LETTERS

Five (15.5%) of the 33 pregnant women had *P. jiroveci* DNA in their nasal swab samples versus none (0%) of the 28 nonpregnant controls (p=0.04 by 1-sided Fisher exact test). Immunologic parameters were not tested. The *P. jiroveci*-positive women were all multiparous with 1 (n=2), 2 (n=2), or 3 (n=1) previous pregnancies.

These results suggest that pregnancy is a host factor that favors asymptomatic nasal carriage of *P. jirovec*. However, PCR detection of *P. jiroveci* DNA in the nares of pregnant women does not necessarily indicate either a mild active pulmonary infection or viable or transmissible organisms. In animal models, detection of *P. carinii* DNA in nasal and oral samples is a good indicator that *Pneumocystis* is in the lungs (8).

These results also support the hypothesis that pregnant women who nasally carry P. jiroveci may play a role as contagious sources for susceptheir tible persons, especially immunologically naive newborn infants. This hypothesis warrants further study. Mother-to-infant transmission may explain the accumulating evidence that the primary infection is widely acquired very early in life (9). Recent animal model studies have documented the early acquisition of P. carinii (within 1 to 2 h after birth) in neonatal rats, likely transmitted by the dams (10). Evidence of mother-offspring transmission would be clinically relevant for infants born to HIVinfected mothers, who currently rely on empiric anti-Pneumocystis chemotherapy started at 1 month of age as their only prophylactic option.

Acknowledgments

We thank Henry N. Claman for critically reviewing the manuscript in aspects related to human pregnancy.

This study was supported by grant PG 51153-26 from the Elizabeth Glaser Pediatric AIDS Foundation; by Fondo Nacional de Desarrollo Científico y Tecnológico (FONDECYT research grant 1011059), Santiago, Chile; and by a 2002 research award from the Hospital Clínico de la Universidad de Chile.

Sergio L. Vargas,* Carolina Angelica Ponce,* Catherine Andrea Sanchez,* Ana Victoria Ulloa,* Rebeca Bustamante,* and Guido Juarez†

*University of Chile School of Medicine, Santiago, Chile; and †University of Chile Hospital, Santiago, Chile

References

- Wakefield AE. Pneumocystis carinii. Br Med Bull 2002;61:175–88.
- Contini C, Villa MP, Romani R, Merolla R, Delia S, Ronchetti R. Detection of Pneumocystis carinii among children with chronic respiratory disorders in the absence of HIV infection and immunodeficiency. J Med Microbiol 1998;47:329–33.
- Sing A, Geiger AM, Hogardt M, Heesemann J. Pneumocystis carinii carriage among cystic fibrosis patients, as detected by nested PCR. J Clin Microbiol 2001;39:2717–8.
- Oz HS, Hughes WT. Search for Pneumocystis carinii DNA in upper and lower respiratory tract of humans. Diagn Microbiol Infect Dis 2000;37:161–4.
- Wegmann TG, Lin H, Guilbert L, Mosmann TR. Biderectional cytokine interactions in the maternal-fetal relationship:is successful pregnancy a TH-2 phenomenon? Immunol Today 1993;14:353–6.
- Claman HN. The immunology of human pregnancy. Totowa (NJ): Humana Press; 1993.
- Ahmad H, Mehta N, Manikal VM, Lamoste TJ, Chapnick EK, Lutwick LI, et al. Pneumocystis carinii pneumonia in pregnancy. Chest 2001;120:666–71.
- Oz HS, Hughes WT. DNA amplification of nasopharyngeal aspirates in rats: a procedure to detect Pneumocystis carinii. Microb Pathog 1999;27:119–21.
- Miller RF, Ambrose HE, Novelli V, Wakefield AE. Probable mother to infant transmission of Pneumocystis carinii f. sp. hominis infection. J Clin Microbiol 2002;40:1555–7.
- Icenhour CR, Rebholz S, Collins MS, Cushion MT. Evidence for early acquisition of Pneumocystis carinii in neonatal rats using PCR and oral swabs. Eukaryotic Cell 2002;1:414–9.

Address for correspondence: Sergio L. Vargas, Respiratory Infections Laboratory, Program in Microbiology and Mycology, Biomedical Sciences Institute, University of Chile School of Medicine, Independencia 1027, Santiago, Chile; fax: +56-2-732 5160; email: svargas@ terra.cl

First Evidence of Aedes albopictus (Skuse) in Southern Chiapas, Mexico

To the Editor: The mosquito Aedes albopictus (Skuse, 1894) was first identified in the Americas in Texas in 1985 (1,2). That year, this newly introduced species had dispersed widely in Texas and was implicated in the transmission of dengue virus (3). Later, the first states in Mexico that were infested by Ae. albopictus were along the northern Mexican border: Coahuila, Nuevo Leon, and Tamaulipas (4,5; J.P. Martínez-Muñoz, thesis). In 1997, this species was reported farther south in Veracruz (6). Although Ae. albopictus was expected to spread to southernmost Mexico, this mosquito has never been reported there until now. We have confirmed Ae. albopictus in the city limits of Tapachula, southern Chiapas, Mexico.

On September 13, 2002, one of the authors, who resides in Tapachula, was bitten by a mosquito. He collected the specimen, which was later identified as Ae. albopictus by the Centro de Investigación de Paludismo (CIP). Nearby larval habitats were then comprehensively searched to collect the immature stages of the species; the sampling area was located at 14° 55' 22.5" north and 92°15' 05.7" west at an altitude of 220 m along the periphery of Tapachula. We found the following containers with larval stages of mosquitos: five water containers, two discarded tires (containing 300-3,000 mL of water), one thermal bottle (250 mL), one plastic bottle (50 mL), and one bucket (2,500 mL). Larvae were placed in plastic bags and transported to CIP laboratories, where they were allowed to emerge to adults during 17 days. The fourth instar larval and pupal exuvias were fixed and identified to species according to Darsie (7) and Superintendência de Campanhas de Saúde Pública (8). Twenty-five female and male Ae. albopictus from these collections are available from CIP laboratory upon request.

Additional field collections are being conducted to establish the distribution range of this species along the Chiapas coastal plain, to determinate the entomologic levels of infestation, and to determine its susceptibility to insecticides. Considering the epidemiologic relevance of this discovery, we have notified the proper health authorities to take necessary control measures to reduce the possibility of increased dengue transmission and to prevent other arboviruses, such as West Nile virus (9), from being spread by this new species in southern Mexico.

Mauricio Casas-Martínez* and José Luis Torres-Estrada*

*Centro de Investigación de Paludismo/ Instituto Nacional de Salud Pública, Chiapas, México

References

- Centers for Disease Control. Aedes albopictus introduction—Texas. MMWR Morb Mortal Wkly Rep 1986;35:141–2.
- Centers for Disease Control. Aedes albopictus infestation—United States, Brazil. MMWR Morb Mortal Wkly Rep 1986;35:493–5.
- Moore CG, Francy DB, Eliason DA, Monath TP. *Aedes albopictus* in the United States: rapid spread of a potential disease vector. J Am Mosq Control Assoc 1988;4:356–61.
- Ibáñez-Bernal S, Martínez-Campos C. *Aedes albopictus* in México. J Am Mosq Control Assoc 1994;10:231–2.

- Rodríguez-Tovar ML, Ortega-Martínez MG. Aedes albopictus in Muzquiz City, Coahuila, México. J Am Mosq Control Assoc 1994;10:587.
- Secretaría de Salud. 2000. Available from: URL: http://www.ssaver.gob.mx/Servicios _de_Salud/BoletinEpidem/ Boletines/2000-6/ page17.html
- Darsie RF Jr. The identification of *Aedes* albopictus in the neartic region. J Am Mosq Control Assoc 1986;2:336–40.
- Superintendência de Campanhas de Saúde Pública. Resumo dos principias caracteres morfológicos diferenciais do *Aedes aegypti e* do *Aedes albopictus*. Brasilia: SUCAM/ Min. da Saúde; 1989.
- Holick J, Kyle A, Ferraro W, Delaney RR, Iwaseczk M. Discovery of *Aedes albopictus* infected with West Nile virus in southeastern Pennsylvania. J Am Mosq Control Assoc 2002;18:131.

Address for correspondence: Mauricio Casas-Martínez, Centro de Investigación de Paludismo, Instituto Nacional de Salud Pública, Apartado Postal 537, Tapachula, Chiapas, C. P. 30700, México; fax: (962) 626 57 82; email: mcasas@insp.mx

Virus Isolation and "Acute" West Nile Virus Encephalitis (Response to Huang et al.)

To the Editor: We read with interest a recent article in your journal, First Isolation of West Nile virus from a Patient with Encephalitis in the United States (1); in the report, we were unable to ascertain indisputable evidence that this patient had indeed acquired acute West Nile virus (WNV) encephalitis. In animals (2,3) and humans (4), West Nile virus can persist in the host even after the host has recovered from an acute WNV infection, presumably more so in the immunocompromised persons. Therefore, in the case described by Huang et al. (1), proving that the patient did not have a history of WNV infection is important, particularly because this patient is from a geographic area where WNV is known to exist. The findings at autopsy of perivascular lymphocyte cuffing in mammillary bodies of the brain are not the classic findings reported during the West Nile encephalitis outbreak in New York City (5). The immunoglobulin (Ig) G antibody against WNV, if it had been present, would have been useful in that IgG antibody in the absence of IgM antibody is indicative of past rather than acute infection.

The WNV copy numbers in clinical samples and clinical indices (leukocyte count) suggest that the virus multiplies in the setting of leukopoenia or immune suppression and cannot be definitive proof that it was an acute infection, unless a negative preillness sample was available. The cause of the transient viremia, whether acutely acquired or from increased proliferation in a chronic infection, needs to be clarified further. In the future, antigen detection will guide patient management decisions; therefore, the possibility of a human chronic carrier state warrants study.

Vijay K. Krishnamoorthy,* Jayashri Bhaskar,* and John N. Sheagren*

*Advocate Illinois Masonic Medical Center, Chicago, Illinois, USA

References

- Huang C, Slater B, Rudd R, Parchuri N, Hull R, Dupuis M, et al. First isolation of West Nile virus from a patient with encephalitis in the United States. Emerg Infect Dis 2002;8:1367–71.
- Pogodina VV, Frolova MP, Malenko GV, Fokina GI, Koreshkova GV, Kiseleva LL, et al. Study on West Nile virus persistence in monkeys. Arch Virol 1983;75:71–86.
- Camenga DL, Nathanson N, Cole GA. Cyclophosphamide-potentiated West Nile viral encephalitis: relative influence of cellular and humoral factors. J Infect Dis 1974;130:634–41.