poorly understood. Therefore, we hypothesized that changes in the early signaling cascades are critically important in ethanol-mediated cell death. In this study, we investigate the role of the MAP kinases during ethanol-induced damage to SK-N-SH neuroblastoma cells. Ethanol caused time- and dose-dependent cell death in SK-N-SH cells. Ethanol increased c-Jun N-terminal protein kinase (JNK) activity in a time- and concentration dependent manner. Within 15 min after ethanol exposure, JNK activity increased and the elevated JNK activity persisted until 16 h after exposure to ethanol. In contrast, p38 kinase activity was transiently increased between 15 min and 4 h after ethanol treatment before it returned to control level. The activation of JNK and p38 kinase was important in cell death since treatment with a respective inhibitor of JNK or p38 kinase significantly reduced the rate of ethanol-induced cell death. Ethanol-induced cell death was accompanied by increased cytochrome C release and caspase 3 activity observed at 12 h in a successive manner. In contrast, the level of anti-apoptotic Bcl-2 protein did not change. In addition to JNK activation, ethanol also increased phosphorylation of p53, which led to accumulation of p53 protein at 1 h after ethanol exposure. The p53 activation was followed by increase in

p21 tumor suppressor protein accompanied by a gradual decrease in phospho-Rb protein. Taken together, these results strongly indicate that ethanol causes apoptosis of SK-N-SH neuroblastoma cells by stimulating p53-related cell cycle arrest and apoptosis possibly mediated through activation of the JNK-related cell death pathway.

COMPLEX BIOLOGICAL OXIDATIONS (622.1-622.11)

622.1

Cloning and expression of ribonucleotide reductase from *Archaeoglobus fulgidus*

Hector Hugo Hernandez¹, Bernard A. Brown II², Catherine L. Drennan¹. ¹Chemistry, Massachusetts Institute of Technology, 56-546, 77 Massachusetts Ave, Cambridge, MA 02139, ²Chemistry, Wake Forest University, Winston-Salem, NC

Ribonucleotide reductases (RNRs) catalyze the conversion of nucleotides to deoxynucleotides in all organisms thus playing a central role in DNA synthesis and repair. RNRs are important targets in biotechnology because drugs that inhibit these enzymes would find ready use as anticancer agents. Although all RNRs catalyze the same reaction, their essential metal cofactors have not been evolutionarily conserved. There are currently three classes of RNRs that are distinguished by their enzymatic cofactors. Class I contains a diferric cluster and an observable tyrosyl radical, Class II contains coenzyme B12 (adenosylcobalamin; AdoCbl), and Class III contains a FeS cluster and an observable glycol radical. We have identified the *Archaeoglobus fulgidus* gene encoding the coenzyme B12 dependent ribonucleotide reductase (afRNR) protein by sequence homology, and this gene has been amplified and cloned. The gene was sequenced and expression of recombinant (His)₆-tagged RNR protein was confirmed by SDS-PAGE and N-terminal sequencing. Progress is being made in the biochemical characterization of this enzyme.

622.2

Engineering Terpene Synthases for Novel Product Specificity

Bryan Thomas Greenhagen¹, Yuxin Zhao², Paul O'Maille³, Joe Noel¹, Robert M Coates², Joe Chappell¹. ¹University of Kentucky, N221W Ag. Sci. Ctr. N, Lexington, KY 40546, ²University of Illinois, Urbana, IL, ³Salk Institute, La Jolla, CA

Terpene cyclases catalyze the stereospecific conversion of simple linear isoprenoid polymers into cyclic terpenes. Engineered chimeras of closely related terpene cyclases (5-epi-aristolochene synthase (EAS) and premaradiene synthase (HPS)) which catalyze reaction pathways that differ only in a penultimate carbon bond migration and terminal proton elimination were subjected to a deductive analysis for structure-function relationships. We developed contact maps based on a well resolved substrate analog positioned within the active site of crystallized EAS enabling us to make mechanistic inferences about residues which might control the regioselectivity of proton elimination. A double mutant, EAS-T402S/V516L, catalyzed formation of 4-epieremophilene, the double bond regioisomer of the native EAS product as the dominant reaction product. These mutations essentially reposition a methyl group in the second tier of residues surrounding the active site. Significantly, this new enzyme, 4-epieremophilene synthase, retains wild type cyclase kinetic parameters, a factor of great importance in mechanistic interpretation of the current results and in the practical application of new terpene cyclases.

622.3

Rat cyt19 encodes arsenic methyltransferase and reductase functions

Stephen Brian Waters¹, Miroslav Styb², David Thomas³. ¹University of North Carolina-Chapel Hill, Manning Drive, Chapel Hill, North Carolina 27599, ²Pediatrics, University of North Carolina-Chapel Hill, Chapel Hill, North Carolina, ³National Health and Environmental Effects Research Laboratory, US Environmental Protection Agency, Research Triangle Park, North Carolina

In many species, biomethylation of inorganic arsenic (iAs) yields methyl- and dimethylarsenic. This enzymatically-catalyzed process involves the reduction of arsenic from the pentavalent to trivalent

Program # 621.11

GADD45 is an Oxidative Stress Response Protein

Jacquelyn Jo Bower, Stephen S. Leonard, Yong Qian, Fei Chen, Xianglin Shi. Pathology and Physiology Research Branch, National Institute for Occupational Safety and Health, 1095 Willowdale Road, Morgantown, West Virginia 26505

Cell cycle arrest is a major mechanism for the repair of DNA damage caused by a variety of insults including UV-induced damage, radiation, and exposure to some transition metals. GADD45 (growth arrest and DNA damage-inducible protein 45) is one of many proteins involved in the regulation of cell cycle arrest. This protein has been shown to interfere with the interaction between the cyclin dependent kinase, cdc2, and Cyclin B1, causing cell cycle arrest at the G2/M phase. In this report we investigated the involvement of reactive oxygen species (ROS) in Cr(VI)- and As(III)-induced GADD45 expression in the human airway epithelial cell line, BEAS2B. Cr(VI) induces ROS generation as demonstrated by ESR and confocal microscopy using specific fluorescent dyes for superoxide ($O_2^{\cdot -}$) and H_2O_2 , the precursor of hydroxyl radical ($\cdot OH$). Western blots indicated an induction of GADD45 by Cr(VI) and As(III), which is correlated with the generation of ROS. Here we report that GADD45 is induced in response to ROS generation, particularly $O_2^{\cdot -}$ and $\cdot OH$. Furthermore, the induction of GADD45 by Cr(VI) was suppressed by the over-expression of superoxide dismutase (SOD) and glutathione peroxidase (GPx). Since the overexpression of SOD and GPx substantially decreased GADD45 induction by Cr(VI), we concluded that GADD45 is an oxidative stress response protein.

An Annual Meeting of Professional Research Scientists

Experimental Biology 2003[®]
San Diego, California

April 11 – April 15, 2003

ABSTRACTS 456.1 – 886.2

PART II

The American Physiological Society
American Society for Biochemistry and
Molecular Biology
American Society for Pharmacology and
Experimental Therapeutics
American Society for Investigative Pathology
American Society for Nutritional Sciences
American Association of Anatomists

American Federation for Medical Research
The American Society for Clinical Nutrition
Latin-American Association of Physiological
Sciences
The Biomedical Engineering Society
Cajal Club
The Histochemical Society
International Society for Stereology
The Microcirculatory Society
North American Vascular Biology Organization
Society for Experimental Biology and Medicine
Society for International Nutrition Research

The abstracts on pages A755–A1381 were prepared by the authors and printed by photo-offset without change. Abstracts are not subject to scientific review; therefore, the scientific validity of the results reported is the responsibility of the authors and sponsors. **Accuracy, form of citation, designation of materials,**

acknowledgment of coauthors and of grant support, terminology, nomenclature, and the like, remain the responsibility of the authors and sponsors. The appearance of an abstract in this issue does not necessarily imply future publication of a scientific paper.

Editor-in-Chief
Vincent T. Marchesi

Editorial Associate
Claire S. Veilleux

Associate Editors
Edward J. Goetzl
Yusuf A. Hannun
Joseph A. Madri

Publications and Communications Committee
Alan G. Goodridge (ASBMB)
Sandra R. Wolman (ASIP)
Susan S. Percival (ASNS)
Eleanor S. Metcalf (AAI)
Suse B. Broyde (BPS)
Donald A. Fischman Chair (AAA)
Mark A. Hermodson (Protein)
Marc K. Drezner (ASBMR)
Stephen J. Weiss (ASCI)
Peter H. Byers (ASHG)
Thomas D. Sargent (SDB)
Sidney H. Golub (non-voting)

Ex Officio
Steven L. Teitelbaum
Vincent T. Marchesi
FASEB Society Executive Officers

Director, FASEB Office of Publications
Nancy J. Rodnan

Senior Editor
Lynn Willis

Copy Editor
Kendall Sites

FJ Express Production Coordinator
Mary Kiorpes Eig

Marketing/Advertising Manager
Jennifer L. Pesanelli

Advertising Account Manager
Susan J. Mergenhagen
301-634-7103
Fax: 301-634-7153

Subscription Manager
Eleanor B. Peebles
301-634-7029
Fax: 301-634-7099

Editorial Board

Kari Alitalo
Mina J. Bissell
Meredith Bond
David A. Brenner
H. Franklin Bunn
George H. Caughey
Pierre Chambon
Thomas O. Daniel
Balz Frei
Martin E. Hemler
Timothy Hla
Tadamitsu Kishimoto
Hynda K. Kleinman
John S. Lazo
John J. Lemasters
George M. Martin

Mark P. Mattson
Linda C. McPhail
Hideaki Nagase
Lina M. Obeid
Jordan S. Pober
Robert E. Pollack
Stanley B. Prusiner
Russel J. Reiter
Noel R. Rose
Charles N. Serhan
William C. Sessa
Solomon H. Snyder
Andrew P. Somlyo
William G. Stetler-Stevenson
Makoto M. Taketo
George D. Yancopoulos

Editor-in-Chief
Boyer Center for Molecular Medicine
Yale University School of Medicine
295 Congress Avenue, (P.O. Box 9812)
New Haven, CT 06519-1418, USA
Phone: 203-737-2334 Fax: 203-737-2267
email: vincent.marchesi@yale.edu

Editorial Office
Boyer Center for Molecular Medicine
Yale University School of Medicine
295 Congress Avenue (P.O. Box 9812)
New Haven, CT 06519-1418, USA
Phone: 203-737-2334 Fax: 203-737-2267
Email: faseb@yale.edu

Publications Office
The FASEB Journal
9650 Rockville Pike
Bethesda, MD 20814-3998, USA
Phone: 301-634-7100
Fax: 301-634-7809
Email: ksites@faseb.org

The FASEB Journal (ISSN-0892-6638) is published 15 times a year (monthly except three times in March and two times in April) by the Federation of American Societies for Experimental Biology, 9650 Rockville Pike, Bethesda, MD 20814-3998, U.S.A. Copyright © 2003 by FASEB. All rights reserved. Requests for copyrighted material should be made in writing to the Copyright Clearance Center, Inc., 222 Rosewood Drive, Danvers, MA 01923, USA. Periodicals postage paid at Bethesda, Maryland, and at additional mailing offices. **Postmaster:** Send change of address to *The FASEB Journal*, 9650 Rockville Pike, Bethesda, MD 20814-3998. The views expressed in articles are those of the authors and not necessarily those of the Federation. Send manuscripts and proposals to the Editor-in-Chief. See **Instructions for Authors** online at <http://www.fasebj.org>.

2003	United States	Canada/Mexico	Rest of World	Online Only
INSTITUTION	\$648.	\$672.	\$710.	\$648.
MEMBER	\$93.	\$113.	\$146.	N/A
INDIVIDUAL	\$159.	\$179.	\$212.	N/A
STUDENT	\$45.	\$65.	\$97.	N/A

Corporate members of FASEB

The American Physiological Society • American Society for Biochemistry and Molecular Biology • American Society for Pharmacology and Experimental Therapeutics
American Society for Investigative Pathology • American Society for Nutritional Sciences • The American Association of Immunologists • Biophysical Society
American Association of Anatomists • The Protein Society • The American Society for Bone and Mineral Research • American Society for Clinical Investigation
The Endocrine Society • The American Society of Human Genetics • Society for Developmental Biology

THE FASEB JOURNAL

Volume 17, Number 5

March 17, 2003

ABSTRACTS

(continued from Part I)

SUNDAY MORNING

APRIL 13, 2003

American Society for Nutritional Sciences

Carotenoids II	A755
Botanicals and Dietary Supplements I	A760
Functional Foods II	A764
Food Composition and Databases	A765
Diet in Reproductive Cancers I	A768

American Association of Anatomists

Teaching Innovations I	A774
Growth and Development	A775
Growth and Development: Development of the Myotone	A777
Imaging and Microscopy	A784
Anatomical Variations	A786
Animal Models of Disease	A787
Anatomical Form and Function	A789

The Histochemical Society

Signal Transduction: Histochemical Techniques for Visualizing Individual Pathways <i>In Vitro</i> and <i>In Vivo</i>	A772
--	------

International Society for Stereology

Sampling and Statistical Applications in Stereology .	A771
The Power of Stereological Measurements for Modeling Function	A773

SUNDAY AFTERNOON

APRIL 13, 2003

American Society for Biochemistry and Molecular Biology

Morphagens and Development	A790
Biological Functions of Glycosphingolipids and Glycosylphosphatidylinositols	A790
Lipid Enzymes—Structure and Function	A790
Fundamental and Emerging Issues in Enzymatic Catalysis	A791
Novel Mechanisms for Insulin Resistance	A791

Signaling Targets for Drug Therapies	A791
Protein Folding and Unfolding	A792
Replication, Recombination and Repair	A792
Protein Unfolding and Refolding on Membranes	A792
Education/Training of Biomedical Scientists Including the Howard K. Schachman Award in Honor of Ruth L. Kirschstein	A793
Schering-Plough Scientific Achievement Award Lecture	A793
Plenary Lecture	A793

American Society for Investigative Pathology

Infection and Immunity	A793
Leukocyte-Endothelial Cell Interactions I	A796
Apoptosis	A798

North American Vascular Biology Organization

Angiogenesis and Vasculogenesis	A801
Vascular Responses	A803

American Society for Nutritional Sciences

Changing Dietary Intakes: Intervention, Choice, and Environment	A805
Conjugated Linoleic Acid: Effects on Metabolism	A807
Food Intake Regulation II	A808
Protein Metabolism II	A810

American Association of Anatomists

Undergraduate and Medical Education Together in the Classroom	A812
R. R. Bensley Award Lecture in Cell Biology	A812
Intraflagellar Transport and Human Health	A813
Growth and Development: Development of the Myotone	A814
AAA Keynote Lecture	A816

Cajal Club

Cajal Club Presidential Symposium: Neurobiology of Rehabilitation	A812
Pinckney J. Harman Memorial Lecture	A815