

30.3 Bladder Cancer

Avima M Ruder, Tania Carreón, Elizabeth M Ward, Paul A Schulte, William Halperin

It is well documented that the etiology of bladder cancer involves environmental risk factors. Occupational risks may account for 21–27% of bladder cancers among men in the United States,^{1,2} an estimated 40,000 cases in 2001,³ and 11% among the estimated 15,000 cases in women in 2001.^{3,4}

Occupational exposure to aromatic amines has been known to cause bladder cancer since Rehn identified the first few cases in workers in the new organic chemical industry in 1895.⁵ Since that time, numerous occupations and specific substances have been associated with an increased risk of bladder cancer (Tables 30.3.1 and 30.3.2). Beyond the certainties that specific aromatic amines have been demonstrated to be human occupational bladder carcinogens and that a broad range of occupations are at risk of bladder cancer, a well-informed approach to the prevention and management of bladder cancer depends on appreciating various controversies involved in its primary and secondary prevention and treatment. The reader is referred to a report of a national conference held in 1989⁶ to delineate these issues. Many of these controversies, such as the relevance to human bladder cancer of findings from animal studies, the line between benign and malignant tumors, the appropriate screening regimen for workers exposed to bladder carcinogens, and whether early detection is worthwhile, remain relevant more than a decade later.

In addition to incidence and survival differences by social class, race, and gender,⁷ new developments in understanding inherited risk factors, such as acetylator status and intermediate biomarkers, influence understanding of etiology, prevention and management.

ETIOLOGY

Epidemiology

Bladder cancer is the eighth most common neoplasm worldwide. An estimated 261,000 new cases of bladder cancer occur each year, with four-fifths of the cases occurring in men.⁸ Incidence rates of bladder cancer vary about 10-fold, with higher rates in Western Europe and North America, and lower rates in Eastern Europe and several Asian countries.⁹

Cigarette smoking accounts for 47% of bladder cancer among men and 37% among women; smokers have twice the risk of non-smokers.³ The contributions of coffee drinking and alcohol consumption to bladder cancer are equivocal. The World Health Organization estimates that bladder cancer cases can be subdivided into two broad categories based on etiology. (1) Those caused by tobacco and industrial carcinogens are predominantly transitional cell carcinomas and are common in industrialized countries. (2) Those due to bilharzia, human papilloma virus, and schistosomiasis infection^{10,11} are more likely to be squamous cell carcinomas and are found chiefly in subtropical

and tropical countries. Thus, both types are, in principle, preventable.¹²

Bladder cancer is historically the neoplastic disease most strongly linked to occupational exposure to chemicals. Several occupations have been suspected to increase the risk of bladder cancer, but strong associations only exist for dye workers, aromatic amine manufacturing workers, leather workers, rubber workers, painters, truck drivers, and aluminum workers.¹³ Table 30.3.1 presents the epidemiologic studies of occupational associations. Occupational exposures to chemicals such as arylamines, polycyclic aromatic hydrocarbons and other industry-related agents may explain some of the risk associated with these occupations.¹⁴ Some of the relative risks for exposed workers are substantially higher than the two-fold increased risk for smokers. Table 30.3.2 includes specific chemicals or processes classified by IARC as carcinogenic to the human or canine bladder, combined with data on working populations potentially at risk. Over 200 chemicals have been confirmed as rodent bladder carcinogens; however, rodent bioassays do not appear to be a good model for human bladder cancer¹⁵ (see 'Animal studies' section below).

A relationship between bladder cancer and exposure to chemical dyes was first established in 1895.⁵ In the late 1930s, oral administration of the industrial arylamine 2-naphthylamine was shown to induce bladder cancer in dogs.¹⁶ In 1954, Case et al. reported a 20-fold excess of bladder cancer in arylamine-exposed individuals, compared to the general population of England and Wales.¹⁷ Since then, the most investigated bladder carcinogens have been 2-naphthylamine, benzidine, and 4-aminobiphenyl. Several studies have reported increased risks of bladder cancer in workers exposed to 2-naphthylamine and 4-aminobiphenyl, and have been reviewed elsewhere.¹⁸

One newly recognized factor that may have an occupational component is fluid intake. In a study of about 50,000 men, it was observed that drinking more fluids was associated with a significantly decreased rate of bladder cancer.¹⁹ These results are consistent with the urogenous contact hypothesis, which proposed that the level of DNA adducts to 4-aminobiphenyl (a carcinogenic amine) could be decreased by increased fluid intake.^{20,20a} Occupations that have exposure to carcinogens and limit the workers' ability to drink liquids could be at risk.

There is no known distinctive histologic feature for occupational bladder cancer. However, in some heavily exposed cohorts, cases regularly occur at ages 15 years younger than the median age at diagnosis of the general population,^{17,21} in which the rate of bladder cancer increases substantially with age. Usually, the interval from first exposure to onset of symptoms is decades long; however, occupational cases have occurred surprisingly early after exposure, which substantiates the argument that cases with only a few years of latency may be occupationally induced. It has been estimated that the latency for chemically induced bladder cancer ranges from 4 to 45 years.^{21,22}

Occupation /Industry	Study site, type, population	Increased morbidity relative risk	Increased mortality relative risk (95% CI)	Controlled for (95% CI)	Reference
Aluminum smelters (10 yrs)	Canada, NCC, men	3.9 (1.6-9.6)*		smoking	Armstrong et al., 1986 ⁵⁸
Armed services	Canada, CC, men	1.8 (1.2-2.7)			Howe et al., 1980 ⁵⁹
Auto workers (>10 yrs)	US, CC, African-American men	4.7 (1.7-10.7)		age, smoking	Silverman et al., 1989 ¹
Butchers	Sweden, CC, men	1.3 (1.0-1.6)			Malker et al., 1987 ⁶⁰
Carpenters	US, CC, men	11.1 (3.3-37.0)		current smokers	Schumacher et al., 1989 ⁶¹
Carpenters	US, CC, white men	1.4 (1.1-1.8)		smoking	Silverman et al., 1989 ²
Chemical mfg. workers	UK, CC, men	2.2 (1.7-3.0)			Boyko et al., 1985 ⁶²
Chemical mfg. workers	US, C, white men	2.6 (1.1-6.4)*		age	Schulte et al., 1985 ⁶³
Chemical workers†	US, C, African-American men	5.0 (2.2-11.3)*		age	
Chemical workers	Sweden, CC, men	1.3 (1.1-1.5)			Malker et al., 1987 ⁶⁰
Chemical workers	US, CC, white women	2.1 (0.9-5.1)*		smoking	Silverman et al., 1990 ⁴
Clerical workers	UK, CC, men	1.5 (1.1-1.9)			Cartwright, 1982 ⁶⁴
Clerical workers	China, CR, men	1.3 (1.0-1.7)			Zheng et al., 1992 ⁶⁵
Clerical workers (>10 yrs)	US, CC, African-American men	2.9 (1.2-6.2)		age, smoking	Silverman et al., 1989 ¹
Construction workers	US, CC, white men	1.6 (1.1-2.5)*		smoking	Silverman et al., 1989 ²
Construction workers	US, C, Latino men		PMR 1.6 (0.9-2.8)		Schultz & Loomis, 2000 ⁶⁶
Crafts workers	China, CR, women	1.2 (1.0-1.4)			Zheng et al., 1992 ⁶⁵
Dental technicians	Sweden, CC, men	2.5 (1.3-4.3)			Malker et al., 1987 ⁶⁰
Drivers	Argentina, CC, men	5.3 (2.3-12.2)			Iscoyich et al., 1987 ⁶⁷
Drivers	Denmark, CC	1.6 (1.1-2.3)*		sex, age, smoking	Jensen et al., 1987 ⁶⁸
Drivers†	US, CC, white men	1.2 (1.1-1.4)*		smoking	Silverman et al., 1989 ^{1,2}
Drivers, railroad	Germany, CC, men	3.0 (1.2-8.8)			Claude et al., 1988 ⁶⁹
Drivers, taxi	US, CC, white men	6.3 (1.6-29.3)*		age, smoking	Silverman et al., 1986 ⁸⁷
Drivers, truck	US, CC, white men	2.1 (1.4-4.4)*		age, smoking	Silverman et al., 1983 ⁸⁸
Drivers, truck	Germany, CC, men	1.8 (1.1-2.8)*			Claude et al., 1988 ⁶⁹
Drivers, truck	US, CC, white men	1.5 (1.1-2.0)		age, smoking	Silverman et al., 1986 ⁸⁷
Drivers, truck	UK, CC, men		PMR 2.0 (P<0.05)		Baxter & McDowall, 1986 ⁷⁰
Dry cleaners	US, CC, African-American men	2.8 (1.1-7.4)			Silverman et al., 1989 ¹
Dry cleaners	US, C, African-American men		5.1 (1.4-13.1)	age	Ruder et al., 2001 ⁷¹
Dye mfg. workers	China, C, men	11.1 (3.6-25.9)		non-smokers	Bi et al., 1992 ⁷²
		31.5 (20.4-46.4)		smokers	
			17.5 (7.5-34.5)	age	
Dye mfg. workers	UK, CC, men	3.5 (2.2-5.3)			Cartwright, 1982 ⁶⁴
Dye mfg. workers	UK, CC, men	2.6 (1.8-3.7)		age, smoking	Boyko et al., 1985 ⁶²
Dye mfg. workers	US, C, white men		5.2 (1.4-13.2)	age, exposure (azo dyes only)	Sathiakumar & Delzell, 2000 ⁷³
				smoking	
Dyers, printers, textile ind.	US, CC, white men	4.4 (1.2-16.8)			Silverman et al., 1989 ²
Dye workers†	Canada, CC, men	4.1 (2.9-5.5)		latency ≥ 8 yrs	Risch et al., 1988 ⁷⁴
Dye workers	Russia, C, men	3.9 (2.7-6.0)	2.8 (1.9-3.9)	age	Bulbulyan et al., 1995 ⁷⁵
Dye workers	Russia, C, women	8.6 (4.6-80.0)	3.1 (1.5-5.7)	age	Bulbulyan et al., 1995 ⁷⁵
Fabricators, assemblers, hand workers	US, C, African-American men		PMR 1.6 (0.9-2.9)		Schultz & Loomis, 2000 ⁶⁶
Fabricators, etc.	US, Latino men		PMR 2.8 (1.0-7.9)		
Farm workers (field crops, vegetables)	Europe, CC, women	1.8 (1.0-3.1) *		age, smoking	't Mannetje et al., 1999 ⁸⁵
Farm workers (nurseries)	Canada, CC, men	5.5 (1.2-51.1)			Howe et al., 1980 ⁵⁹
Food counter workers	US, white men	2.6 (1.4-5.1)		age, smoking	Schoenberg et al., 1984 ⁷⁶
Food counter workers	US, CC, white men	1.4 (0.9-2.1)*		smoking	Silverman et al., 1989 ²
Guards	Canada, CC, men	4.0 (1.3-16.4)			Howe et al., 1980 ⁵⁹
Guards	Germany, CC, men	3.5 (1.2-9.9)*			Claude et al., 1988 ⁶⁹
Janitors and cleaners	US, CC, white men	3.5 (1.6-7.7)		age, alcohol, smoking	Brownson et al., 1987 ⁷⁷
Janitors	Germany, CC, men	3.5 (1.2-9.9)*			Claude et al., 1988 ⁶⁹
Laborers in mfg. ind.	US, CC, white men	12.3 *			Silverman et al., 1989 ²
Laborers in metal ind.	US, African-American women		PMR 4.2 (1.1-16.8)		Schulz & Loomis, 2000 ⁶⁶
Leatherworkers	US, CC, men	6.3 (3.1-11.3)*			Decoufle, 1979 ⁷⁸
Leatherworkers	US, CC, women	4.4 (1.2-12.1)			Decoufle, 1979 ⁷⁸
Lumber jacks	US, CC, white men	1.3 (1.0-1.5)*		smoking	Silverman et al., 1989 ²
Machine operators, tenders	US, C, Latino men		PMR 1.7 (1.0-3.1)		Schultz & Loomis, 2000 ⁶⁶
Machinists	Sweden, CC, men	1.2 (1.1-1.3)			Malker et al., 1987 ⁶⁰

Table 30.3.1 Occupations associated with increased risk of bladder cancer

Occupation /Industry	Study site, type, population	Increased morbidity relative risk	Increased mortality relative risk (95% CI)	Controlled for (95% CI)	Reference
Machinists, metal	Canada, CC, men	2.7 (1.1–7.6)			Howe et al., 1980 ⁵⁹
Machinists†	US, CC, white men	1.3 (1.0–1.7)		smoking	Silverman et al., 1989 ²
Mail sorting clerks	Europe, CC, women	4.4 (1.0–19.5)		age, smoking	't Mannetje et al., 1999 ⁸⁵
Mfg. workers	China, CR, women	1.2 (1.0–1.5)			Zheng et al., 1992 ⁶⁵
Mfg. checkers, examiners, and inspectors	US, CC, white men	1.4 (1.1–1.8)*		smoking	Silverman et al., 1989 ²
Mfg. checkers	US, CC, white women	1.5 (1.0–2.3)		smoking	Silverman et al., 1990 ⁴
Mechanics	US, CC, white men	3.5 (1.4–9.1)		age, alcohol, smoking	Brownson et al., 1987 ⁷⁷
Mechanics	Spain, CC, men	1.8 (1.2–2.7)		smoking, other jobs	Gonzalez et al., 1989 ⁸⁹
Mechanics†	US, CC, white men	1.2 (1.0–1.4)		smoking	Silverman et al., 1989 ²
Mechanics, auto	US, CC, white men	10.2 (2.1–68.6)		smoking	Silverman et al., 1989 ²
Mechanics & repairers	US, C, Latino men		PMR 1.6 (0.7–3.7)		Schultz & Loomis, 2000 ⁶⁶
Metal workers†	US, CC, white men	1.2 (1.0–1.4)		smoking	Silverman et al., 1989 ²
Metal workers	China, CR, men	1.4 (1.0–2.0)			Zheng et al., 1992 ⁶⁵
Metal workers	Europe, CC, women	1.9 (1.1–3.6)*		age, smoking	't Mannetje et al., 1999 ⁸⁵
Metal workers	US, women	1.4 (1.0–1.9)		smoking	Silverman et al., 1990 ⁴
Mining machine ops	US, CC, white men	2.9 (1.1–7.5)		age, alcohol, smoking	Brownson et al., 1987 ⁷⁷
Mining workers	Germany, CC, men	2.0 (1.2–3.3)*			Claude et al., 1988 ⁶⁹
Painters	Switzerland, C, men	1.7 (1.0–2.7)	2.1 (1.0–3.9)		Guberan et al., 1988 ⁷⁹
Painters	Denmark, CC	2.5 (1.1–5.7)*		sex, age, smoking	Jensen et al., 1987 ⁶⁸
Painters, artistic	US, CC, C, men	2.5 (1.1–5.7)*	PMR 3.5 (2.1–5.7)	smoking (incidence)	Miller et al., 1986 ⁸⁰
Painters	US, CC, white men	1.5 (1.2–2.0)*		smoking	Silverman et al., 1989 ²
Painters	US, C, white men	1.2 (1.1–1.4)*		age	Steenland & Palu, 2000 ⁸¹
Paper pulp workers†	Sweden, CC, men	1.1 (1.0–1.3)			Malker et al., 1987 ⁶⁰
Pesticide mfg. workers	Europe, C, men	35 (14–66)		smoking	Popp et al., 1992 ⁸²
Petroleum processors†	US, CC, white men	2.4 (1.1–5.5)		smoking	Silverman et al., 1989 ²
Printers	Sweden, CC, men	1.2 (1.0–1.3)			Malker et al., 1987 ⁶⁰
Printers	UK, CC, men	3.1 (1.4–6.8)			Cartwright, 1982 ⁶⁴
Printers	US, CC, white men	2.1 (1.0–4.3)		smoking	Silverman et al., 1989 ²
Print machine operators	US, CC, white men	3.1 (1.1–8.9)		age, alcohol, smoking	Brownson et al., 1987 ⁷⁷
Produce graders, packers	US, CC, white men	3.2 (1.1–9.3)		smoking, educ.	Silverman et al., 1989 ²
Precision prod. workers	US, C, African–American women		PMR 1.8 (1.0–3.3)		Schultz & Loomis, 2000 ⁶⁶
Professional specialists	US, C, African–American men		PMR 1.4 (1.0–1.9)		Schultz & Loomis, 2000 ⁶⁶
Rubber & plastics workers	China, CR, men	2.1 (1.2–3.4)			Zheng et al., 1992 ⁶⁵
Rubber additive worker†	US, C	3.6 (1.9–6.3)			Ward et al., 1991 ⁹⁰
Rubber processing workers	US, CC, white women	4.5 (1.1–21.9)		smoking	Silverman et al., 1990 ⁴
Salespeople	US, C, Asian men		PMR 2.1 (0.8–5.6)		Schultz & Loomis, 2000 ⁶⁶
Salespeople, service, construction industries	US, CC, white men	2.2 (1.2–4.1)*		smoking	Silverman et al., 1989 ²
Salespeople	US, CC, white women	2.5 (1.0–6.0)*		smoking	Silverman et al., 1990 ⁴
Salespeople	Europe, CC, women	2.6 (1.0–6.9)		age, smoking	't Mannetje et al., 1999 ⁸⁵
		4.8 (1.2–18.7) (≥ 10 yrs)			
Services, personal	US, Asian women		PMR 5.3 (1.6–16.8)		Schulz & Loomis, 2000 ⁶⁶
Switchboard ops > 10 yrs	Europe, CC, women	8.1 (2.1–32.0)		age, smoking	't Mannetje et al., 1999 ⁸⁵
Tailors	Germany, CC, men	2.7 (1.1–6.6)			Claude et al., 1988 ⁶⁹
Tailors	Canada, CC, men	3.9 (1.3–14.2)		latency ≥ 8 yrs	Risch et al., 1988 ⁷⁴
Tailors, dressmakers	Europe, CC, women	1.4 (1.0–2.1)		smoking	't Mannetje et al., 1999
Technicians	US, C, African–American women		PMR 1.8 (0.8–3.8)		Schultz & Loomis, 2000 ⁶⁶

Table 30.3.1 (Cont'd) Occupations associated with increased risk of bladder cancer

Occupation /Industry	Study site, type, population	Increased morbidity relative risk	Increased mortality relative risk (95% CI)	Controlled for (95% CI)	Reference
Telephone & telegraph ops	US, CC, white men	1.9 (0.9–4.0)*		smoking	Silverman et al., 1989 ²
Textile workers	Spain, CC, women	6.4 (1.3–30.0)		smoking, other jobs	Gonzalez et al., 1989 ⁸⁹
Textile workers	Spain, CC, men	1.9 (1.1–3.1)		smoking, other jobs	Gonzalez et al., 1989 ⁸⁹
Textile workers	Denmark, CC	1.7 (1.1–2.4)		sex, age, smoking	Jensen et al., 1987 ⁶⁸
Textile workers†	Italy, CC, women	1.9 (0.9–4.2)			Maffi & Vineis, 1986 ⁸⁶
Tobacco processors	Europe, CC, women	3.1 (1.1–9.3)		age, smoking	't Mannetje et al., 1999 ⁸⁵
Turners (lathe operators)	UK, CC, men	1.5 (1.2–1.8)			Cartwright, 1982 ⁶⁴
Turners (lathe operators)	Germany, CC, men	2.3 (1.0–5.6)*			Claude et al., 1988 ⁶⁹
Upholsterers	Germany, CC, men	2.7 (1.1–6.6)			Claude et al., 1988 ⁶⁹
Weavers	Germany, CC, men	2.7 (1.1–6.6)			Claude et al., 1988 ⁶⁹
Weavers	Spain, CC, men	3.5 (1.3–9.3)		smoking, other jobs	Gonzalez et al., 1989 ⁸⁹
Weavers	Spain, CC, women	21.2 (1.5–298)		smoking, other jobs	Gonzalez et al., 1989 ⁸⁹
Weavers	US, CC, white men	3.5 (1.3–9.3)		smoking	Silverman et al., 1989 ²
Welders	Canada, CC, men	2.8 (1.1–8.8)			Howe et al., 1980 ⁵⁹
Welders, oxyacetylene	Italy, C, men		3.7 (1.2–8.6)	age	Merlo et al., 1989 ⁸³

Only occupations explicitly mentioned in a study are included. Study types: CC (case-control), C (cohort), NCC (nested case-control), CR (linkage of case registry and census data)

* Dose-response demonstrated

† Higher risks were found for some subcategories of workers

Table 30.3.1 (Cont'd) Occupations associated with increased risk of bladder cancer

Based on mortality data from England and Wales, it was estimated that bladder cancer due to occupational exposures was responsible for half of the rate difference between high and low social classes because these exposures are concentrated in blue-collar jobs.²³ However, no evidence was provided associating the blue-collar jobs with exposure to bladder carcinogens.

Metabolic polymorphisms and bladder cancer

Interindividual variation is common for many metabolic enzymes. In some cases, the variability has been attributed to inherited polymorphisms.²⁴ Phenotypic and genotypic tests have shown that variation in xenobiotic metabolizing enzymes is associated with cancer risk and may have an influence on human susceptibility to genotoxic agents.

A limited number of studies of bladder cancer genetic susceptibility in populations exposed occupationally to arylamines has been published.^{25–29} The results of a meta-analysis of all studies of acetylation status and bladder cancer in the general population suggest that certain groups with the NAT2 slow-acetylation phenotype are at greater risk of bladder cancer.³⁰ Additional studies are needed to establish if individuals could be at higher risk of bladder cancer given the presence of certain alleles that make them more susceptible. In the workplace, various metabolic polymorphisms could be acting in combination with occupational toxicants to produce risk. For occupational bladder cancer, polymorphic genotypes in the NAT (N-acetyltransferase) and GST (glutathione S-transferase) families of genes have been explored. Their joint effect, together with the effect of other genotypes, has not yet

been investigated. Moreover, the metabolic differences between monoarylamines and diarylamines, such as benzidine, warrant careful attention to the specific compounds to which each worker is exposed.

Animal models

Use of animal models to predict human bladder carcinogens has been problematic. Rats and mice are not susceptible to bladder cancer by most aromatic amines, including some highly potent occupational carcinogens. For aromatic amines, the Syrian hamster and dog were found to be better predictors for human bladder cancer than mice or rats, but this may not necessarily be the case for other chemical classes.¹⁵ There has been considerable debate about the relevance to human bladder cancer of bladder tumors associated with urinary calculi in mice or rats or calcium phosphate-containing precipitates in rats. The International Agency for Research on Cancer (IARC), which has a formal program for identification of carcinogenic hazards to humans, has issued a consensus report on this subject.³¹ Based on this consensus report, the IARC monograph program has recently classified saccharin in Group 3 (not classifiable as to carcinogenicity in humans) because the mechanism through which it is thought to cause bladder cancer in rats, formation of a calcium phosphate-containing precipitate, is not relevant to humans.³²

Numerous classes of genotoxic chemicals have been identified as bladder carcinogens in rodents and some of these have been identified in humans, most notably, aromatic amines, nitrosamines, and cyclophosphamide. In contrast, non-genotoxic chemicals appear to be highly specific with regard to species strain, diet, agent, dose, and mechanism.

Compound name [variant name]	CAS#	IARC group	Source(s)	Potentially exposed occupations, numbers*
Aluminum (production)	7429-90-5†	1	Boffetta	al production workers
Arsenic & arsenic compounds	7440-38-2	1	Wilbourn	arsenical pesticide manufacturing workers; pesticide users
Auramine dye manufacturing	492-80-8†	1	Boffetta	dye mfg. workers
Benzidine	92-87-5	1	Boffetta	1,554 (NOES)
Benzidine-based dyes		2A	IARC 29; IARC Supp 7	28,442 (NOES)
Direct Black 38 [2,7-Naphthalenedisulfonic acid, 4-amino-3-[[4'-((2,4-diaminophenyl)azo)(1,1'-biphenyl)-4-yl]azo]-5-hydroxy-6-(phenylazo)-disodium salt]	1937-37-7			dye manufacturing workers; dye-using workers - 44,500 (BLS)
Direct Blue 6 [2,7-Naphthalenedisulfonic acid, 3,3'-[(1,1'-biphenyl)-((4,4'-diylbis(azo))bis(5-amino-4-hydroxy)-, tetrasodium salt]	2602-46-2			
Benzidine, 3,3'-dichloro-	91-94-1	2A	Wilbourn	
4-Biphenylamine [4-Aminobiphenyl]	92-67-1	1	Boffetta; Wilbourn	
Chemotherapy agents				
Chlornaphazine [(N,N-Bis(2-chloro-ethyl)-2-naphthylamine)]	494-03-1	1	Wilbourn	pharmaceutical mfg. workers, oncology nurses, pharmacists
Cyclophosphamide [2H-1,3,2-Oxazaphosphorine, 2-(bis(2-chloroethyl)amino)tetrahydro, 2-oxide]	50-18-0	1		27,171 (NOES)
Coal-tar pitches	65996-93-2	1	Wilbourn; Boffetta	roofers - 142,600 (BLS)
Coal tars	65996-89-6	1	Wilbourn; IARC Supp 7	roofers - 142,600 (BLS)
Diesel engine exhaust	various	2A	Wilbourn	truck drivers - 2,500,000+ (BLS)
p-Dimethylaminoazobenzene [Brilliant yellow]	60-11-7	2B	Wilbourn	dye-using workers - 44,500 (BLS)
Magenta dye manufacture	632-99-5†	1	Boffetta	dye mfg. workers
4,4'-Methylene bis (2-chloroaniline) [MOCA]	101-14-4	2A	Wilbourn	
Mineral oils, untreated and mildly treated	various	1	Wilbourn	metal machinists - 1 million+ (BLS)
2-Naphthylamine	91-59-8	1	Boffetta	275 (NOES)
Phenacetin [p-Acetophenetidide]	62-44-2	2A	Wilbourn	17,658 (NOES) pharmaceutical mfg.
Analgesic mixtures containing phenacetin	various	1	Wilbourn	workers, oncology nurses, pharmacists
Rubber industry (certain occupations)	various†	1	Boffetta	rubber additives workers
Tobacco smoke		1	Wilbourn	‡
p-Chloro-o-toluidine	95-69-2	2A	IARC, 2000; Wilbourn	
o-Toluidine [o-Aminoazotoluene]	97-56-3	2A	IARC, 2000; Wilbourn	rubber additives workers & dye workers

An RTECS search for substances associated with bladder tumors produced the original list, adapted from Ruder et al., 1990.⁸⁴

IARC classifications are adapted from Wilbourn et al. 1999¹⁵ and Boffetta et al 1997.²³ Updates through Monograph 84 by the authors of this chapter. IARC ratings: 1, definite human bladder carcinogen; 2A, probable human bladder carcinogen. Compounds rated by Wilbourn et al.¹⁵ as canine bladder carcinogens are included, but not those rated as exclusively rodent bladder carcinogens.

* Estimated numbers are from two sources: 1. (NOES) Chemical-specific numbers of workers potentially exposed are from the 1981-1983 National Occupational Exposure Survey. Note that all these workers would be reaching > 20 years latency about 2001-3; 2. (BLS) Estimates of numbers of workers in occupational categories are from the Bureau of Labor Statistics 1999 National Occupational Employment and Wage Estimates (<http://stats.bls.gov/oes>). These data do not consider turnover among employees, which would lead to a greater number of persons exposed than is suggested by estimates from specific points in time.

† These manufacturing processes involve exposure to a number of chemicals.

‡ No study to date has linked passive smoking to bladder cancer; several have found no association.

Table 30.3.2 Known and suspected human bladder carcinogens and estimated numbers of potentially exposed US workers

Use of molecular and genetic mechanism information may be helpful in identifying possible mechanisms involved for these non-genotoxic chemicals and, therefore, can be important for a rational evaluation of human risk.

CLINICAL ASPECTS

Pathology

Transitional cell carcinomas (TCC) are graded by the World Health Organization histologically by the degree of abnormality of the tissues³³ and staged by the American Committee on Cancer - Union Internationale Contre

le Cancer by the extent to which they have spread.³⁴ One problem in the pathology of bladder tumors is the somewhat ambiguous line between the benign and the malignant, distinguishing between a papilloma, a papillary tumor with delicate fibrovascular stroma covered by a layer of epithelial cells indistinguishable from normal bladder mucosa, and a papillary carcinoma. This distinction should be viewed as a region where borders shift between pathologists, between institutions, and given the circumstances, even successive biopsies of the patient.

Some so-called benign papillomas display effects of inflammation and reactive or regenerative conditions so that they are classified by some pathologists as anaplastic,

although most of them do not behave as malignant tumors. To make the situation more complicated, some papillary tumors do become aggressive. Robinson and Hall³³ summarized the results of several studies: about 2–5% of 'benign' papillomas progress to carcinoma. The use of biochemical, molecular and genetic characteristics of cells is beginning to provide pathologists with a way to reduce these uncertainties. Strong associations of various markers with progression, invasiveness, and metastatic potential may provide a way to distinguish between pathologic subtypes of bladder cancer in the future.³⁵

Therapy

Strategies for diagnosis and therapy of occupational bladder cancer do not differ from those for bladder cancer resulting from non-occupational etiologies. A comprehensive guide to current treatment options by stage and grade has been assembled by the National Cancer Institute.³⁶ Treatment strategies do not appear to differ by histological type. Occupational bladder cancer does have a public health component: putative bladder carcinogens and high-risk populations that need to be followed with screening have been identified when exceptional cases of bladder cancer (i.e., in young non-smokers) were diagnosed.

Survival

The survival of patients with bladder cancer depends on the grade of anaplasia of the tumor and the stage of tumor invasion at time of diagnosis. SEER 1992–1997 5-year survival rates range from 94.5% for localized disease in white males to 0% for distant disease in African-American males. At each stage, women fare more poorly than men (except African-American women with distant disease) and African-Americans more poorly than whites. From 1974 to 1997 overall survival has improved from 48% to 65% in African-Americans and from 74% to 82% in whites. During the same period, incidence increased slightly overall from 14.6 to 16.7/100,000, in men from 25.6 to 29.0, and in women from 6.3 to 7.4.³⁷

Survival also appears to depend on social class and race, which, of course, are somewhat correlated. A review of five studies on bladder cancer survival found about a 20% discrepancy overall between higher and lower income patients.³⁸ African-Americans are less likely than whites to develop bladder cancer; however, once diagnosed, African-Americans experience poorer survival: 64% 5-year survival vs. 82% for whites.³

PREVENTION

Techniques for early detection

Cytoscopy is effective in identifying visible tumors in the bladder. A cytoscope is a slender tube with a lens and a light that is inserted through the urethra, allowing the physician to visually inspect the urethra and bladder.

Cytoscopy is invasive and not employed for asymptomatic individuals.³⁹

Urine cytology is the accepted technique for detection of bladder cancer in asymptomatic individuals. Urine cytology microscopically identifies the presence of abnormal, malignant cells, which are shed into the urine of patients with bladder cancer.³⁹ Cytologic screening for bladder cancer has a sensitivity of about 70% and a specificity of 90–95%, depending on the grade and stage of the tumor,⁴⁰ which is comparable to screening tests for cervical cancer, breast cancer, and colon cancer.⁴¹ Bladder cytology is effective in detecting preclinical stages of aggressive tumors and is substantially less effective in detecting low-grade tumors. There is widespread agreement that superficial well-differentiated papillary tumors rarely can be diagnosed definitively from voided urine cytology. In summary, cytology may be used to detect aggressive tumors, but these tumors may be advanced by the time they are discovered by this method. Cytology is less effective for low-grade tumors, which, although they are less aggressive, it would be desirable to find.⁴²

The greatest determinant of the sensitivity of urine cytology is the level of cytopathologist expertise. Ancillary techniques have been tested to improve the sensitivity of urine cytology. Of the large variety of methods, the most promising techniques appear to be DNA flow cytometry and image analysis for the detection of nuclear aneuploidy. Other sensitive methods include immunocytochemistry to detect the presence of antigens that are commonly expressed in neoplastic urothelium but not in the normal urothelium, such as the Lewis X antigen, and immunohistochemical analysis for the detection of p53 overexpression.⁴³

Hematuria screening, by urinalysis or by dipstick – a positive reaction for blood on urine-reagent-strip testing of asymptomatic people⁴⁴ – may be a more effective method than cytology for detecting low-grade early stage bladder tumors. The dilemmas with testing for hematuria are: (1) although almost all bladder tumors eventually cause hematuria, an infrequent examination may not be adequately sensitive, and (2) hematuria due to bladder cancer may be intermittent.⁴⁵ Although more frequent examinations increase the sensitivity of the test for bladder cancer, this method decreases the specificity of the test because it will detect other, non-malignant conditions causing hematuria, including cystitis, kidney disease, and urinary calculi.³ The debate then focuses on the predictive value of a positive test result for hematuria or the probability that a positive test result will reflect bladder cancer rather than another problem. It has been suggested that 5–10% of patients with hematuria have bladder cancer and 10–20% have some other serious urinary tract disease. As a condition becomes more prevalent in a population, the predictive value of a positive test increases. Exposure of an individual to an occupational carcinogen, as well as the individual's age and other risk factors for bladder cancer, should ensure a higher underlying prevalence of bladder cancer and thus increase the predictive value of a screen for hematuria.⁴⁵ The American Urological

Association recommends that asymptomatic microhematuria be evaluated only when associated with a risk (such as occupational exposure to carcinogens) of disease.⁴⁶

Tests of genetic factors are now being developed or assessed for evaluation of risk factors for bladder cancer. In addition to variations in metabolic phenotypes such as *N*-acetyltransferase, there are other genetic factors and acquired factors, such as recessive alleles for oncogenes, mutated tumor suppressor genes, and growth factors, that may place individuals at increased risk for bladder cancer independent of occupational exposure.⁴⁷ These genetic factors could add to any occupational risks for bladder cancer or multiply those risks. It is likely that the rapid pace of research will result in the identification of new predictive or prognostic markers in the near future.

A number of new techniques are being tested for use in bladder cancer screening. Most markers appear to have an advantage over urine cytology in terms of sensitivity, especially for detecting low-grade superficial tumors. However, most markers tend to be less specific than cytology, yielding more false-positive results. This scenario is more common in patients with concurrent bladder inflammation or other benign bladder conditions. A summary of the sensitivity, specificity and limitations of these methods is presented in Table 30.3.3.

The nuclear matrix protein (NMP) 22 test detects and measures urinary levels of a particular NMP called NuMA (nuclear mitotic apparatus). The NMP22 assay appears to be useful only to monitor, with high accuracy, for recurrence in patients with a past history of bladder cancer.⁴⁸

The bladder tumor antigen (BTA) test detects the presence of a bladder tumor antigen in the urine of patients with bladder cancer. The BTA stat and the BTA trak assays are qualitative and quantitative assays, respectively. Both detect a human complement factor H-related protein in the urine, and both accurately identify two-thirds of patients with bladder cancer. Both are limited because of the high number of false-positive reactions compared to urine cytology in low-grade bladder tumors.³⁹

The fibrin and fibrinogen degradation products (FDP) test is positive in two-thirds of patients with bladder cancer. The FDP test detects the degradation product of an

extravascular fibrin clot produced by tumors. It is more sensitive than urine cytology and has a high specificity.⁴⁹

Telomerase is an essential enzyme for cellular immortality and tumorigenesis. The telomeric repeat amplification protocol assay for telomerase in exfoliated cells can be used as a tumor marker. However, the low stability of telomerase in urine affects test sensitivity.⁵⁰ Inflammatory cells and stem cells have telomerase activity, and may be the source of false-positive tests.

Hyaluronic acid is a glycosaminoglycan that promotes tumor metastasis. High levels are detectable in the urine of patients with bladder cancer. Patients with high-grade TCC have elevated urinary hyaluronidase activity. A combination of both tests (HA-HAase test) yields a higher sensitivity than the sensitivity of individual tests,⁵¹ but it has no better sensitivity than urine cytology for detecting low-grade lesions.

Recently, detection of survivin in urine has been suggested as a predictive molecular marker of bladder cancer. Survivin is an enzyme inhibitor of apoptosis that is selectively overexpressed in human cancers, but undetectable in most normal adult tissues. In a patient series, the sensitivity of the urine survivin test for new or recurrent bladder cancer was 100%, and the specificity was 90–100%, depending on the population tested.⁵²

Test batteries

Combining tests can increase their sensitivity and specificity.⁴¹ Series testing is used to increase specificity and reduce the number of false-positive results. Parallel testing is used to increase sensitivity and reduce the number of false-negative results. A new US/European research consortium wants to create a simple, cost-effective, non-invasive diagnostic test to replace cystoscopy and cytology, initiating multicenter trials to find which of seven molecular markers, alone or in combination, is the most accurate detector of bladder cancer.⁵³

Screening programs

There are two reasons for screening a population exposed to a known or suspect bladder carcinogen. First, individu-

Test	Sensitivity	Specificity	Limitations
Urine cytology	17–70%	90–95%	Poor criteria to identify low-grade TCC*
BTA test	29–40%	68–91%	Low detection of grade I TCC. Poorer predictive value than urine cytology
BTA stat test	67–87%	40–70%	High false positive with gross hematuria, prostate cancer, BCG
BTA trak test	72%	43–48%	High false positive with UTI, stones, instrumentation
NMP22	48–80%	64–80%	High false positive with gross hematuria
FDP test	40–68%	80–96%	High false positive with gross hematuria
Telomerase	70–86%	60–90%	False negatives with gross hematuria, false positives with inflammation, complicated assay not widely available
HA-HAase test	90–92%	80–84%	No detection of grade I TCC

Modified from Brown, 2000⁴³

* Abbreviations: BCG = bacillus Calmette-Guérin, BTA = bladder tumor antigen, FDP = fibrin/fibrinogen degradation products, HA-HAase = hyaluronic acid/hyaluronidase, NMP = nuclear matrix protein, TCC = transitional cell carcinoma, UTI = urinary tract infection

Table 30.3.3 Sensitivity and specificity of non-invasive bladder tumor markers

als may be screened so that their tumors can be detected early when they are more readily treated, resulting in less morbidity and higher survival rates. This type of screening is for the personal benefit of the individuals. The second rationale for screening is to detect disease in a population at the earliest time possible in order to ensure that more primary methods of disease prevention, such as engineering controls and use of personal protective devices, are effectively incorporated to prevent exposure.⁵⁴ The two motivations should be kept in mind in appreciating a consensus view that was reached at the 1989 conference on screening for bladder cancer in high-risk groups. For populations exposed to known carcinogens at high levels, cytologic examination and testing for hematuria was recommended at 6-month intervals. The rationale for including hematuria was to ensure the acceptability of the screening program by ensuring that low-grade tumors would be detected that otherwise may be missed by cytology.

For low-exposure groups, as may be found in patients suffering from conditions as a result of environmental exposures, cytology was recommended 2 years after the first exposure, then every 5 years thereafter. For a suspect carcinogen, at high-exposure levels, cytology was recommended every 6 months, as well as measurement of hematuria to detect low-grade tumors. The argument for detecting low-grade tumors, even though there may be limited personal benefit for the individual because most such tumors are less aggressive, was to provide information that exposure had not been adequately controlled. The panel was not enthusiastic about any recommendations for a suspect carcinogen at low levels of exposure.⁵⁵

When weighing the benefits of a strategy of early detection, be it for the personal benefit of the worker or for the benefit of the workforce, it is necessary to consider the extent to which false-positive findings will be involved. A screening modality that leads to a disproportionate number of unnecessary follow-up and diagnostic procedures may not be cost effective or personally desirable. Moreover, the lengthening of the lead time, although possibly providing an extended opportunity for therapeutic intervention, also could provide a longer period of anxiety and distress for the worker.⁵⁴

In contrast to the recommendations of the bladder cancer conference consensus panel, the US Preventive Services Task Force concluded that there was insufficient evidence that hematuria and cytology screening improved the prognosis for those found to have cancer, even within high-risk groups.⁵⁶ If the efficacy of hematuria and cytology screening is not established, then in monitoring high-risk populations, cystoscopy should be reserved as a diagnostic test in individuals who had positive results on cytology and hematuria. However, it should be remembered that in at least one high-risk group, the MBOCA cohort, bladder cancer was diagnosed in two individuals who had had negative hematuria by dipstick and cytology screening.⁵⁷ Perhaps the non-invasive screening batteries now under development will end this dilemma.

SUMMARY

Despite substitution and process changes to prevent or reduce worker exposure, a substantial number of workers continue to be exposed to bladder carcinogens. Much larger numbers have been exposed to bladder carcinogens in the past. Some of these workers may still be at risk for bladder cancer. Improved screening options for high-risk groups, as well as better treatment options, should continue to improve survival and quality of life for these individuals.

References

1. Silverman DT, Levin LI, Hoover RN. Occupational risks of bladder cancer in the United States: II. Non-white men. *J Natl Cancer Inst* 1989; 81:1480-3.
2. Silverman DT, Levin LI, Hoover RN, Hartge P. Occupational risks of bladder cancer in the United States: I. White men. *J Natl Cancer Inst* 1989; 81:1472-9.
3. American Cancer Society. *Cancer facts and figures 2001*. Atlanta: ACS, 2001.
4. Silverman DT, Levin LI, Hoover RN. Occupational risks of bladder cancer among white women in the United States. *Am J Epidemiol* 1990; 132:453-61.
5. Rehn L. Blasengeschwülste bei fuchsin-arbeitern [Urinary bladder tumors in dye workers]. *Arch Klin Chir* 1895; 50:588-600.
6. Schulte P, Halperin W, Ward E, Ruder A. Bladder cancer screening in high risk groups. *J Occup Med* 1990; 32:787-945.
7. Hartge P, Harvey ED, Linehan WM, et al. Unexplained excess risk of bladder cancer in men. *J Natl Cancer Inst* 1990; 82: 1636-40.
8. Parkin DM, Pisani P, Ferlay J. Global cancer statistics. *CA Cancer J Clin* 1999; 49:33-64.
9. Silverman DT, Hartge P, Morrison AS, Devesa SS. Epidemiology of bladder cancer. *Hematol Oncol Clin North Am* 1992; 6:1-30.
10. EL Mawla MD, EL Bolkainy MN, Khaled HM. Bladder cancer in Africa: update. *Sem Oncol* 2001; 28:174-8.
11. Lopez-Beltran A, Escudero AL. Human papillomavirus and bladder cancer. *Biomed Pharmacother* 1997; 51:252-7.
12. Koroltchouk V, Stanley K, Sthernward J, Mott K. Bladder cancer: approaches to prevention and control. *Bull WHO* 1987; 65:513-20.
13. Silverman DT, Morrison AS, Devesa SS. Bladder cancer. In: *Cancer epidemiology and prevention*, 2nd edn. New York: Oxford University Press 1996;1156-79.
14. Zhang Z-F, Steineck G. Epidemiology and etiology of bladder cancer. In: Raghavan D, Scher HI, Leibel SA, Lang PH, eds. *Principles and practice of genitourinary oncology*. Philadelphia: Lippincott-Raven, 1997;215-22.
15. Wilbourn JD, Partensky C, Rise JM. Agents that induce epithelial neoplasms of the urinary bladder, renal cortex and thyroid follicular lining in experimental animals and humans: summary of data from IARC Monographs volumes 1-69. In: Capen CC, Dybing E, Rice JM, Wilbourn JD, eds. *Species differences in thyroid, kidney and urinary bladder carcinogenesis*. IARC Scientific Publications No. 147. Lyon: IARC, 1999;191-209.
16. Hueper WC, Wiley FH, Wolfe HD. Experimental production of bladder tumors in dogs by administration of beta-naphthylamine. *J Industr Hyg Toxicol* 1938; 20:46-84.
17. Case RAM, Hosker ME, MacDonald DB, Pearson JT. Tumors of the urinary bladder in workmen engaged in the manufacture and use of certain dyestuff intermediaries in the British chemical industry. *Br J Ind Med* 1954; 11:75-104.
18. Vineis P, Pirastu R. Aromatic amines and cancer. *Cancer Causes Control* 1997; 8:346-55.

19. Michaud DS, Spiegelman D, Clinton SK, et al. Fluid intake and the risk of bladder cancer in men. *N Engl J Med* 1999; 340:1390-7.
20. Oyasu R, Hupp ML. The etiology of cancer of the bladder. *Surg Gynecol Obstet* 1974; 138:97-108.
- 20a. Melicow MM. Tumors of the bladder: a multifaceted problem. *J Urol* 1974; 112: 467-78.
21. Schulte PA, Ringen K, Hemstreet GP, Ward E. Occupational cancer of the urinary tract. *Occup Med* 1987; 2:85-107.
22. Wallace OMA. Occupational urothelial cancer. *Br J Urol* 1988; 61:175-82.
23. Boffetta P, Kogevinas M, Westerholm P, Saracci R. Exposure to occupational carcinogens and social class differences in cancer occurrence. *IARC Sci Publ* 1997; 138:331-41.
24. Hayes RB. Genetic susceptibility and occupational cancer. *Med Lav* 1995; 86: 206-13.
25. Cartwright RA, Glashan RW, Rogers HJ, et al. Role of N-acetyltransferase phenotype in bladder cancer. *Lancet* 1982; ii(8303):842-5.
26. Hanke J, Krajewska B. Acetylation phenotypes and bladder cancer. *J Occup Med* 1990; 32: 917-18.
27. Hayes RB, Bi W, Rothman N, et al. N-Acetylation phenotype and genotype and risk of bladder cancer in benzidine-exposed workers. *Carcinogenesis* 1993; 14:675-8.
28. Rothman N, Hayes RB, Zenser TV, et al. The glutathione S-transferase M1 (GSTM1) null genotype and benzidine-associated bladder cancer, urine mutagenicity, and exfoliated urothelial cell DNA adducts. *Cancer Epidemiol Biomark Prev* 1996; 5:979-81.
29. Shinka T, Ogura H, Morita T, Nishikawa T, Fujinaga T, Ohkama T. Relationship between glutathione S-transferase M1 deficiency and urothelial cancer in dye workers exposed to aromatic amines. *J Urol* 1998; 159:380-3.
30. Marcus PM, Vineis P, Rothman N. NAT2 slow acetylation and bladder cancer risk: a meta-analysis of 22 case-control studies conducted in the general population. *Pharmacogenetics* 2000; 10:115-22.
31. Capen CC, Dybing E, Rice JM, Wilbourn JD, and Workshop Participants. Consensus report. In: Capen CC, Dybing E, Rice JM, Wilbourn JD, eds. Species differences in thyroid, kidney and urinary bladder carcinogenesis. IARC Scientific Publications No. 147. Lyon: IARC, 1999;1-14.
32. International Agency for Research on Cancer. Saccharin and its salts. Lyon: IARC, 2003. <http://www.wcie.iarc.fr/htdocs/monographs/vol73/73-19.html>
33. Robinson MC, Hall RR. Histopathology of urothelial cancer: consensus or controversy? In: Hall RR, ed. *Clinical management of bladder cancer*. New York: Oxford University Press, 1999;25-65.
34. Droller MJ. Clinical presentation, investigation, and staging of bladder cancer. In: Raghavan D, Scher HI, Leibel SA, Lang PH, eds. *Principles and practice of genitourinary oncology*. Philadelphia: Lippincott-Raven 1997;249-59.
35. Fradet Y, Cordon-Cardo C. Tumor markers in the management of bladder cancer. In: Raghavan D, Scher HI, Leibel SA, Lang PH, eds. *Principles and practice of genitourinary oncology*. Philadelphia: Lippincott-Raven 1997;231-8.
36. National Cancer Institute. PDQ website, December 15, 2003. <http://www.nci.nih.gov/cancerinfo/pdq/cancerdatabase>.
37. Ries LAG, Eisner MP, Kosary CL, et al., eds. *SEER Cancer Statistics Review, 1973-1998*. Bethesda, MD: National Cancer Institute, 2001.
38. Kogevinas M, Porta M. Socioeconomic differences in cancer survival: a review of the evidence. *IARC Sci Publ* 1997; 138:177-206.
39. Pirtskalaishvili G, Konety BR, Getzenberg RH. Update on urine-based markers for bladder cancer. *Postgrad Med J* 1999; 106:85-94.
40. Hemstreet GP, Bonner RB, Hurst RE, Rao JY. Cytology of bladder cancer. In: Vogelzang NJ, Scardino PT, Shipley WV, Coffey DS, eds. *Comprehensive textbook of genitourinary oncology*. Philadelphia: Lippincott Williams & Wilkins, 2000;322-32.
41. Hulka B. Principles of bladder cancer screening in an intervention trial. *J Occup Med* 1990; 32:812-6.
42. Farrow G. Urine cytology in the detection of bladder cancer: a critical approach. *J Occup Med* 1990; 32:817-21.
43. Brown FM. Urine cytology: is it still the gold standard for screening? *Urol Clin North Am* 2000; 27:25-37.
44. Hall RR. 1999 The diagnosis of bladder cancer. In: Hall RR, ed. *Clinical management of bladder cancer*. New York: Oxford University Press 1999;1-20.
45. Messing E, Vaillancourt A. Hematuria screening for bladder cancer. *J Occup Med* 1990; 32:838-46.
46. Grossfeld GD, Wolf JS, Litwan MS, et al. Asymptomatic microscopic hematuria in adults: summary of the AUA best practice policy recommendations. *Am Fam Phys* 2001; 63:114-54.
47. Koenig F, Jung K, Schnorr D, Loening SA. Urinary markers of malignancy. *Clin Chim Acta* 2000; 297:191-205.
48. Soloway MS, Briggman JV, Carpinito GA, et al. Use of a new tumor marker, urinary NMP22, in the detection of occult or rapidly recurring transitional cell carcinoma of the urinary tract following surgical treatment. *J Urol* 1996; 156:363-7.
49. Schmetter BS, Habicht KK, Lamm DL, et al. A multicenter trial evaluation of the fibrin/fibrinogen degradation products test for detection and monitoring of bladder cancer. *J Urol* 1997; 158:801-5.
50. Lokeshwar VB, Soloway MS. Current bladder tumor tests: does their projected utility fulfill clinical necessity? *J Urol* 2001; 165:1067-77.
51. Lokeshwar VB, Block NL. HA-HAase urine test. A sensitive and specific method for detecting bladder cancer and evaluating its grade. *Urol Clin North Am* 2000; 27:53-61.
52. Smith SD, Wheeler MA, Plescia J, Colberg JW, Weiss RM, Altieri DC. Urine detection of survivin and diagnosis of bladder cancer. *J Am Med Assoc* 2001; 285:324-8.
53. Agres T. Finding a better way to identify bladder cancer. *The Scientist* 2001; 15 (June 25): 21.
54. Halperin WE, Ratcliffe J, Frazier TM, Wilson L, Becker SP, Schulte PA. Medical screening in the workplace: proposed principles. *J Occup Med* 1986; 28:547-52.
55. Halperin W, Cartwright RA, Farrow GM, et al. Where do we go from here? *J Occup Med* 1990; 32: 936-45.
56. United States Preventive Services Task Force. *Guide to Clinical Preventive Services*, 2nd edn, 1996. December 15, 2003. <http://www.ahcpr.gov/clinic/cpsix.htm>
57. Ward E, Halperin W, Thun M, et al. Screening workers exposed to 4,4'-methylenebis(2-chloroaniline) for bladder cancer by cystoscopy. *J Occup Med* 1990; 32:865-68.
58. Armstrong BG, Tremblay CG, Cyr D, Theriault GP. Estimating the relationship between exposure to tar volatiles and the incidence of bladder cancer in aluminium smelter workers. *Scand J Work Environ Health* 1986; 12:483-93.
59. Howe GR, Burch JD, Miller AB, et al. Tobacco use, occupation, coffee, various nutrients and bladder cancer. *J Natl Cancer Inst* 1980; 64:701-3.
60. Malker HSR, McLaughlin JK, Silverman DT, et al. Occupational risks for bladder cancer among men in Sweden. *Cancer Res* 1987; 47:6763-6.
61. Schumacher MC, Slattery ML. Occupation and bladder cancer in Utah. *Am J Ind Med* 1989; 16:89-102.
62. Boyko RW, Cartwright RA, Glashan RW. Bladder cancer in dye manufacturing workers. *J Occup Med* 1985; 27:799-803.
63. Schulte PA, Ringen K, Hemstreet GP, et al. Risk assessment of a cohort exposed to aromatic amines. Initial results. *J Occup Med* 1985; 27:115-21.
64. Cartwright R. Occupational bladder cancer and cigarette smoking in West Yorkshire. *Scand J Work Environ Health* 1982; 8:79-82.
65. Zheng W, McLaughlin JK, Gao YT, Silverman DT, Gao RN, Blot WJ. Bladder cancer and occupation in Shanghai, 1980-1984. *Am J Ind Med* 1992; 21:877-85.

66. Schulz M, Loomis D. Occupational bladder cancer mortality among racial and ethnic minorities in 21 states. *Am J Ind Med* 2000; 38:90-8.
67. Isovich J, Castellrito R, Esteve J, et al. Tobacco smoking, occupational exposure and bladder cancer in Argentina. *Int J Cancer* 1987; 49:734-40.
68. Jensen OM, Wahrendorf J, Knudsen JB, et al. The Copenhagen case-referent study on bladder cancer. Risks among drivers, painters and certain other occupations. *Scand J Work Environ Health* 1987; 13:129-34.
69. Claude JC, Prenzal-Beyme RR, Kunze E. Occupation and risk of cancer of the lower urinary tract among men. A case-control study. *Int J Cancer* 1988; 41:371-9.
70. Baxter PJ, McDowall ME. Occupation and cancer in London. An investigation into nasal and bladder cancer using the Cancer Atlas. *Br J Ind Med* 1986; 43:44-9.
71. Ruder AM, Ward EM, Brown DP. Mortality in dry cleaning workers: an update. *Am J Ind Med* 2001; 39:121-32.
72. Bi W, Hayes RB, Feng P, et al. Mortality and incidence of bladder cancer in benzidine-exposed workers in China. *Am J Ind Med* 1992; 21:481-9.
73. Sathiakumar N, Delzell E. An updated mortality study of workers at a dye and resin manufacturing plant. *J Occup Environ Med* 2000; 42:762-71.
74. Risch HA, Burch JD, Miller AB, et al. Occupational factors and the incidence of cancer of the bladder in Canada. *Br J Ind Med* 1988; 45:361-7.
75. Bulbulyan MA, Figgs LW, Zahm SH, et al. Cancer incidence and mortality among beta-naphthylamine and benzidine dye workers in Moscow. *Int J Epidemiol* 1995; 24:266-75.
76. Schoenberg JB, Stemhagen A, Mogielnicki AP, Altman R, Abe T, Mason TJ. Case-control study of bladder cancer in New Jersey. I Occupational exposure in white males. *J Natl Cancer Inst* 1984; 72:973-81.
77. Brownson RC, Chang JC, Davis JR. Occupation, smoking and alcohol in the epidemiology of bladder cancer. *Am J Public Health* 1987; 77:1298-300.
78. Decoufle P. Cancer risks associated with employment in the leather and leather products industry. *Arch Environ Health* 1979; 34:33-7.
79. Guberan E, Usel M, Raymond L, et al. Disability, mortality and incidence of cancer among Geneva painters and electricians: a historical prospective study. *Br J Ind Med* 1988; 46:16-23.
80. Miller BA, Silverman DT, Hoover RN, Blair A. Cancer risk among artistic painters. *Am J Ind Med* 1986; 9:281-7.
81. Steenland K, Palu S. Cohort mortality study of 57,000 painters and other union members: a 15 year update. *Occup Environ Med* 1999; 56:315-21.
82. Popp W, Schmieding W, Speck M, Vahrenholz C, Norpoth K. Incidence of bladder cancer in a cohort of workers exposed to 4-chloro-o-toluidine while synthesizing chlordimeform. *Br J Ind Med* 1992; 49:529-31.
83. Merlo F, Costantini M, Doria M. Cause specific mortality among workers exposed to welding fumes and gases: a historical prospective study. *Sangyo I Daigaku Zasshi* 1989; 11 (Suppl):302-15.
84. Ruder A, Fine L, Sundin D. National estimates of occupational exposure to animal bladder tumorigens. *J Occup Med* 1990; 32:797-805.
85. 't Mannetje A, Kogevinas M, Chang-Claude J, et al. Occupation and bladder cancer in European women. *Cancer Causes Control* 1999; 10:209-17.
86. Maffi L, Vineis P. Occupation and bladder cancer in females. *Med Lav* 1986; 77:511-4.
87. Silverman DT, Hoover RN, Mason TJ, Swanson GM. Motor exhaust-related occupations and bladder cancer. *Cancer Res* 1986; 46:2113-6.
88. Silverman DT, Hoover RN, Albert S, Graff KM. Occupation and cancer of the lower urinary tract in Detroit. *J Natl Cancer Inst* 1983; 70:237-45.
89. Gonzalez CA, Lopez-Abente G, Errezola M, et al. Occupation and bladder cancer in Spain: a multi-centre case-control study. *Int J Epidemiol* 1989; 18: 569-77.
90. Ward E, Carpenter A, Markowitz S, Roberts D, Halperin W. Excess number of bladder cancers in workers exposed to ortho-toluidine and aniline. *J Natl Cancer Inst* 1991; 83:501-6.

Textbook of Clinical Occupational and Environmental Medicine

Second Edition

Edited by

Linda Rosenstock MD MPH

Dean, School of Public Health

Professor of Medicine and Environmental Health Sciences

University of California, Los Angeles

Los Angeles, CA

USA

Mark R Cullen MD

Professor of Medicine and Public Health Director

Occupational and Environmental Medicine Program

Yale University School of Medicine

New Haven, CT

USA

Carl Andrew Brodtkin MD MPH

Clinical Associate Professor of Medicine and

Environmental and Occupational Health Sciences

University of Washington

Seattle, WA

USA

Carrie A Redlich MD MPH

Professor of Medicine

Occupational and Environmental Medicine Program

and Pulmonary Critical Care Section

Yale University School of Medicine

New Haven, CT

USA



University of Louisville
Kornhauser Library