



Published in final edited form as:

*Chemosphere*. 2021 January ; 262: 128309. doi:10.1016/j.chemosphere.2020.128309.

## ASSOCIATIONS OF PCBS, DIOXINS AND FURANS WITH FOLLICLE-STIMULATING HORMONE AND LUTEINIZING HORMONE IN POSTMENOPAUSAL WOMEN: NATIONAL HEALTH AND NUTRITION EXAMINATION SURVEY 1999–2002

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### Abstract

**Background:** The general population is exposed to the group of endocrine disrupting chemicals persistent organic pollutants (POPs), that includes polychlorinated biphenyls (PCBs), polychlorinated dibenzo-p-dioxins (PCDDs) and polychlorinated dibenzofurans (PCDFs).

**Objectives:** The aim of this research was to evaluate the associations of serum levels of PCB, PCDD, and PCDF congeners with follicle stimulating hormone (FSH) and luteinizing hormone (LH) in postmenopausal women not taking exogenous hormones. We hypothesized that associations of POPs with these gonadotropins could be modified by factors affecting endogenous hormones.

**Methods:** Cross-sectional analyses were conducted on data from 89 postmenopausal women using data from the National Health and Nutrition Examination Survey (NHANES). POPs were summarized based on classification schemes thought to reflect toxicological properties.

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Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Associations of POPs and gonadotropin hormones were modeled with multivariable regression models. When evidence of interaction was found, conditional effects were estimated.

**Results:** We found inverse associations of LH, but not FSH, with exposure to anti-estrogenic and/or dioxin-like POPs, but not with non dioxin-like PCBs. A doubling of dioxin-like toxic equivalents (TEQs) was associated with a decrease in LH of 11.9% (95% CI = -21.3%, -1.4%, p=0.03). Inverse associations were enhanced by potential effect modifiers related to both direct and indirect estrogenicity, including obesity and the obesity-related condition inflammation.

**Conclusions:** These investigations support a pattern of endocrine-disrupting effects by dioxin-like POPs among postmenopausal women, especially those with conditions related to peripheral estrogenicity. Elucidation of the complex relationship of gonadotropin hormones with thyroid hormones await further investigation.

## Keywords

Follicle stimulating hormone; luteinizing hormone; PCBs; dioxins; endocrine disruption

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## 1. Introduction

Polychlorinated biphenyls (PCBs) are a class of heat-resistant, oily liquids that were used as insulating fluids in capacitors and transformers (ATSDR, 2000). Polychlorinated dibenzo-p-dioxins (PCDDs) and polychlorinated dibenzofurans (PCDFs), commonly known as dioxins, are compounds formed from a variety of sources, including as unintentional by-products of industrial processes (Fiedler, 1996). Like PCBs, dioxins are resistant to abiotic and biotic degradation in the environment and bioaccumulate and magnify in animals and humans (Safe, 1994; Van den Berg et al., 1998). Human body burdens of these chemicals have declined over time (Aylward et al., 2002; Turyk et al., 2012), and with continued exposure through inhalation as well as consumption of contaminated food items (van Larebeke et al., 2001).

Persistent organic pollutants (POPs), including PCBs, PCDFs and dioxins, exhibit a broad range of toxic effects including disruption of sex steroid hormone homeostasis. Dioxin-like compounds, acting through the AhR, are thought generally to be antiestrogenic, although data showing positive associations with estrogen dependent tumors (van Larebeke et al., 2001, Ohtake et al., 2003)) and the upregulation of genes related to enzyme CYP 19, which encodes aromatase activity (Warner et al., 2012), also suggest estrogen-like activity. Effects of PCBs on sex steroid hormones are varied, with some congeners thought to be estrogenic and some, more dioxin-like, antiestrogenic. Additional mechanisms suggested for effects of PCBs and their metabolites on steroid hormones relate to inhibition of estrogen sulfotransferases (Kester et al, 2000, Kester et al 2002) and decreased sex hormone binding globulin (SHBG), leading to greater bioavailability of peripheral sex hormones as well as direct action on hypothalamic gonadotropin releasing hormone gene expression (Gore et al., 2002)

Studies in male and female animals and in in vitro investigations have also shown direct effects of dioxins on the pituitary hormones follicular stimulating hormone (FSH) and

luteinizing hormone (LH) as well as associations between PCB exposure and FSH and LH levels, although results have been inconsistent and differed by PCB congener or Aroclor (Desaulniers et al., 1999; Wade et al., 2002; Oskam et al., 2005; Uslu et al., 2013; Taketoh et al 2007, Cao et al 2011).

To date, there is only one known study in occupationally exposed postmenopausal women that found an inverse association between PCBs and FSH (Persky et al., 2011). In one of two other investigations of premenopausal women during days 1-5 of the menstrual cycle, PCBs were inversely associated with FSH (Pan et al 2019). In the other study there were no associations with individual gonadotropins, but a positive association with the FSH:LH ratio (Gallo et al 2018). Relationships of PCBs with gonadotropins in men have been inconsistent (Emeville et al., 2013, Ferguson et al., 2012, Richthoff et al., 2003, Hagmar et al., 2001, Persky et al 2001, Haugen 2011, Vested 2014, Giwercman et al., 2006. Persky et al 2012, Sweeney et al., 1997, Petersen et al 2018, Vitku et al 2016).

In this investigation, we explored associations of POP exposure with FSH and LH levels in postmenopausal women using the National Health and Nutrition Examination Survey (NHANES), a cross-sectional survey examining a representative sample of the US population. The present study uses PCB, PCDD, PCDF, FSH, and LH measurements obtained in the 1999–2000 and 2001–2002 survey cycles. We examined the effects of POP exposures, grouped into categories with similar structure or a common mechanism of action, on FSH and LH in a subgroup of postmenopausal women not taking glucocorticoids or sex hormones. In addition, we examined the hypothesis that associations of POPs with FSH and LH are modified by factors that may influence endogenous hormones.

## 2. Methods

### 2.1 Participants

Data from the NHANES survey cycles conducted in 1999–2000 and 2001–2002 were obtained online. Each survey is a nationally representative sample of the US civilian, noninstitutionalized population based on a complex probability sampling design. The University of Illinois at Chicago Institutional Review Board has determined that analysis of NHANES data does not meet the definition of human subject research as defined by 45 CFR 46.102(f).

Any associations between POPs and gonadotropins may be more clearly observed among postmenopausal women who do not require precise timing with the menstrual cycle. Menopause is defined as one year after the permanent cessation of menstrual periods, which women experience at the average age of 51 years. Menopause can occur naturally or be induced through a medical intervention such as bilateral oophorectomy. Normal FSH levels for premenopausal women are 4.7–21.5 mIU/mL, while normal FSH levels for postmenopausal women are 25.8–134.8 mIU/mL (Lobo, 2007). Standards used to define menopause were based on a prior report (Kalkwarf et al., 2003) and were applied consecutively so that the rules were applied only to women not already in a previous category. The following are inclusion categories for postmenopausal women:

1. Any age and last period 12 months without hysterectomy or with hysterectomy and bilateral oophorectomy
2. 56–59 years of age and last period 12 months with hysterectomy and without bilateral oophorectomy, and FSH 25.8 mIU/mL
3. <56 years of age and last period 12 months with hysterectomy, without bilateral oophorectomy, and FSH 50 mIU/mL

In the present investigation, we focused on the 1,847 postmenopausal women 40 years of age with questionnaire data that included hysterectomy, bilateral oophorectomy, and prescription medications. Serum levels of gonadotropins were originally used by NHANES investigators to classify women according to menopausal status; therefore, FSH and LH tests were performed only on women aged 35–60 years. We excluded postmenopausal participants who were <40 years of age (n=3), were <60 years of age with FSH <25.8 mIU/mL (n=10), if they did not have data on exposure and hormone measures (n=1,697), those with missing cotinine data (n=1), those who reported taking glucocorticoids (n=3), and those who specified taking sex hormones (estrogen, progestins, sex hormone combinations, miscellaneous sex hormones, gonadotropin-releasing hormones and analogs, androgens and anabolic steroids, and contraceptives) or other hormones/hormone modifiers, including selective estrogen receptor modulators, aromatase inhibitors, antiandrogens, and antigonadotropic agents (n=44). Data for analysis of the associations of POPs with FSH and LH were available for 89 participants

## 2.2 Follicle-stimulating hormone, luteinizing hormone, and other physiological measurements

Details of the NHANES laboratory measurements are available online for 1999–2000 (CDC, 2020a) and 2001–2002 (CDC, 2020b). Briefly, serum FSH and LH concentrations for 1999–2000 were measured by a microparticle enzyme immunoassay technology. The sensitivity for FSH was 0.2 IU/L and the sensitivity for LH was 0.5 IU/L. Serum FSH and LH concentrations for 2001–2002 were measured by a paramagnetic particle, chemiluminescent two-step enzyme assay. The sensitivity for FSH and LH was 0.02 IU/L. Measurements below the limit of detection (LOD) were assigned a value of 0.2 or 0.5 IU/L divided by the square root of two by the CDC. The inter-assay coefficient of variation (CV) for 1999–2000 varied from 2.37 to 7.95 for FSH and from 1.65 to 7.59 for LH, and the CV for 2001–2002 varied from 3.2 to 7.2 for FSH and from 3.3 to 10.1 for LH. C-reactive protein (CRP) levels were measured by latex-enhanced nephelometry with high sensitivity by using a Dade Behring Nephelometer II Analyzer System (Dade Behring Diagnostics, Inc., Somerville, New Jersey). Serum total cholesterol was measured enzymatically after hydrolyzation and oxidation, while triglycerides were analyzed enzymatically after hydrolyzation into glycerol.

## 2.3 Polychlorinated biphenyl, polychlorinated dibenzo-p-dioxin, and polychlorinated dibenzofuran measurements

All POPs were measured in serum by high-resolution gas chromatography/isotope-dilution high-resolution mass spectrometry (Organic Toxicology Branch, National Center for Environmental Health, CDC, Atlanta, Georgia). The congener groupings used in the current

study are listed in Table 1. Compounds able to bind to the AhR (PCDDs, PCDFs, and dioxin-like PCBs) were used to calculate TEQs by multiplying the TEQ factor by the congener concentration (Van den Berg et al., 2006) and then summing the values to yield  $\Sigma$ TEQs. PCB congeners were summed to yield  $\Sigma$ PCBs. The PCB congeners were grouped according to structure, including  $\Sigma$ non-dioxin-like PCBs,  $\Sigma$ mono-*ortho* PCBs, and  $\Sigma$ dioxin-like PCBs (consisting of non-*ortho* and mono-*ortho* PCBs), and were also grouped according to mechanism of action, including estrogenic and anti-estrogenic activity (Wolff et al., 1997; Cooke et al., 2001). Polychlorinated biphenyl 126 is listed in both  $\Sigma$ Cooke estrogenic PCBs and  $\Sigma$ Cooke anti-estrogenic PCBs. For congeners with results below the LOD, the measurement was imputed by CDC as the LOD for that specific congener divided by the square root of two. In the first study cycle, more of the individual congener measurements were below the LOD than in the second study cycle. Only congeners that had >10% of results >LOD for each of the two study cycles were included in the analysis. When results for more than one congener were not reported by CDC for a participant, the participant was coded as missing for each summary exposure mentioned previously. However,  $\Sigma$ Wolff estrogenic PCBs comprised two congeners; therefore, the participant was coded as missing if one of the congeners was not reported by CDC.

## 2.4 Covariates

Potential confounders and effect modifiers evaluated in this study included age, alcohol consumption, BMI, CRP, cotinine level, lipids, race/ethnicity, study cycle, antidiabetic medications, and thyroid hormone medications. Alcohol consumption was dichotomized as <12 drinks/year and  $\geq$  12 drinks/year (“Had at least 12 alcohol drinks/1 year?”). Serum cotinine was dichotomized as  $\leq$  10 ng/mL and >10 ng/mL, a cutoff previously used as a marker for both active smoking and high environmental tobacco smoke exposure (Pirkle et al., 1996). Participants were classified as Caucasian, African American, or other. We calculated total serum lipids using the formula: lipids=[total cholesterol (mg/dL) x 2.27] + triglycerides (mg/dL) + 62.3 (Phillips et al. 1989). Lipids and CRP were analyzed as continuous measures. Finally, we evaluated medications that can affect hormone homeostasis including antidiabetic and thyroid hormones.

## 2.5 Statistical analyses

Statistical analyses were performed with SAS 9.2 (SAS Institute Inc., Cary, North Carolina) without the use of sample weights due to the limited sample size. Similar results were obtained when statistical analyses were repeated with SAS-Callable SUDAAN 11.0 (Research Triangle Institute, Research Triangle Park, North Carolina) using a four-year sample weight provided by NHANES for the dioxins subsample and variables that accounted for the complex survey design (not shown). Results were considered significant at  $p < 0.05$  and borderline significant at  $0.05 < p < 0.10$ .

Natural log transformations of exposures, CRP and lipids were used to approximate a normal distribution, with geometric means presented for descriptive purposes. Differences in demographics, health and lifestyle factors, medication use, and study cycle by POPs and hormones were examined using Student’s t-tests for continuous variables or Chi-square tests for dichotomous variables. We used analysis of variance with Tukey post hoc testing to

evaluate differences in exposure or outcome measures among race categories. Associations between continuous variables were tested with Pearson's correlation coefficients.

Associations of POPs with gonadotropins were estimated using linear regression. For all analyses, we used natural log transformations of continuous measurements of wet weight PCB, PCDD, and PCDF congeners rather than lipid-standardized measurements (Schisterman et al., 2005). FSH and LH were log transformed for regression analysis since model fit, as judged by r-square, was improved compared with non-transformed models. Age, BMI, and lipids were included in all adjusted models. We estimated the percent change in FSH and LH levels for a doubling of serum POPs as  $(e^{(\ln 2 \times \beta)} - 1) \times 100$ , with 95% confidence intervals (CIs) calculated as  $(e^{[\ln 2 \times (\beta \pm 1.96 \times SE)]} - 1) \times 100$ .

To assess confounding, additional covariates were added individually to the adjusted model. Confounding was identified by a change in the exposure beta coefficient of more than 10% after the addition of a potential confounder. Effect modification was evaluated using variables indicating the product of the potential effect modifier (CRP, BMI, thyroid medication use, diabetes medication use, serum cotinine) with the exposure. When significant interaction was found ( $p < 0.05$ ), conditional effects at the 25<sup>th</sup> and 75<sup>th</sup> percentile of the modifier were estimated.

### 3. Results

#### 3.1 Descriptive statistics

The mean age was 54.3 years (range 42–60 years) and mean BMI was 29.5 kg/m<sup>2</sup> (range 18.4–46.9 kg/m<sup>2</sup>) (Table 2). About 36% were Caucasian, 25% African American, and the remainder other or multiple race/ethnicities; 53% reported having 12 or more alcohol drinks/year; 34% had elevated cotinine levels, 11.2% were taking thyroid hormones and 18% using antidiabetic medications (data not shown). For participants with elevated cotinine (>10 ng/mL), 86.7% reported smoking cigarettes every day or some days (data not shown). The mean FSH level was 68.6 and ranged from 27.9 to 196.1, and one participant had an FSH (mIU/mL) level notably above the reference range for postmenopausal women. The mean LH level was 37.6 (mIU/mL) and ranged from 10.2 to 91.0 (Table 2). There were no significant differences between the 1999–2000 and 2001–2002 study cycles for continuous or categorical variables.

#### 3.2 Bivariate analyses

In unadjusted analyses, POP exposures and CRP were significantly higher in African Americans than Caucasians (data not shown). Body mass index and CRP were also significantly higher, while mean FSH was significantly lower, in antidiabetic medication users (data not shown). CRP was significantly lower in participants who specified an alcohol consumption of 12 drinks/year. Table 3 shows unadjusted Pearson's correlation coefficients among continuous measures. Luteinizing hormone was significantly and positively related to FSH ( $r=0.77$ ), and was inversely associated with  $\Sigma$ TEQs ( $r=-0.26$ ); somewhat weaker associations were noted with  $\Sigma$ mono-*ortho* PCBs ( $r=-0.22$ ) and  $\Sigma$ Wolff anti-estrogenic

PCBs ( $r=-0.20$ ). In general, POP groupings were significantly and positively associated with lipids.

### 3.3 Associations of PCBs, dioxins and furans with gonadotropins

In analyses adjusted for age, BMI, and lipids (Table 4), LH was significantly and inversely associated with  $\Sigma$ TEQs,  $\Sigma$ mono-ortho PCBs and  $\Sigma$ Cooke antiestrogenic PCBs. A doubling of  $\Sigma$ TEQs was associated with a decrease in LH of 11.9% (95% CI=-21.3%, -1.4%,  $p=0.03$ ). The inverse associations of  $\Sigma$ dioxin-like and Wolff anti-estrogenic PCBs with LH were of borderline significance. No significant or borderline significant associations were found between POPs and FSH.

### 3.4 Effect modification

Effect modification by factors that may influence endogenous hormones, namely BMI, CRP, thyroid medication use, diabetes medication use and serum cotinine levels, was evaluated in adjusted regression models by including a variable indicating the product of the modifier with the exposure. When significant interaction was identified ( $p<0.05$ ), conditional estimates were generated for associations of POPs with gonadotropins at the 25<sup>th</sup> and 75<sup>th</sup> percentiles of the modifier (Table 5). We found evidence for modification of POP/gonadotropin associations by BMI and CRP, even when a main effect was not present. There were stronger inverse associations in women with elevated CRP for LH with non-dioxin-like, mono-ortho, dioxin-like, Cooke antiestrogenic and both Cooke and Wolff estrogenic PCBs; similarly there were stronger associations in women with higher CRP for FSH with TEQs and with total, mono-ortho, dioxin-like, Cooke and Wolff anti-estrogenic PCBs. Women with higher BMI had stronger inverse associations of LH with total, dioxin-like and Cooke anti-estrogenic PCBs; women with higher BMI had stronger inverse associations of FSH with TEQs, total PCBs and both Cooke and Wolff antiestrogenic PCBs. There was a lack of support for modification of POP/gonadotropin associations by cotinine, thyroid medication use, and diabetes medication use.

### 3.5 Sensitivity analyses of associations of POPs with gonadotropins

The majority of participants with the absence of menstrual periods for 12 months or more specified the reason for not having regular periods as “going/gone through menopause.” Only three postmenopausal women specified “medical conditions/treatments” as the reason for not having regular menstrual periods (two of the three were 60 years of age). One of these participants reported the use of thiazolidinediones (TZDs), which are a class of insulin-sensitizing agents used to treat diabetes and may affect estrogen metabolism as a result of inhibition of aromatase activity (Seto-Young et al., 2011). Exclusion of these three participants in a sensitivity analysis yielded somewhat increased effect estimates. Associations of LH with total, dioxin-like PCBs and Wolff’s antiestrogenic PCBs were significant ( $p<0.05$ ) as were associations of FSH with  $\Sigma$ TEQs and  $\Sigma$ mono-ortho PCBs.

Compared with natural menopause, oophorectomy in postmenopausal women has been shown to lower androgen levels (Laughlin et al., 2000; Labrie et al., 2011); therefore, we repeated the current analysis excluding 10 postmenopausal participants with bilateral oophorectomy. This exclusion decreased effect estimates, and only associations of LH with

$\Sigma$ TEQs and  $\Sigma$ Cooke anti-estrogenic PCBs remained borderline significant, which may be the result of reduced sample size. Repeating the analysis with exclusion of an FSH level notably outside of the laboratory range produced results that were unchanged.

## 4. Discussion

### 4.1 Follicle-stimulating hormone and luteinizing hormone

In this investigation, we found inverse associations of anti-estrogenic and/or dioxin-like POP groupings with LH, but not FSH, in postmenopausal women not taking glucocorticoids or sex hormones. Inverse associations of dioxin-like POPs with LH and/or FSH were stronger in participants with elevated CRP or BMI.

In a previous investigation of postmenopausal women with occupational exposures, PCBs were inversely associated with FSH (Persky et al., 2011). In one study of premenopausal women without ovarian insufficiency in whom blood was obtained in the first five days of the menstrual cycle there was also a negative association of PCBs with FSH (Pan et al 2019). In another study of premenopausal women in which blood was obtained at day 3 of the menstrual cycle there was no association with either FSH or LH, although there was a positive association of estrogenic PCBs with the FSH:LH ratio (Gallo et al 2018) Studies in men have also not been consistent. In several studies representing a range of exposure levels, PCBs were not associated with FSH or LH. These include healthy men from the West Indies, (Emeville et al., 2013), men from fertility clinics (Ferguson et al., 2012, Vitku et al 2016), young Swedish men from the general population, (Richthoff et al., 2003), men exposed to high levels of POPs through fish consumption (Hagmar et al., 2001, Persky et al 2001), and men living in Norway (Haugen 2011). Similarly, there were no associations of in utero exposure to PCBs with adult FSH or LH (Vested 2014). In contrast, among Inuits and several European cohorts, there were positive associations of PCB 153 with LH in some but not all cohorts (Giwercman et al., 2006). Among men with occupational exposures to PCBs at a capacitor manufacturing plant there was an inverse association of dioxin like PCBs with LH of borderline significance (Persky et al 2012). In another study of occupationally exposed men, TCDD was significantly and positively associated with FSH and LH (Sweeney et al., 1997). In the Faroe Islands, among men with a large range of exposure, PCBs were positively associated with LH but not FSH (Petersen et al 2018). Variations in associations of POPs with gonadotropins could be indicative of gender differences or a threshold of bioavailable estrogen necessary for impacts on circulating hormones. Further, the effect of mixtures of PCBs, PCDDs, and PCDFs might be additive or antagonistic, depending on the population, dose, mixture components and endpoints.

Surprisingly, we found that anti-estrogenic and/or dioxin-like POP groupings were negatively associated with LH. The literature has shown, among multiple factors that contribute to LH secretion, that estrogen plays an important role by exerting feedback to the pituitary in the normal functioning of the HPG axis (Clarke, 2002; Christian et al., 2005); however, the mechanism by which estrogen controls these events has not been delineated. Because control of FSH secretion is more complex than LH and includes stimulus by inhibins and activins, LH has been thought by some to be a better marker for estrogen-negative feedback control of gonadotropin secretion although there is debate on this issue.



(Weiss et al., 2004; Cosma et al., 2008; Shaw et al., 2010). In our primary analyses in this study, associations were stronger for LH than FSH.

Because the 1999–2002 NHANES data sets do not provide measurements of other estrogen-related hormones for women, it is difficult to postulate potential mechanisms. Dioxins and related compounds that bind to the AhR are generally thought to elicit antiestrogenic responses (Safe et al., 1998). The findings of associations between antiestrogenic and/or dioxin-like POP groupings with LH appear counterintuitive, given that LH levels were decreased, which implies an estrogenic effect. However, our findings may be consistent with previous research suggesting cross-talk between the estrogen receptor and the AhR signaling pathways (Cooke et al., 2008; Swedenborg and Pongratz, 2010). Further, dioxin-like and potentially anti-estrogenic, PCBs have been associated with increased gene expression of estrogen receptor beta and CYP19 coding for aromatase, an enzyme involved in estrogen synthesis (Warner et al., 2012). Inverse relationships of dioxin-like compounds and LH could also reflect direct inhibition of LH synthesis and/or release from the pituitary (Cao et al 2011, Taketoh 2007)

## 4.2 Effect modification

Inverse associations of anti-estrogenic and/or dioxin-like PCB groupings with FSH and LH in the present study were stronger in postmenopausal participants with higher BMI. There is evidence that body composition may play an important role in steroid hormones and SHBG concentrations. In general, the aromatization of androstenedione to estrone in adipose tissue correlates positively with weight (Bulun et al., 1994). Inverse correlations have also been reported between BMI and SHBG for postmenopausal women, resulting in increased bioavailable estradiol (Cauley et al., 1989).

Inverse associations of POPs with LH and FSH were larger in postmenopausal women with higher CRP levels. Some investigators have suggested that PCBs may act through increased inflammatory responses (Hennig et al., 2002). C-reactive protein is a general marker of systematic inflammation and in the current study, CRP was associated with antidiabetic medication use, and higher BMI and triglycerides after adjusting for potential confounders (data not shown). Several studies have also noted associations of CRP with factors relating to estrogen status, including higher levels of estradiol and lower levels of SHBG (Stork et al., 2008; Maggio et al., 2011), as well as stimulation of aromatase (Zhao et al., 1996).

## 4.3 Limitations and strengths

The present investigation has a number of limitations including a small sample size that may have decreased our ability to identify significant associations of POPs with gonadotropins. The cross-sectional design of the study does not allow us to establish a temporal relationship of POP exposures with changes in gonadotropin levels. We adjusted for age, BMI, and lipids, but there may be other important potential confounders for which we have not controlled. Additional congener measurements for complete congener groupings might have helped to better elucidate mechanisms related to associations between PCB groupings and hormones. Estrogenic and anti-estrogenic classes of PCB congeners can express diverse and sometimes conflicting effects (Warner et al., 2012). Finally, the findings might be due to

chance, as multiple comparisons were made in the statistical analysis. We did not adjust for multiple comparisons; however, the purpose of these exploratory analyses was to inform and guide future research that could be subject to further rigorous testing (Goldberg et al., 2011).

Despite the limitations, this study has several strengths. To our knowledge, only one other study has evaluated the association of POPs with gonadotropins in postmenopausal women (Persky et al 2011). The consistency of our findings with that study suggest that our results are not due to chance alone. The use of continuous data and centering potential effect modifiers ensured that we did not further limit the power of our analysis. Finally, the consistency among multiple exposures groupings with control for confounders support the biologic plausibility of our results.

#### 4.4 Conclusion

In this investigation, we found significant and borderline significant inverse associations of anti-estrogenic and/or dioxin-like PCB groupings, but not non dioxin-like PCBs, with LH in postmenopausal women, with stronger associations in participants with elevated CRP or BMI. Although not statistically significant in the overall analyses, PCBs were also inversely associated with FSH in the effect modification analysis, with stronger inverse associations in women who were with greater BMI or CRP levels. Adiposity primarily impacted associations of gonadotropins with dioxin-like and anti-estrogenic POPs. Additional studies will be important in delineating specific effects of POPs on sex hormone homeostasis in postmenopausal women.

#### Acknowledgments

A. Lambertino was supported in part by the National Institute for Occupational Safety and Health training grant #T42/OH008672.

#### References

- ATSDR. "Public Health Statement." Toxicological Profile for Polychlorinated Biphenyls (PCBs) (2000): 1–12. <http://www.atsdr.cdc.gov/ToxProfiles/tp17.pdf>. Accessed [July 23, 2020].
- Aylward LL, and Hays SM. "Temporal Trends in Human TCDD Body Burden: Decreases over Three Decades and Implications for Exposure Levels." *Journal of Exposure Analysis and Environmental Epidemiology* 12, no. 5 (2002): 319–328. [PubMed: 12198580]
- Bulun SE, and Simpson ER. "Competitive Reverse Transcription-Polymerase Chain Reaction Analysis Indicates that Levels of Aromatase Cytochrome P450 Transcripts in Adipose Tissue of Buttocks, Thighs, and Abdomen of Women Increase with Advancing Age." *The Journal of Clinical Endocrinology and Metabolism* 78, no. 2 (1994): 428–432. [PubMed: 8106632]
- Bulun Serdar E., and Adashi Eli Y. "The Physiology and Pathology of the Female Reproductive Axis" In *Williams Textbook of Endocrinology*, edited by Melmed Shlomo, Polonsky Kenneth S., Larsen P. Reed, and Kronenberg Henry M., 644–647. Philadelphia: Saunders Elsevier, 2008.
- Cao J, Patisaul HB, Petersen SL: Aryl hydrocarbon receptor activation in lactotropes and gonadotropes interferes with estradiol-dependent and -independent preprolactin, glycoprotein alpha and luteinizing hormone beta gene expression. *Mol Cell Endocrinol* 333: 151–159, 2011 [PubMed: 21187122]
- Cauley JA, Gutai JP, Kuller LH, LeDonne D, and Powell JG. "The Epidemiology of Serum Sex Hormones in Postmenopausal Women." *American Journal of Epidemiology* 129, no. 6 (1989): 1120–1131. [PubMed: 2729251]

- CDC. "1999–2000 Lab Methods." (2020a). <https://wwwn.cdc.gov/nchs/nhanes/ContinuousNhanes/labmethods.aspx?BeginYear=1999>. Accessed July 23, 2020.
- CDC. "2001–2002 Lab Methods." (2020b). <https://wwwn.cdc.gov/nchs/nhanes/ContinuousNhanes/labmethods.aspx?BeginYear=2001> Accessed July 23, 2020.
- Christian CA, Mobley JL, and Moenter SM. "Diurnal and Estradiol-Dependent Changes in Gonadotropin-Releasing Hormone Neuron Firing Activity." *Proceedings of the National Academy of Sciences of the United States of America* 102, no. 43 (2005): 15682–15687. [PubMed: 16230634]
- Clarke IJ "Multifarious Effects of Estrogen on the Pituitary Gonadotrope with Special Emphasis on Studies in the Ovine Species." *Archives Physiology and Biochemistry* 110, no. 1–2 (2002): 62–73.
- Cooke PS, Sato T, and Buchanan DL. "Disruption of Steroid Hormone Signaling by PCBs" In *Recent Advances in Environmental Toxicology and Health Effects*, edited by Larry Robertson and Larry Hansen, 257–263. Lexington, Kentucky: The University Press of Kentucky, 2001.
- Cooke PS, Mukai M, and Buchanan DL. "Cross-Talk between Aryl Hydrocarbon Receptor and Estrogen Receptor Signaling Pathways" In *PCBs: Human and Environmental Disposition and Toxicology*, edited by Larry Robertson and Larry Hansen, 139–148. Urbana and Chicago: University of Illinois Press, 2008.
- Cosma M, Bailey J, Miles JM, Bowers CY, and Veldhuis JD. "Pituitary and/or Peripheral Estrogen-Receptor Alpha Regulates Follicle-Stimulating Hormone Secretion, Whereas Central Estrogenic Pathways Direct Growth Hormone and Prolactin Secretion in Postmenopausal Women." *The Journal of Clinical Endocrinology and Metabolism* 93, no. 3 (2008): 951–958. [PubMed: 18089703]
- Desaulniers D, Leingartner K, Wade M, Fintelman E, Yagminas A, and Foster WG. "Effects of Acute Exposure to PCBs 126 and 153 on Anterior Pituitary and Thyroid Hormones and FSH Isoforms in Adult Sprague Dawley Male Rats." *Toxicological Sciences* 47, no. 2 (1999): 158–169. [PubMed: 10220852]
- Emeville E, Giton F, Giusti A, Oliva A, Fiet J, Thome JP, Blanchet P, and Multigner L. "Persistent Organochlorine Pollutants with Endocrine Activity and Blood Steroid Hormone Levels in Middle-Aged Men." *PLoS One* 8, no. 6 (2013): e66460. [PubMed: 23785499]
- Ferguson KK, Hauser R, Altshul L, and Meeker JD. "Serum Concentrations of p, p'-DDE, HCB, PCBs and Reproductive Hormones among Men of Reproductive Age." *Reproductive Toxicology* 34, no. 3 (2012): 429–435. [PubMed: 22564984]
- Fiedler H "Sources of PCDD/PCDF and Impact on the Environment." *Chemosphere* 32, no. 1 (1996): 55–64. [PubMed: 8564435]
- Gallo MV, Ravenscroft J, Carpenter DO, Schell LM, Akwesasne Task Force on the Environment. "Persistent organic pollutants as predictors of increased FSH:LH ratio in naturally cycling, reproductive age women." *Environmental Research* 164 (2018): 556–564. [PubMed: 29621723]
- Giwercman AH, Rignell-Hydbom A, Toft G, Rylander L, Hagmar L, Lindh C, Pedersen HS, Ludwicki JK, Lesovoy V, Shvets M, Spano M, Manicardi GC, Bizzaro D, Bonfeld-Jorgensen EC, and Bonde JP. "Reproductive Hormone Levels in Men Exposed to Persistent Organohalogen Pollutants: A Study of Inuit and Three European Cohorts." *Environmental Health Perspectives* 114, no. 9 (2006): 1348–1353. [PubMed: 16966087]
- Goldberg M, and Silbergeld E. "On Multiple Comparisons and on the Design and Interpretation of Epidemiological Studies of Many Associations." *Environmental Research* 111, no. 8 (2011): 1007–1009. [PubMed: 21906734]
- Gore AC, Wu TJ, Oung T, Lee JB, and Woller MJ. "A Novel Mechanism for Endocrine-Disrupting Effects of Polychlorinated Biphenyls: Direct Effects on Gonadotropin-Releasing Hormone Neurones." *Journal of Neuroendocrinology* 14, no. 10 (2002): 814–823. [PubMed: 12372006]
- Hagmar L, Bjork J, Sjodin A, Bergman A, and Erfurth EM. "Plasma Levels of Persistent Organohalogenes and Hormone Levels in Adult Male Humans." *Archives of Environmental Health* 56, no. 2 (2001): 138–143. [PubMed: 11339677]
- Haugen TB, Tefre T, Malm G, Jonsson BAG, Rylander L, Hagmar L, Bjorsvik C, Henriksen T, Saether T, Figenschau Y, Giwercman A. "Differences in serum levels of CB-153 and p,p'-DDE,

- and reproductive parameters between men living south and north in Norway” *Reproductive Toxicology* 32 (2011): 261–267. [PubMed: 21736938]
- Hennig B, Meerarani P, Slim R, Toborek M, Daugherty A, Silverstone AE, and Robertson LW. “Proinflammatory Properties of Coplanar PCBs: In Vitro and in Vivo Evidence.” *Toxicology and Applied Pharmacology* 181, no. 3 (2002): 174–183. [PubMed: 12079426]
- Kalkwarf HJ, Khoury JC, and Lanphear BP. “Milk Intake During Childhood and Adolescence, Adult Bone Density, and Osteoporotic Fractures in US Women.” *The American Journal of Clinical Nutrition* 77, no. 1 (2003): 257–265. [PubMed: 12499350]
- Kester MH, Bulduk S, Tibboel D, Meinel W, Glatt H, Falany CN, Coughtrie MW, Bergman A, Safe SH, Kuiper GG, Schuur AG, Brouwer A, and Visser TJ. “Potent Inhibition of Estrogen Sulfotransferase by Hydroxylated PCB Metabolites: A Novel Pathway Explaining the Estrogenic Activity of PCBs.” *Endocrinology* 141, no. 5 (2000): 1897–1900. [PubMed: 10803601]
- Kester MH, Bulduk S, van Toor H, Tibboel D, Meinel W, Glatt H, Falany CN, Coughtrie MW, Schuur AG, Brouwer A, and Visser TJ. “Potent Inhibition of Estrogen Sulfotransferase by Hydroxylated Metabolites of Polyhalogenated Aromatic Hydrocarbons Reveals Alternative Mechanism for Estrogenic Activity of Endocrine Disrupters.” *The Journal of Clinical Endocrinology and Metabolism* 87, no. 3 (2002): 1142–1150. [PubMed: 11889178]
- Kim MJ, Pelloux V, Guyot E, Tordjman J, Bui LC, Chevallier A, Forest C, Benelli C, Clement K, and Barouki R. “Inflammatory Pathway Genes Belong to Major Targets of Persistent Organic Pollutants in Adipose Cells.” *Environmental Health Perspectives* 120, no. 4 (2012): 508–514. [PubMed: 22262711]
- Labrie F, Martel C, and Balsler J. “Wide Distribution of the Serum Dehydroepiandrosterone and Sex Steroid Levels in Postmenopausal Women: Role of the Ovary?” *Menopause* 18, no. 1 (2011): 30–43. [PubMed: 20683211]
- Laughlin GA, Barrett-Connor E, Kritiz-Silverstein D, and von Muhlen D. “Hysterectomy, Oophorectomy, and Endogenous Sex Hormone Levels in Older Women: The Rancho Bernardo Study.” *The Journal of Clinical Endocrinology and Metabolism* 85, no. 2 (2000): 645–651. [PubMed: 10690870]
- Lee DH, Ha MH, Kim JH, Christiani DC, Gross MD, Steffes M, Blomhoff R, and Jacobs DR Jr. “Gamma-Glutamyltransferase and Diabetes—a 4-Year Follow-up Study.” *Diabetologia* 46, no. 3 (2003a): 359–364. [PubMed: 12687334]
- Lee DH, Jacobs DR Jr., Gross M, Kiefe CI, Roseman J, Lewis CE, and Steffes M. “Gamma-Glutamyltransferase Is a Predictor of Incident Diabetes and Hypertension: The Coronary Artery Risk Development in Young Adults (CARDIA) Study.” *Clinical Chemistry* 49, no. 8 (2003b): 1358–1366. [PubMed: 12881453]
- Lee DH, and Jacobs DR Jr. “Association between Serum Concentrations of Persistent Organic Pollutants and Gamma Glutamyltransferase: Results from the National Health and Examination Survey 1999–2002.” *Clinical Chemistry* 52, no. 9 (2006): 1825–1827. [PubMed: 16940464]
- Lobo Rogerio A. “Menopause: Endocrinology, Consequences of Estrogen Deficiency, Effects of Hormone Replacement Therapy, Treatment Regimens” In *Comprehensive Gynecology*, edited by Katz Vern L., Lentz Gretchen M., Lobo Rogerio A., and Gershenson David M., 1039–1079. Philadelphia: Mosby Elsevier, 2007.
- Maggio M, Ceda GP, Lauretani F, Bandinelli S, Corsi AM, Giallauria F, Guralnik JM, Zuliani G, Cattabiani C, Parrino S, Ablondi F, Dall’aglio E, Ceresini G, Basaria S, and Ferrucci L. “SHBG, Sex Hormones, and Inflammatory Markers in Older Women.” *The Journal of Clinical Endocrinology and Metabolism* 96, no. 4 (2011): 1053–1059. [PubMed: 21239514]
- Ohtake F, Takeyama K, Matsumoto T, Kitagawa H, Yamamoto Y, Nohara K, Tohyama C, Krust A, Mimura J, Chambon P, Yanagisawa J, Fujii-Kuriyama Y, and Kato S. “Modulation of Oestrogen Receptor Signaling by Association with the Activated Dioxin Receptor.” *Nature* 423, no. 6939 (2003): 545–550. [PubMed: 12774124]
- Oskam IC, Lyche JL, Krogaas A, Thomassen R, Skaare JU, Wiger R, Dahl E, Sweeney T, Stien A, and Ropstad E. “Effects of Long-Term Maternal Exposure to Low Doses of PCB126 and PCB153 on the Reproductive System and Related Hormones of Young Male Goats.” *Reproduction* 130, no. 5 (2005): 731–742. [PubMed: 16264102]

- Pan W, Ye X, Yin S, Ma X, Li C, Zhou J, Liu W, Liu J. "Selected persistent organic pollutants associated with the risk of primary ovarian insufficiency in women." *Environment International*. 129 (2019) :51–58 [PubMed: 31108393]
- Persky V, Turyk M, Anderson HA, Hanrahan LP, Falk C, Steenport DN, Chatterton R Jr., and Freels S. "The Effects of PCB Exposure and Fish Consumption on Endogenous Hormones." *Environmental Health Perspectives* 109, no. 12 (2001): 1275–1283. [PubMed: 11748036]
- Persky V, Piorowski J, Turyk M, Freels S, Chatterton R Jr., Dimos J, Bradlow HL, Chary LK, Burse V, Unterman T, Sepkovic D, and McCann K. "Associations of Polychlorinated Biphenyl Exposure and Endogenous Hormones with Diabetes in Post-Menopausal Women Previously Employed at a Capacitor Manufacturing Plant." *Environmental Research* 111, no. 6 (2011): 817–824. [PubMed: 21684538]
- Persky V, Piorowski J, Turyk M, Freels S, Chatterton R Jr., Dimos J, Bradlow HL, Chary LK, Burse V, Unterman T, Sepkovic DW, and McCann K. "Polychlorinated Biphenyl Exposure, Diabetes and Endogenous Hormones: A Cross-Sectional Study in Men Previously Employed at a Capacitor Manufacturing Plant." *Environmental Health* 11 (2012): 57. [PubMed: 22931295]
- Petersen MS, Halling J, Jorgensen N, Nielsen F, Grandjean P, Jensen TK, Weihe P. "Reproductive function in a population of young Faroese men with elevated exposure to polychlorinated biphenyls (PCBs) and perfluorinated alkylate substances (PFAS)". *International Journal of Environmental Research and Public Health*. 2018, 15, 1880.
- Phillips DL, Pirkle JL, Burse VW, Bernert JT Jr., Henderson LO and Needham LL. "Chlorinated hydrocarbon levels in human serum: effects of fasting and feeding." *Arch Environ Contam Toxicol*. 1989 18(4): 495–500. [PubMed: 2505694]
- Pirkle JL, Flegal KM, Bernert JT, Brody DJ, Etzel RA, and Maurer KR. "Exposure of the US Population to Environmental Tobacco Smoke: The Third National Health and Nutrition Examination Survey, 1988 to 1991." *The Journal of the American Medical Association* 275, no. 16 (1996): 1233–1240. [PubMed: 8601954]
- Poland A, Knutson J, and Glover E. "Studies on the Mechanism of Action of Halogenated Aromatic Hydrocarbons." *Clinical Physiology and Biochemistry* 3, no. 2–3 (1985): 147–154. [PubMed: 2988846]
- Richthoff J, Rylander L, Jonsson BA, Akesson H, Hagmar L, Nilsson-Ehle P, Stridsberg M, and Giwercman A. "Serum Levels of 2,2',4,4',5,5'-Hexachlorobiphenyl (CB-153) in Relation to Markers of Reproductive Function in Young Males from the General Swedish Population." *Environmental Health Perspectives* 111, no. 4 (2003): 409–413. [PubMed: 12676591]
- Rignell-Hydbom A, Rylander L, Giwercman A, Jonsson BA, Nilsson-Ehle P, and Hagmar L. "Exposure to CB-153 and p,p'-DDE and Male Reproductive Function." *Human Reproduction* 19, no. 9 (2004): 2066–2075. [PubMed: 15284211]
- Safe SH "Polychlorinated Biphenyls (PCBs): Environmental Impact, Biochemical and Toxic Responses, and Implications for Risk Assessment." *Critical Reviews in Toxicology* 24, no. 2 (1994): 87–149. [PubMed: 8037844]
- Safe S, Bandiera S, Sawyer T, Robertson L, Safe L, Parkinson A, Thomas PE, Ryan DE, Reik LM, Levin W, Denomme MA, and Fujita T. "PCBs: Structure-Function Relationships and Mechanism of Action." *Environmental Health Perspectives* 60 (1985): 47–56. [PubMed: 2992927]
- Safe S, Wang F, Porter W, Duan R, and McDougal A. "Ah Receptor Agonists as Endocrine Disruptors: Antiestrogenic Activity and Mechanisms." *Toxicology Letters* 102–103 (1998): 343–347.
- Schisterman EF, Whitcomb BW, Louis GM, and Louis TA. "Lipid Adjustment in the Analysis of Environmental Contaminants and Human Health Risks." *Environmental Health Perspectives* 113, no. 7 (2005): 853–857. [PubMed: 16002372]
- Seto-Young D, Avtanski D, Parikh G, Suwandhi P, Strizhevsky M, Araki T, Rosenwaks Z, and Poretsky L. "Rosiglitazone and Pioglitazone Inhibit Estrogen Synthesis in Human Granulosa Cells by Interfering with Androgen Binding to Aromatase." *Hormone and Metabolic Research* 43, no. 4 (2011): 250–256. [PubMed: 21321839]
- Shaw ND, Histed SN, Srouji SS, Yang J, Lee H, and Hall JE. "Estrogen Negative Feedback on Gonadotropin Secretion: Evidence for a Direct Pituitary Effect in Women." *The Journal of Clinical Endocrinology and Metabolism* 95, no. 4 (2010): 1955–1961. [PubMed: 20133465]

- Stork S, Bots ML, Grobbee DE, and van der Schouw YT. "Endogenous Sex Hormones and C-Reactive Protein in Healthy Postmenopausal Women." *Journal of Internal Medicine* 264, no. 3 (2008): 245–253. [PubMed: 18341528]
- Swedenborg E, and Pongratz I. "AhR and ARNT Modulate ER Signaling." *Toxicology* 268, no. 3 (2010): 132–138. [PubMed: 19778576]
- Sweeney MH, Calvert GM, Egeland GA, Fingerhut MA, Halperin WE, and Piacitelli LA. "Review and Update of the Results of the NIOSH Medical Study of Workers Exposed to Chemicals Contaminated with 2,3,7,8-Tetrachlorodibenzodioxin." *Teratogenesis, Carcinogenesis, and Mutagenesis* 17, no. 4–5 (1997): 241–247.
- Taketoh J, Muto J, Takeda T, Ogishima T, Takeda S, Ishii Y, Ishida T, Yamada H: Suppression of fetal testicular cytochrome p450 17 by maternal exposure to 2,3,7,8-tetrachlorodibenzo-p-dioxin: a mechanism involving an initial effect on gonadotropin synthesis in the pituitary. *Life Sci* 80: 1259–1267, 2007 [PubMed: 17291543]
- Turyk ME, Anderson HA, Freels S, Chatterton R Jr., Needham LL, Patterson DG Jr., Steenport DN, Knobeloch L, Imm P, and Persky VW. "Associations of Organochlorines with Endogenous Hormones in Male Great Lakes Fish Consumers and Nonconsumers." *Environmental Research* 102, no. 3 (2006): 299–307. [PubMed: 16563369]
- Turyk ME, Bhavsar SP, Bowerman W, Boysen E, Clark M, Diamond M, Mergler D, Pantazopoulos P, Schantz S, and Carpenter DO. "Risks and Benefits of Consumption of Great Lakes Fish." *Environmental Health Perspectives* 120, no. 1 (2012): 11–18. [PubMed: 21947562]
- Tyrrell J, Melzer D, Henley W, Galloway TS, and Osborne NJ. "Associations between Socioeconomic Status and Environmental Toxicant Concentrations in Adults in the USA: NHANES 2001–2010." *Environment International* 59 (2013): 328–335. [PubMed: 23892225]
- Uslu U, Sandal S, Cumbul A, Yildiz S, Aydin M, and Yilmaz B. "Evaluation of Estrogenic Effects of Polychlorinated Biphenyls and Organochlorinated Pesticides Using Immature Rat Uterotrophic Assay." *Human & Experimental Toxicology* 32, no. 5 (2013): 476–482. [PubMed: 23515497]
- Van den Berg M, Birnbaum L, Bosveld AT, Brunstrom B, Cook P, Feeley M, Giesy JP, Hanberg A, Hasegawa R, Kennedy SW, Kubiak T, Larsen JC, van Leeuwen FX, Liem AK, Nolt C, Peterson RE, Poellinger L, Safe S, Schrenk D, Tillitt D, Tysklind M, Younes M, Waern F, and Zacharewski T. "Toxic Equivalency Factors (TEFs) for PCBs, PCDDs, PCDFs for Humans and Wildlife." *Environmental Health Perspectives* 106, no. 12 (1998): 775–792. [PubMed: 9831538]
- Van den Berg M, Birnbaum LS, Denison M, De Vito M, Farland W, Feeley M, Fiedler H, Hakansson H, Hanberg A, Haws L, Rose M, Safe S, Schrenk D, Tohyama C, Tritscher A, Tuomisto J, Tysklind M, Walker N, and Peterson RE. "The 2005 World Health Organization Reevaluation of Human and Mammalian Toxic Equivalency Factors for Dioxins and Dioxin-Like Compounds." *Toxicological Sciences: An Official Journal of the Society of Toxicology* 93, no. 2 (2006): 223–241. [PubMed: 16829543]
- van Larebeke N, Hens L, Schepens P, Covaci A, Baeyens J, Everaert K, Bernheim JL, Vlietinck R, and De Poorter G. "The Belgian PCB and Dioxin Incident of January–June 1999: Exposure Data and Potential Impact on Health." *Environmental Health Perspectives* 109, no. 3 (2001): 265–273. [PubMed: 11333188]
- Vested A, Ramlau-Hansen CH, Olsen SF, Bonde JP, Stovring H, Kristensen SL, Halldorsson TI, Rantakokko P, Kiviranta H, Ernst EH, Toft G. "*In utero* exposure to persistent organochlorine pollutants and reproductive health in the human male" *Reproduction* 148 (2014): 635–646. [PubMed: 25190505]
- Vitku J, Heracek J, Sosvorova L, Hampel R, Chlupacova T, Hill M, Sobotka V, Bicikova M, Starka L. "Associations of biphenol A and polychlorinated biphenyls with spermatogenesis and steroidogenesis in two biological fluids from men attending an infertility clinic." *Environment International* 89-90 (2016): 166–173 [PubMed: 26863184]
- Wade MG, Foster WG, Younglai EV, McMahan A, Leingartner K, Yagminas A, Blakey D, Fournier M, Desaulniers D, and Hughes CL. "Effects of Subchronic Exposure to a Complex Mixture of Persistent Contaminants in Male Rats: Systemic, Immune, and Reproductive Effects." *Toxicological Sciences: An Official Journal of the Society of Toxicology* 67, no. 1 (2002): 131–143. [PubMed: 11961226]

- Warner J, Osuch JR, Karmaus W, Landgraf JR, Taffe B, O'Keefe M, Mikucki D, and Haan P. "Common Classification Schemes for PCB Congeners and the Gene Expression of CYP17, CYP19, ESR1 and ESR2." *The Science of the Total Environment* 414 (2012): 81–89. [PubMed: 22119029]
- Weiss G, Skurnick JH, Goldsmith LT, Santoro NF, and Park SJ. "Menopause and Hypothalamic-Pituitary Sensitivity to Estrogen." *The Journal of the American Medical Association* 292, no. 24 (2004): 2991–2996. [PubMed: 15613667]
- Wolff MS, Camann D, Gammon M, and Stellman SD. "Proposed PCB Congener Groupings for Epidemiological Studies." *Environmental Health Perspectives* 105, no. 1 (1997): 13–14. [PubMed: 9074863]
- Zhao Y, Nichols JE, Valdez R, Mendelson CR, and Simpson ER. "Tumor Necrosis Factor-Alpha Stimulates Aromatase Gene Expression in Human Adipose Stromal Cells through Use of an Activating Protein-1 Binding Site Upstream of Promoter 1.4." *Molecular Endocrinology* 10, no. 11 (1996): 1350–1357. [PubMed: 8923461]

**HIGHLIGHTS**

- Lower luteinizing hormone with exposure to anti-estrogenic and dioxin-like POPs
- No overall association of follicle-stimulating hormone with PCBs, dioxins or furans
- Stronger impact postmenopausal women with elevated C-reactive protein or adiposity
- Adiposity primarily impacted associations with dioxin-like and anti-estrogenic POPs



TABLE 1.

## CONGENER GROUPINGS FOR EXPOSURE MEASUREMENTS

Grouping	Congeners <sup>b</sup>
$\Sigma$ TEQs <sup>b</sup>	PCB congeners <b>105, 118, 126, 156, 169</b> ; PCDD congeners <b>1,2,3,7,8-PentaPCDD, 1,2,3,6,7,8-HexaPCDD, 1,2,3,7,8,9-HexaPCDD, 1,2,3,4,6,7,8-HeptaPCDD, 1,2,3,4,6,7,8,9-OctaPCDD</b> ; PCDF congeners <b>2,3,4,7,8-PentaPCDF, 1,2,3,4,7,8-HexaPCDF, 1,2,3,6,7,8-HexaPCDF, 1,2,3,4,6,7,8-HeptaPCDF</b>
$\Sigma$ PCBs <sup>c</sup>	<b>74, 99, 105, 118, 126, 138, 146, 153, 156, 169, 170, 177, 178, 180, 183, 187</b>
$\Sigma$ Non-dioxin-like PCBs <sup>c</sup>	<b>74, 99, 138, 146, 153, 170, 177, 178, 180, 183, 187</b>
$\Sigma$ Mono-ortho PCBs <sup>b</sup>	<b>105</b> , 114, <b>118</b> , 123, <b>156</b> , 157, 167, 189
$\Sigma$ Dioxin-like PCBs <sup>b</sup>	77, 81, <b>105</b> , 114, <b>118</b> , 123, <b>126, 156</b> , 157, <b>169</b> , 189
$\Sigma$ Cooke estrogenic PCBs <sup>b</sup>	1, 3, 4, 8, 15, 18, 21, 31, 44, 47, 48, 49, 52, 54, 61, 70, 75, 77, 80, 95, <b>99</b> , 101, 104, 110, <b>126</b> , 136, <b>153</b> , 155, 184, 188
$\Sigma$ Wolff estrogenic PCBs <sup>b</sup>	101, 174, <b>177, 187</b> , 201
$\Sigma$ Cooke anti-estrogenic PCBs <sup>b</sup>	37, 77, 81, <b>105</b> , 114, <b>126<sup>d</sup></b> , 155, <b>156, 169</b>
$\Sigma$ Wolff anti-estrogenic PCBs <sup>b</sup>	66, <b>74, 77, 105, 118, 126<sup>d</sup></b> , 155, <b>156, 169</b>

<sup>a</sup>Warner et al., 2012.

<sup>b</sup>Congeners in bold were included in the grouping.

<sup>c</sup>Only measured congeners in the grouping are shown.

<sup>d</sup>PCB 126 is listed in both  $\Sigma$ Cooke estrogenic PCBs and  $\Sigma$ Cooke anti-estrogenic PCBs.

**TABLE 2:**  
DEMOGRAPHIC CHARACTERISTICS AND BIOMARKERS IN POSTMENOPAUSAL WOMEN

Characteristic	n	Mean <sup>a</sup>	95% CI	Percentile		
				25 <sup>th</sup>	50 <sup>th</sup>	75 <sup>th</sup>
LnΣTEQs (pg/g)	66	0.11	0.09–0.13	0.07	0.10	0.14
LnΣPCBs (ng/g)	70	1.2	1.2–1.8	0.9	1.4	2.2
LnΣNon-dioxin-like PCBs (ng/g)	87	1.3	1.1–1.5	0.8	1.3	1.9
LnΣMono- <i>ortho</i> PCBs (ng/g)	89	0.19	0.17–0.23	0.12	0.18	0.29
LnΣDioxin-like PCBs (ng/g)	70	0.20	0.16–0.24	0.12	0.18	0.30
LnΣCooke estrogenic PCBs (ng/g)	89	0.39	0.33–0.46	0.26	0.42	0.63
LnΣWolff estrogenic PCBs (ng/g)	87	0.11	0.09–0.12	0.06	0.10	0.16
LnΣCooke anti-estrogenic PCBs (ng/g)	70	0.08	0.07–0.10	0.05	0.08	0.11
LnΣWolff anti-estrogenic PCBs (ng/g)	70	0.28	0.24–0.34	0.17	0.26	0.39
FSH (mIU/mL) <sup>b</sup>	89	68.6	62.8–74.4	50.0	64.9	85.7
LH (mIU/mL) <sup>c</sup>	89	37.6	34.4–40.9	26.4	35.4	45.6
Age (years)	89	54.3	53.3–55.3	51	55	59
BMI (kg/m <sup>2</sup> )	89	29.5	28.3–30.7	25.3	29.4	32.9
LnLipids (mg/dL)	89	675.1	647.3–704.0	606.4	663.4	735.0
LnCRP (mg/dL)	89	0.32	0.26–0.39	0.16	0.34	0.73
Family PIR	89	2.5	2.2–2.8	1.3	2.2	3.5

<sup>a</sup>Mean or geometric mean.

<sup>b</sup>Normal FSH levels for postmenopausal women are 25.8–134.8 mIU/mL.

<sup>c</sup>Normal LH levels for postmenopausal women are 10.0–54.7 mIU/mL.

**TABLE 3:**  
**PEARSON'S CORRELATION COEFFICIENTS AMONG BIOMARKERS AND DEMOGRAPHIC CHARACTERISTICS IN 89 POSTMENOPAUSAL WOMEN**

	$\Sigma$ TEQs	$\Sigma$ PCBs	$\Sigma$ NDL	$\Sigma$ MO	$\Sigma$ DL	$\Sigma$ CE	$\Sigma$ WE	$\Sigma$ CA	$\Sigma$ WA	FSH	LH	Age	BMI	Lipids	CRP	PIR
$\Sigma$ TEQs	0.72*	0.70*	0.81*	0.81*	0.68*	0.79*	0.75*	0.79*	-0.12	-0.26*	0.018	0.070	0.46*	0.21	-0.11	
$\Sigma$ PCBs	0.72*	0.99*	0.92*	0.92*	0.99*	0.94*	0.92*	0.92*	0.00	-0.13	0.00	-0.05	0.24*	0.01	-0.03	
$\Sigma$ NDL	0.70*	0.99*	0.89*	0.89*	0.99*	0.93*	0.91*	0.90*	-0.02	-0.11	0.05	-0.07	0.17	-0.07	0.03	
$\Sigma$ MO	0.81*	0.92*	0.89*	0.89*	0.87*	0.82*	0.94*	0.99*	-0.14	-0.22*	0.06	0.04	0.29*	0.08	-0.08	
$\Sigma$ DL	0.81*	0.92*	0.89*	0.89*	0.89*	0.83*	0.93*	0.99*	-0.08	-0.19	0.03	0.04	0.33*	0.14	-0.12	
$\Sigma$ CE	0.68*	0.99*	0.99*	0.87*	0.89*	0.89*	0.89*	0.90*	-0.04	-0.14	0.02	-0.04	0.17	-0.06	0.08	
$\Sigma$ WE	0.79*	0.94*	0.93*	0.82*	0.89*	0.85*	0.85*	0.82*	-0.07	-0.13	0.03	-0.05	0.16	-0.04	-0.10	
$\Sigma$ CA	0.75*	0.92*	0.91*	0.94*	0.89*	0.85*	0.93*	0.93*	0.00	-0.17	-0.02	-0.08	0.33*	0.04	-0.01	
$\Sigma$ WA	0.79*	0.92*	0.90*	0.99*	0.90*	0.82*	0.93*	0.93*	-0.06	-0.20	0.06	0.02	0.31*	0.13	-0.09	
FSH	-0.12	0.01	-0.02	-0.14	-0.08	-0.04	-0.07	0.00	-0.06	0.77*	-0.16	-0.38*	-0.02	-0.16	0.23*	
LH	-0.26*	-0.13	-0.11	-0.22*	-0.19	-0.14	-0.13	-0.17	-0.20	0.77*	-0.17	-0.26*	-0.09	-0.15	0.07	
Age	0.02	0.00	0.05	0.07	0.02	0.03	-0.02	0.06	-0.16	-0.17	0.01	0.01	-0.10	-0.08	0.03	
BMI	0.07	-0.05	-0.07	0.04	-0.04	-0.04	-0.05	-0.08	0.01	-0.38*	-0.26*	0.01	0.03	0.46*	-0.08	
Lipids	0.46*	0.24*	0.17	0.29*	0.33*	0.17	0.16	0.33*	0.31*	-0.02	-0.09	-0.10	0.03	0.19	-0.10	
CRP	0.21	0.01	-0.07	0.08	0.14	-0.06	-0.04	0.04	0.13	-0.16	-0.15	-0.08	0.46*	0.19	-0.26*	
PIR	-0.11	-0.03	0.03	-0.08	-0.12	0.08	-0.10	-0.01	-0.09	0.23*	0.07	0.03	-0.10	-0.26*		

\* p<0.05

Exposures, lipids and CRP were natural-log transformed.

NDL=non-dioxin-like PCBs; MO=mono-ortho PCBs; CE=Cooke estrogenic PCBs; WE=Wolff estrogenic PCBs; CA=Cooke anti-estrogenic PCBs; WA=Wolff anti-estrogenic PCBs

**TABLE 4:**

## ASSOCIATIONS OF PCBs, DIOXINS AND FURANS WITH GONADOTROPINS IN POSTMENOPAUSAL WOMEN

LnPOP	n	LnFSH			LnLH		
		Effect estimate <sup>a</sup>	95% CI	p-value	Effect estimate <sup>a</sup>	95% CI	p-value
ΣPCBs	70	-2.5	-9.5, 5.1	0.51	-6.5	-14.0, 1.6	0.11
ΣNDL PCBs	87	-2.5	-9.5, 5.1	0.51	-5.5	-12.9, 2.5	0.17
ΣMO PCBs	89	-6.1	-13.1, 1.5	0.11	<b>-8.6</b>	<b>-16.1, -0.4</b>	<b>0.04</b>
ΣDL PCBs	70	-4.2	-11.5, 3.6	0.27	<b>-7.7</b>	<b>-15.5, 0.8</b>	<b>0.07</b>
ΣTEQs	66	-7.6	-16.6, 2.5	0.13	<b>-11.9</b>	<b>-21.3, -1.4</b>	<b>0.03</b>
ΣCA PCBs	70	-4.5	-12.9, 4.8	0.32	<b>-10.7</b>	<b>-19.4, -1.1</b>	<b>0.03</b>
ΣWA PCBs	70	-3.5	-11.0, 4.5	0.37	<b>-8.2</b>	<b>-16.0, 0.3</b>	<b>0.06</b>
ΣCE PCBs	89	-2.8	-8.9, 3.7	0.39	-5.5	-12.0, 1.5	0.12
ΣWE PCBs	87	-4.1	-11.4, 3.7	0.29	-5.8	-13.6, 2.7	0.17

<sup>a</sup>Models adjusted for age, BMI, and Inlipids. Effect estimates are the percent change in the gonadotropin with a doubling in POPs exposure.

NDL=non-dioxin-like PCBs; MO=mono-*ortho* PCBs; DL=dioxin-like PCBs; CE=Cooke estrogenic PCBs; WE=Wolff estrogenic PCBs; CA=Cooke anti-estrogenic PCBs; WA=Wolff anti-estrogenic PCBs

TABLE 5:

## MODIFICATION OF ASSOCIATIONS OF PCBs, DIOXINS AND FURANS WITH GONADATROPINS BY CRP AND BMI

Gonadotropin	POP	Percentile of CRP or BMI	Conditional association of gonadotropin and POP at CRP percentile		Conditional association of gonadotropin and POP at BMI percentile	
			effect	95% CI	effect	95% CI
LH	ΣPCBs	25th	na <sup>c</sup>		0.9	-9.6, 12.6
		75th	na		-13.0	-21.9, -3.2
FSH	ΣPCBs	25th	3.8	-5.7, 14.1	5.4	-4.4, 16.2
		75th	-8.2	-16.5, 0.9	-9.4	-17.6, -0.3
LH	ΣNDL PCBs	25th	3.1	-7.1, 7.4	na	
		75th	-13.7	-22.1, -4.4	na	
LH	ΣMO PCBs	25th	-7.2	-14.7, 1.0	na	
		75th	-14.5	-22.6, -5.5	na	
FSH	ΣMO PCBs	25th	-4.8	-11.8, 2.8	na	
		75th	-11.4	-19.1, -3.1	na	
LH	ΣDL PCBs	25th	-0.1	-10.9, 11.9	1.0	-11.2, 14.7
		75th	-13.0	-21.7, -3.5	-13.2	-22.1, -3.3
FSH	ΣDL PCBs	25th	4.0	-5.9, 15.0	na	
		75th	-10.0	-17.9, -1.3	na	
FSH	ΣTEQs	25th	4.5	-10.6, 22.1	3.0	-9.3, 17.0
		75th	-12.9	-22.7, -1.9	-15.7	-25.2, -4.9
LH	ΣCA PCBs	25th	0.8	-12.1, 15.6	3.0	-5.5, 12.2
		75th	-18.1	-27.3, -7.6	-10.4	-18.3, -1.8
FSH	ΣCA PCBs	25th	7.8	-4.6, 21.8	6.0	-1.7, 14.3
		75th	-12.3	-21.2, -2.3	-9.4	-16.2, -2.0
FSH	ΣWA PCBs	25th	4.5	-5.6, 15.7	7.8	-3.6, 20.6
		75th	-9.3	-17.5, -0.3	-10.6	-18.8, -1.7
LH	ΣCE PCBs	25th	1.8	-7.1, 11.6	na	
		75th	-21.5	-20.1, -4.2	na	
LH	ΣWE PCBs	25th	2.7	-8.4, 15.1	na	
		75th	-31.1	-22.0, -3.1	na	

<sup>a</sup> Models adjusted for age, BMI, and Inlipids.

<sup>b</sup> Conditional associations are shown for significant interaction terms ( $p < 0.05$ ) and can be interpreted as the percent change in the gonadotropin with a doubling in POPs exposure at the indicated level of the effect modifier.

<sup>c</sup>na=not applicable since interaction term p-value>0.05

NDL=non-dioxin-like PCBs; MO=mono-*ortho* PCBs; DL=dioxin-like PCBs; CE=Cooke estrogenic PCBs; WE=Wolff estrogenic PCBs; CA=Cooke anti-estrogenic PCBs; WA=Wolff anti-estrogenic PCBs

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