Global Distribution of Rubella Virus Genotypes

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Phylogenetic analysis of a collection of 103 E1 gene sequences from rubella viruses isolated from 17 countries from 1961 to 2000 confirmed the existence of at least two genotypes. Rubella genotype I (RGI) isolates, predominant in Europe, Japan, and the Western Hemisphere, segregated into discrete subgenotypes; international subgenotypes present in the 1960s and 1970s were replaced by geographically restricted subgenotypes after ~1980. Recently, active subgenotypes include one in the United States and Latin America, one in China, and a third that apparently originated in Asia and spread to Europe and North America, starting in 1997, indicating the recent emergence of an international subgenotype. A virus that potentially arose as a recombinant between two RGI subgenotypes was discovered. Rubella genotype II (RGII) showed greater genetic diversity than did RGI and may actually consist of multiple genotypes. RGII viruses were limited to Asia and Europe; RGI viruses were also present in most of the countries where RGII viruses were isolated.

Rubella virus infection during the first trimester of pregnancy can lead to severe birth defects (congenital rubella syndrome) (1). Live attenuated vaccines, available since the late 1960s (2), are currently in use in roughly half of the countries in the world, including all industrialized countries, although vaccine coverage varies widely (3). Concentration on comprehensive rubella vaccination has recently increased in developing countries in conjunction with measles elimination efforts, particularly in Latin America (4,5). As part of the surveillance component of these efforts, an understanding of the worldwide molecular epidemiology of rubella virus, which is limited, is necessary.

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Rubella virus is an RNA virus that is the sole member of the Rubivirus genus, within the Togaviridae family (6). The rubella virus genome is ~10,000 nucleotides and encodes five protein products, including three virion proteins: the C or capsid protein and two envelope glycoproteins, E1 and E2. The E1 gene sequence has been used for genotyping and phylogenetic analysis of rubella virus isolates (7–10). Thus far, rubella viruses from Europe, Asia, and North America have been found for the most part to group in a single genotype (Rubella Genotype I or RGI) that has a maximum diversity at the nucleotide level of ~5%. However, a limited number of viruses from Asia (China and India), and more recently Italy, formed a distant phylogenetic branch, differing from RGI viruses by 8% to 10%, which was designated Rubella Genotype II (RGII) (8,9,11,12). These two genotypes belong to the single rubella virus serotype (11). Because of limited sampling, the geographic range of RGII has not been determined.

This study was designed to increase information and understanding on worldwide molecular epidemiology of rubella virus. We have performed combined phylogenetic analysis on viruses from earlier studies (8–10) and, to gain further information on RGII viruses, we included viruses collected from the Eastern Hemisphere, namely Russia, South Korea, China, New Zealand, and Israel.

Materials and Methods

Rubella Isolates and Sequences

A total of 103 rubella virus E1 gene nucleotide sequences were used in this study; "new" sequences not reported in previous studies are shown in Table 1, a complete list is available online (http://www.cdc.gov/ncidod/EID/vol09no12/03-0242.htm#table1). The length of the sequence was 1179 nt, which covered 8291–9469 nt

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| Isolate | Isolation site and y | GenBank no. |
|--------------------------------------|--------------------------------------|-----------------------|
| China | | |
| AH2/AH-CHN99 | Anhui, China 1999 AY326 | |
| AH5/AH-CHN99 | Anhui, China 1999 | AY326351 |
| Germany | | |
| BCM/-GER91 | Germany 1991 | AY326341 |
| G696/-GER98 | Germany 1998 | AY326342 |
| Israel | | |
| I11/TA-ISR68 | Tel Aviv, Israel 1968 | AY326335 |
| I19/HF-ISR72 | Haifa, Israel 1972 | AY326338 |
| I9/JS-ISR75 | Jerusalem, Israel 1975 | AY326334 |
| I13/BB-ISR79 | Bene-Berak, Israel 1979 | AY326336 |
| I15/JF-ISR78 | Jaffa, Israel 1978 | AY326337 |
| I34/TB-ISR88 | Tiberias, Israel 1988 | AY326339 |
| I76/EV-ISR92 | Ein-Vered, Israel 1992 | AY326340 |
| India | | |
| MTS/-IND00 | India 2000 | AY326343 ^b |
| Japan | | |
| J05/TK-JPN93 | Tokyo, Japan 1993 | AB072382 |
| J91/GM-JAP94 | Gunma, Japan 1994 | AB072384 |
| J86/ST-JAP95 | Saitama, Japan 1995 | AB072387 |
| J13/HS-JAP97 | Hiroshima, Japan 1997 | AY397695 |
| Korea | | |
| AN1/SO-KOR95 | Seoul, Korea 1995 | AY326345 |
| AN3/SO-KOR96 | Seoul, Korea 1996 | AY326346 |
| AN5/SO-KOR96 | Seoul, Korea 1996 | AY326347 |
| AN6/SO-KOR96 | Seoul, Korea 1996 | AY326348 |
| New Zealand | | |
| JC1/AL-NEZ81 | Auckland, NZ 1981 | AY326331 |
| JC2/AL-NEZ91 | Auckland, NZ 1991 | AY326332 |
| JC5C/AL-NEZ91 | Auckland, NZ 1991 | AY326333 |
| Russia | | |
| C4/MO-RUS67 | Moscow, Russia 1967 | AY247015 |
| C19/MO-RUS68 | Moscow, Russia 1968 | AY247016 |
| C44/MO-RUS69 | Moscow, Russia 1969 | AY247017 |
| C68/MO-RUS73 | Moscow, Russia 1973 | AY247018 |
| C74/MO-RUS97 | Moscow, Russia 1997 | AY247019 |
| ^a New E1 gene sequences n | ot previously reported in earlier st | udies (8-12) are |

Table 1. "New" rubella virus E1 gene sequences used in this study $\ensuremath{\mathsf{study}^\mathsf{a}}$

"New E1 gene sequences not previously reported in earlier studies (8-12) are listed in this Table. A complete list of sequences used in this study is available online (http://www.cdc.gov/ncidod/EID/vol09no12/03-0242.htm#table1). "Isolated in Seattle, WA, USA.

of the rubella virus genome (the complete E1 gene is between nts 8258–9700 [13]). This collection consisted of sequences from new isolates from China, Israel, Japan, Korea, New Zealand, and Russia and representative sequences from previous studies (8–10). Methods of isolate propagation and E1 gene sequence determination were as previously described (9).

Genetic Sequence Similarities and Distances

Distance matrix tables were computed by using the Old distance with Simple correction method in the GCG software package (Wisconsin package version 10.0, Genetics Computer Group, Madison, WI). These tables were used to calculate the average genetic distance among viruses in a single genotypic group or between viruses in two genotypic groups.

Phylogenetic Analysis

The phylogenetic trees (Figures 1 and 2) were made using Tree-Puzzle 5.0 (14) (maximum likelihood [ML] criterion with 10,000 or 25,000 quartet puzzling steps and HKY85 model of substitution [15]) and viewed by using TreeView (16). To test the consistency of branching, additional software packages were used to construct trees, including PAUP (David L Swofford. 2001. ver 4.0, Sinauer Associates, Sunderland, MA) and PHYLIP (Joseph Felsenstein, ver 3.6 α , July 2000, WA). Both ML and maximum parsimony (MP) algorithms in these packages were used. A histogram of the ML distances computed by Tree Puzzle 5.0 (14) was constructed with Excel (Microsoft, Redmond, WA).

Results

The sequences used in this study consist of representative sequences from three earlier studies (Europe, North America, and Asia, 1961–1997 [8]; Italy, 1991–1997 [9]; and the United States, 1997-2001 [10]) and sequences of viruses collected in Russia, South Korea, New Zealand, Israel, China, Japan, and the United States that have not been reported (these new sequences are listed in Table 1; a complete sequence list is available online (http://www. cdc.gov/ncidod/EID/vol09no12/03-0242.htm#table1). The sequences from the three earlier studies have not previously been amalgamated into a single phylogenetic study. Phylogenetic trees produced with different programs yielded the same major branches, and only representative trees are shown (Figure 1). Branches were defined as reproducible clusterings of sequences by these different programs.

Most of the new sequences belonged to RGI, which emanate from a single node in the tree shown in Figure 1. For simplicity, the tree shown in Figure 1A contains only half of the RGI sequences; the RGI node with all of the RGI sequences is shown in Figure 1B. New sequences belonging to RGI included both from China, all three from New Zealand, all four from Japan, five of six from Israel, two of four from South Korea, and one of five from Russia. Additionally, all of the sequences from the U.S. study (10) and 19 of 21 from the Italy study (9) were RGI. Fourteen sequences were grouped on branches removed from the RGI node, including the three viruses originally used to define RGII, two RGII viruses isolated from Italy (1994) [9], and 9 of the new sequences: 4 from Russia (3 from 1967 to 1969 and 1 from 1997), 2 from South Korea (1996), 1 from Israel (1968), 1 from China (1979), and 1 isolated in the United States from a case contracted in India (2000).

RGI Viruses

The sequences of viruses isolated before 1980 are colored black in the expanded RGI node (Figure 1B). These



Figure 1. Phylogenetic trees. Unrooted tree was made by the maximum likelihood method in the Tree-Puzzle 5.0 program (25,000 puzzling steps for the tree in A; 10,000 puzzling steps for the tree in B) using the complete E1 gene sequence (1179 nt). Bootstrapping values (out of 100) for each node are given. The tree in A was constructed with half of the rubella genotype I (RGI) and all of the RGII sequences (to allow the reader to read the RGI virus designations); the tree in B is a blowup of the RGI node from a tree constructed with all of the sequences. In B, sequences used in the previous study (8) are designated by an (*), and sequences of viruses isolated before 1980 are in black. Branches are color-coded as follows: RGI International 1961–1986 and 1964–1981, black; RGI Europe 1972–1991, light green; RGI Europe 1986–1994 and Europe 1991–1998, green; RGI China, 1999, gold; RGI USA, 1990–2000, light blue; and a branch containing sub-branches from Japan 1987–1991, International 1997–2000, Japan, Korea 1994–1996, New Zealand, 1991, and Japan-Philippines, 1997, dark blue. Of these, the black International, green Europe, light-blue USA, and dark-blue branches were recognized in the previous study (the light-blue branch as US-Japan and the dark-blue branch as Japan-Hong Kong).

sequences, which were from viruses isolated in Europe, North America, and Asia, congregated around the central node (International 1964-1981) or segregated into one of two branches, denoted International 1961-1986 and Europe 1972–1991. Many of these sequences were used in our previous study (8; marked with an asterisk) in which the sequences in the International 1961-1986 branch formed three related branches, the sequences in the International 1964–1981 nodal congregation were spread along the baseline of the dendogram, and the sequences in the Europe 1972-1991 branch were at the base of the Europe branch. Additions to the International 1964–1981 nodal congregation included sequences from a virus isolated in Russia in 1973 and a virus isolated in New Zealand in 1981; a sequence from a virus from Israel 1978 was added to the International 1961-1986 branch. The Europe 1972–1991 branch (colored gold in Figure 1) contained viruses from Israel (1972, 1975, 1979, and 1988), Italy (1991), and the United Kingdom (1978 and 1986; both

used in the previous study [8]). The TCRB vaccine strain (Japan, 1967) sequence segregated onto this branch; in our previous study (8); the TCRB sequence was at the base of the European branch. In summary, while the branching pattern of sequences of viruses isolated internationally before ~1980 (8) was basically preserved, the addition of sequences led to definition of a new branch of viruses from Europe and Israel, isolated between 1972 and 1991.

Newly added sequences of viruses isolated after 1990 segregated primarily onto the three continent-restricted branches defined in our earlier study (8), generally in a geographically consistent manner. These branches, designated Europe, Japan-Hong Kong, and US-Japan in the earlier study (8), are colored green, dark blue, and light blue (the US-Japan branch was redesignated USA 1990–2000; see below). The Europe branch was augmented by sequences of viruses from Italy, Germany, and Israel (1991–1998) and subdivided into two sub-branches, one containing sequences of viruses isolated in Europe from



Figure 2. Phylogenetic trees. Trees were made by the maximum likelihood method in the Tree-Puzzle 5.0 program (1,000 puzzling steps) using the 5' half (640 nt: 8291–8930) or 3' half (539 nt: 8931–9469) of the E1 gene sequence. Bootstrapping values (out of 100) for each node are given.

1986 to 1994 (Europe 1986-1994) and a second containing sequences of viruses isolated in Europe and Israel from 1991 to 1995 (Europe 1991-1998). Several newly added sequences of viruses from the United States and Latin America (1997-2000) enlarged the USA 1990-2000 branch; these viruses are thought to be endemic in regions bordering the United States and to seed rubella outbreaks and clusters in that country (10). The only two viruses from Japan in this branch were isolated from a single province in 1991; it was previously hypothesized that this finding represented transport of viruses from the United States to Japan or vice versa (8). In light of the isolation of several viruses in this group subsequently from the United States or Latin America and no viruses from Japan (either before or after 1991), the former transport pathway likely occurred; thus, this branch has been redesignated USA 1990-2000.

The Japan-Hong Kong branch expanded considerably to include sequences of a pair of viruses from Japan and the Philippines (1997) that were related to each other, two sequences of viruses from New Zealand (1991) that were related to each other, and sequences of a group of five related viruses isolated in Japan and South Korea from 1995 to 1996. Another distinct sub-branch was formed by a closely related group of sequences of viruses associated with outbreaks in Europe, the United States, and the Caribbean from 1997 to 2000 (International, 1996–2000). The viruses in this branch caused several outbreaks in the northeastern United States and on cruise ships in Florida (10) and were isolated during a rubella epidemic in Italy in 1997 (9); the relatedness of these viruses was not previously recognized. A novel branch contained sequences of two viruses from China (1999) (China, 1999, gold in Figure 1B).

RGII Viruses

The tree depicted in Figure 1A indicated that sequence divergence was greater among RGII viruses than among RGI viruses. RGII viruses segregated into three distinct clusters (RGIIA, RGIIB, and RGIIC), all of which were supported by bootstrapping values of >90. Only the RGIIA cluster contained more than one virus isolated within the past decade, and this active cluster contained viruses from diverse locations (India, China, Korea, Italy). The other two clusters contained viruses from a single location (RGIIB-Russia; RGIIC-Beijing, China). The RGII clusters were more distant from each other than were the RGI branches. Variability between RGI and the three RGII clusters is shown in Table 2. Maximal variability among RGI viruses was ~5.8%, however, RGII viruses varied by up to $\sim 8.0\%$. Viruses within each of the three RGII clusters varied by up to 5.5% and the clusters differed from each other by an average of ~7% (range of variability between viruses in the RGII clusters: RGIIA-RGIIB: 6.35%-7.78%; RGIIA-RGIIC: 6.86%-7.95%; RGIIB-RGIIC: 6.85%-7.53%). Average variation of the RGII clusters from RGI was from $\sim 6\%$ to 8% (range 5.5%-10.3%, which translates to a range of 0.8% to 2.1% variation at the amino acid level).

Evidence for Recombinant Virus

Use of the entire E1 gene resulted in excessive time being required to run phylogenetic programs with the large number of sequences included in this study (e.g., trees in Figure 1 took days to compute). We therefore investigated using smaller segments within the E1 gene (five windows of ~400 nt encompassing nt 8291–8640, 8491–8890, 8687–9088, 8891–9290, and 9100–9469 of the genome) to construct phylogenetic trees. All major branches shown in Figure 1 were preserved in trees constructed from each of

| Table 2. Intra- and | intergoup genetic distances ar | nong rubella genoty | pe I (RGI) and RGII clu | isters ^a | |
|---------------------|--------------------------------|---------------------|-------------------------|---------------------|-------|
| | _ | | Mean distance from | | |
| Genotype/cluster | Intragroup variability | RGII | RGIIA | RGIIB | RGIIC |
| RGI | 0.08-5.75 | 7.28 | 7.59 | 6.20 | 8.21 |
| RGII | 0-7.95 | | | | |
| RGIIA | 0-5.41 | | | 7.24 | 7.13 |
| RGIIB | 0.42-1.95 | | | | 7.19 |
| RGIIC | 2.54 | | | | |

^aRanges and mean genetic distances (% nucleotide difference) were determined from all pairwise combinations from viruses in these groups (Figure 1).

these windows, although placement and joining of the branches varied at the base of the tree. During this investigation, the sequence of one isolate, FAR-GER89, was found to group with three contemporaneous viruses in the Europe 1986-1994 branch (TOM-UK86, FEL-UK88, and D075-GER92) in the three 5'-most windows within the E1 gene, but with three International 1964-1981 viruses (RA27-USA64, JC1-NEZ81, and THE-USA64) in the two 3'-most windows (illustrated in Figure 2). When these seven sequences were aligned, over the 5' 630 nt, 14 nt characteristics of the TOMI/FEL/D075 or THE/JC1/RA27 sequences were identified; of these, FAR had 12 characteristics to the TOMI/FEL/D075 sequences, 1 characteristic to the THE/JC1/RA27, sequences and 1 unique nucleotide (data not shown). In contrast, over the 3' 540 nt of the E1 gene, 11 nt characteristics of the TOM/FEL/D075 or THE sequence were identified and of these 11 characteristic nts, FAR shared all 11 with the THE sequence and none with the TOM/FEL/D075 sequences. These results indicate that FAR may be a recombinant between viruses from these two Genotype I groups. The FAR-GER89 sequence is on its own branch, emanating from the RGI node (Figure 1); similar observations have been made with poliovirus recombinants (17).

Discussion

In this study, we extended phylogenetic analysis of rubella viruses collected worldwide. The baseline for this study was an analysis (8) of viruses collected from Europe, North America, and Asia, 1961–1997; we found that an international genotypic group existed until ~1980 and was replaced by continent-restricted genotypic groups after ~1980. In this study, this analysis was augmented by inclusion of comprehensive collections from Italy (9) and the United States (10) as well as viruses collected from new locations. In the previous analysis (8), a second genotype was identified among a limited number of specimens collected from Asia and a specific goal of this study was to analyze viruses from new locations in hopes of learning more about this second genotype.

Use of the entire E1 gene resulted in excessive time being required to run phylogenetic programs with the large number of sequences included in this study. We therefore investigated using ~400-nt segments within the E1 gene. While the major genotypic and subgenotypic grouping were preserved in these segmental trees, the trees produced from the nt 8687-9088 window were most similar to those produced from the entire E1 gene. This window was named the "molecular epidemiology window." As shown in Table 3, this window had a similar to somewhat higher intersequence variability than the E1 gene and preserved the GC content and intersequence transition/transversion ratio exhibited by the E1 gene. Although virus isolates were used in this study, amplification and sequencing of E1 gene segments directly from clinical specimens using reverse transcription-polymerase chain reaction (RT-PCR) are at hand (18-22). Our findings indicate that RT-PCR products from any region of the E1 gene will produce phylogenetic trees consistent with those from the entire E1 gene. However, thus far the regions amplified by in these studies have not been standardized.

Analysis of the segmental trees indicated that the FAR-GER89 isolate might have arisen by a recombination event within the E1 gene between a virus from the Europe 1986-1994 branch (TOM-UK86, FEL-UK88, D075-GER92) and a virus from the International 1964-1981 nodal cluster (THE-US64, RA27-USA64, JC1-NEZ81). Such an event is temporally consistent because viruses related to the International 1964-1981 nodal cluster have been isolated in Italy as late as 1991 (9) but were not included in this study. Although rubella virus recombination in cell culture has been documented (23), this report is the first of a natural recombinant. Although the FAR-GER89 isolate was related to the RA27/3 vaccine strain in the 3' end of the E1 gene, the FAR-GER89 sequence contains none of the nucleotides characteristic of the RA27/3 sequence (7); thus, the recombination event did not involve a vaccine virus.

In our previous analysis (8), all of the viruses isolated in Europe, North America, and Japan were RGI. This finding was maintained with the expanded collections from all three regions used in this study with the exception of two RGII viruses isolated in Italy. Additionally, most of the viruses in the expanded RGI collection from these regions segued into the previously defined RGI branches (8), with the exception of a newly defined Europe 1972–1991

| Table 3. Comparison of genotypic statistics using the E1 gene and the molecular epidemiology window (MEW) | | | | | | | | | |
|---|----------------------|---------------------------|------------|--|-------------|-----------|--------------|----------------------------|------|
| | | | Inter | Intergenotypic distance ^{a,b} | | 1 | ntragenotypi | c distances ^{a,c} | |
| | G+C | Transition/ | | | Mean | RG | [| R | GII |
| Window | content ^a | transversion ^a | Range | Mean | RGI vs RGII | Range | Mean | Range | Mean |
| E1 | 66.6 | 6.34 | 0.08~10.32 | 4.92 | 7.28 | 0.08~5.75 | 3.55 | 0~7.95 | 5.66 |
| MEW | 66.1 | 6.15 | 0~11.69 | 4.97 | 8.32 | 0~5.97 | 3.49 | 0~8.71 | 6.61 |

^aStatistics were determined from all sequences listed in Table 1.

^bThe range and mean of genetic distances (% difference) were determined by using all sequences. The mean rubella genotype I (RGI) vs. RGII was determined from all of the pairwise RGI-RGII combinations.

"The range and mean of genetic distances were determined for RGI and RGII viruses separately.

branch that contained viruses at the base of the Europe branch defined in the previous study and a novel branch consisting of two viruses from China (1999). In the case of viruses from Europe, with a larger number of viruses representing a longer time span, these viruses belonged to two branches, the second of which divided into two subbranches. The RGI viruses from Israel were related to the European viruses and fit into the temporal pattern of isolation of the European viruses. The temporal pattern of isolation of viruses in the European branches and sub-branches (1972-1991, 1986-1994, and 1991-1998) indicated that temporal displacement of genotypic groups occurred, as had been noted in our previous study on viruses from Italy (9). These European branches and sub-branches have been more recently displaced by the International 1997-2000 sub-branch of the Japan-Hong Kong branch. The addition of viruses isolated recently in the United States (10) clarified that the US-Japan branch in the previous study (8) was a United States branch, viruses from which, when transported to Japan, caused an outbreak in 1991. Viruses in the USA 1990-2000 branch were related to the limited number of viruses available from Latin America.

The branch termed a Japan-Hong Kong branch in our earlier study (8) contained viruses from Japan and Hong Kong isolated between 1976 and 1991. Ten representative viruses were selected from that branch for this study, including 2 viruses from Japan (1976 and 1977) and 1 virus from Hong Kong (1985) that grouped at the base of the branch as well as several viruses from Japan (1987–1991) that formed a sub-branch. This branch was expanded by the addition of four sub-branches containing viruses from New Zealand, 1991; Japan and Korea, 1994-1996; Japan and the Philippines, 1997; and an international sub-branch that contained viruses associated with epidemics and outbreaks in Europe and the United States during 1997 to 2000 that were unrelated to viruses previously isolated from those regions. Recent evidence indicates that these viruses are closely related to viruses from China (Z. Zhen et al., unpub. data). Thus, the appearance of this sub-branch in Europe and the United States was likely due to intercontinental transport. Considering that viruses from this sub-branch have been predominant in Europe since 1997 and have been one of the two genotypic groups isolated recently in the United States (USA 1990–2000 is the other), an international genotypic branch of rubella virus appears to have emerged after a ~20-year hiatus since the previous demonstrable international branch. Along with this branch, the two other currently active branches are the USA 1990-2000 branch and the China 1999 branch, only represented by two viruses from the collection used in this study.

With its expanded virus collection, this study demonstrates that RGI rubella virus isolates segregate into discrete subgenotypic groups (i.e., branches) that exhibit geographic and temporal consistency. These groups could provide the basis for a standard classification scheme, as has been developed for other viruses. In this study, the number of available RGII viruses also increased to 14 in contrast to the 3 available in the previous analysis; the RGII viruses segregated into three discrete clusters (RGIIA, RGIIB, and RGIIC). RGI viruses formed a discrete cluster clearly distinguishable from the most closely related RGII cluster (RGIIB) with no indication of intermediate viruses. The maximum diversity within RGI was 5.8%, and RGI viruses were an average of >7% different from the RGII viruses (Table 2); when one considers the extensive nature of the RGI virus collection, this is a working definition of a rubella virus genotype. RGII was originally defined to contain a limited number of sequences (three) that distinctly differed from the RGI viruses that made up most of the collection (8). The sequences in each of the RGII clusters vary by up to 5.4%, and the average distance between viruses in each of the clusters is >7% (Table 2). Thus, each of these clusters could represent a genotype. This hypothesis is strengthened by a histogram of the intersequence distances that show a bimodal distribution with peaks from 0.5%-6.0% to 6.5%-11.0%, corresponding to intra- and intergenotypic distances (Figure 3). The bimodal distribu-



Figure 3. Histogram of genetic distances between rubella virus sequences. The histogram, showing the distribution of all of the pairwise distances between the rubella virus sequences in the study, was constructed from the maximum likelihood distance matrix computed by Tree Puzzle 5.0 program

tion also indicated equivalency of genotypic groups (24). However, in light of the limited number of RGII specimens, more specimens are needed to fully characterize the extent of the diversity within non-RGI viruses, and criteria need to be established for definition of additional genotypes. In this regard, as represented by the viruses in this collection, only one of the RGII clusters (RGIIA) has been repeatedly active in the past decade. However, viruses belonging to the RGII C cluster (China) have been recently isolated (Z. Zhen et al., unpub. data).

Figure 4 shows the countries from which RGI and RGII viruses have been isolated. RGII has not been isolated from an indigenous case outside of the Eastern Hemisphere. Only RGI viruses have been isolated from the United Kingdom, Belgium, Germany, Japan, and New Zealand; other studies have shown RGI viruses in Brazil (10) and Greece (23). Only RGII viruses have been isolated from India, and both genotypes have been isolated from Russia, Italy, Israel, China, and Korea. In Italy and Korea, the two genotypes were isolated in the same year. In both Italy and Israel, isolation of RGII viruses was only during a single year. By contrast, RGI viruses were isolated in both previous and subsequent years. This finding indicates that these RGII viruses were imports, although the relative distant relatedness of the two Italy isolates suggests at least two importation events, albeit in a single year (9). In China, RGII viruses were isolated in 1979 and 1980; the most recent isolates (1999) were RGI, although RGIIC viruses were recently isolated (Z. Zhen et al., unpub. data). These data indicate that RGI has a wider worldwide distribution than does RGII and that in much of the world RGI is the sole genotype. Recent RGII activity is confined to

Asia and overlaps with RGI; however, the dataset from areas in which RGII viruses appear to circulate is limited. Additionally, as shown in Figure 4, large regions of the world remain to be sampled to complete the rubella virus genotypic picture.

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References

 Chantler J, Wolinsky JS, Tingle A. Rubella virus. In: Knipe DM, Howley PM, editors. Fields virology. 4th ed. Philadelphia: Lippincott, Williams, and Wilkins; 2001. p. 963–90.



Figure 4. Distribution of rubella genotypes.

- Plotkin SA. Rubella. In: Plotkin SA, Orenstein WA, editors. Vaccines. 3rd ed. Philadelphia: Saunders;1999. p. 409–40.
- World Health Organization. Preventing congenital rubella syndrome. Wkly Epidemiol Rec 2000;75:289–96.
- Castillo-Solorzano C, Carrasco P, Tambini G, Reef S, Brana M, De Quadros CA. New horizons in the control of rubella and prevention of congenital rubella syndrome in the Americas. J Infect Dis 2003;187(Suppl 1):S146–52.
- Venczel L, Rota J, Dietz V, Morris-Glasgow V, Siqueira M, Quirogz E, et al. The measles laboratory network in the region of the Americas. J Infect Dis 2003;187(Suppl 1):S140–5.
- Frey TK. Molecular biology of rubella virus. Adv Virus Res 1994;44:69–160.
- Frey TK, Abernathy ES. Identification of strain-specific nucleotide sequences in the RA27/3 rubella virus vaccine. J Infect Dis 1993;168:854–64.
- Frey TK, Abernathy ES, Bosma TJ, Starkey WG, Corbett KM, Best JM, et al. Molecular analysis of rubella virus epidemiology across three continents, North America, Europe, and Asia, 1961–1977. J Infect Dis 1998;178:642–50.
- Zheng DP, Zhu H, Revello MG, Gerna G, Frey TK. Phylogenetic analysis of rubella virus isolated during a period of epidemic transmission in Italy, 1991–1997. J Infect Dis. 2003;187:1587–97.
- Reef SE, Frey TK, Theall K, Abernathy E, Burnett CL, Icenogle J, et al. The changing epidemiology of rubella in the 1990s, on the verge of elimination and new challenges for control and prevention. JAMA 2002;287:464–72.
- Katow S, Minahara H, Fukushima M, Yamaguchi Y. Molecular epidemiology of rubella by nucleotide sequences of the rubella virus E1 gene in three East Asia countries. J Infect Dis 1997;176:602–16.
- Bosma TJ, Best JM, Corbett KM, Banatvala JE, Starkey WG. Nucleotide sequence analysis of a major antigenic domain of the E1 glycoprotein of 22 rubella virus isolates. J Gen Virol 1996;77:2523–30.
- Pugachev KV, Abernathy ES, Frey TK. Genomic sequence of the RA27/3 vaccine strain of rubella virus. Arch Virol 1997;142:1165–80.

- Schmidt HA, Strimmer K, Vingron M, von Haeseler A. TREE-PUZ-ZLE: maximum likelihood phylogenetic analysis using quartets and parallel computing. Bioinformatics 2002;18:502–4.
- Hasegawa M, Kishino H, Yano K. Dating of the human-ape splitting by a molecular clock of mitochondrial DNA. J Mol Evol 1985;22:160–74.
- Page RDM. TreeView: an application to display phylogenetic trees on personal computers. Comp Appl Biosci 1996;12:357–8.
- Zheng DP, Zhang LB, Fang ZY, Yang CF, Mulders M, Pallansch MA, et al. Distribution of wild type 1 poliovirus genotypes in China. J Infect Dis 1993;168:1361–7.
- Jin L, Vyse A, Brown DW. The role of RT-PCR assay of oral fluid for diagnosis and surveillance of measles, mumps and rubella. Bull World Health Organ 2002;80:76–7.
- Vyse AJ, Lin L. An RT-PCR assay using oral fluid samples to detect rubella virus genome for epidemiological surveillance. Mol Cell Probes 2002;16:93–7.
- Bosma TJ, Corbett KM, Eckstein MB, O'Shea S, Vijayalakshmi P, Banatvala JE, et al. Use of PCR for prenatal and postnatal diagnosis of congenital rubella. J Clin Microbiol 1995;33:2881–7.
- Bosma TJ, Corbett KM, O'Shea S, Banatvala JE, Best JM. PCR for detection of rubella virus RNA in clinical samples. J Clin Microbiol 1995;33:1075–9.
- 22. Tanemura M, Suzumori K, Yagami Y, Katow S. Diagnosis of fetal rubella infection with reverse transcription and nested polymerase chain reaction: a study of 34 cases diagnosed in fetuses. Am J Obstet Gynecol 1996;174:578–82.
- Adams SD, Tzeng W-P, Chen M-H, Frey TK. Analysis of intermolecular RNA-RNA recombination by rubella virus. Virology 2003;309:258–71.
- Ando T, Noel JS, Fankhauser L. Genetic classification of "Norwalklike viruses." J Infect Dis 2000;181(Suppl 2):336–48.

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