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# Antiviral susceptibility of clade 2.3.4.4b highly pathogenic avian influenza A(H5N1) viruses isolated from birds and mammals in the United States, 2022

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#### Abstract

Clade 2.3.4.4b highly pathogenic avian influenza (HPAI) A(H5N1) viruses that are responsible for devastating outbreaks in birds and mammals pose a potential threat to public health. Here, we evaluated their susceptibility to influenza antivirals. Of 1,015 sequences of HPAI A(H5N1) viruses collected in the United States during 2022, eight viruses (~0.8%) had a molecular marker of drug resistance to an FDA-approved antiviral: three adamantane-resistant (M2-V27A), four oseltamivir-resistant (NA-H275Y), and one baloxavir-resistant (PA-I38T). Additionally, 31 viruses contained mutations that may reduce susceptibility to inhibitors of neuraminidase (NA) (n = 20) or cap-dependent endonuclease (CEN) (n = 11). A panel of 22 representative viruses was tested phenotypically. Overall, clade 2.3.4.4b A(H5N1) viruses lacking recognized resistance mutations were susceptible to FDA-approved antivirals. Oseltamivir was least potent at inhibiting NA activity, while the investigational NA inhibitor AV5080 was most potent, including against NA mutants. A novel NA substitution T438N conferred 12-fold reduced inhibition by zanamivir, and in combination with the known marker N295S, synergistically affected susceptibility to all

Declaration of competing interest

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The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the US Centers for Disease Control and Prevention.

A.A.I. and A.V.I. are the founders of ChemDiv, managing members of ASAVI LCC. A.V.I. is the author of patent application US 8895613. The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.antiviral.2023.105679.

five NA inhibitors. In cell culture-based assays HINT and IRINA, the PA-I38T virus displayed 75- to 108-fold and 37- to 78-fold reduced susceptibility to CEN inhibitors, baloxavir and the investigational AV5116, respectively. Viruses with PA-I38M or PA-A37T showed 5- to 10-fold reduced susceptibilities. As HPAI A(H5N1) viruses continue to circulate and evolve, close monitoring of drug susceptibility is needed for risk assessment and to inform decisions regarding antiviral stockpiling.

#### **Keywords**

Neuraminidase inhibitors; CEN inhibitor; Baloxavir; AV5080; Antiviral resistance

Highly pathogenic avian influenza (HPAI) A(H5N1) viruses persist in wild birds in many countries and cause deadly outbreaks in poultry and mammals worldwide. They are also known to cause sporadic human infections of various severity. Their circulation in wild birds is of public health concern as they may acquire the ability to effectively infect humans and transmit person-to-person (Krammer and Schultz-Cherry, 2023). HPAI A(H5N1) viruses display remarkable genetic heterogeneity due to genetic drift and reassortment that has resulted in multiple phylogenetically distinct hemagglutinin clades and various genotypes. In December 2021, clade 2.3.4.4b HPAI A(H5N1) viruses of Eurasian-origin were first detected on the Atlantic Coasts of North America (Alkie et al., 2023; Bevins et al., 2022). These viruses then underwent reassortment where they exchanged genes with those from North American lineage low pathogenic avian influenza viruses (Alkie et al., 2023; Elsmo et al., 2023; Kandeil et al., 2023). By the end of 2022, clade 2.3.4.4b HPAI A(H5N1) viruses were found in wild birds, domestic poultry, or wild mammals in nearly all continental states of the United States (US) (CDC, 2023).

Vaccines and antivirals are used to control seasonal influenza infections and are key components of pandemic preparedness. Antivirals can mitigate the impact of outbreaks in humans caused by zoonotic viruses for which vaccines may not be readily available. However, influenza viruses can become less susceptible to antivirals by acquiring genomic changes due to spontaneous mutations, gene reassortment, or selective pressure from antiviral treatment. To manage human influenza A virus infections, Food and Drug Administration (FDA) has approved antivirals from three classes: M2 blockers (adamantanes), neuraminidase (NA) inhibitors, and the cap-dependent endonuclease (CEN) inhibitor of the polymerase acid (PA) protein (Jones et al., 2023). Due to resistance of seasonal influenza A viruses to M2 blockers, Centers for Disease Control and Prevention (CDC) recommends the use of NA inhibitors oseltamivir, zanamivir, and peramivir and a CEN inhibitor, baloxavir (CDC, 2022). Laninamivir is another NA inhibitor which is approved in Japan (Ikematsu and Kawai, 2011).

Evaluation of antiviral susceptibility is an integral part of the risk assessment conducted on influenza viruses that pose a threat to public health (Cox et al., 2014; Global Influenza Programme WEP, 2020). In this study, clade 2.3.4.4b HPAI A(H5N1) viruses collected from birds and mammals in the US during 2022 were assessed for their susceptibility to FDA-approved and two investigational antivirals. First, virus genome sequences (n = 1,015)

obtained from the Global Initiative on Sharing All Influenza Data (GISAID) database were examined for known markers of resistance with perceived clinical relevance. Of the known markers of adamantane resistance (Gubareva and Hayden, 2006), M2-V27A was detected in three viruses collected from domestic birds in South Dakota on the same day (Table S1). Four viruses collected from birds in New England states had NA-H275Y, the common marker of oseltamivir resistance which may also confer resistance to peramivir (Table S1) (Lee and Hurt, 2018). Furthermore, a virus from a chicken contained PA-I38T, the main determinant for baloxavir resistance (Gubareva et al., 2019). Hence, viruses containing common markers of resistance to any FDA-approved antivirals (M2-V27A, NA-H275Y, PA-I38T) were detected at a low frequency (0.8%, 8 of 1015). Additionally, sequences were also analyzed for other amino acid substitutions previously reported to confer reduced inhibition by NA inhibitor(s) in NA inhibition (NI) assays or reduced baloxavir susceptibility in cell culture-based assays (WHO, 2023). Two viruses, from a chicken and a skunk, containing NA-I117T were detected in different states. In Florida, a virus collected from a dolphin had NA-S247N, while two viruses from vultures had NA-N295S (Table S1). We also flagged viruses that contained NA sequence changes with unknown effects on drug susceptibility because these occurred at residues previously implicated in reduced inhibition. For example, an NA-V149A induced 8-fold reduced inhibition by zanamivir in a clade 1 A(H5N1) virus (Naughtin et al., 2011). Therefore, we flagged viruses with V149F (n = 2) and V149I (n =8). Similarly, the following NA substitutions were flagged: Q136P (n = 1), D199E (n = 1), S247G (n = 1), and N295D (n = 1) (Table S1). For substitutions in PA, we flagged viruses with changes previously reported to confer reduced baloxavir susceptibility in seasonal influenza viruses: K34R (n = 1), A36V (n = 2), A37T (n = 4), I38M (n = 1), and I38V (n =  $\frac{1}{2}$ ) 3) (Table S1) (WHO, 2023).

To conduct phenotypic testing, we assembled a panel of clade 2.3.4.4b A(H5N1) viruses (n = 22) that were collected from different animal species and different genotypes (Elsmo et al., 2023), including viruses with flagged mutations that were available for testing (Table S2). A/American wigeon/South Carolina/2021, collected at the beginning of the US outbreak, was also included. Viruses were tested in a fluorescence-based NI assay (Okomo-Adhiambo et al., 2010) with four marketed NA inhibitors and AV5080, an oral NA inhibitor currently undergoing clinical trials (Ivachtchenko et al., 2014; Jones et al., 2023). The baseline susceptibility was determined by calculating the median  $IC_{50}$  (50% inhibitory concentration) of clade 2.3.4.4b viruses lacking any of the known or suspected NA markers of reduced inhibition (n = 15, Table 1). The median  $IC_{50}$ s were in a sub-nanomolar range for all NA inhibitors, except for oseltamivir, which was 13- to 27-fold less effective than other marketed NA inhibitors. Notably, AV5080 was the most potent at inhibiting the NA activity of clade 2.3.4.4b A(H5N1) viruses, especially compared to oseltamivir (67-fold). Next, we evaluated the effect of each NA mutation on drug susceptibility by comparing the  $IC_{50}$  to the determined baseline. Kode et al. (2019) reported that NA-I117T in a clade 2.2 A(H5N1) virus conferred 12- to 19-fold reduced inhibition by oseltamivir and zanamivir. However, we did not observe any effect on susceptibility to NA inhibitors, whether comparing to the baseline or IC<sub>50</sub> of NA sequence-matched wildtype virus (Table S2). This apparent discrepancy may point to a clade-specific effect or differences in calculating fold-change. NA-S247N reportedly reduced oseltamivir inhibition by 24-fold in a clade 2.3.4 virus (Boltz

et al., 2010), whereas in a clade 2.3.4.4b virus tested here, NA-S247N only conferred 4- to 6-fold reduction by oseltamivir and peramivir. NA-V149I was found in a clade 7.1 A(H5N1) virus but was not evaluated for its effect on drug susceptibility (Creanga et al., 2017). Here, NA-V149I did not alter susceptibility to NA inhibitors. A previously unreported NA-N295D mutation conferred 4-fold reduced inhibition by zanamivir and AV5080. In different clades of A (H5N1) viruses, NA-N295S was reported to confer 12- to 126-fold reduced inhibition by oseltamivir (Earhart et al., 2009; Le et al., 2005; McKimm-Breschkin et al., 2018; Sood et al., 2018). The two A(H5N1) viruses collected from vultures in Florida that had NA-N295S additionally contained a threonine-to-asparagine substitution at residue 438 (Table 1), which is highly conserved among influenza A viruses (Yasuhara et al., 2022). Notably, this dual N295S+T438N mutation was detected in the original specimens and their respective isolates. However, sequence analysis showed the presence of mixed virus populations at both residues in one of the bird viruses, A/black vulture/Florida/ 22-012333-001/2022 (Table 1). This isolate was subjected to conventional limiting dilution procedure in MDCK-SIAT1 cells to separate NA variant, yielding three distinct virus clones containing N295S, T438N, or both N295S+T438N (Table 1). By itself, NA-N295S conferred 14-fold reduced inhibition by oseltamivir and 3- to 7-fold reduced inhibition by the other NA inhibitors, whereas NA-T438N conferred ~12-fold reduced inhibition by zanamivir only. The effect of the dual NA mutation N295S+T438N was synergistic, causing 16- to 76-fold reduced inhibition by all NA inhibitors; this is similar to the inhibition profile of the virus with no detected mixture (Table 1).

The A(H5N1) viruses were then tested for their susceptibility to the CEN inhibitors baloxavir (acid) and AV5116 (Ivashchenko et al., 2021) using cell culture-based assays HINT (high-content imaging neutralization test) (Gubareva et al., 2019) and IRINA (influenza replication inhibition neuraminidase-based assay) (Patel et al., 2022). To minimize the effect of different growth kinetics between viruses, the time of virus replication was limited to 8–10 h (Supp. Fig. 1). The baseline susceptibilities to CEN inhibitors were determined by calculating the median  $EC_{50}$  (50% effective concentration) for viruses (n = 17) that lack flagged PA mutations (Table 2). The median  $EC_{50}$ s fell into a sub-nanomolar range for both baloxavir and AV5116. Overall, clade 2.3.4.4b A (H5N1) viruses seem to be slightly (1.2- to 5-fold) more susceptible to these antivirals than the reference human influenza A viruses. EC<sub>50</sub>s determined by HINT tended to be higher compared to those determined by IRINA, but this difference was small (~2-fold) (Table 2 and S2). The clade 2.3.4.4b A(H5N1) virus with PA-I38T displayed 75- to 108-fold reduced susceptibility to baloxavir and 37- to 78-fold reduced susceptibility to AV5116. These results were similar to those obtained for the reference baloxavir-resistant A(H1N1)pdm09 virus with PA-I38T (Table 2). Expectedly, the effect of PA-I38M was less pronounced. This change caused 6to 10-fold reduction in baloxavir and AV5116 susceptibility, which is similar to the human A(H3N2) virus with PA-I38M. Two available viruses with PA-A37T showed 5- to 6-fold reduced susceptibility to the CEN inhibitors.

Sequence-based analysis provides a high-throughput means to monitor susceptibility of emerging and circulating viruses and is the primary tool in virological surveillance. This approach is especially valuable for HPAI viruses as sequencing does not require enhanced biosafety containment and can be carried out on original specimens using complete

genome PCR primers universal to all influenza A virus subtypes. However, sequence-based assessment has its limitations, particularly with respect to susceptibility to newer antivirals with limited or no phenotypic data. It is also well-recognized that effects of amino acid substitutions can be subtype- and clade-specific. Thus, phenotypic testing serves two main purposes: to establish baseline susceptibility for newly emerged viruses and to assess the effect of identified mutations on drug phenotype. We and others (Kandeil et al., 2023) showed that clade 2.3.4.4b viruses collected in the US in 2021–2022 were susceptible to NA and CEN inhibitors. Our study demonstrated that the oseltamivir susceptibility baseline for clade 2.3.4.4b viruses was somewhat higher compared to human A(H1N1) pdm09 viruses. Similar observations were made in the past for certain genetic clades of A(H5N1) viruses (Creanga et al., 2017; Le et al., 2008; McKimm-Breschkin et al., 2007, 2018; Nguyen et al., 2013). Here, we also report the detection of sporadic mutations in clade 2.3.4.4b viruses that may confer resistance or reduced susceptibility to FDA-approved antivirals. The four identified oseltamivir-resistant viruses with NA-H275Y were not available for phenotypic testing, however, they are expected to display highly reduced inhibition in NI assays (>100-fold) (Lee and Hurt, 2018). Notably, two viruses with the dual NA mutation N295S+T438N displayed oseltamivir IC50s similar to that of the human A(H1N1)pdm09 virus with NA-H275Y. Not only did the dual mutation confer reduced inhibition by all NA inhibitors, but it also appeared to have lowered the NA activity of these viruses (Table S4) which may reduce virus fitness in mucin-rich environment, such as the upper respiratory tract of humans. Because these dual-mutant viruses were collected from vultures on the same day in Florida, it is possible that they acquired these drug-resistant viruses from the same source (e.g., consuming the same carcass of an infected animal). However, bird-to-bird transmission also cannot be ruled out. Notably, limited transmission of oseltamivir-resistant viruses with NA-H275Y was recently reported in birds (Skog et al., 2023). Clusters of drugresistant viruses can serve as indicators of local transmission. The four oseltamivir-resistant viruses with NA-H275Y identified in this study did not form a cluster as they were collected in different places and at different times. The detection of a cluster of adamantane-resistant viruses, however, is not surprising as such viruses are known to transmit well (Gubareva and Hayden, 2006). We also detected a small cluster of three viruses with PA-A37T that displayed reduced baloxavir susceptibility. To the best of our knowledge, this is the first report of a potential local transmission of such viruses among domestic poultry. To improve risk assessment, studies are needed to evaluate the replicative fitness of A(H5N1) viruses with PA-A37T and other PA mutations affecting susceptibility to CEN inhibitors. Compared to the currently marketed antivirals, we also showed that the investigational drugs AV5080 and AV5116 demonstrated better inhibitory activity against viruses bearing markers for drug resistance. Relative to their analogue counterparts, these drugs contain functional moiety modifications which may contribute to their improved inhibitory activities (Ivachtchenko et al., 2014; Ivashchenko et al., 2021). Such results indicate their potential to expand our current arsenal of influenza antivirals.

The current threshold (3-fold) for baloxavir susceptibility is arbitrary and more data are needed to improve laboratory data interpretation. A new cell culture-based assay, IRINA, provides improved throughput and its robustness was evident from the ability to test viruses displaying different growth kinetics. As clade 2.3.4.4b A(H5N1) viruses continue

to circulate and evolve, close monitoring of their drug susceptibility is needed to inform clinical management in the event of human infections and to make informed decisions on the national stockpiling of antiviral medications.

#### Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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#### Data availability

Data will be made available on request.

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## Table 1

Neuraminidase inhibitor susceptibility of NGS-flagged clade 2.3.4.4b HPAI A(H5N1) viruses assessed in a neuraminidase inhibition assay.

Influenza A virus	Amino acid change in NA head <sup>a</sup>		Mean IC <sub>50</sub>	± SD, nM (fold-chan	ge)b	
		Oseltamivir	Zanamivir	Peramivir	Laninamivir	AV5080
Median IC <sub>50</sub> A(H5N1) <sup>c</sup>		2.66	0.21	0.10	0.18	0.04
chicken/ID/22	1117T	$3.00 \pm 0.37$ (1)	$0.29 \pm 0.02$ (1)	$0.08 \pm 0.02$ (1)	$0.16 \pm 0.01$ (1)	$0.05 \pm 0.01$ (1)
skunk/WA/22	I117T	$2.80 \pm 0.09$ (1)	$0.25 \pm 0.02$ (1)	$0.09 \pm 0.00$ (1)	$0.15\pm 0.00~(1)$	$0.04 \pm 0.00 \ (1)$
fox/WI-30/22	V149I, N355S	$1.30\pm0.35\;(0.5)$	$0.21 \pm 0.01$ (1)	$0.09 \pm 0.00$ (1)	$0.16\pm 0.02~(1)$	$0.05 \pm 0.00 \ (1)$
dolphin/FL/22	S247N	$15.49 \pm 0.45$ (6)	$0.32 \pm 0.02$ (1)	$0.38 \pm 0.03$ (4)	$0.30 \pm 0.01$ (2)	$0.09 \pm 0.01$ (2)
vulture/FL-58/22	N295D	$3.56 \pm 0.77$ (1)	$0.84 \pm 0.10$ (4)	$0.20\pm0.05~(2)$	$0.39\pm0.08~(2)$	$0.15 \pm 0.02$ (4)
vulture/FL-31/22	N295S, T438N	196.54 ± 30.59 (74)	$18.01 \pm 4.52$ (86)	$9.03 \pm 2.22$ (90)	$3.36 \pm 1.06$ (19)	$2.19 \pm 0.11$ (55)
vulture/FL-33/22d	N/S295, T/N438	$11.05 \pm 1.60$ (4)	$4.20\pm 0.36\ (20)$	$0.55 \pm 0.05$ (6)	$0.64 \pm 0.10$ (4)	$0.14 \pm 0.02$ (4)
clone 1	N295S	37.11 ± 5.27 (14)	$0.85\pm 0.15~(4)$	$0.59 \pm 0.10$ (6)	$0.50\pm0.05~(3)$	$0.28 \pm 0.00$ (7)
clone 2	T438N	$4.24 \pm 0.26$ (2)	$2.55\pm 0.25~(12)$	$0.18\pm 0.02$ (2)	$0.34 \pm 0.04$ (2)	$0.06 \pm 0.01$ (2)
clone 3	<b>N295S</b> , T438N	$135.15\pm39.93~(51)$	$15.97 \pm 0.62$ (76)	$7.34 \pm 1.03$ (73)	$2.84\pm 0.28~(16)$	$2.03 \pm 0.45$ (51)
Reference <sup>e</sup>						
IL/45/19 A (H1N1)pdm09	I	$0.17 \pm 0.03$	$0.16\pm0.01$	$0.06 \pm 0.01$	$0.20 \pm 0.02$	$0.09\pm0.01$
AL/03/20 A (H1N1)pdm09	H275Y	229.67 ± 25.29 (1351)	$0.22 \pm 0.01$ (1)	15.25 ± 2.11 (254)	$0.43 \pm 0.04$ (2)	$0.76 \pm 0.07$ (8)

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and laninamivir were purchased from BioSynth, Berkshire, United Kingdom; AV5080 was provided by ChemDiv, San Diego, CA. Dash lines (-) indicate the absence of NA substitution. Antiviral testing Susceptibility of HPAI A(H5N1) viruses, grown in 10-day-old embryonated chicken eggs, was assessed using a fluorescence-based NI assay. NA inhibitors oseltamivir carboxylate, zanamivir, peramivir, was conducted in biosafety level 3 enhanced facility.

<sup>a</sup>Encompasses the head domain (amino acid residues 82–470) of NA (also see Supplemental Table S2 for complete virus list and information). NA amino acid substitutions previously associated with reduced drug susceptibility are shown in **bold**.

 $b_{\rm Each}$  virus was tested in 3 independent experiments to determine IC50 value (50% inhibitory concentration, nM); SD, standard deviation.

c(H5N1) viruses lacking the flagged NA substitutions (n = 15) were used to determine the median IC50 (the baseline susceptibility). A fold change in IC50s of flagged A (H5N1) viruses relative to the median IC50 is shown.

<sup>d</sup>/black vulture/Florida/22-012,333-001/2022 (vulture/FL-33/22) was subjected to limiting dilution procedure in MDCK-SIAT1 cells and resulting virus clones containing either a single or dual amino acid substitution at residues 295 and 438 were recovered and tested.

e From the CDC Neuraminidase Inhibitor Susceptibility Reference Virus Panel version 3.0 (International Reagent Resource, IRR: FR-1755). For reference viruses, IC50 fold change was determined by

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comparing to NA sequence-matched wildtype reference virus IC50.

#### Table 2

Susceptibility of NGS-flagged clade 2.3.4.4b HPAI A(H5N1) viruses to PA cap-dependent endonuclease inhibitors baloxavir and AV5116 in cell culture-based assays.

Influenza A Virus	Amino acid substitution in PA <sup>a</sup>	Mean EC <sub>50</sub> $\pm$ SD, nM (fold-change) <sup>b</sup>				
		Baloxavir <sup>c</sup>		AV5116		
		HINT	IRINA	HINT	IRINA	
Median EC <sub>50</sub> A(H5N1) <sup>d</sup>		0.57	0.34	0.52	0.25	
chicken/PA-6/22	A37T	3.21 ± 0.82 (6)	$2.06 \pm 0.59$ (6)	3.15 ± 0.28 (6)	$1.48 \pm 0.16$ (6)	
chicken/PA-10/22	A37T	$3.15 \pm 0.74$ (6)	$1.84 \pm 0.41$ (5)	$2.40 \pm 0.35 \ (5)$	$1.62 \pm 0.68$ (6)	
hawk/MN-1/22	I38M	$3.34 \pm 0.89$ (6)	$2.94 \pm 0.62 \ (9)$	$3.12 \pm 1.07$ (6)	$2.45 \pm 0.21 \; (10)$	
chicken/MI/22	<b>I38T</b> <sup>e</sup>	$42.51 \pm 9.40$ (75)	36.84 ± 10.21 (108)	$19.36 \pm 4.94$ (37)	19.53 ± 3.21 (78)	
Reference <sup>f</sup>						
IL/08/18 A (H1N1) pdm09	-	$1.65\pm0.44$	$1.12\pm0.32$	$1.89\pm0.55$	$1.35\pm0.48$	
IL/08/18 A (H1N1) pdm09	138T	$\underline{128.66 \pm 34.68\ (78)}$	$\underline{112.84 \pm 27.26\ (101)}$	<u>61.87 ± 18.2 (33)</u>	$\underline{52.59 \pm 10.33}~(39)$	
LA/50/17 A (H3N2)	-	$1.04\pm0.43$	$0.76\pm0.4$	$1.10\pm0.37$	$0.81\pm0.21$	
LA/49/17 A (H3N2)	I38M	10.58 ± 3.79 (10)	8.70 ± 3.09 (11)	12.21 ± 3.53 (11)	$10.32 \pm 1.94 \ (13)$	

Susceptibility to CEN inhibitors baloxavir acid (baloxavir) (Shionogi and Co., Ltd., Osaka, Japan) and AV5116 (ChemDiv, San Diego, CA) was determined in MDCK-SIAT1 cells using HINT (High-content Imaging Neutralization Test) and IRINA (Influenza Replication Inhibition Neuraminidase-based Assay). Dash lines (–) indicate the absence of PA substitution. Antiviral testing was conducted in biosafety level 3 enhanced facility.

 $a^{4}$ Encompasses the endonuclease active site (amino acid residues 1–200) of the PA protein (also see Supplemental Table S3 for complete virus list and information). PA amino acid substitutions previously associated with reduced drug susceptibility are shown in **bold**.

<sup>b</sup>Each virus was tested in 3 independent runs to determine EC50 value (50% effective concentration, nM); SD, standard deviation.

<sup>c</sup>Baloxavir acid, active metabolite form of the pro-drug baloxavir marboxil.

 $d_{A(H5N1)}$  viruses lacking the flagged PA substitutions (n = 17) were used to determine the median EC50 (baseline susceptibility). Fold-change in EC50s of flagged A(H5N1) virus relative to the median EC50 is shown.

 $e^{A/chicken/Michigan/22-013,961-001-original/2022}$  (chicken/MI/22) contains two additional amino acid substitutions M61I and A85T within the PA endonuclease active site.

<sup>*f*</sup> From the CDC Baloxavir Susceptibility Reference Virus Panel version 1.1 (IRR: FR-1678). For reference viruses, EC50 fold-change was determined by comparing to PA sequence-matched wildtype reference virus EC50. An arbitrary threshold (3-fold) is used to report PA amino acid substitutions that confer reduced susceptibility to baloxavir.