### Dispatches

# New *Rickettsiae* in Ticks Collected in Territories of the Former Soviet Union

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Dermacentor nuttallii from Siberia, Rhipicephalus sanguineus from Crimea, and Rh. pumilio from the Astrakhan region were infected with Rickettsia sibirica (12%), R. conorii (8%), and the Astrakhan fever agent (3%), respectively. Three new Rickettsiae of the R. massiliae genogroup were identified in ticks by 16S rDNA, gltA, and ompA sequencing.

Rickettsiae are obligate intracellular gramnegative bacteria associated with arthropod vectors, ticks, mites, and insects that, while feeding, can transmit Rickettsiae to animals and humans. The rickettsioses have characteristic clinical features, including fever, headache, maculopapular eruption, and sometimes eschar formation (primary lesion). The number of representatives of the genus *Rickettsia* has increased over the last decades as a result of improved cell culture isolation and agent identification techniques (1). Sequence comparison of gene coding for citrate synthase (gltA)(2), rOmpA outer membrane protein (ompA) (3), and 16S rRNA (4) has become the most reliable method of identifying Rickettsiae (5-8). We describe polymerase chain reaction (PCR) amplification and sequence determination to identify Rickettsiae in naturally infected ixodid ticks in three regions of Russia endemic for tickborne rickettsioses.

*Rhipicephalus pumilio* ticks (65 adults) were collected in 1996 from dogs in the Astrakhan region. *Dermacentor nuttallii* ticks (101 adults) were collected in 1994 in the village of Verhnyi Kouus, the Altay Mountains, Siberia. In 1997, *Rh. sanguineus* ticks (2 adults and 35 nymphs) were collected in the town of Saki, Crimea region, from dogs whose owners had serologic evidence of Mediterranean spotted fever (Figure 1). The ticks were kept at room temperature before being washed in iodized alcohol (10 minutes) just before testing, rinsed in distilled water, and dried on sterile filter paper. DNA was extracted from ticks by using the QIAmp Tissue Kit (QIAGEN, Hilden, Germany). Rickettsial DNA was detected by PCR with primers specific for *Rickettsiae*: RpCS.877p-RpCS.1273r, which amplify a 396-bp fragment of *gltA* (2), and Rr190.70p-190-701 (3), which amplify a fragment of *ompA* from 629 to 632 bp. For all positive ticks, 587 to 590 bp of *ompA* were sequenced by using the ABI PRISM Dye



Figure 1. Areas from which ticks in the study were collected.

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Terminator Cycle Sequencing Kit with Amplitap Polymerase FS (PE Applied Biosystems, Warrington WA1 4SR, UK). Sequences were analyzed with the Applied Biosystem 377 automatic sequencing system. For newly detected genotypes, sequences of 16S rRNA encoding gene, *gltA*, and *ompA* were determined as previously described (2-4) (see Figure 2 for GenBank codes).

We detected two different *Rickettsiae* in *Rh. pumilio* (Astrakhan fever agent and RpA4 genotype); two *Rickettisiae* from *D. nutallii* in Siberia (*Rickettsia sibirica* and DnS14 and DnS 28 genotypes); and *R. conorii* from *Rh. sanguineous* ticks in Crimea (Table).

Our results confirm previous data of high epidemic activity of the Altay focus for North Asian tick typhus and the crucial role of D. nuttallii as a reservoir of R. sibirica infection (9). Our results are also consistent with those of a study in 1991 based on hemolymph testing (10), in which 3.2% of ticks from the Astrakhan region were demonstrated to be infected with Rickettsiae.

An outbreak of Mediterranean spotted fever due to infection with R. conorii occurred in Crimea from 1947 to 1957. Only sporadic cases of the disease were reported (11) until 1995, when the incidence of Mediterranean spotted fever increased in central Crimea, with 40 cases in 1996 and more than 70 in 1997. Most cases occurred in the summer, when the Rh. sanguineus nymphs (principal vectors of *R. conorii*) (1) were active. Our results, showing that 8% of the Rh. sanguineus studied contained R. conorii DNA, provide further evidence of the Mediterranean spotted fever outbreak in the region. To date, only the *R. conorii* strain M-1, isolated in the territories of the former Soviet Union (the Black Sea coast of Georgia), has been genetically characterized. This strain is genetically distinct from the other strains of R. conorii, i.e., Indian tick typhus and the Moroccan and Malish strains (3). Our detection of the R. conorii strain identical to the Malish strain is the first evidence of the genetic heterogeneity of R. conorii in the region.

The *ompA* sequences obtained from PCRamplified products were different from those described for the known *Rickettsiae* for one DNA sample extracted from *Rh. pumilio* from the Astrakhan region (RpA4) and four DNA samples from *D. nuttallii* collected in Siberia (DnS14, DnS28, DnS79, DnS94). The sequences for the samples from *D. nuttalii* (DnS28, DnS79, and DnS94) were identical but differed from those of DnS14 and *Rh. pumilio* RpA4/2.

The three new rickettsial agents were closely related and branched with members of the *R. massiliae* group, together with *R. rhipicephali*, Bar 29, *R. aeschlimannii*, and *R. montanensis* (Figure 2). Comparison of the sequences obtained by using the program BLAST demonstrated that they also differed from those of the Cadiz agent characterized from *Ixodes ricinus* in Spain (6), those of the *Cooleyi* genotype characterized from *I. scapularis* (5), MOA and WB-8-2 isolated from *Amblyomma americanum* and *I. scapularis*, respectively (8), and *R. peacockii* (7) in the United States.

The pathogenicity of the members of the *R. massiliae* group is unknown, and their main reservoirs are regarded as ticks of the genus *Rhipicephalus* for *R. massiliae* and Bar 29. *R. aeschlimannii* has been isolated from *Hyalomma marginatum* and *R. montanensis* from ticks of the genus *Dermacentor*. *R. rhipicephali* has been demonstrated in ticks of the genus *Dermacentor* and in *Rh. sanguineus* (1). The similarity of *gltA*, *ompA*, and 16S rRNA gene sequences indicates that these three new

Tick species	Location	No. positive ticks/total examined	% infected ticks	Rickettsial species
Rhipicephalus pumilio	Astrakhan region	2/65	3	Astrakhan fever agent
Rh. pumilio	Astrakhan region	1/65	1.5	RpA4 genotype
Dermacentor nutallii	Siberia	12/101	12	Rickettsia sibirica
D. nutallii	Siberia	4/101	4	DnS14 and DnS28 genotypes
Rh. sanguineus	Crimea	3/37	8	R. conorii (Malish strain)

Table. Ticks infected in regions of the former Soviet Union

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Figure 2.<sup>1</sup> Phylogenetic tree of *Rickettsiae* inferred from comparison of the *ompA* sequences. The known tick vectors for the bacteria presented on the dendrogram are indicated on the right. The *ompA* sequences were aligned by the multisequence alignment program CLUSTAL in the BISANCE software package. Phylogenetic relationships were inferred by using version 3.4 of the PHYLIP software package. Evolutionary distance matrices, generated by DNADIST, were determined by the Kimura method. Matrices were used to construct dendrograms by the neighbor-joining method. Data were also examined by using parsimony analysis (DNAPARS in PHYLIP), and bootstrap analyses were performed to investigate the stability of the trees obtained. Bootstrap values were obtained for a consensus tree based on 100 randomly generated trees by using SEQBOOT and CONSENSE in the same package. The percentage of similarity between strains was determined by using the PC/GENE software package.

 $\label{eq:sequences} ^{1}\mbox{The received sequences of the new rickettsial agents have been deposited in GenBank. Sequences of $ompA$ were deposited as two parts under accession numbers: DnS28, AF120018 and AF120019; DnS14, AF120021, and AF120020; RpA4, AF120022, and AF120023. The 16S rRNA gene and $gltA$ sequences have been deposited in GenBank under accession numbers DnS28, $gltA$ - AF120027, 16S rRNA gene - AF120024; DnS14, $gltA$ - AF120028, 16S rRNA gene - AF120025; RpA4, $gltA$ - AF120029, 16S rRNA gene - AF120026.$ 

agents are close to each other (from 99.7% to 99.9%) and could constitute a new rickettsial species.

In the United States, various tickborne Rickettsiae occur in areas endemic for R. rickettsii, the agent of Rocky Mountain spotted fever (7). Similarly, in Mediterranean countries, several recently described Rickettsiae have been found in ticks of the Rh. sanguineus complex in the regions endemic for Mediterranean spotted fever caused by R. conorii (6). The effects of the presence of different Rickettsiae on the prevalence of infection rates of ticks with individual Rickettsiae and on the epidemiology of

infections in humans have yet to be determined. R. sibirica and the Astrakhan fever agent are prevalent in Siberia and the Astrakhan region, respectively, but the pathogenicity of the new rickettsial genotypes has yet to be investigated.

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