

## **HHS Public Access**

Ticks Tick Borne Dis. Author manuscript; available in PMC 2023 November 02.

Published in final edited form as:

Author manuscript

Ticks Tick Borne Dis. 2022 March ; 13(2): 101886. doi:10.1016/j.ttbdis.2021.101886.

### Inter-annual variation in prevalence of *Borrelia burgdorferi* sensu stricto and *Anaplasma phagocytophilum* in host-seeking *Ixodes scapularis* (Acari: Ixodidae) at long-term surveillance sites in the upper midwestern United States: Implications for public health practice

Erik Foster<sup>a,\*</sup>, James Burtis<sup>a</sup>, Jennifer L. Sidge<sup>b,c</sup>, Jean I. Tsao<sup>d</sup>, Jenna Bjork<sup>e</sup>, Gongping Liu<sup>e</sup>, David F. Neitzel<sup>e</sup>, Xia Lee<sup>f</sup>, Susan Paskewitz<sup>f</sup>, Diane Caporale<sup>g</sup>, Rebecca J. Eisen<sup>a</sup> <sup>a</sup>Division of Vector-Borne Diseases, National Center for Emerging and Zoonotic Infectious Diseases, Centers for Disease Control and Prevention, Fort Collins, CO 80521, USA

<sup>b</sup>Comparative Medicine and Integrative Biology, Michigan State University, East Lansing, MI 48824, USA

<sup>c</sup>Michigan Department of Agriculture and Rural Development, Lansing, MI 48933, USA

<sup>d</sup>Department of Fisheries and Wildlife, Michigan State University, East Lansing, MI 48824, USA

eVectorborne Diseases Unit, Minnesota Department of Health, St. Paul, MN 55164, USA

<sup>f</sup>Department of Entomology, University of Wisconsin-Madison, Madison, WI 53706, USA

<sup>g</sup>Department of Biology, University of Wisconsin-Stevens Point, Stevens Point, WI 54481, USA

#### Abstract

The geographic range of the blacklegged tick, *Ixodes scapularis*, and its associated human pathogens have expanded substantially over the past 20 years putting an increasing number of persons at risk for tick-borne diseases, particularly in the upper midwestern and northeastern

Disclaimers

<sup>&</sup>lt;sup>\*</sup>Corresponding author at: Division of Vector-Borne Diseases, National Center for Emerging and Zoonotic Infectious Diseases, Centers for Disease Control and Prevention, 3156 Rampart Road, Fort Collins, CO 80521, USA. owm1@cdc.gov (E. Foster). Declaration of competing interests

None

The findings and conclusions of this study are by the authors and do not necessarily represent the views of the Centers for Disease Control and Prevention, Michigan State University, Minnesota Department of Health, University of Wisconsin-Madison, or University of Wisconsin-Stevens Point.

CRediT authorship contribution statement

Erik Foster: Conceptualization, Data curation, Methodology, Visualization, Formal analysis, Writing – original draft, Writing – review & editing. James Burtis: Conceptualization, Data curation, Methodology, Visualization, Formal analysis, Writing – original draft, Writing – review & editing. Jennifer L. Sidge: Data curation, Methodology, Writing – review & editing. Jean I. Tsao: Data curation, Methodology, Writing – review & editing. Jennifer L. Sidge: Data curation, Methodology, Writing – review & editing. Jean I. Tsao: Data curation, Methodology, Writing – review & editing. Jean I. Tsao: Data curation, Methodology, Writing – review & editing. Jean Bjork: Data curation, Methodology, Writing – review & editing. Conceptualization, Methodology, Writing – review & editing. Susan Paskewitz: Data curation, Methodology, Writing – review & editing. Diane Caporale: Data curation, Methodology, Writing – review & editing. Diane Caporale: Data curation, Formal analysis, Writing – original draft, Writing – review & editing. Methodology, Visualization, Data curation, Methodology, Visualization, Formal analysis, Writing – original draft, Writing – review & editing.

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.ttbdis.2021.101886.

Page 2

United States. Prevention and diagnosis of tick-borne diseases rely on an accurate understanding by the public and health care providers of when and where persons may be exposed to infected ticks. While tracking changes in the distribution of ticks and tick-borne pathogens provides fundamental information on risk for tick-borne diseases, metrics that incorporate prevalence of infection in ticks better characterize acarological risk. However, assessments of infection prevalence are more labor intensive and costly than simple measurements of tick or pathogen presence. Our objective was to examine whether data derived from repeated sampling at longitudinal sites substantially influences public health recommendations for Lyme disease and anaplasmosis prevention, or if more constrained sampling is sufficient. Here, we summarize inter-annual variability in prevalence of the agents of Lyme disease (Borrelia burgdorferi s.s.) and anaplasmosis (Anaplasma phagocytophilum) in host-seeking I. scapularis nymphs and adults at 28 longitudinal sampling sites in the Upper Midwestern US (Michigan, Minnesota, and Wisconsin). Infection prevalence was highly variable among sites and among years within sites. We conclude that monitoring infection prevalence in ticks aids in describing coarse acarological risk trends, but setting a fixed prevalence threshold for prevention or diagnostic decisions is not feasible given the observed variability and lack of temporal trends. Reducing repeated sampling of the same sites had minimal impact on regional (Upper Midwest) estimates of average infection prevalence; this information should be useful in allocating scarce public health resources for tick and tick-borne pathogen surveillance, prevention, and control activities.

#### Keywords

Tick surveillance; Tick-borne disease; *Ixodes scapularis*; *Borrelia burgdorferi*; *Anaplasma phagocytophilum* 

#### Introduction

Tick-borne diseases are an increasing public health burden in the United States. Of the nearly 650,000 cases of vector-borne diseases reported in the United States from 2004 to 2016, more than 75% were tick-borne and a majority of those were associated with the blacklegged tick, Ixodes scapularis (Rosenberg et al., 2018). In addition to transmitting Borrelia burgdorferi sensu stricto, the primary causative agent of Lyme disease, which is the most commonly reported vector-borne disease in the United States, the tick also transmits a less common agent of Lyme disease (Borrelia mayonii) and agents of anaplasmosis (Anaplasma phagocytophilum), babesiosis (Babesia microti), hard tick relapsing fever (Borrelia miyamotoi), ehrlichiosis (Ehrlichia muris eauclairensis), and a viral neuroinvasive disease (Powassan virus) (Eisen and Eisen 2018). Although the reported geographic range of each pathogen varies across the tick's range, all have been identified in host-seeking *I*. scapularis in the upper midwestern United States (Johnson et al., 2018). In the past two decades, the geographic range of *I. scapularis* and its associated human pathogens have expanded dramatically, resulting in an increase in reported tick-borne disease cases, most notably in the Upper Midwest, Northeast, and Mid-Atlantic regions (Kugeler et al., 2015; Eisen et al., 2016, 2017; Eisen and Paddock, 2021).

Prevention and diagnosis of tick-borne diseases rely on an accurate understanding by the public and health care providers of when and where persons are at risk for exposure to human-biting ticks and their associated pathogens. However, national maps showing the distribution and abundance of medically important ticks and their associated pathogens are often incomplete, not current, or lack data entirely (Eisen and Paddock 2021). Efforts to generate data to inform such maps have been hampered by a lack of standardized routine tick-based surveillance. A recent survey of vector-borne disease professionals in the U.S. revealed that fewer than half of respondents were engaged in routine active tick surveillance. Most of those engaged in tick surveillance were focused on describing the distribution of ticks, with fewer aiming to describe pathogen presence or prevalence within the targeted tick populations. Cited barriers to conducting tick surveillance and pathogen testing included a lack of guidance and funding constraints (Mader et al., 2021).

In 2018, the U.S. Centers for Disease Control and Prevention (CDC) issued guidance aimed at standardizing tick and tick-borne pathogen surveillance and increased support to public health partners to conduct tick surveillance (CDC, 2018; Eisen and Paddock 2021). The recommendations describe a set of objectives that progressively increase the amount of data available to support assessments of human risk of exposure to ticks and tick-borne pathogens. Objectives range from describing the distribution of medically important ticks to identifying the presence of human pathogens in ticks, and progress to quantifying tick densities and the prevalence of pathogens in host-seeking ticks. While the utility of the data increases with each escalating objective, the resources required to conduct tick surveillance also intensify with those requiring pathogen detection being among the most costly and time-consuming.

Tick and tick-borne pathogen surveillance data are commonly used to explain epidemiological trends (primarily at coarse spatial scales), to guide tick bite prevention recommendations and to establish a prior probability of exposure when diagnosing a tickborne disease (Pepin et al., 2012; Stromdahl and Hickling 2012; Dahlgren et al., 2016; Moore et al., 2016; Bisanzio et al., 2020; Kugeler and Eisen 2020; O'Connor et al., 2021; Eisen and Paddock 2021; Lantos et al., 2021). Recognizing resources are limited for conducting tick and tick-borne pathogen surveillance, we sought to assess the feasibility of scaling back tick testing without seriously compromising data used in public health practice. Here, we describe spatial and temporal variation in the prevalence of the two most common pathogens (*B. burgdorferi* s.s. and *A. phagocytophilum*) in host-seeking *I. scapularis* nymphs and adults in the Upper Midwest (for the purposes of this study, the Upper Midwest is defined as a region including Michigan, Minnesota, and Wisconsin). Additionally, we sought to determine if a less intensive approach yielded comparable regional (Michigan, Minnesota, and Wisconsin) estimates of infection prevalence in host-seeking ticks compared with multiple-year sampling of the same sites.

Specifically, in this study we analyzed historic *I. scapularis* nymphal and adult surveillance records among sites in the Upper Midwest with multiple years of collections and pathogen testing. We summarized inter-annual variability in infection prevalence of each pathogen in host-seeking *I. scapularis* nymphs and adults at sites that were sampled at least three years. We also assessed whether pathogen prevalence in one year is predictive of future

years within the same site and whether pathogen prevalence changes significantly over time. We further estimated regional and state averages and ranges in infection prevalence of each pathogen by tick life stage and created random subsets of the data to assess the impacts of a reduced sampling regime for estimating regional averages in infection prevalence.

#### **Methods**

#### **Collection sites**

Retrospective tick collection and pathogen testing records from three states in the Upper Midwest were provided by state public health agencies or their academic partner institutions. These data were used originally for public health tick surveillance or research, and in many instances have been published in part or fully (Hamer et al., 2010, 2012, 2014; Pritt et al., 2016; Bjork and Schiffman 2020), but not previously as a combined data set. From 2000 through 2019 host-seeking I. scapularis nymphs and adults were collected by dragging at 34 forested sites, including edge habitat, in areas considered by the collectors to be of public health concern. Drag sampling is recommended in areas where I. scapularis is endemic or emerging, as the method reliably quantifies tick density and yields a highly correlated measure of the human risk of contact with infected host-seeking ticks (Falco and Fish 1992; Mather et al., 1996). Sites included novel areas of potential human exposure to *I. scapularis*; areas where *I. scapularis* is newly established; areas where incidence of I. scapularis-borne illnesses have changed over time; heavily used recreational areas; areas where novel pathogens are suspected to be circulating; and representative habitat types in areas where I. scapularis-borne infections are prevalent. Sites were sampled one or more times per year during peak nymphal and/or adult activity periods. When sampling was conducted more than once per year, the highest observed density per life stage was considered the peak value.

Data elements included site location, year of collection, peak number of nymphal and adult I. scapularis collected per area sampled, number of nymphal and adult I. scapularis tested for B. burgdorferi s.s. and A. phagocytophilum, and number of nymphal and adult I. scapularis positive for B. burgdorferi s.s. and A. phagocytophilum by site and year. For inclusion of records in this study, site selection, tick collection and pathogen identification methods had to conform to *I. scapularis* surveillance guidance published by the CDC (CDC, 2018). Data were screened to exclude sites with less than three years of repeated sampling within a sequential five-year period. One additional site in which sampling was conducted for three consecutive years was excluded because sample sizes were extremely low (*n*= one, two and five ticks tested per year), yielding consistently unreliable estimates of infection prevalence. After screening, 28 sampling sites met the criteria for inclusion in the study for one or more pathogen and life stage combinations. The geographic range of sites meeting all data inclusion criteria is shown in Fig. 1. Within included sites, years where only one tick or no ticks were tested were excluded from analyses. The inclusion of years where low numbers of ticks were collected was done to ensure that sites with emerging tick or pathogen populations were not excluded from our data set.

#### Pathogen detection

Pathogen detection methods varied by state and entity performing the testing but met minimum criteria for acceptability according to CDC *I. scapularis* surveillance guidance (CDC 2018). Briefly, collected nymphal and adult ticks were tested individually using molecular assays specific to *B. burgdorferi* s.s. or *A. phagocytophilum*. Assays were demonstrated to be species-specific by testing against genetically similar species or designed according to previously published assays meeting the same criteria. While all assays specifically targeted *B. burgdorferi* sensu stricto, *A. phagocytophilum* assays did not discriminate human-active (ha) variant or variant 1 (v1). Specific pathogen detection methods used are listed in Supplemental Table 1.

#### Statistical analysis

To generate descriptive statistics, pathogen infection prevalence was calculated for *B. burgdorferi* s.s. and *A. phagocytophilum* by *I. scapularis* life stage, sampling site, and year. Site specific 95% confidence intervals (95% C.I.) were calculated as Wilson score intervals, which are applied to binomial data including small sample sizes, or point estimates close to one or zero (Wilson 1927). State and regional (all states combined) averages were based on these site-specific point estimates of infection prevalence and 95% confidence intervals were derived assuming a t-distribution to account for small sample sizes (< 30 sites).

The resulting annual site-level point estimates were used in mixed effects models to determine if infection prevalence increased or decreased over time. Only sites with at least five years of continuous pathogen testing data were included. First, qualifying sites were classified as 'emerging,' or 'established' where an 'emerging' site was defined as any site where the prevalence point estimates for the first three years of sampling were below the lower 95% CI for the Upper Midwest region. Data were analyzed separately for each of the four pathogen and life stage combinations, and each of these groups were split into 'emerging' and 'established' analyses for a total of eight models. Each model included 'year' as a fixed effect and 'site' as a random effect, if more than one site was included in the analysis. Recognizing that pathogen detection methods varied among sites and over time, we included pathogen testing method as a second random effect. However, it did not significantly improve Akaike information criterion (AIC) scores, indicating testing method did not explain observed differences, and the variable was not included in the final models.

In addition to general linear trends that evaluated consistent increases or decreases in infection prevalence over time, we applied an autocorrelation function (ACF) to determine if the annual prevalence of pathogens was generally temporally autocorrelated within each site which would indicate that infection prevalence in one year is predictive of observed prevalence the following year.

We aimed to determine if limiting observations on each site to a single year significantly affected estimates of regional infection prevalence compared against estimates that were generated using the full dataset. We subsampled the full data set ten times. In these subsamples, each site was limited to a single year that was selected randomly with replacement from those available. For nymphs, the full dataset contained 25 sites that

were sampled over multiple years (156 total yearly prevalence point estimates). Each nymph subset contained all 25 sites which included approximately 16% of the observations present in the full dataset. These subsamples were compared against estimates generated using all 156 prevalence point estimates (i.e. the full dataset). For adults, the full dataset contained 14 sites that were sampled over multiple years (117 total yearly prevalence point estimates). Each adult subset contained all 14 sites which included approximately 12% of the observations present in the full dataset. These subsamples were compared against estimates generated using all 117 prevalence point estimates. Differences between the regional point estimates of the subsamples and full dataset were analyzed using analysis of variance (ANOVA) and pairwise comparisons were made using post-hoc Tukey tests. These analyses were only conducted with data for *B. burgdorferi* infected nymphs and adults as fewer sites were sampled for *A. phagocytophilum* infected ticks.

All data analyses were conducted in R version 4.0.3 (R Foundation for Statistical Computing, Vienna, Austria) or JMP v. 13.2.1 (SAS Institute, Cary, N.C.). Mixed effects models were constructed using the lme4 package.

#### Results

#### Prevalence of B. burgdorferi s.s. in I. scapularis nymphs

From 2004 to 2019 a total of 12,594 host-seeking *I. scapularis* nymphs were collected from 25 sites across three states in the Upper Midwest. Sampling was conducted from three to 12 years (median: six years) per site with two to 817 nymphs tested per site per year (median: 69.5 nymphs tested per site per year). The mean site-specific prevalence of *B. burgdorferi* s.s. was as low as 1.40% (95% CI: 0.60–3.23%) at the Fenner Nature Center in Michigan and as high 28.18% (95% CI: 23.57–32.80%) at Tower Hill State Park in Wisconsin. State specific mean prevalence of *B. burgdorferi* s.s. ranged from 13.63% (95% CI: 5.72–21.54%) in Michigan to 18.54% (95% CI: 14.32–22.76%) in Wisconsin. The regional mean prevalence of *B. burgdorferi* s.s. in host-seeking *I. scapularis* nymphs across all sampling sites and years was 16.97% (95% CI: 13.96–19.98%) (Table 1, Supplemental Table 2). Overall, 80% (20 of 25) and 60% (15 of 25) of site estimates were statistically similar to state-specific and regional averages, respectively (Table 1).

Among 10 established sites for which we had at least five years of contiguous data, the mixed effects model for *B. burgdorferi* s.s. infected nymphs showed no statistically significant temporal trend (t = -1.7, df = 84, p = 0.10) in infection prevalence indicating that infection prevalence was not consistently increasing or decreasing over time. Only Fenner Nature Center in MI met our criteria for an emerging site and we detected no statistically significant temporal trend (t = 0.34, df = 3, p = 0.76), although data were limited for this analysis with only five years of observations (Fig. 2). The ACF plots revealed no temporal autocorrelation between sampling years for any site, meaning that prevalence in one year was not predictive of prevalence in the next (Supplemental Figure 1).

There was no statistically significant difference when comparing the regional point estimates of *B. burgdorferi* s.s infection prevalence in nymphs generated using the full data set (16.97% [95% CI: 14.12–19.83%]) to the subsets where each site was limited to a single

year of data (d.f. = 10,264; F = 1.26; p = 0.253). In pairwise comparisons, none of the regional point estimates from the ten subsamples differed significantly (p >0.05) from the regional point estimate derived from the full data set (Supplemental Figure 5).

#### Prevalence of B. burgdorferi s.s. in I. scapularis adults

From 2000 to 2019, a total of 8262 host-seeking *I. scapularis* adults were collected from 14 sites in three states. In Minnesota, all sites sampled for nymphs were also sampled for adults, but in Michigan two additional sites with limited nymph data were included, and in Wisconsin 14 sites were sampled for nymphs and an independent site (Stevens Point) was sampled only for adults. Sampling years for adults ranged from three to 20 years (median 12 years) with two to 232 adults tested for *B. burgdorferi* s.s. per site per year (median: 92.5 adults tested per site per year). Across all sampling sites and years, the regional mean prevalence of *B. burgdorferi* s.s. in host-seeking *I. scapularis* adults was 29.53% (95% CI: 22.08–36.98%). Ionia Recreation Area in Michigan yielded the lowest infection prevalence in adult ticks (3.57% [95% CI: 0.18–17.71%]), while Richard J. Dorer Memorial Hardwood State Forest in Minnesota yielded the highest prevalence (45.07% [95% CI: 42.30–47.88%]) (Table 2, Supplemental Table 3). In total 85% (11 of 13 sites) and 64% (9 of 14 sites) of site-specific estimates were statistically similar to state and regional estimates, respectively (Table 2).

Among the five established sites with five years of contiguous sampling there was no statistically significant temporal trend (t= 0.66, df = 55, p = 0.51) in *B. burgdorferi* s.s. infection prevalence, indicating infection prevalence was stable over time. However, the model that included the three emerging sites showed a statistically significant positive temporal trend (t= 3.1, df = 30, p = 0.004) or consistent increase in infection prevalence over time (Fig. 3). The autocorrelation function plots revealed no statistically significant temporal autocorrelation between sampling years for any sites regardless of its status as an emerging or established site (Supplemental Figure 2).

There was no statistically significant difference when comparing the regional point estimates of *B. burgdorferi* s.s infection prevalence in adults generated using the full data set (29.53% [95% CI: 22.77–36.29%]) to the subsets where each site was limited to a single year of data (df = 10,143; F = 0.383, p = 0.952). In pairwise comparisons, none of the regional point estimates from the ten subsamples differed significantly (p > 0.05) from the regional point estimate derived from the full data set (Supplemental Figure 5).

#### Prevalence of A. phagocytophilum in I. scapularis nymphs

From 2005 to 2019, 7562 nymphs collected from 10 sites in Minnesota and Wisconsin were tested for *A. phagocytophilum*. Among sites included in estimates of *A. phagocytophilum* prevalence, the number of years included per site ranged from four to 12 (median: eight years). From each site and year, the number of nymphs tested ranged from six to 738 (median: 84.25 nymphs tested). The regional mean prevalence of *A. phagocytophilum* in host-seeking *I. scapularis* nymphs across all sampling sites and years was 6.57% (95% CI: 4.47–8.66%) and was as low as 2.67% (95% CI: 1.23–5.69%) at McCaslin Brook in Wisconsin, and as high as 9.98% (95% CI: 8.18–12.12%) at Camp Ripley in Minnesota

(Table 3, Supplemental Table 4). Site-specific estimates were statistically similar to state specific averages for 90% of sites (9 of 10) and 80% (8 of 10 sites) were statistically similar to the regional average (Table 3).

Among the five established sites for which five years of contiguous data were available, results of the mixed effect model for *A. phagocytophilum* infected nymphs showed no statistically significant temporal trend (t = -0.05, df = 44, p = 0.96) in infection prevalence, indicating that infection prevalence was not increasing or decreasing consistently over time. Only American Legion Northern Highland in Wisconsin met our criteria for an emerging site and we detected no statistically significant temporal trend (t = 1.03, df = 3, p = 0.38), although data were limited for this analysis with only five years of observations (Fig. 4). Autocorrelation function plots revealed no significant temporal autocorrelation between sampling years by site (Supplemental Figure 3).

#### Prevalence of A. phagocytophilum in I. scapularis adults

From 2005 to 2019, 6381 adult ticks were collected from five sites in Minnesota and Wisconsin. The number of years sampled per site ranged from 10 to 20 (median 16 years per site) and the number of adults tested per site per year ranged from eight to 232 (median 93.7 adults tested per year). The regional mean prevalence of *A. phagocytophilum* in host-seeking *I. scapularis* adults across all sampling sites and years was 8.59% (95% CI: 5.01–12.17) (Table 4, Supplemental Table 5). All site-specific prevalence estimates were statistically similar to the state and regional averages (Table 4).

Results of the mixed effect model for four established sites showed a marginally statistically significant positive temporal trend (t = 1.9, df = 42, p = 0.06) in infection prevalence. Only Stevens Point in Wisconsin was classified as 'emerging' and data were analyzed in a linear model which detected a statistically significant positive trend (t = 3.1, df = 18, p = 0.007) in infection prevalence (Fig. 5). Autocorrelation function plots revealed no significant temporal autocorrelation between sampling years by site (Supplemental Figure 4).

#### Discussion

Consistent with previous studies from other endemic regions, the prevalence of *B. burgdorferi* s.s. and *A. phagocytophilum* were both highly variable in ticks among sites and among years within individual sites in the upper Midwest (Piesman et al., 1999; Eisen et al., 2004; Diuk-Wasser et al., 2012; Keesing et al., 2014; Prusinski et al., 2014; Feldman et al., 2015; Johnson et al., 2018). At sites considered "established," prevalence of *B. burgdorferi* s.s. exhibited high interannual variability, but there were no discernable increasing or decreasing trends over time. Prevalence of *A. phagocytophilum* in host-seeking nymphs remained stable over time at 'established' sites, but a slight marginally significant increase in infection prevalence was noted across sites where host-seeking adults were tested. Similarly, no temporal trends for either *B. burgdorferi* or *A. phagocytophilum* infection prevalence were detected at the 'emerging' nymphal sites, although only a limited number of observations were analyzed. However, a significant positive temporal trend in infection prevalence was detected in adults for both pathogens in sites classified as 'emerging'. At

all sites regardless of pathogen or tick life stage, infection prevalence in one year was not predictive of the next, according to ACF analysis.

In addition to sharing a common vector, *B. burgdorferi* and *A. phagocytophilum* share a common primary reservoir host, the white-footed mouse (*Peromyscus leucopus*). Compared with *B. burgdorferi* the infectious period for *A. phagocytophilum* in white-footed mice is transient (Telford et al., 1996; Stafford et al., 1999; Levin and Ross, 2004). This contributes to explaining why prevalence of *A. phagocytophilum* is generally lower than *B. burgdorferi* in host-seeking nymphs and adults. Neither pathogen is transmitted transovarially (Piesman, 1989; Teglas and Foley, 2006). Thus, acquisition is limited to single blood feeding events per life stage, with adults having two opportunities to acquire infection and nymphs only one. As a result, prevalence of infection is typically higher in adults. With higher prevalence of infection in adults, we were more likely to detect significant trends in adults than nymphs. However, in most cases due to differences in contiguous yearly sampling data, both life stages were not assessed for temporal trends at the same sites. Therefore, it is not clear if observed positive temporal trends observed in adults reflects the higher prevalence of infection, or differences in sites included in the nymphal compared with adult tick mixed-effect models.

The high degree of spatial and temporal variability in pathogen prevalence in ticks suggests that identifying and adhering to a fixed and precise prevalence threshold for prevention or diagnostic decisions is not feasible. However, coarse level estimates of pathogen prevalence (e. g., state or regional estimates) provide sufficient data for most public health purposes. We showed that sampling the included sites for as little as a single year yielded similar regional estimates of infection prevalence to multi-year sampling of the same sites. This implies that Upper Midwest regional estimates based on reduced sampling effort (i.e., as little as a single year of sampling per site) are comparable with more extensive longitudinal sampling of sites. Resampling sites with low infection prevalence may provide useful information regarding an emerging site but is unlikely to strongly impact regional estimates or public health messaging at larger scales. This suggests that tick sampling and testing efforts can be scaled to optimize scarce public health resources.

In addition to providing valuable data explaining ecological drivers of variation in acarological risk indices (e.g., host-seeking tick densities, infection prevalence, densities of infected host-seeking ticks) (Schulze and Jordan 1996; Jones and Kitron 2000; Ostfeld et al., 2001, 2006; Ginsberg et al., 2004; Elias et al., 2011; Ogden et al., 2018; Larson et al., 2021), longitudinal sampling of ticks and tick-borne pathogens from fixed sites provides insights into the complexity of characterizing acarological risk. Our long-term sampling data show that at any given location, the peak abundances of nymphs or adults is highly variable, as is the prevalence of infection in host-seeking ticks. Specifically, within a single site, we observed up to a 160-fold difference among years in the density of host-seeking nymphs and up to a 6.9-fold difference of *B. burgdorferi* s.s. in host-seeking nymphs varied as much as 6.9-fold among years within a single site.

Factors that influence variation in estimates of host-seeking tick density derived from drag or flag sampling at a single site include (1) seasonal and diel timing of tick collections (Schulze and Jordan 1996, 2003; Diuk-Wasser et al., 2006; Thomas et al., 2020), (2) number of sampling occasions that are used to estimate the seasonal peak (Dobson et al., 2014), (3) host composition (Daniels et al., 1993; VanBuskirk and Ostfeld 1995; Ostfeld et al., 2001, 2006; Ginsberg et al., 2020), and (4) weather conditions at the time of sampling and preceding sampling (Eisen, Eisen, Ogden and Beard, 2016). Infection prevalence estimates should be less sensitive to error introduced by timing or frequency of tick sampling compared with tick density estimates because the cohort of nymphs or adults being examined was infected over a long duration (months) when the previous life stage (larvae or nymphs, respectively) was active. Therefore, the absolute proportion of nymphs or adults infected with *B. burgdorferi* or *A. phagocytophilum* is expected to be constant during the sampling season; interannual variability in infection prevalence is explained mainly by host composition when the prior life stage was active (Ostfeld et al., 2001; Vuong et al., 2017). While the product of host-seeking tick density and infection prevalence is believed to be a more accurate correlate of human risk of exposure to infected ticks than either measure alone (Mather et al., 1996; Pepin et al., 2012), in this study, we focused primarily on assessing variability in infection prevalence because this is the costliest measure to assess. Our intent was to evaluate if less intensive testing to support tick surveillance activities could yield useful data for public health action.

Tick surveillance data are typically used to 1) explain epidemiological trends (Pepin et al., 2012; Stromdahl and Hickling 2012; Dahlgren et al., 2016; Bisanzio et al., 2020; Kugeler and Eisen 2020; O'Connor et al., 2021), 2) inform public health messaging for tick-bite prevention by identifying areas posing a risk for exposure to infected hostseeking ticks (Eisen and Paddock 2021), and 3) assess a likelihood of human exposure to pathogens following a tick bite (Lantos et al., 2021). Several studies have demonstrated a positive association between the density of *B. burgdorferi*-infected host-seeking nymphs and occurrence of Lyme disease (Mather et al., 1996; Stafford et al., 1998; Connally et al., 2006; Pepin et al., 2012). Although some of these analyses have focused on county or sub-county spatial scales, owing in part to the high degree of variability in both acarological and epidemiological data, these reported trends are generally more consistent when comparing between rather than within regions. Variation in pathogen prevalence between regions influences the epidemiology of tick-borne diseases. This is evident in the contrasting risk of acquiring Lyme disease in the southeastern U.S. versus other regions where I. scapularis is currently established. Despite presence of *I. scapularis* in southern states, the prevalence of B. burgdorferi s.s. in host-seeking ticks is significantly lower than the Northeast, Mid-Atlantic, and Upper Midwest where Lyme disease incidence is significantly higher than in southeastern states (Diuk-Wasser et al., 2012; Schwartz et al., 2017; Lehane et al., 2021). Therefore, determining prevalence of tick-borne pathogens provides greater insights into regional risk of acquiring tick-borne disease than tick presence or density alone.

Prevention of tick-borne diseases, including Lyme disease and anaplasmosis, relies primarily on education promoting the use of personal protection measures. In general, persons who perceive their risk of encounters with infected ticks or of acquiring a tick-borne disease to be higher are more likely to take precautions against tick bites or pathogen exposure

(e.g., wearing repellents, checking for and removing ticks) than those with lower perceived risks (Herrington et al., 1997; Niesobecki et al., 2019). Tick surveillance data aid in raising awareness of locations where risk of exposure to infected ticks is elevated. However, public health education or personal protection strategies are not likely to differ based on data suggesting a moderately low (e.g., Fenner Nature Center in Michigan) compared with a moderately high prevalence of infection in ticks (e.g., Tower Hill State Park in Wisconsin). Therefore, coarse (state or regional scale) data-driven estimates of infection prevalence in host-seeking ticks by life stage are generally adequate for public health messaging. While some have advocated for prevention strategies (use of antibiotic prophylaxis to prevent Lyme disease) based on a high likelihood of exposure to B. burgdorferi-infected I. scapularis where highly endemic areas are generally defined as > 20% *B. burgdorferi* prevalence in host-seeking *I. scapularis* nymphs (Wormser et al., 2006; Lantos et al., 2021), our data indicate oscillation above or below that 20% prevalence threshold across years. Such variation within single sites was observed both in states considered high incidence for Lyme disease (Wisconsin and Minnesota) or not (Michigan). The high degree of spatio-temporal variation in our data set demonstrates the difficulty of gaging such precise estimates across localities for public health action.

Nonetheless, we show that site-specific estimates of *B. burgdorferi* infection prevalence in host-seeking nymphs or adults were statistically similar to state averages for 80% of sites, and statistically similar to regional averages for 60% of sites. Although fewer sites were included, site specific estimates of A. phagocytophilum prevalence in host-seeking nymphs or adults was statistically similar to state or regional averages for 90% or 80% of sites, respectively. Where site estimates of *B. burgdorferi* s.s. prevalence differed significantly from state or regional averages, in most instances site estimates were lower than state or regional averages. Some of the lower-than-average estimates may have arisen because site specific estimates included a period of introduction or emergence of *B. burgdoferi* s.s. Significant increases in *B. burgdorferi* s.s. prevalence over time were observed more commonly in longitudinal sampling sites classified as emerging compared with those classified as established, suggesting that if lower than expected prevalence is observed, resampling is indicated. However, in some cases at established sites, specifically Saugatuck Dunes State Park in Michigan where *I. scapularis* has been present since 2004, prevalence of *B. burgdorferi* s.s. remained stable at low prevalence. This could be explained by host composition (a factor not examined in this study) contributing to a stable low prevalence of infection, or perhaps other site-level factors slowing the establishment of the I. scapularis population at this site.

The data presented here demonstrate the high degree of variability in estimates of infection prevalence at fine spatial and temporal scales. However, they also demonstrate that, in general, after *B. burgdorferi* s.s. or *A. phagocytophilum* become established in an area, their prevalence of infection in *I. scapularis* nymphs and adults typically reaches stable and predictable levels as noted elsewhere (Hamer et al., 2014; Keesing et al., 2014; Prusinski et al., 2014; Feldman et al., 2015). Here, we estimate that in the Upper Midwest, regional infection prevalence of *B. burgdorferi* s.s. and *A. phagocytophilum* in nymphal *I. scapularis*, the most epidemiologically important life stage, averaged 16.97% (95% CI: 13.96–19.98%) and 6.57% (95% CI: 4.47–8.66%) respectively. This is consistent with estimates from a

separate data set presented recently by Lehane et al. (2021) which found similar rates of *B. burgdorferi* s.s. in *I. scapularis* nymphs (17.99% [16.82–19.22%]) in the Midwest (IN, MI, MN, WI). However, the estimate of *A. phagocytophilum* in *I. scapularis* nymphs (4.03% [3.46–4.69%]) was slightly but not significantly lower than shown in our study, but differences are not likely to impact public health action and might be attributable to inclusion of more sites in the Lehane et al. (2021) study along the leading edge of *A. phagocytophilum* expansion. Similarly, in New York, Prusinski et al. (2014) presented a regional prevalence of *B. burgdorferi* s.s. infection in nymphs as 14.4%, again consistent with state estimates derived from an independent surveillance data set (Lehane et al., 2021). Although there are relatively fewer studies focused on *A. phagocytophilum*, Keesing et al. (2014) showed an 8.3% ( $\pm$ 0.6% SEM) infection prevalence in questing *I. scapularis* nymphs in Dutchess County, NY, (an estimate similar to the New York estimate presented by Lehane et al. (2021)) and demonstrated stability of infection prevalence with no discernable temporal trends.

Although our data represent many years of repeated, systematic sampling of *I. scapularis* at sites in the Upper Midwest, there are some significant time breaks at select sites in the data. We accounted for this in our analysis by only running the mixed effect model and ACF on those sites with five years of contiguous sampling which limited the data included in the analyses, and therefore, our ability to draw broader conclusions.

Optimizing effort and resource allocation for tick surveillance is important because public health resources are limited. Designing optimal sampling strategies depends on local factors and goals of public health agencies. For sites where prevalence of *B. burgdorferi* s.s. and/or A. phagocytophilum are consistent with regional averages in local I. scapularis populations, our study suggests extending the interval between sampling events is likely sufficient to maintain up-to-date estimates of infection prevalence for the public and health care providers. Moreover, our subset analyses where reduced sampling (infection prevalence estimates based on as little as a single year per site) yielded similar infection prevalence results to multiple-year sampling estimates at a regional level, suggests single year sampling across a broad spatial area yields estimates of infection prevalence that are similar to more labor-intensive and costly longitudinal sampling efforts. However, because the data are a convenience sample of previous tick surveillance activities and not a designed study, we are unable to make evidence-based recommendations regarding the optimal number of sampling sites or site placement. Future efforts to refine tick surveillance to improve efficiency and cost-effectiveness should focus on optimal placement of sampling locations, and the minimum number of sites required to generate reliable risk estimates. Our study was limited in scope to assessing estimates of infection prevalence. However, we recognize a need for similar assessments that address other surveillance metrics, including tick densities and describing host-seeking phenology.

Given the observed variability, lack of temporal trends, and consistency of site-specific estimates with regional estimates of *B. burgdorferi* and *A. phagocytophilum* prevalence, we conclude that monitoring infection prevalence in ticks aids in describing coarse acarological risk trends, but setting a fixed prevalence threshold for prevention or diagnostic decisions is not feasible. Additionally, we show that reducing repeated sampling of the same sites

has minimal impact on calculation of regional estimates of average infection prevalence, information that might be useful in allocating scarce public health resources for tick and tick-borne disease surveillance and control activities.

#### **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

#### Acknowledgments

We thank all our public health partners who participate in proposing, designing, funding, and conducting tick and tick-borne disease surveillance activities in Michigan (Michelle Clayson, Emily Dinh, Sarah Hamer, Vishvipali Kobbekaduwa, Megan Porter, Kimberly Signs, Mary Grace Stobierski, Rebecca Wong), Minnesota (Jordan Mandli, Molly Peterson, Elizabeth Schiffman), and Wisconsin (Rebecca Osborn, Ryan Wozniak).

#### References

- Bjork J, Schiffman E, 2020. The changing landscape of tickborne disease in Minnesota: a spotlight on emerging diseases. Minn. Med. Assoc 24–26. March/April.
- Bisanzio D, Fernández MP, Martello E, Reithinger R, Diuk-Wasser MA, 2020. Current and future spatiotemporal patterns of Lyme disease reporting in the northeastern United States. JAMA Netw. Open. 3, e200319. [PubMed: 32125426]
- CDC, 2018. Surveillance for *Ixodes scapularis* and Pathogens Found in this Tick Species in the United States (Accessed 04 June 2021). https://www.cdc.gov/ticks/surveillance/BlackleggedTick.html.
- Connally NP, Ginsberg HS, Mather TN, 2006. Assessing peridomestic entomological factors as predictors for Lyme disease. J. Vector Ecol. 31, 364–370. [PubMed: 17249354]
- Dahlgren FS, Paddock CD, Springer YP, Eisen RJ, Behravesh CB, 2016. Expanding Range of *Amblyomma americanum* and Simultaneous Changes in the Epidemiology of Spotted Fever Group Rickettsiosis in the United States. Am. J. Trop. Med. Hyg. 94, 35–42. [PubMed: 26503270]
- Daniels TJ, Fish D, Schwartz I, 1993. Reduced abundance of Ixodes scapularis (Acari: ixodidae) and Lyme disease risk by deer exclusion. J. Med. Entomol. 30, 1043–1049. [PubMed: 8271246]
- Dobson AD, 2014. History and complexity in tick-host dynamics: discrepancies between 'real' and 'visible' tick populations. Parasit. Vectors. 19, 231.
- Diuk-Wasser MA, Gatewood AG, Cortinas MR, Yaremych-Hamer S, Tsao J, Kitron U, Hickling G, Brownstein JS, Walker E, Piesman J, Fish D, 2006. Spatiotemporal patterns of host-seeking Ixodes scapularis nymphs (Acari: ixodidae) in the United States. J. Med. Entomol. 43, 166–176. [PubMed: 16619595]
- Diuk-Wasser MA, Hoen AG, Cislo P, et al., 2012. Human risk of infection with *Borrelia burgdorferi*, the Lyme disease agent, in eastern United States. Am. J. Trop. Med. Hyg. 86, 320–327. [PubMed: 22302869]
- Eisen L, Eisen RJ, Chang CC, Mun J, Lane RS, 2004. Acarologic risk of exposure to *Borrelia burgdorferi* spirochaetes: long-term evaluations in north-western California, with implications for Lyme borreliosis risk-assessment models. Med Vet Entomol 18, 38–49. [PubMed: 15009444]
- Eisen RJ, Eisen L, Beard CB, 2016. County-scale distribution of Ixodes scapularis and Ixodes pacificus (Acari: ixodidae) in the continental United States. J. Med. Entomol. 53, 349–386. [PubMed: 26783367]
- Eisen RJ, Eisen L, Ogden NH, Beard CB, 2016. Linkages of Weather and Climate With Ixodes scapularis and Ixodes pacificus (Acari: ixodidae), Enzootic Transmission of *Borrelia burgdorferi*, and Lyme Disease in North America. J. Med. Entomol. 53, 250–261. [PubMed: 26681789]
- Eisen RJ, Kugeler KJ, Eisen L, Beard CB, Paddock CD, 2017. Tick-Borne Zoonoses in the United States: persistent and emerging threats to human health. ILAR J 58, 319–335. [PubMed: 28369515]

- Eisen RJ, Eisen L, 2018. The blacklegged tick, *Ixodes scapularis*: an increasing public health concern. Trends. Parasitol. 34, 295–309. [PubMed: 29336985]
- Eisen RJ, Paddock CD, 2021. Tick and tick-borne pathogen surveillance as a public health tool in the United States. J. Med. Entomol. 58, 1490–1502. [PubMed: 32440679]
- Elias SP, Smith RP Jr., Morris SR, Rand PW, Lubelczyk C, Lacombe EH, 2011. Density of *Ixodes scapularis* ticks on Monhegan Island after complete deer removal: a question of avian importation? J. Vector Ecol. 36, 11–23. [PubMed: 21635637]
- Falco RC, Fish D, 1992. A comparison of methods for sampling the deer tick, Ixodes *dammini*, in a Lyme disease endemic area. Exp. Appl. Acarol. 14, 165–173. [PubMed: 1638929]
- Feldman KA, Connally NP, Hojgaard A, Jones EH, White JL, Hinckley AF, 2015. Abundance and infection rates of *Ixodes scapularis* nymphs collected from residential properties in Lyme diseaseendemic areas of Connecticut, Maryland, and New York. J Vector Ecol 40, 198–201. [PubMed: 26047204]
- Ginsberg HS, Zhioua E, Mitra S, Fischer J, Buckley PA, Verret F, Underwood HB, Buckley FG, 2004. Woodland type and spatial distribution of nymphal *Ixodes* scapularis (Acari: ixodidae). Environ. Entomol. 33, 1266–1273.
- Ginsberg HS, Rulison EL, Miller JL, Pang G, Arsnoe IM, Hickling GJ, Ogden NH, LeBrun RA, Tsao JI, 2020. Local abundance of Ixodes scapularis in forests: effects of environmental moisture, vegetation characteristics, and host abundance. Ticks Tick Borne Dis 11, 1–11.
- Hamer SA, Tsao JI, Walker ED, Hickling GJ, 2010. Invasion of the lyme disease vector *Ixodes scapularis*: implications for *Borrelia burgdorferi* endemicity. Ecohealth 7, 47–63. [PubMed: 20229127]
- Hamer SA, Hickling GJ, Sidge JL, Walker ED, Tsao JI, 2012. Synchronous phenology of juvenile Ixodes scapularis, vertebrate host relationships, and associated patterns of Borrelia burgdorferi ribotypes in the midwestern United States. Ticks Tick Borne Dis 3, 65–74. [PubMed: 22297162]
- Hamer SA, Hickling GJ, Walker ED, Tsao JI, 2014. Increased diversity of zoonotic pathogens and *Borrelia burgdorferi* strains in established versus incipient *Ixodes scapularis* populations across the Midwestern United States. Infect. Genet. Evol. 27, 531–542. [PubMed: 24953506]
- Herrington JE Jr., Campbell GL, Bailey RE, Cartter ML, Adams M, Frazier EL, Damrow TA, Gensheimer KF, 1997. Predisposing factors for individuals' Lyme disease prevention practices: connecticut, Maine, and Montana. Am. J. Public Health. 87, 2035–2038. [PubMed: 9431299]
- Johnson TL, Graham CB, Maes SE, Hojgaard A, Fleshman A, Boegler KA, Delory MJ, Slater KS, Karpathy SE, Bjork JK, Neitzel DF, Schiffman EK, Eisen RJ, 2018. Prevalence and distribution of seven human pathogens in host-seeking Ixodes scapularis (Acari: ixodidae) nymphs in Minnesota, USA. Ticks Tick Borne Dis 9, 1499–1507. [PubMed: 30055987]
- Jones CJ, Kitron UD, 2000. Populations of Ixodes scapularis (Acari: ixodidae) are modulated by drought at a Lyme disease focus in Illinois. J. Med. Entomol. 37, 408–415. [PubMed: 15535585]
- Keesing F, McHenry DJ, Hersh M, Tibbetts M, Brunner JL, Killilea M, LoGiudice K, Schmidt KA, Ostfeld RS, 2014. Prevalence of human-active and variant 1 strains of the tick-borne pathogen *Anaplasma phagocytophilum* in hosts and forests of eastern North America. Am. J. Trop. Med. Hyg. 91, 302–309. [PubMed: 24865688]
- Kugeler KJ, Eisen RJ, 2020. Challenges in predicting Lyme disease risk. JAMA Netw. Open. 3, e200328. [PubMed: 32125424]
- Kugeler KJ, Farley GM, Forrester JD, Mead PS, 2015. Geographic distribution and expansion of human Lyme disease, United States. Emerg. Infect. Dis. 21, 1455–1457. [PubMed: 26196670]
- Lantos PM, Rumbaugh J, Bockenstedt LK, Falck-Ytter YT, Aguero-Rosenfeld ME, Auwaerter PG, Baldwin K, et al., 2021. Clinical Practice Guidelines by the Infectious Diseases Society of America, American Academy of Neurology, and American College of Rheumatology: 2020 Guidelines for the Prevention, Diagnosis, and Treatment of Lyme Disease. Neurology 96, 262– 273. [PubMed: 33257476]
- Larson S, Sabo A, Kruger E, Jones P, Paskewitz SM, 2021. Ixodes Scapularis Density in US Temperate Forests Shaped by deer, earthworms, and Disparate Factors at Two Scales. Ecosphere press.

- Lehane A, Maes SE, Graham CB, Jones E, Delorey M, Eisen RJ, 2021. Prevalence of single and coinfections of human pathogens in Ixodes ticks from five geographical regions in the United States, 2013–2019. Ticks Tick Borne Dis 12, 101637. [PubMed: 33360805]
- Levin ML, Ross DE, 2004. Acquisition of different isolates of *Anaplasma phagocytophilum* by *Ixodes scapularis* from a model animal. Vector Borne Zoonotic Dis 4, 53–59. [PubMed: 15018773]
- Mader EM, Ganser C, Geiger A, Harrington LC, Foley JE, Smith RL, Mateus-Pinilla N, Teel PD, Eisen RJ, 2021. A survey of tick surveillance and control practices in the United States. J. Med. Entomol. 58, 1503–1512. [PubMed: 34270770]
- Mather TN, Nicholson MC, Donnelly EF, Matyas BT, 1996. Entomologic index for human risk of Lyme disease. Am. J. Epidemiol. 144, 1066–1069. [PubMed: 8942438]
- Moore A, Nelson C, Molins C, Mead P, Schriefer M, 2016. Current guidelines, common clinical pitfalls, and future directions for laboratory diagnosis of Lyme Disease, United States. Emerg. Infect. Dis. 22, 1169–1177. [PubMed: 27314832]
- Niesobecki S, Hansen A, Rutz H, Mehta S, Feldman K, Meek J, Niccolai L, Hook S, Hinckley A, 2019. Knowledge, attitudes, and behaviors regarding tick-borne disease prevention in endemic areas. Ticks Tick Borne Dis 10, 101264. [PubMed: 31431351]
- O'Connor C, Prusinski MA, Jiang S, Russell A, White J, Falco R, Kokas J, Vinci V, Gall W, Tober K, Haight J, Oliver J, Meehan L, Sporn LA, Brisson D, Backenson PB., 2021. A Comparative Spatial and Climate Analysis of Human Granulocytic Anaplasmosis and Human Babesiosis in New York State (2013–2018). J. Med. Entomol. 1–14. [PubMed: 32772108]
- Ogden NH, Pang G, Ginsberg HS, Hickling GJ, Burke RL, Beati L, Tsao JI, 2018. Evidence for Geographic Variation in Life-Cycle processes affecting phenology of the Lyme disease vector Ixodes scapularis (Acari: ixodidae) in the United States. J. Med. Entomol. 55, 1386–1401. [PubMed: 29986046]
- Ostfeld RS, Schauber EM, Canham CD, Keesing F, Jones CG, Wolff JO, 2001. Effects of acorn production and mouse abundance on abundance and *Borrelia burgdorferi* infection prevalence of nymphal *Ixodes scapularis* ticks. Vector Borne Zoonotic Dis 1, 55–63. [PubMed: 12653136]
- Ostfeld RS, Canham CD, Oggenfuss K, Winchcombe RJ, Keesing F, 2006. Climate, deer, rodents, and acorns as determinants of variation in lyme-disease risk. PLoS Biol 4, 1058–1068.
- Pepin KM, Eisen RJ, Mead PS, Piesman J, Fish D, Hoen AG, Barbour AG, Hamer S, Diuk-Wasser MA, 2012. Geographic variation in the relationship between human Lyme disease incidence and density of infected host-seeking *Ixodes scapularis* nymphs in the Eastern United States. Am. J. Trop. Med. Hyg. 86, 1062–1071. [PubMed: 22665620]
- Piesman J, 1989. Transmission of Lyme disease spirochetes (Borrelia burgdorferi). Exp. Appl. Acarol. 7, 71–80. [PubMed: 2667921]
- Piesman J, Clark KL, Dolan MC, Happ CM, Burkot TR, 1999. Geographic survey of vector ticks (Ixodes scapularis and Ixodes pacificus) for infection with the Lyme disease spirochete, Borrelia *burgdorferi*. J. Vector Ecol. 24, 91–98. [PubMed: 10436883]
- Pritt BS, Respicio-Kingry LB, Sloan LM, Schriefer ME, Replogle AJ, Bjork J, Liu G, Kingry LC, Mead PS, Neitzel DF, Schiffman E, Johnson Hoang, Davis DK, Paskewitz JP, Boxrud SM, Deedon D, Lee A, Miller X, Feist TK, Steward MA, Theel CR, Patel ES, Irish R, Petersen JM, C.L., 2016. *Borrelia mayonii* sp. nov., a member of the *Borrelia burgdorferi* sensu lato complex, detected in patients and ticks in the upper midwestern United States. Int. J. Syst. Evol. Microbiol. 66, 4878–4880. [PubMed: 27558626]
- Prusinski MA, Kokas JE, Hukey KT, Kogut SJ, Lee J, Backenson PB, 2014. Prevalence of Borrelia burgdorferi (Spirochaetales: spirochaetaceae), Anaplasma phagocytophilum (Rickettsiales: anaplasmataceae), and Babesia microti (Piroplasmida: babesiidae) in Ixodes scapularis (Acari: ixodidae) collected from recreational lands in the Hudson Valley Region, New York State. J. Med. Entomol. 51, 226–236. [PubMed: 24605473]
- Rosenberg R, Lindsey NP, Fischer M, Gregory CJ, Hinckley AF, Mead PS, Paz-Bailey G, Waterman SH, Drexler NA, Kersh GJ, Hooks H, Partridge SK, Visser SN, Beard CB, Petersen LR, 2018. Vital Signs: trends in reported vectorborne disease cases - United States and territories, 2004– 2016. Morb. Mortal. Wkly. Rep. 67, 496–501.

- Schulze TL, Jordan RA, 1996. Seasonal and long-term variations in abundance of adult Ixodes scapularis (Acari:ixodidae) in different coastal plain habitats of New Jersey. J. Med. Entomol. 33, 963–970. [PubMed: 8961647]
- Schulze TL, Jordan RA, 2003. Meteorologically mediated diurnal questing of Ixodes scapularis and Amblyomma americanum (Acari: ixodidae) nymphs. J. Med. Entomol. 40, 395–402. [PubMed: 14680102]
- Schwartz AM, Hinckley AF, Mead PS, Hook SA, Kugeler KJ, 2017. Surveillance for Lyme disease— United States, 2008–2015. MMWR Surveill. Summ. 66, 1–12.
- Stafford KC, Cartter ML, Magnarelli LA, Ertel SH, Mshar PA, 1998. Temporal correlations between tick abundance and prevalence of ticks infected with *Borrelia burgdorferi* and increasing incidence of Lyme disease. J. Clin. Microbiol. 36, 1240–1244. [PubMed: 9574684]
- Stafford KC 3rd, Massung RF, Magnarelli LA, Ijdo JW, Anderson JF, 1999. Infection with agents of human granulocytic ehrlichiosis, Lyme disease, and babesiosis in wild white-footed mice (Peromyscus leucopus) in Connecticut. J. Clin. Microbiol. 37, 2887–2892. [PubMed: 10449470]
- Stromdahl EY, Hickling GJ, 2012. Beyond Lyme: aetiology of tick-borne human diseases with emphasis on the south-eastern United States. Zoonoses Public Health 59 (Suppl 2), 48–64. [PubMed: 22958250]
- Teglas MB, Foley J, 2006. Differences in the transmissibility of two *Anaplasma* phagocytophilum strains by the North American tick vector species, Ixodes pacificus and Ixodes scapularis (Acari: ixodidae). Exp. Appl. Acarol. 38, 47–58. [PubMed: 16550334]
- Telford SR 3rd, Dawson JE, Katavolos P, Warner CK, Kolbert CP, Persing DH, 1996. Perpetuation of the agent of human granulocytic ehrlichiosis in a deer tick-rodent cycle. Proc Natl Acad Sci 93 (12), 6209–6214. [PubMed: 8650245]
- Thomas CE, Burton ES, Brunner JL, 2020. Environmental Drivers of Questing Activity of Juvenile Black-Legged Ticks (Acari: ixodidae): temperature, Desiccation Risk, and Diel Cycles. J. Med. Entomol. 57, 8–16. [PubMed: 31370063]
- Van Buskirk J, Ostfeld RS, 1995. Controlling Lyme disease by modifying the density and species composition of tick hosts. Eco. Appl. 5, 1133–1140.
- Vuong HB, Chiu GS, Smouse PE, Fonseca DM, Brisson D, Morin PJ, Ostfeld RS, 2017. Influences of Host Community Characteristics on Borrelia burgdorferi Infection Prevalence in Blacklegged Ticks. PLoS ONE 12, e0167810. [PubMed: 28095423]
- Wilson EB, 1927. Probable inference, the law of succession, and statistical inference. J. Am. Stat. Assoc. 22, 209–212.
- Wormser GP, Dattwyler RJ, Shapiro ED, Halperin JJ, Steere AC, Klempner MS, Krause PJ, Bakken JS, Strle F, Stanek G, Bockenstedt L, Fish D, Dumler JS, Nadelman RB, 2006. The clinical assessment, treatment, and prevention of lyme disease, human granulocytic anaplasmosis, and babesiosis: clinical practice guidelines by the Infectious Diseases Society of America. Clin. Infect. Dis. 43, 1089–1134. [PubMed: 17029130]



#### Fig. 1.

Geographic locations of tick surveillance sites in Michigan (N= 9), Minnesota (N= 4), and Wisconsin (N= 15), meeting study inclusion criteria. Numbered labels correspond to site identification numbers referenced in subsequent tables and figures.



#### Fig. 2.

Point estimates with bars showing 95% confidence intervals for the annual proportion of *I. scapularis* nymphs infected with *B. burgdorferi* s.s. at sites with 5 contiguous years of data. Breaks in the lines connecting dots represent years where data were not collected. For 10 established sites, the mixed effects model for *B. burgdorferi* s.s. infected nymphs showed no significant temporal trend (t = -1.7, df = 84, p = 0.10) in infection prevalence. Additionally, at a single site classified as emerging, no significant temporal trend in infection prevalence was detected (t = 0.34, df = 3, p = 0.76). Points with solid 95% CI lines were included in the autocorrelation function (ACF) plots (Supplemental Figure 1).



#### Fig. 3.

Point estimates with bars showing 95% confidence intervals for the annual proportion of *I. scapularis* adults infected with *B. burgdorferi* ss at sites with 5 contiguous years of data. Breaks in the lines connecting dots represent years where data were not collected. For 5 established sites, the mixed effects model for *B. burgdorferi* s.s. infected adults showed no significant temporal trend (t = 0.66, df = 55, p = 0.51) in infection prevalence. For 3 emerging sites, a significant positive temporal trend in infection prevalence was detected (t = 3.1, df = 30, p = 0.004). Points with solid 95% CI lines were included in the autocorrelation function (ACF) plots (Supplemental Figure 2).



#### Fig. 4.

Point estimates with bars showing 95% confidence intervals for the annual proportion of *I. scapularis* nymphs infected with *A. phagocytophilum* at sites with 5 contiguous years of data. Breaks in the lines connecting dots represent years where data were not collected. For 5 established sites, the mixed effects model for *A. phagocytophilum*. infected nymphs showed no significant temporal trend (t = -0.05, df = 44, p = 0.96) in infection prevalence. Additionally, at a single site classified as emerging, no significant temporal trend in infection prevalence was detected (t = 1.03, df = 3, p = 0.38). Points with solid 95% CI lines were included in the autocorrelation function (ACF) plots (Supplemental Figure 3).



#### Fig. 5.

Point estimates with bars showing 95% confidence intervals for the annual proportion of *I. scapularis* adults infected with *A. phagocytophilum* at sites with 5 contiguous years of data. Breaks in the lines connecting dots represent years where data were not collected. For 4 established sites, the mixed effects model for *A. phagocytophilum* infected adults showed no significant temporal trend (t = 1.9, df = 42, p = 0.06) in infection prevalence. At a single site classified as emerging, a significant positive temporal trend in infection prevalence was detected (t = 3.1, df = 18, p = 0.007). Points with solid 95% CI lines were included in the autocorrelation function (ACF) plots (Supplemental Figure 4).

1	τ	5
	-	+
	٦	
	2	
	2	
	2	-
	-	2
	C	2
	-	
	_	-
	<	<
	5	5
	뇓	2
	-	2
	C	
	ā	5
	۲	5
	2	4
	-	
1	C	5
	ē	÷

Author Manuscript

# Table 1

Abundance and prevalence estimates for B. burgdorferis.s. in host-seeking nymphal I. scapularis at 25 sites, surveyed multiple years, in the Upper Midwestern United States.

		Survey site information		<i>I. scapulari</i> s nymphal abundance <sup>a</sup>	I. scapularis 1 burgdorferi s.	nymphs assayed for $B$ .	B. burgdorf (95% CI) <sup>c</sup>	feri s.s. prevaleı	nce estimate
State	Site ID	Site Name	Years Sampled (range)	Median peak abundance, ticks/100m <sup>2</sup> (range)	Total Ticks Tested	Median # Ticks Assayed / Year (range)	Mean	Lower	Upper
W	-	Duck Lake State Park	7	0.40 (0.03 – 1.03)	77	11 (3 – 22)	0.1429	0.0817	0.2380
	2	Fenner Nature Center	S	3.00(1.00 - 8.12)	358	65 (28 – 122)	0.0140	0.0060	0.0323
	3	Fort Custer Recreation Area	5	$0.40\ (0.05 - 2.00)$	82	8 (2 – 51)	0.2683	0.1844	0.3730
	5	Ludington State Park	ε	$0.70 \ (0.44 - 0.75)$	62	14 (7 – 41)	0.1613	0060.0	0.2721
	9	Saugatuck Dunes State Park	9	$0.88 \ (0.17 - 6.25)$	96	11 $(3 - 37)$	0.0521	0.0224	0.1162
	7	SLBE Platte-Eldorado	9	0.44 (0.07 – 3.10)	133	13.5 (3 – 66)	0.1955	0.1370	0.2710
	6	Van Buren State Park	12	$2.00\ (0.17-5.00)$	1116	27.5 (3 – 817)	0.1201	0.1023	0.1405
		MI Summary Data	6 (3–12)	0.70 (0.40 – 3.00)	1924	13.5 (8 – 65)	0.1363	0.0572	0.2154
Į	10	Camp Ripley	11	1.54 (0.38 – 12.50)	892	83 (49 – 125)	0.2119	0.1863	0.2399
	11	Richard J. Dorer Memorial Hardwood State Forest	8	$0.46\ (0.06 - 1.54)$	310	22.5 (6 - 106)	0.1419	0.1075	0.1852
	12	Itasca State Park	12	$0.83 \ (0.46 - 3.33)$	723	56 (35 – 102)	0.1618	0.1368	0.1904
	13	St. Croix State Park	11	2.04 (0.59 – 12.00)	1132	105 (51 – 170)	0.1776	0.1564	0.2009
		MN Summary Data	11 (8–12)	1.18 (0.46 – 2.04)	3057	69.5 (22.5 – 105)	0.1733	0.1262	0.2204
И	14	American Legion Northern Highland	6	2.50 (1.62 – 6.44)	624	84 (56 – 232)	0.1795	0.1514	0.2115
	15	Big Eau Pleine County Park	9	7.29 (0.41 – 65.70)	1960	273 (50 – 708)	0.2327	0.2145	0.2519
	16	Black River Falls State Forest	6	4.40 (2.10 – 12.10)	835	106 (6 – 128)	0.2359	0.2084	0.2659
	17	Camp Phillips	3	2.30 (2.00 – 7.90)	268	101 (50 - 117)	0.2239	0.1781	0.2775
	18	Devil's Lake State Park	3	0.22~(0.20-0.40)	146	45 (10 – 91)	0.0959	0.0580	0.1545
	19	Flambeau State Forest	5	1.79 (0.92 – 3.83)	263	43 (22 – 92)	0.2548	0.2059	0.3107
	20	Hartman Creek State Park	Э	$2.80\ (0.50 - 17.80)$	393	166 (50 – 177)	0.2697	0.2282	0.3157
	21	Kettle Moraine State Forest-Southern Unit	10	5.60 (0.33 – 12.40)	894	111 (7 – 130)	0.2248	0.1987	0.2533
	22	Kohler-Andrae State Park	4	$11.45\ (5.00 - 15.10)$	333	82 (50 - 119)	0.2072	0.1671	0.2540

		Survey site information		<i>I. scapulari</i> s nymphal abundance <sup>a</sup>	I. scapularis n burgdorferi s.	ymphs assayed for $B$ .	B. burgdorf (95% CI) <sup>c</sup>	<i>eri</i> s.s. prevalen	ce estimate
State S	lite ID	Site Name	Years Sampled (range)	Median peak abundance, ticks/100m <sup>2</sup> (range)	Total Ticks Tested	Median # Ticks Assayed / Year (range)	Mean	Lower	Upper
2	ŝ	McCaslin Brook	4	2.44 (0.96 – 2.80)	225	58.5 (23 – 85)	0.1244	0.0875	0.1740
6	4	Mirror Lake State Park	з	3.23 (1.20 – 4.72)	358	77 (53 – 228)	0.0866	0.0617	0.1203
2	5	Tower Hill State Park	4	3.40 (1.70 – 5.20)	369	99.5 (50 – 120)	0.2818	0.2357	0.3280
2	9	UW-Arboretum	7	$0.40\ (0.06-0.81)$	626	74 (20 – 239)	0.0879	0.0656	0.1101
2	Ľ	Wildcat Mt. State Park	з	3.60(3.50-5.00)	319	101 (98 - 120)	6060.0	0.0640	0.1275
		WI Summary Data	4 (3–10)	3.02 (0.22 – 11.45)	7613	91.75 (43 – 273)	0.1854	0.1432	0.2276
		Regional Summary Data	6 (4–11)	1.18 (0.70 – 3.02)	12,594	69.5 (13.5 – 91.75)	0.1697	0.1396	0.1998

<sup>a</sup>Ticks were collected via drag cloth during peak nymphal activity periods. When sites were sampled multiple times per year, the highest value was denoted as the peak. Median tick abundance and range by state, and region calculated on site medians.

Centers for Disease Control and Prevention Guidelines: "Surveillance for *Ixodes scapularis* and pathogens found in this tick species in the United States" (2018). https://www.cdc.gov/ticks/resources/ <sup>b</sup> to identify *B. burgdorferi*ss in *I. scapularis* nymphs, ticks were tested individually using species specific molecular assays which met the minimum criteria for acceptability according to the TickSurveillance\_Iscapularis-P.pdf. Median ticks assayed and range by state, and region calculated on site medians.

Ticks Tick Borne Dis. Author manuscript; available in PMC 2023 November 02.

<sup>c</sup>. The proportion of ticks infected per site and Wilson score 95% confidence intervals are shown; state and regional averages were based on these site-specific point estimates and 95% confidence intervals were derived assuming a t-distribution to account for small sample sizes (<30).

Author Manuscript

Author Manuscript

-
_
_
<b>+</b>
-
-
C
$\mathbf{U}$
_
_
~
$\leq$
Ň
Ma
Mai
Mar
Man
Manu
Manu
Manus
Manus
Manus
Manusc
Manusci
Manuscr
Manuscri
Manuscrip
Manuscrip
Manuscript

S	
tate	
S T	2
tec	
L	
n I	
ter	
ves	
q	5
Ī	
er	
Jnr	1
еI	,
Ę	
.E	
LS.	
vea	-
é	ľ
ti.	ļ
- Inc	
ļ	
vec	5
ve.	
SUIT	
5	1
site	
4	
at 1	
.i.s.L	
lar	
IUE	4
SC	
1	
ac	1
0 LL	p
eki	
-se	
St	
, ř	
·=	
S	
eri	
tro	
рø.	i D
Inc	ľ
8	
, T	
fc	
tes	
m	
5Sti	
je e	
enc	
vale	
rei	

Foster et al.

				abundance <sup>a</sup>	burgdorferi s.s	<i>q</i> 's	CI)¢	4	
State	Site ID	Site Name	Years Sampled (range)	Median peak abundance, ticks/100m <sup>2</sup> (range)	Total Ticks Tested	Median # Ticks Tested/ Year (range)	Mean	Lower	Upper
IM	1	Duck Lake State Park	5	0.48 (0.11 – 3.06)	123	23 (3 – 49)	0.3008	0.2268	0.3869
	2	Fenner Nature Center	6	2.35 (0.50 – 4.29)	652	113 (20 – 229)	0.0997	0.0790	0.1251
	б	Fort Custer Recreation Area	5	1.64 (0.08 – 3.93)	136	24 (4 – 51)	0.5149	0.4311	0.5979
	4	Ionia Recreation Area	4	0.49~(0.22 - 3.93)	28	4(4-16)	0.0357	0.0018	0.1771
	5	Ludington State Park	3	$0.44\ (0.31-0.70)$	38	8 (8 – 22)	0.2368	0.1299	0.3921
	9	Saugatuck Dunes State Park	4	0.60 (0.33 – 0.75)	56	10.5(2-18)	0.3036	0.1990	0.4334
	7	SLBE Platte-Eldorado	7	$0.58\ (0.40-0.77)$	116	19 (2 – 34)	0.2500	0.1801	0.3360
	8	SLBE Pyramid Point	4	$0.22\ (0.10-0.37)$	26	6.5(2-11)	0.1923	0.0851	0.3788
	6	Van Buren State Park	12	$1.50\ (0.82 - 5.10)$	574	50.5 (6 – 97)	0.3763	0.3376	0.4166
		MI Summary Data	5 (3 - 12)	0.58 (0.22 – 2.35)	1749	19 (4 - 113)	0.2567	0.1469	0.3665
MN	10	Camp Ripley	13	3.67 (1.00 – 7.21)	1145	101 (25 – 119)	0.3790	0.3514	0.4075
	11	Richard J. Dorer Memorial Hardwood State Forest	12	1.57 (0.70 – 3.92)	1218	101.5 (27 – 177)	0.4507	0.4230	0.4788
	12	Itasca State Park	10	$1.73 \ (0.54 - 4.54)$	912	105(44 - 115)	0.3827	0.3517	0.4146
	13	St. Croix State Park	12	2.92 (0.75 – 5.58)	1291	108.5(40 - 171)	0.3261	0.3011	0.3522
		MN Summary Data	12 (10 – 13)	2.33 (1.57 – 3.67)	4566	103.25 (101 - 108.5)	0.3846	0.3034	0.4659
IW	28	Stevens Point	20	NA	1947	92.5 (8 – 232)	0.2861	0.2664	0.3066
		WI Summary Data					0.2861	NA	NA
		Regional Summary Data	12 (5 – 20)	1.46 (0.58 – 2.33)	8262	92.5 (19 – 103.25)	0.2953	0.2208	0.3698

Ticks Tick Borne Dis. Author manuscript; available in PMC 2023 November 02.

b. To identify *B. burgdorferiss* in *I. scapularis* adults, ticks were tested individually using species specific molecular assays which met the minimum criteria for acceptability according to the Centers for Disease Control and Prevention Guidelines: "Surveillance for *Ixodes scapularis* and pathogens found in this tick species in the United States" (2018). https://www.cdc.gov/ticks/resources/ TickSurveillance\_Iscapularis-P.pdf. Median tick abundance and range by state, and region calculated on site medians.

Author Manuscript

<sup>C</sup>The proportion of ticks infected per site and Wilson score 95% confidence intervals are shown; state and regional averages were based on these site-specific point estimates and 95% confidence intervals were derived assuming a t-distribution to account for small sample sizes (<30).

Foster et al.

-	
_	
-	
_	
_	
_	
-	
_	
$\sim$	
$\mathbf{U}$	
_	
_	
-	
_	
5	
0	
۵	
lar	
lan	
lan	
lanu	
lanu	
lanus	
lanus	
lanus	
lanuso	
lanusc	
lanusci	
lanuscr	
lanuscri	
lanuscri	
lanuscrip	
lanuscrip	
lanuscript	

Prevalence estimates for A. phagocytophilum in host-seeking nymphal I. scapularis at 10 sites, surveyed multiple years, in the Upper Midwestern United States.

Survey	y site infor.	mation		<i>I. scapularis</i> nymphal abundance <sup>a</sup>	I. scapularis n phagocytophili	ymphs assayed for <i>A.</i> <i>um<sup>b</sup></i>	A. phagocyt. (95% CI) <sup>c</sup>	<i>ophilum</i> preval	ence estimate
State	Site ID	Site Name	Years Sampled (range)	Median peak abundance, ticks/100m <sup>2</sup> (range)	Total Ticks Tested	Median # Ticks Tested/ Year (range)	Mean	Lower	Upper
MN	10	Camp Ripley	11	1.54 (0.38 - 12.50)	892	83 (49 – 125)	8660.0	0.0818	0.1212
	11	Richard J. Dorer Memorial Hardwood State Forest	×	0.46 (0.06 – 1.54)	310	22.5 (6 – 106)	0.0419	0.0247	0.0704
	12	Itasca State Park	12	$0.83 \ (0.46 - 3.33)$	723	56(20-102)	0.0733	0.0565	0.0946
	13	St. Croix State Park	11	$2.04\ (0.59 - 12.00)$	1132	105 (51 - 170)	0.0557	0.0437	0.0706
		MN Summary Data	11 (8 – 12)	1.18 (0.46 – 2.04)	3057	69.5 (56 – 105)	0.0677	0.0279	0.1074
IW	14	American Legion Northern Highland	S,	2.50 (1.62 – 6.44)	567	90 (56 – 232)	0.0406	0.0272	0.0601
	15	Big Eau Pleine County Park	4	10.33 (4.72 – 65.70)	1827	519.5 (50 – 738)	0.0996	0.0867	0.1142
	16	Black River Falls State Forest	8	3.70 (3.30 – 12.10)	734	108 (6 – 128)	0.0572	0.0426	0.0764
	19	Flambeau State Forest	5	$1.79\ (0.92 - 3.83)$	263	43 (22 – 92)	0.0494	0.0291	0.0827
	21	Kettle Moraine State Forest-Southern Unit	10	5.60 (0.33 – 12.40)	889	111 (7 – 130)	0.1125	0.0934	0.1349
	23	McCaslin Brook	4	2.44 (0.96 – 2.80)	225	58.5 (23 – 85)	0.0267	0.0123	0.0569
		WI Summary Data	5(4-10)	3.10 (1.79 – 10.33)	4505	99 (43 – 519.5)	0.0643	0.0285	0.1001
		Regional Summary Data	8 (5 – 11)	2.14 (1.18 – 3.10)	7562	84.25 (69.5 – 99)	0.0657	0.0447	0.0866

Ticks Tick Borne Dis. Author manuscript; available in PMC 2023 November 02.

Centers for Disease Control and Prevention Guidelines: "Surveillance for Ixodes scapularis and pathogens found in this tick species in the United States" (2018). https://www.cdc.gov/ticks/resources/ <sup>b</sup> to identify A. phagocytophilum in I. scapularis nymphs, ticks were tested individually using species specific molecular assays which met the minimum criteria for acceptability according to the TickSurveillance\_Iscapularis-P.pdf. Median tick abundance and range by state, and region calculated on site medians. <sup>c</sup>The proportion of ticks infected per site and Wilson score 95% confidence intervals are shown; state and regional averages were based on these site-specific point estimates and 95% confidence intervals were derived assuming a t-distribution to account for small sample sizes (<30).

$\mathbf{r}$
~
=
-
ō
U.
<
-
0
la
lan
lanu
lanu
lanus
lanus
lanusc
lanuscr
lanuscri
lanuscrip
<b>Nanuscrip</b>

## Table 4

Prevalence estimates for A. phagocytophilum in host-seeking adult Ixodes scapularis at 5 sites, surveyed multiple years, in the Upper Midwestern United States.

Surve	y site infor	mation		<i>I. scapularis</i> adult abundance <sup>a</sup>	I. scapularis ad phagocytophilu	ults assayed for $A$ . $n^b$	A. phagocytı CI) <sup>c</sup>	<i>ophilum</i> prevalend	ce estimate (95%
State	Site ID	Site Name	Years Sampled	Median peak abundance, ticks/100m <sup>2</sup> (range)	Total Ticks Tested	Median # Ticks Tested/ Year (range)	Mean	Lower	Upper
NM	10	Camp Ripley	12	3.67 (1.00 – 7.21)	1218	101.5 (27 – 177)	0.0944	0.0792	0.1121
	11	Richard J. Dorer Memorial Hardwood State Forest	10	1.57 (0.70 – 3.92)	912	103.5 (44 – 115)	0.0428	0.0314	0.0579
	12	Itasca State Park	13	$1.73\ (0.54-4.54)$	1145	101 (25 – 119)	0.1223	0.1045	0.1425
	13	St. Croix State Park	12	2.92 (0.75 – 5.58)	1291	108.5(40 - 171)	0.0790	0.0655	0.0950
		MN Summary Data	12 (10 – 13)	2.33 (1.57 – 3.67)	4566	102.5 (101 – 108.5)	0.0846	0.0319	0.1374
IV	28	Stevens Point	20	NA	1815	85 (8 – 232)	0.0909	0.0785	0.1050
		WI Summary Data					0.0909	NA	NA
		Regional Summary Data	16 (12 – 20)	NA	6381	93.7 (85 – 102.5)	0.0859	0:0201	0.1217

Ticks Tick Borne Dis. Author manuscript; available in PMC 2023 November 02.

Centers for Disease Control and Prevention Guidelines: "Surveillance for Ixodes scapularis and pathogens found in this tick species in the United States" (2018). https://www.cdc.gov/ticks/resources/ <sup>b</sup>To identify A. phagocytophilum in I. scapularis adults, ticks were tested individually using species specific molecular assays which met the minimum criteria for acceptability according to the TickSurveillance\_Iscapularis-P.pdf. Median tick abundance and range by state, and region calculated on site medians. <sup>c</sup>The proportion of ticks infected per site and Wilson score 95% confidence intervals are shown; state and regional averages were based on these site-specific point estimates and 95% confidence intervals were derived assuming a t-distribution to account for small sample sizes (<30).