

during the acute phase of a traumatic brain injury a variety of growth factors, cytokines and inflammatory mediators are released. Here we investigated the temporal effect of cerebral tissue extract from traumatically-injured rats on the differentiation of cultured embryonic stem cells.

Methods: Sprague-Dawley rats were subjected to lateral fluid percussion traumatic brain injury (TBI). Brain extracts were derived by pooling and homogenising the respective hemispheres of three traumatised or healthy rats. Mouse embryonic stem cells (CGR8) were grown under standard culture conditions for two days on gelatine-coated 6 well plates. Subsequently, FCS was omitted (control, CTRL) and cerebral tissue extract from traumatised (trauma extract, TE) or healthy rats (healthy extract, HE) was added at a concentration of 20%. Cells grown under standard culture conditions were used as standard (STD). All experimental results obtained are presented as percentage of standard values. After 3, 5 and 7 days total RNA was isolated and the differentiation status of the co-cultured stem cells was assessed by amplifying stem cell specific marker OCT4, pluripotency marker nestin and neuronal marker MAP2 using real-time PCR. **Results:** We observed a significant increase of OCT4 expression after 5 ($0,85 \pm 0,011$; $p < 0,05$) and 7 days ($0,87 \pm 0,013$; $p < 0,01$) in TE treated cells as compared to standard values, indicating an onset of differentiation. Reduced Oct4 expression observed at any time point in control or HE treated cells was not statistically significant. An increase of nestin expression could be observed in all groups after only 3 days, though a significant increase was only demonstrated in TE treated cells ($1,92 \pm 0,059$; $p < 0,01$). Nestin expression in HE treated cells peaked on day 5 ($1,68 \pm 0,036$; $p < 0,05$). On day 7, nestin expression declined in all groups. In TE treated cells expression of neuronal marker MAP2 was initially increased at day 3 and remained constant at later time points. However, values remained below the values of spontaneous differentiation in control cells. In HE treated cells MAP2 expression was elevated as compared to spontaneous differentiation in control cells after 5 days of incubation. However, values obtained at this time point were not significant. **Conclusion:** A decrease in Oct4 expression and increase in nestin expression is associated with the onset of differentiation of stem cells into pluripotency cells. TE dependent reduction of OCT4 expression and rapid TE dependent upregulation of nestin expression indicates that trauma extract strongly induces stem cell differentiation. This is in accordance with the significant decrease of nestin after 7 days of TE treatment, as nestin is only transiently expressed during early differentiation of stem cells. However, our results also suggest that TE does not seem to induce neuronal differentiation, since MAP2 expression was hardly altered during long term TE stimulation of stem cells. MAP2 expression was elevated during incubation of stem cells with extract from healthy rats, indicating healthy brain to support neuronal differentiation pathways. These results indicate a strong micro-environmental affect on stem cell

differentiation pathways. Such observations should be taken into account when considering therapeutic cell replacement strategies. Presently, the impact of injured environment on stem cell differentiation pathways is being examined in more detail.

177

PULMONARY OXIDATIVE STRESS, INFLAMMATION, AND FIBROSIS INDUCED BY CARBON NANOTUBES. E. Kisin¹, A.R. Murray³, V. Johnson², O. Gorelik^{4,5}, S. Arepalli^{4,5}, V.Z. Gandelman^{4,5}, A.F. Hubbs¹, R.R. Mercer^{1,2}, P. Baron⁶, V.E. Kagan⁷, A.I. Potapovich⁷, V. Castranova^{1,3,7} and A.A. Shvedova^{1,2}.

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Society is currently amidst a revolutionary development of remarkable new technologies based on novel applications of nanomaterials. From drug delivery tools and micro-circuitry elements to microcomputer networks and super-durable composite materials – this is just an illustration of the unprecedentedly broad range of applications and approaches using nanomaterials. One of the most interesting examples of nanomaterials are carbon nanotubes (CNT), new members of carbon allotropes similar to fullerenes and graphite. Previously, we reported that exposure of human bronchial epithelial cells to CNT induced oxidative stress, depletion of antioxidants, morphological changes, cytotoxicity, and apoptosis. In the current study, we investigated pulmonary toxicity of CNT after pharyngeal aspiration by C57BL/6 mice. We found that CNT caused dose-dependent formation of granulomatous bronchointerstitial pneumonia, fibrosis (collagen accumulation), and changed pulmonary function. Administration of carbon nanotubes to C57BL/6 mice also resulted in a dose-dependent augmentation of biomarkers of inflammation quantified by bronchoalveolar lavage (BAL) cell counts, total protein, lactate dehydrogenase (LDH) and g-glutamyltranspeptidase (GGT) activities in BAL fluids and accumulation of pro-inflammatory and pro-fibrotic cytokines. CNT induced release of pro-inflammatory and pro-fibrotic cytokines from macrophages in vitro. Overall, our data suggest that exposure to CNT leads to pulmonary toxicity realized through synergized interactions of inflammatory response and oxidative stress culminating in the development of multifocal granulomatous pneumonia and fibrosis.