Feral Cats and Risk for Nipah Virus Transmission

To the Editor: Nipah virus (NiV) emerged in peninsular Malaysia in 1998 and 1999 as a respiratory and neurologic disease of domestic pigs and an acute febrile encephalitic disease in humans (1). Nipah virus infection is associated with a case-fatality ratio of 40% to 76% in humans (1,2). Cats (Felis catus) were infected with NiV at the site of the outbreak in northern Malaysia (3). Experimental studies have shown that cats are susceptible to Hendra virus and NiV (4,5). Infected cats shed NiV through the nasopharynx and in urine while viremic, and 1 (of 2) recovered from experimental NiV infection with a high neutralizing antibody titer (>256) within 21 days (5).

Fruit bats of the genus Pteropus are believed to be the reservoir for NiV in Malaysia (6). In June 2000, NiV was isolated from partially eaten fruit and from the urine of Pteropus hypomelanus in the village of Air Batang on Tioman Island, Peninsular Malaysia (7). Although humans live in close proximity to these bats, no evidence for local human exposure to NiV has been seen (8). In contrast, epidemiologic evidence from recent NiV outbreaks in Bangladesh suggests that direct infection from pteropid bats may occur, possibly when bats are pregnant (2,9).

Despite limited contact with bats, residents and visitors to Air Batang have ample opportunity for close contact with feral cats, which are often fed and sometimes housed by residents. Cats have been observed under trees that are occupied by roosting fruit bats in Air Batang. NiV could be transmitted from bats to cats through urine and then among cats oronasally, given their gregarious nature, which frequently includes mutual grooming. Cats are also frequently seen in close contact with humans in restaurants, on the tables, and in food preparation areas, where they are fed. If NiV is also present in bat fetal tissues, cats could become infected through contact with or by eating these tissues after mass births among bats.

We tested feral cats from Air Batang for neutralizing antibodies to NiV to determine whether cats might play a role in the zoonotic transmission of Nipah virus. Fifty bats were captured from Air Batang and tested for NiV and neutralizing antibodies to NiV as part of a long-term NiV surveillance study (A. Rahman, unpub. data). Thirty-two cats were caught July 12–19, 2004, in a 200-m radius of a bat colony. Cats were anesthetized, and 3.0 mL blood was collected from the jugular vein or medial saphenous vein. Serum was allowed to separate at 4°C for 24 hours and was then further separated and frozen in liquid nitrogen. Serum was tested by serum neutralization test (SNT), which is considered the reference standard for serologic assays, at the Australian Animal Health Laboratory, Geelong, Australia, as described (5,10).

The time of year was similar to the time when NiV was isolated from bats in 2000; however, none of the 32 cats (18 males, 14 females; 25 adults, 7 juveniles [<1 year of age]) had detectable antibodies to NiV on SNT. All cats appeared healthy except for 1 adult that was markedly jaundiced. The period of the study did not overlap the seasonal gestation period of P. hypomelanus, and none of the adult female bats tested (n = 20) were pregnant. Although attempts to isolate virus from bat urine and saliva were unsuccessful (A. Rahman, unpub. data), 7 (14%) of 50 bats, including 1 (8%) of 13 post-weaning juveniles (4 months to 2 years of age) had neutralizing antibodies (all >32) to NiV on SNT, which suggests that virus had circulated in the colony since 2000.

Our finding of no seropositive cats may be explained in 3 ways: 1) feral cats are rarely, if at all, exposed to NiV in nature; 2) the death rate from NiV infection in cats is so high that few or none survive with immunity; or 3) our sample size was too small to detect a seropositive cat. We believe that the first hypothesis is most likely. A low incidence of NiV infection in this population of bats (95% confidence interval for 0 of 50 bats, 0.00–0.71), combined with a short viremic period, would make transmission between bats and cats unlikely. However, if transmission occurred, we would expect to find some cats with a detectable titer (5). While the exact age of the cats in this survey was unknown, 25 (78%) of 32 were adults (>1 year of age) and may have been in Air Batang either in 2000, when NiV was isolated from bats, or during a more recent outbreak. We conclude that exposure of feral or peridomestic cats to Nipah virus on Tioman Island is rare and that the risk for zoonotic transmission is low.

Acknowledgments

We thank Amir Nordin Bin Harun, Abdul Karim Bin Abdul Hamid, Mohd Jeffril, Mohd Johan, and the residents of Air Batang, Tioman Island, for their assistance and A. Marm Kilpatrick for critical comments on this manuscript.

This work was supported by a National Institutes of Health/National Science Foundation “Ecology of Infectious Diseases” (R01-TW05869) award from the John E. Fogarty International Center and by core funding to the Consortium for Conservation Medicine from the V. Kann Rasmussen Foundation. This article is published as part of a collaboration with the Australian Biosecurity Cooperative Research Center. Ms Zambriski was funded by an award from The Center for Conservation Medicine at the Cummings School of Veterinary Medicine, Tufts University, and The Consortium for Conservation Medicine, New York.
Jonathan H. Epstein,*
Sohayati Abdul Rahman,†
Jennifer A. Zambriski,‡
Kim Halpin,§ Greer Meehan,§
Abdul Aziz Jamaluddin,¶
Sharifah Syed Hassan,†
Hume E. Field,# Alex. D. Hyatt,§
Peter Daszak,* and the Henipavirus
Ecology Research Group1

*The Consortium for Conservation
Medicine, New York, New York, USA; †Veterinary Research Institute, Ipoh,
Malaysia; ‡Cummings School of Veterinary
Medicine at Tufts University, North Grafton,
Massachusetts, USA; §Commonwealth
Scientific and Industrial Research
Corporation Australian Animal Health
Laboratory, Geelong, Victoria, Australia;
¶Ministry of Agriculture, Kuala Lumpur,
Malaysia; and #Department of Primary
Fisheries and Industry, Brisbane,
Queensland, Australia

1A complete list of Henipavirus Ecology
Research Group members can be found at
http://www.henipavirus.org

References

1. Chua KB, Bellini W, Rota P, Harcourt B,
Tamin A, Lam S, et al. Nipah virus: A
recently emergent deadly paramyxovirus.

2. Hsu VP, Hossain MJ, Parashar UD,
Mohammed MA, Ksiazek TG, Kuzmin I, et
al. Nipah virus encephalitis reemergence,

3. Mohd Nor MN, Gan CH, Ong BL. Nipah
virus infection of pigs in peninsular

4. Westbury HA, Hooper PT, Brouwer SL,
Selleck PW. Susceptibility of cats to equine

5. Middleton DJ, Westbury HA, Morrissey CJ,
van der Heide BM, Russell GM, Braun
MA, et al. Experimental Nipah virus infec-
tion in pigs and cats. J Comp Pathol.

6. Yob JM, Field H, Rashdi AM, Morrissey C,
van der Heide B, Rota P, et al. Nipah virus
infection in bats (order Chiroptera) in
peninsular Malaysia. Emerg Infect Dis.

7. Chua KB, Koh C, Hooi P, Wee K, Khong J,
Chua B, et al. Isolation of Nipah virus from
Malaysian island flying foxes. Microbes
Infect. 2002;4:145–51.

8. Chong HT, Chong TT, Goh KJ, Lam SK,
Chua KB. The risk of human Nipah virus
infection directly from bats (Pteropus
hypsomelaus) is low. Neurology Asia [serial
on the Internet]. 2003 [cited 2006 May
16]. Available from http://www.neurology-
asia.org/articles/20031_031.pdf

9. ICDDR,B. Nipah virus outbreak from date
palm juice. Health and Science Bulletin
[serial on the Internet]. 2005 Dec [cited
classificationID=56&pubID=6590

10. Daniels P, Ksiazek T, Eaton BT. Laboratory
diagnosis of Nipah and Hendra virus infec-

Address for correspondence: Jonathan H.
Epstein, The Consortium for Conservation
Medicine, 460 West 34th St, 17th Floor,
New York, NY 10001, USA; email:
epstein@conservationmedicine.org