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Early priming with inactivated poliovirus vaccine (IPV) and intradermal fractional dose IPV administered by a microneedle device: A randomized controlled trial

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Abstract

Introduction: Inactivated poliovirus vaccine (IPV) introduction and phased oral poliovirus vaccine (OPV) cessation are essential for eradication of polio.

Methods: Healthy 6-week old infants in Bangladesh were randomized to one of five study arms: receipt of trivalent OPV (tOPV) or bivalent OPV (bOPV) at ages 6, 10 and 14 weeks, intramuscular IPV or intradermal one-fifth fractional dose IPV (f-IPV) at ages 6 and 14 weeks, or f-IPV at ages 6 and 14 weeks with bOPV at age 10 weeks (f-IPV/bOPV). All participants received tOPV at age 18 weeks.

Results: Of 975 infants randomized, 95% (922) completed follow-up. Type 1 seroconversion after 3 doses at 6, 10 and 14 weeks was higher with bOPV compared with tOPV (99% vs 94%, $p = 0.019$). Seroconversions to types 1 and 3 after 2 IPV doses at ages 6 and 14 weeks were no different than after 3 doses of tOPV or bOPV at ages 6, 10 and 14 weeks. A priming response, seroconversion 1 week after IPV at 14 weeks among those who did not seroconvert after IPV at 6 weeks, was observed against poliovirus types 1, 2 and 3 in 91%, 84% and 97%, respectively.

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Contributors

AA prepared the first draft of the manuscript and all authors reviewed and approved the manuscript. AA, CFE, HG, MAP, SW, MSO and WW contributed to the design of the study. The design team jointly developed the trial implementation strategy with KZ, SPL, JDH, MY and TBI.

WW and MSO contributed to laboratory testing. AA and HG contributed to data analysis. All authors contributed to interpretation of study results.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.vaccine.2015.09.039>.

Conflict of interest

All authors declare that they have no conflict of interest.

Compared with IPV, f-IPV failed non-inferiority tests for seroconversion with 1 or 2 doses and priming after 1 dose.

Discussion: The findings demonstrate considerable priming with IPV at age 6 weeks, comparable immunogenicity of tOPV and bOPV, and inferior immunogenicity of one-fifth f-IPV compared with IPV. If IPV induced priming at age 6 weeks is similar to that at age 14 weeks, IPV could be administered at a younger age and possibly with a higher coverage.

Keywords

Oral polio vaccine; Inactivated polio vaccine; Priming; Polio immunogenicity

1. Introduction

Oral poliovirus vaccines (OPV) consist of live attenuated poliovirus strains that can revert and cause paralysis, that is indistinguishable from paralysis caused by wild polioviruses (WPV), either due to vaccine-associated paralytic polio (VAPP) or circulating vaccine-derived polioviruses (cVDPV), in which the reverted vaccine virus also acquires the ability to circulate [1]. Since the last type 2 WPV (WPV2) was reported in 1999 in India [2] and about 87% of VDPVs during 2000–2013 were type 2 [3], the strategic advisory group of experts on immunization (SAGE) has recommended a phased cessation of OPV starting with type 2 OPV [4]. In countries using trivalent OPV (tOPV), a mixture of types 1, 2 and 3 OPV, in routine immunization (RI), SAGE has recommended a switch to bivalent OPV (bOPV), a mixture of OPV types 1 and 3 following introduction of 1 dose of inactivated poliovirus vaccine (IPV) generally at age 14 weeks [5]. It is expected that delaying IPV administration to age 14 weeks is likely to maximize IPV immunogenicity [5]; however, compared with vaccinating at age 6 weeks, vaccination at age 14 weeks is likely to be associated with lower vaccination coverage in some high-risk countries [6].

The principal objective of introducing IPV with bOPV is to mitigate the risk associated with increased susceptibility to WPV2 or cVDPV2. For IPV, priming is defined as a seroconversion response 1 week after a second dose of IPV among those who did not seroconvert after the first IPV dose. One clinical trial in Cuba reported considerable immunogenicity (seroconversion [63%] and priming [35%]) with 1 dose of IPV at age 4 months [7]. The absence of immunogenicity data by age, including priming response after IPV, is the chief limitation in assessing the optimal age for IPV administration in RI. In 2012, SAGE also recommended collecting additional immunogenicity data on intradermal (ID) one-fifth dose of IPV (0.1 ml fractional IPV [f-IPV]) as a potential substitute for intramuscular IPV (0.5 ml) [8].

2. Methods

2.1. Randomization and masking

We conducted an open-label 5-arm randomized controlled trial from 27 November 2012 to 30 November 2013 in Mirpur, an urban neighborhood in Dhaka, Bangladesh. The trial enrolled participants from 5 different sections of Mirpur. During the duration of the trial no polio vaccination campaigns were conducted in or around the study site. Infants were

assigned randomly to one of five arms using a block randomization scheme of 65 blocks with a block size of 18 and an allocation ratio of 4:4:3:3:4 (Fig. 1). The tOPV arm received tOPV at ages 6, 10 and 14 weeks; the bOPV arm received bOPV at ages 6, 10 and 14 weeks; the IPV arm received IPV at ages 6 and 14 weeks; the f-IPV arm received f-IPV at ages 6 and 14 weeks; and the f-IPV/bOPV arm received f-IPV at ages 6 and 14 weeks with bOPV at age 10 weeks. All participants received tOPV at age 18 weeks (Table 1 in Supplementary Appendix).

2.2. Study objectives

The study's three primary objectives were to compare immunogenicity of (1) f-IPV and bOPV with bOPV alone; (2) 3 doses of tOPV with 3 doses of bOPV; and (3) 2 doses of intramuscular IPV with 2 doses of f-IPV.

2.3. Study design and procedures

Infants were recruited at age 6–7 weeks (42–51 days), if the parents were willing to participate, comply with study procedures, and provide written informed consent. Exclusion criteria included (1) receipt of any polio vaccine before enrollment; (2) diagnosis or suspicion of immunodeficiency or a bleeding disorder; (3) known allergy to polio vaccines or constituents; (4) any acute illness such as vomiting, diarrhea or infection immediately before enrollment; and (5) an infant who was part of a multiple birth. Enrolled participants were withdrawn from the study if requested by their parents or if they received polio vaccine outside of the study.

Study physicians administered all study vaccines and routine non-polio vaccines for infants as recommended by the Bangladesh Ministry of Health and Family Welfare. Intramuscular IPV (0.5 ml) was administered using a standard needle and syringe. Intradermal f-IPV (0.1 ml) was administered using NanoPass MicronJet 600 (MJ600), a microneedle device with three microneedles (0.6 mm in length) that attaches to an intradermal syringe. Multiple clinical trials have been conducted using MJ600 [9–12]. IPV and f-IPV were administered in the anterolateral thigh, opposite the side used for routine immunization of injectable vaccines.

Blood samples (1 ml) were obtained by venipuncture at ages 6, 14, and 18 weeks from all participants and at age 15 weeks from participants assigned to IPV or f-IPV arms before administering any scheduled study vaccine. Sera were stored at -20°C and tested for antibodies to poliovirus types 1, 2 and 3 at the Centers for Disease Control and Prevention (CDC), Atlanta, USA using microneutralization assay. Titers below a dilution of 1:8 were considered negative for presence of poliovirus antibodies and the highest measurable titer was 1:1448. Parents were asked to collect a stool specimen (8 g) from participants 1 week after tOPV administration at age 18 weeks. Stool specimens were stored at -20°C and tested at CDC for presence of poliovirus by type [13].

2.4. Analysis

No published studies were found to have administered f-IPV with bOPV, or 3 doses of bOPV. Therefore, for sample size calculations based on limited evidence, we assumed

seroconversions of 85% for types 1 and 3 with f-IPV and bOPV and 95% with 3 doses of bOPV [14,15]. For tOPV, we assumed sero-conversions of 75% for type 1 and 65% for type 3 [16]. Therefore, a sample size of 207 per arm would be sufficient to obtain a power of 90% with two-sided α of 0.05 to detect a difference in seroconversion of at least 10% when comparing 3 doses of bOPV with 3 doses of tOPV, and 2 doses of f-IPV and 1 dose of bOPV with 3 doses of bOPV. No published studies were found to have reported immunogenicity of IPV or f-IPV with two doses 8 weeks apart at ages 6 and 14 weeks. For a non-inferiority comparison, we assumed a sero-conversion of 90% with both IPV and f-IPV with a non-inferiority margin of 10% [14,17]. For this comparison a sample size of 155 per arm is required for a power of 90% with one-sided α of 0.05. Hence, the effective sample size for the trial was 931, with an enrollment target of 1170 assuming 20% attrition (Table 1 in Supplementary Appendix).

Seroconversion was defined as either conversion from seronegative to seropositive or a four-fold increase in antibody titers between two specimens after adjusting for decay of maternal antibodies. The half-life of maternal antibodies was assumed to be 28 days [14,18]. The primary analytical approach was intent-to-treat for participants with serological results. The primary end-point was seroconversion at age 18 weeks. To compare immunogenicity across study arms, the proportion of participants who sero-converted were compared using Fisher's exact test (two-tailed). Priming was defined as a seroconversion response at age 15 weeks after receipt of the second IPV/f-IPV dose among those who did not seroconvert by age 14 weeks after one IPV/f-IPV dose at age 6 weeks. Reverse cumulative distribution curves, which are constructed by representing on the vertical axis the percent of subjects with antibody titers equal to or greater than that marked in x -axis, were used to compare distribution of antibody titers by study arms [19].

2.5. Study oversight

The study protocol was reviewed by icddr,b's Institutional Review Board (IRB). The study was conducted in compliance with good clinical practice guidelines. UNICEF assisted in the procurement of vaccines used in this study. OPV was manufactured by Sanofi Pasteur and IPV was manufactured by the Netherlands Vaccine Institute (NVI). NanoPass Technologies Ltd. donated the supplies of MJ600. UNICEF, Sanofi Pasteur, NanoPass, and NVI had no role in the study design, implementation, data analysis, or interpretation of study results. The study was registered with [Clinicaltrials.gov](https://clinicaltrials.gov/ct2/show/study/NCT01813604) (NCT01813604). Adverse events data were reviewed by the Data Safety Monitoring Board (DSMB) of icddr,b.

2.6. Role of funding source

The study was funded by the Global Immunization Division of the Centers for Disease Control and Prevention. CDC staff participated in the study design, sample testing, data analysis and decision to submit for publication.

3. Results

3.1. Baseline characteristics

The study enrolled and randomized 975 participants and of these, 922 (95%) with blood specimens available at ages 6 and 18 weeks were included in the primary end-point analysis (Fig. 1). Enrollment was stopped after enrolling 975 participants as the study had achieved its effective sample size due to lower than anticipated study attrition. No statistically significant differences were observed at baseline among participants who completed the study compared with those who did not (data not shown) except that median type 2 antibody titers at baseline were lower for those who completed the study (1:28 vs 1:41, Kruskal–Wallis = 0.036). No other significant differences in baseline characteristics, including seroprevalence to polioviruses, were observed among study arms (Table 1).

3.2. Humoral immunogenicity

The median bleb diameter after intradermal injection with MJ600 was 10 mm and 99% of the participants had no residual liquid present on the skin following the injection.

Seroconversion to poliovirus type 1 (PV1) after 2 and 3 doses was higher in the bOPV arm compared with the tOPV arm (2 doses: 93% vs 87%, $p = 0.047$; 3 doses: 99% vs 94%, $p = 0.019$; Table 2). PV1 sero-conversion with 2 doses of IPV (95%) was statistically no different from that observed with 3 doses of tOPV or bOPV. PV1 seroconversion with 2 doses of f-IPV and 1 dose of bOPV was higher than that observed with 2 doses of f-IPV alone ($p = 0.005$) and no different from that with 3 doses of tOPV or bOPV.

Seroconversion at 18 weeks to PV2 was higher with 3 doses of tOPV compared with 2 doses of IPV ($p = 0.002$) or f-IPV in either f-IPV arms ($p < 0.001$). Seroconversion to PV3 was statistically no different with 3 doses of tOPV (95%) compared with 3 doses of bOPV (94%), 2 doses of IPV (97%) or f-IPV (89%), or 2 doses of f-IPV with 1 dose of bOPV (94%).

Compared with IPV, f-IPV failed the non-inferiority test for all serotypes for seroconversion observed with 1 or 2 doses (Fig. 2). Additionally, compared with IPV, f-IPV failed the non-inferiority test for all serotypes for priming response observed at 15 weeks.

Reverse cumulative distribution curves for antibody titers by study arm at age 18 weeks show that the highest titers were reached for PV1 in the bOPV arm, PV2 in the tOPV arm and PV3 in the IPV arm (Fig. 3). f-IPV was associated with the lowest titers for all three poliovirus types among those receiving type specific vaccines. One dose of IPV or f-IPV was not associated with a substantial change in distribution of antibody titers, despite the high degree of priming with 1 dose; however, within a week of the second dose of IPV or f-IPV, a rapid rise in antibody titers was observed (Fig. 1 in Supplementary Appendix).

3.3. Intestinal mucosal immunity

One week after receiving tOPV at age 18 weeks, 15%, 6%, and 8% of participants in the tOPV arm were excreting PV 1, 2, and 3, respectively (Table 2). Among participants in the bOPV arm, 61% were excreting PV2 1 week after receiving tOPV. The percent of

participants excreting type 1 poliovirus was statistically lower in the bOPV arm compared with the f-IPV/bOPV arm (4% vs 13%, Fischer's exact = 0.001). The percent excreting PV3 was statistically lower in the bOPV arm compared with f-IPV/bOPV arm (6% vs 14%, Fischer's exact = 0.013). No statistically significant differences in percent excreting polioviruses by type were observed between IPV and f-IPV arms.

3.4. Adverse events

No adverse events (AE) were reported among participants 30 min after receiving the study vaccine. During follow-up (age 6–19 weeks), 68 AE were reported among participants; 11 were considered serious AE (SAE), including hospitalization or death (Table 2 in Supplementary Appendix). Three infants died during follow-up: two in the sequential f-IPV/bOPV arm and one in the f-IPV arm. No AE/SAE were attributed to trial vaccines or MJ600 by the DSMB.

4. Discussion

The study demonstrated that considerable priming can be achieved with 1 dose of IPV at age 6 weeks. Cumulatively, 90% of children had either seroconverted or were primed against type 2 poliovirus with 1 dose of IPV at age 6 weeks. These results are particularly relevant for current policy considerations regarding global polio eradication. In November 2013, SAGE recommended introduction of at least 1 dose of IPV at age 14 weeks in RI in countries where IPV has not been introduced, in advance of a global implementation of the switch from tOPV to bOPV. With removal of type 2 OPV, the objective of IPV introduction is to maximize type 2 population immunity, which is a product of IPV immunogenicity and coverage. If the considerable priming noted in this study at age 6 weeks is similar to the priming noted at age 14 weeks, IPV vaccination at age 6 weeks will likely lead to higher population immunity compared with vaccination at age 14 weeks as vaccination coverage in many high-risk countries is higher at age 6 weeks compared with age 14 weeks [6].

The study confirms that bOPV is more immunogenic than tOPV for poliovirus types 1 and 3 [15]; however, after 3 doses, the differences in seroconversion are small and high titers of antibodies were observed after administration of both vaccines. Prior field assessments of tOPV have reported substantially lower effectiveness though those estimates have been based on parental report of the number of vaccine doses received [20,21]. This study demonstrates a high immunogenicity of tOPV in a developing country with a tropical climate [22–24].

IPV demonstrated a higher immunogenicity compared with f-IPV for priming with one dose and seroconversion with one or two doses. These results address a prior identified information need by SAGE to collect more evidence on the comparative immunogenicity of f-IPV and IPV [8]. Also these results are consistent with other studies that have reported lower immunogenicity of a one-fifth IPV dose compared with IPV [7,14,25]. The findings of this study confirm the safety of NanoPass MJ-600 in intradermal f-IPV administration, a device that had not been previously used for f-IPV administration.

The stool excretion results demonstrate a minimal reduction in type 2 excretion with IPV and f-IPV recipients compared with bOPV recipients, who did not receive any type 2 vaccine. Also a vaccination schedule of f-IPV/bOPV reduced the percent of participants who excreted type 1 or 3 polioviruses 1 week after receiving tOPV compared to the use of IPV or f-IPV alone. Although the percent excreting poliovirus in the f-IPV/bOPV arm was significantly higher than those in the bOPV arm, the absolute difference was not large. A prior study with tOPV demonstrated the substantial reduction in excretion of polioviruses with 1–2 doses of tOPV with minimal reduction with additional doses [26]. These findings taken together with noteworthy priming associated with IPV at age 6 weeks support evaluating polio vaccination schedules with IPV only as the first poliovirus vaccine followed by OPV.

This study has notable limitations. First, transmission of OPV received by other children in the community was observed. However, the effect of community transmission was low with only 14% type 2 seroconversion over 12 weeks in the bOPV arm [23,27]. Second, in the assessment of priming, the primary as well as secondary (challenge at 14 weeks) vaccines had different routes of administration and dosage between IPV and f-IPV arms, which limits comparison. Lastly, assessment of MJ600 performance was limited to safety and injection quality associated with the device and we could not compare immunogenicity of IPV administered by MJ600 with standard needle and syringe for intradermal administration.

Overall, findings from this study address several previously identified information gaps with regard to primary routine polio vaccine performance and could help simplify and expand polio vaccination policy options. The study supports the safety and comparable immunogenicity of tOPV and bOPV for types 1 and 3 poliovirus and demonstrates the lack of non-inferiority of one-fifth f-IPV to IPV. Most importantly, the study shows the promising degree of priming with an early (6 week) dose of IPV. A useful next step would be to compare priming at age 6 weeks to that with the SAGE-recommended IPV schedule at age 14 weeks.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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References

- [1]. Sutter RW, Kew OM, Cochi SL, Aylward RB. Poliovirus vaccine-live. In: Plotkin SA, Orenstein WA, Offit PA, editors. *Vaccines*. 2012. p. 1576.

- [2]. Centers for Disease Control Prevention. Apparent global interruption of wild poliovirus type 2 transmission. MMWR Morb Mortal Wkl Rep 2001;50(March 12):222–4.
- [3]. The Global Polio Eradication Initiative. Circulating vaccine-derived poliovirus; 2014. Available from: <http://www.polioeradication.org/Dataandmonitoring/Poliothisweek/Circulatingvaccinederivedpoliovirus.aspx>.
- [4]. World Health Organization. Meeting of the strategic advisory group of experts on immunization april 2013 – conclusions and recommendations. Wkly Epidemiol Rec 2013;88:201–16. [PubMed: 23696983]
- [5]. World Health Organization. Meeting of the strategic advisory group of experts on immunization october 2014 – conclusions and recommendations. Wkly Epidemiol Rec 2014;89(50):561–76. [PubMed: 25513671]
- [6]. Anand A, Pallansch MA, Estivariz CF, Gary H, Wassilak SG. Estimating the likely coverage of inactivated poliovirus vaccine in routine immunization: evidence from demographic and health surveys. J Infect Dis 2014;210(November (Suppl. 1)):S465–74. [PubMed: 25316869]
- [7]. Resik S, Tejeda A, Sutter RW, Diaz M, Sarmiento L, Alemani N, et al. Priming after a fractional dose of inactivated poliovirus vaccine. N. Engl J Med 2013;368(January (5)):416–24. [PubMed: 23363495]
- [8]. World Health Organization. Meeting of the strategic advisory group of experts on immunization, april 2012 – conclusions and recommendations, The Weekly Epidemiological Record; 2012. p. 201–16. Geneva, Switzerland.
- [9]. Van Damme P, Oosterhuis-Kafeja F, Van der Wielen M, Almagor Y, Sharon O, Levin Y. Safety and efficacy of a novel microneedle device for dose sparing intradermal influenza vaccination in healthy adults. Vaccine 2009;27(January (3)):454–9. [PubMed: 19022318]
- [10]. Hung IF, Levin Y, To KK. Quantitative and qualitative analysis of antibody response after dose sparing intradermal 2009 H1N1 vaccination. Vaccine 2012;30(April (17)):2707–8. [PubMed: 22210225]
- [11]. Hung IF, Levin Y, To KK, Chan KH, Zhang AJ, Li P, et al. Dose sparing intradermal trivalent influenza (2010/2011) vaccination overcomes reduced immunogenicity of the 2009 H1N1 strain. Vaccine 2012;30(October (45)):6427–35. [PubMed: 22910287]
- [12]. Levin Y, Kochba E, Kenney R. Clinical evaluation of a novel microneedle device for intradermal delivery of an influenza vaccine: are all delivery methods the same? Vaccine 2014;32(July (34)):4249–52. [PubMed: 24930715]
- [13]. World Health Organization. Polio laboratory manual; 2004. Geneva, Switzerland.
- [14]. Resik S, Tejeda A, Lago PM, Diaz M, Carmenates A, Sarmiento L, et al. Randomized controlled clinical trial of fractional doses of inactivated poliovirus vaccine administered intradermally by needle-free device in Cuba. J Infect Dis 2010;201(May (9)):1344–52. [PubMed: 20350164]
- [15]. Sutter RW, John TJ, Jain H, Agarkhedkar S, Ramanan PV, Verma H, et al. Immunogenicity of bivalent types 1 and 3 oral poliovirus vaccine: a randomised, double-blind, controlled trial. Lancet 2010;376(November (9753)):1682–8. [PubMed: 20980048]
- [16]. Zaman K, Sack DA, Yunus M, Arifeen SE, Podder G, Azim T, et al. Successful co-administration of a human rotavirus and oral poliovirus vaccines in Bangladeshi infants in a 2-dose schedule at 12 and 16 weeks of age. Vaccine 2009;27(February (9)):1333–9. [PubMed: 19162114]
- [17]. The Cuba IPV Study Collaborative Group. Randomized, placebo-controlled trial of inactivated poliovirus vaccine in Cuba. N Engl J Med 2007;356(April (15)):1536–44. [PubMed: 17429085]
- [18]. Albrecht P, Enterline JC, Boone EJ, Klutch MJ. Poliovirus and polio antibody assay in HEp-2 and Vero cell cultures. J Biol Stand 1983;11(April (2)):91–7. [PubMed: 6306012]
- [19]. Reed GF, Meade BD, Steinhoff MC. The reverse cumulative distribution plot: a graphic method for exploratory analysis of antibody data. Pediatrics 1995;96(September (3 Pt 2)):600–3. [PubMed: 7659485]
- [20]. Grassly NC, Wenger J, Durrani S, Bahl S, Deshpande JM, Sutter RW, et al. Protective efficacy of a monovalent oral type 1 poliovirus vaccine: a case-control study. Lancet 2007;369(April (9570)):1356–62. [PubMed: 17448821]
- [21]. O'Reilly KM, Durry E, ul Islam O, Quddus A, Abid N, Mir TP, et al. The effect of mass immunisation campaigns and new oral poliovirus vaccines on the incidence of poliomyelitis

- in Pakistan and Afghanistan, 2001–11: a retrospective analysis. *Lancet* 2012;380(August (9840)):491–8. [PubMed: 22766207]
- [22]. World Health Organization Collaborative Study Group. Factors affecting the immunogenicity of oral poliovirus vaccine: a prospective evaluation in Brazil and the Gambia. World Health Organization Collaborative Study Group on Oral Poliovirus Vaccine. *J Infect Dis* 1995;171(May 5):1097–106.
- [23]. WHO Collaborative Study Group. Combined immunization of infants with oral and inactivated poliovirus vaccines: results of a randomized trial in the Gambia Oman, and Thailand. *Bull World Health Org* 1996;74(3):253–68. [PubMed: 8789924]
- [24]. Patriarca PA, Wright PF, John TJ. Factors affecting the immunogenicity of oral poliovirus vaccine in developing countries: review. *Rev Infect Dis* 1991;13(September–October (5)):926–39. [PubMed: 1660184]
- [25]. Mohammed AJ, AlAwaidy S, Bawikar S, Kurup PJ, Elamir E, Shaban MM, et al. Fractional doses of inactivated poliovirus vaccine in Oman. *N Engl J Med* 2010;362(June (25)):2351–9. [PubMed: 20573923]
- [26]. Modlin JF, Halsey NA, Thoms ML, Meschievitz CK, Patriarca PA. Humoral and mucosal immunity in infants induced by three sequential inactivated poliovirus vaccine-live attenuated oral poliovirus vaccine immunization schedules. Baltimore Area Polio Vaccine Study Group. *J Infect Dis* 1997;175(February (Suppl. 1)):S228–34. [PubMed: 9203721]
- [27]. Fine PE, Carneiro IA. Transmissibility and persistence of oral polio vaccine viruses: implications for the global poliomyelitis eradication initiative. *Am J Epidemiol* 1999;150(November (10)):1001–21. [PubMed: 10568615]

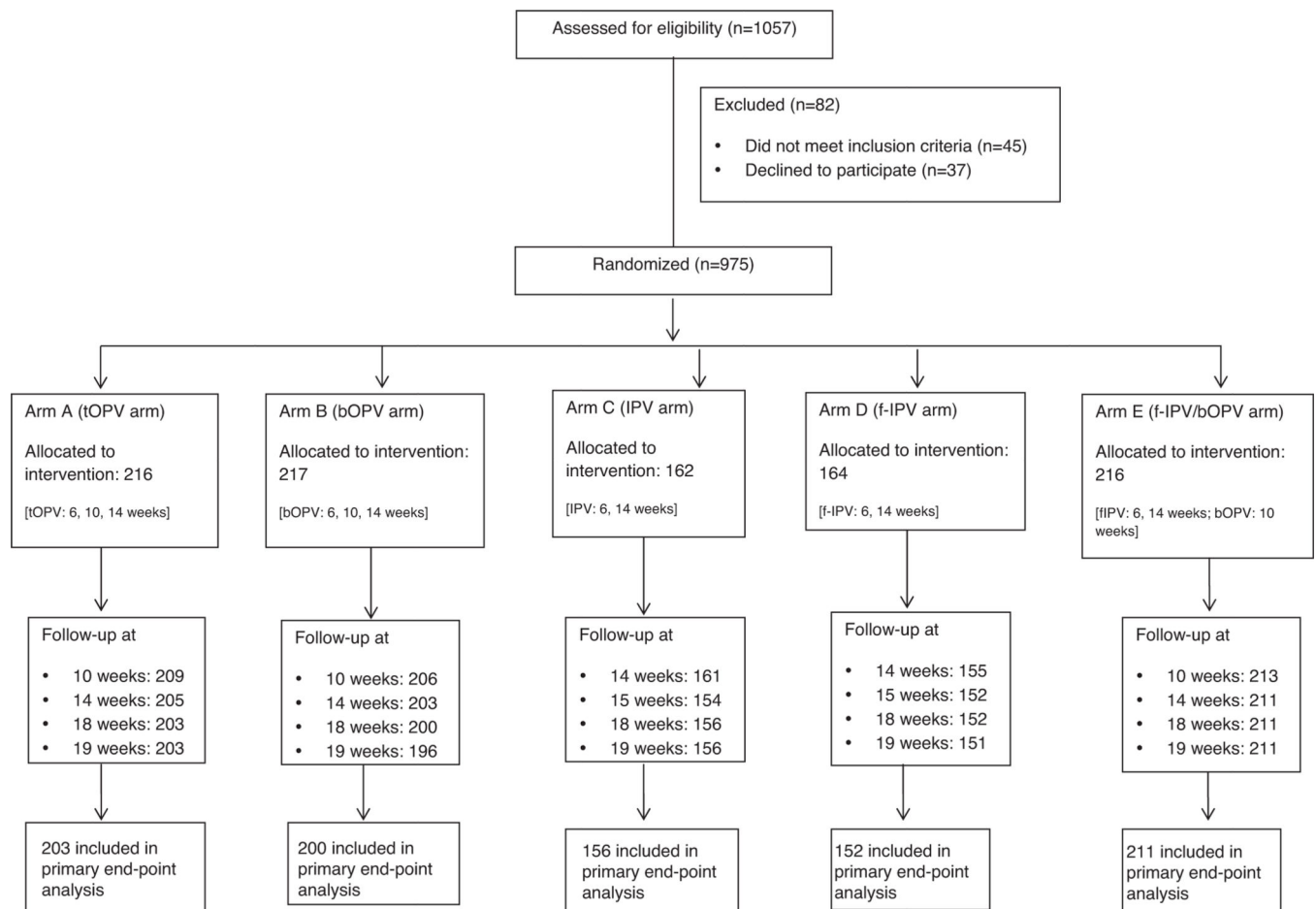
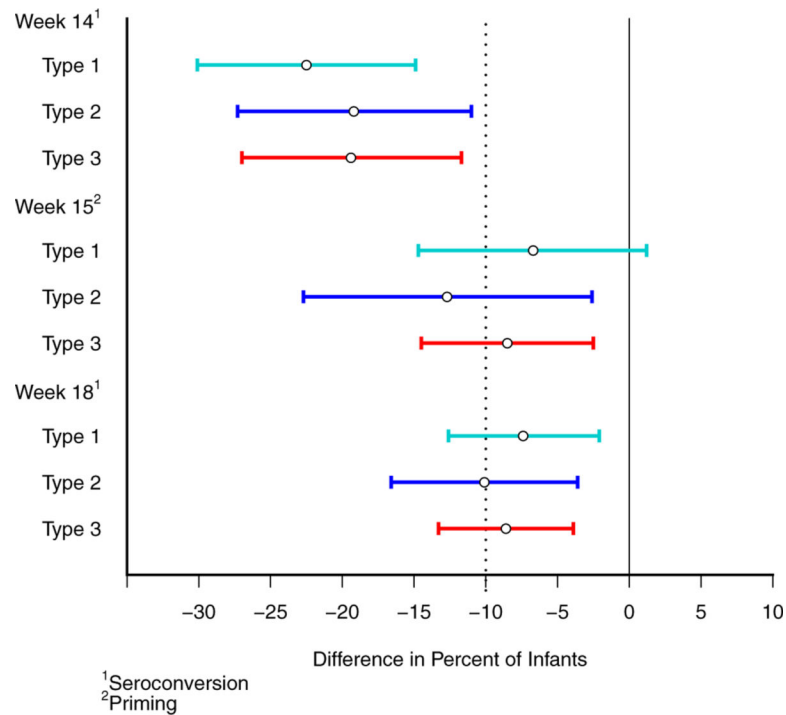


Fig. 1.
Trial profile with number of subjects followed by study time-point.

**Fig. 2.**

Differences in seroconversion and priming between fractional intradermal inactivated poliovirus vaccine (f-IPV) arm and intramuscular IPV arm by poliovirus type. f-IPV fails to pass the test of non-inferiority if the lower limit of the 90% confidence interval crosses -10%.

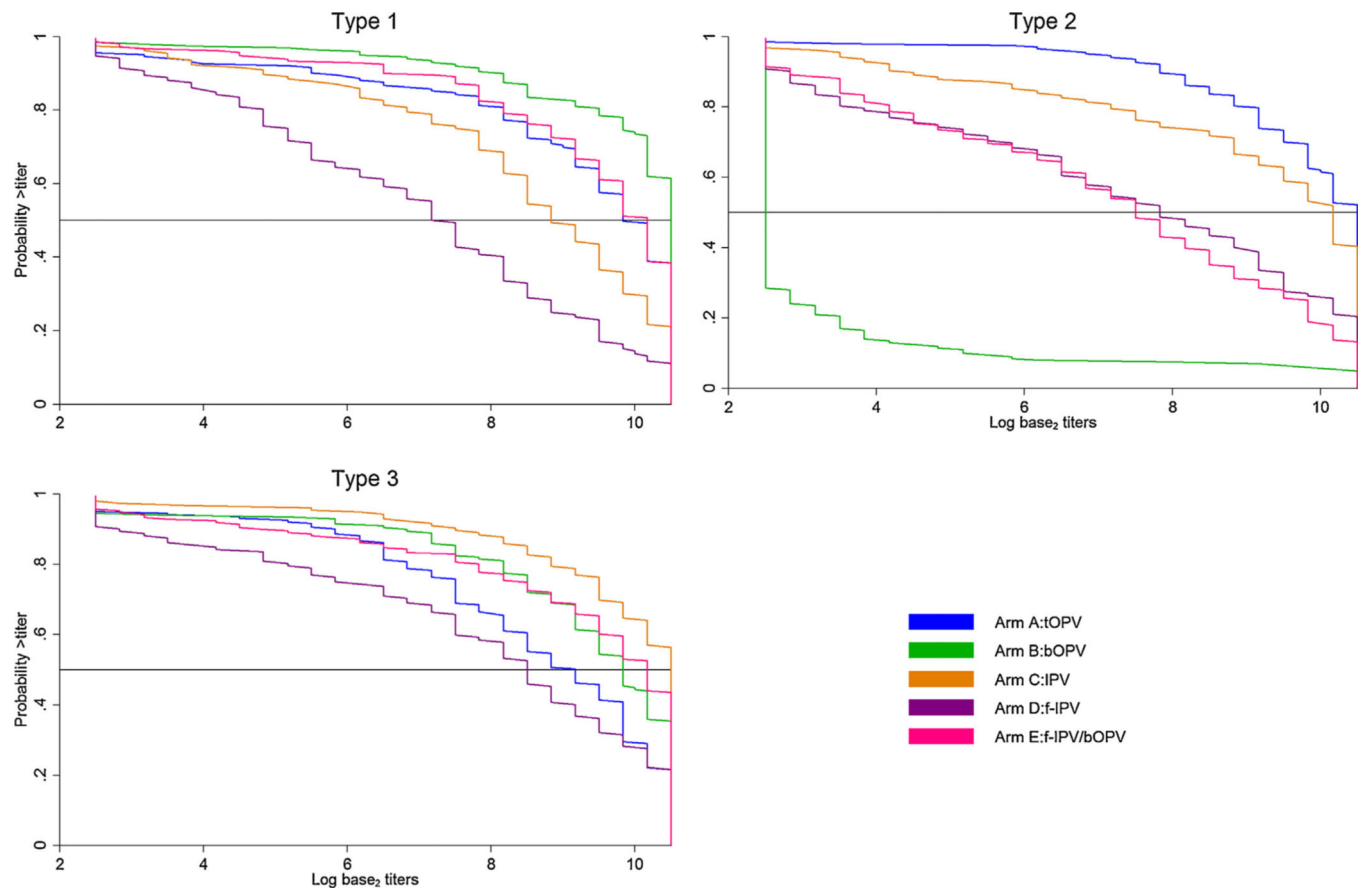


Fig. 3.
Reverse cumulative antibody titers at 18 weeks of age by study arm.

Table 1

Baseline characteristics among those who completed the study by study arms.

Baseline characteristics	A tOPV (<i>n</i> = 203)	B bOPV (<i>n</i> = 200)	C IPV (<i>n</i> = 156)	D f-IPV (<i>n</i> = 152)	E f-IPV/bOPV (<i>n</i> = 211)	<i>p</i> -value ^a
Median age in days (range)	44 (41,53)	44 (41,53)	44 (42, 53)	44(41,52)	44 (41,53)	0.662
Male <i>n</i> (%)	95	46.8%	49.5%	50.6%	52.0%	52.1% 0.828
Mother's education <i>n</i> (%)						
No formal school	36	17.7%	13.5%	18.6%	15.8%	19.0%
Primary school	88	43.4%	46.0%	43.6%	46.1%	41.2% 0.869
Middle school	36	17.7%	21.0%	19.2%	16.5%	23.2%
High school	33	16.3%	17.0%	17.3%	18.4%	15.2%
Graduate	10	4.9%	2.5%	1.3%	3.3%	1.4%
Type 1 serop revalence						
<i>n</i> (%)	99	48.8%	48.5%	50.0%	50.7%	47.4% 0.976
Median (range) ^b	28 (9,1448)	18(9,1448)	23(9, 1448)	23(6,1448)	23 (9, 724)	0.947
Type 2 serop revalence						
<i>n</i> (%)	118	58.1%	59.0%	60.9%	54.0%	58.8% 0.796
Median (range) ^b	28 (9,1448)	36 (9,1448)	28 (9, 1448)	28(9,1448)	23 (9,1448)	0.884
Type 3 serop revalence						
<i>n</i> (%)	51	25.1%	28.0%	27.6%	25.0%	30.8% 0.698
Median (range) ^b	23 (9,1448)	23 (9,1024)	23(9, 1448)	18(9,362)	18(9,1448)	0.643
Wasting present <i>n</i> (%)	31	15.3%	19.0%	18.0%	21.7%	16.1% 0.545
Stunting present <i>n</i> (%)	40	19.7%	18.5%	18.0%	20.4%	23.7% 0.661
Exclusive breastfeeding <i>n</i> (%)	180	88.7%	85.0%	85.9%	85.5%	82.9% 0.465

tOPV, trivalent oral poliovirus vaccine (OPV); bOPV, bivalent OPV; IPV, inactivated poliovirus vaccine; f-IPV, fractional IPV.

^aFisher's exact test. Kruskal-Wallis test used for mother's education and rank test for medians.^bAmong those with titers ≥ 8.

Table 2

Humoral and intestinal immunogenicity by study arm.

	A tOPV	B bOPV	C tPV*	D t-IPV*	E t-IPV/ bOPV	Fisher's exact test (a priori)	Fisher's exact test (post hoc)**					
Type 1												
Seroconversion by 14 weeks: <i>n</i> (%)	178/205	86.8% ^{b,e}	189/203	93.1% ^{a,b,f}	57/161	35.4% ^{e,f}	20/155	12.9% ^{c,d,f}	173/211	82.0% ^{a,d}	^a <i>p</i> = 0.001 B vs E; ^b <i>p</i> = 0.047 A vs B; ^c <i>p</i> <0.001 A vs D; ^d <i>p</i> <0.001 D vs E; NS: A vs E	^e <i>p</i> <0.001 A vs C; ^f <i>p</i> <0.001 B vs C
Priming response by 15 weeks: <i>n</i> (%) [†]	-	-	-	-	78/86	90.7%	91/109	83.5%	-	-		
Cumulative effect of one dose (seroconversion and priming): <i>n</i> (%) [†]	-	-	-	-	124/132	93.9%	110/128	85.9%	-	-		
Seroconversion by 18 weeks: <i>n</i> (%)	190/203	93.6% ^a	197/200	98.5% ^a	148/156	94.9%	133/152	87.5% ^b	202/211	95.7% ^b	^a <i>p</i> = 0.019 A vs B; ^b <i>p</i> = 0.005 D vs E; NS: B vs E; A vs E; A vs D	NS: A vs C; B vs C
Poliovirus shedding at 19 weeks: <i>n</i> (%)	31/203	15.3%	7/196	3.6% ^a	77/156	49.4%	73/151	48.3%	28/211	13.3% ^a	^a <i>p</i> = 0.001 B vs E; NS: C vs D	
Type 2												
Seroconversion by 14 weeks: <i>n</i> (%)	190/205	92.7% ^{b,c,d,e}	14/203	6.9% ^{a,b,f}	62/161	38.5% ^{e,f}	30/155	19.4% ^d	53/211	25.1% ^{a,c}	^a <i>p</i> <0.001 B vs E; ^b <i>p</i> <0.001 A vs B; ^c <i>p</i> <0.001 A vs E; ^d <i>p</i> <0.001 A vs D; NS: D vs E	^e <i>p</i> <0.001 A vs C; ^f <i>p</i> <0.001 B vs C
Priming response by 15 weeks: <i>n</i> (%) [†]	-	-	-	-	66/79	83.5%	73/101	72.3%	-	-		
Cumulative effect of one dose (seroconversion and priming): <i>n</i> (%) [†]	-	-	-	-	119/132	90.2%	100/128	78.1%	-	-		
Seroconversion by 18 weeks: <i>n</i> (%)	200/203	98.5% ^{b,c,d,e}	28/200	14% ^{a,b,f}	142/156	91% ^{e,f}	123/152	80.9% ^d	172/211	81.5% ^{a,c}	^a <i>p</i> <0.001 B vs E; ^b <i>p</i> <0.001 A vs B; ^c <i>p</i> <0.001 A vs E; ^d <i>p</i> <0.001 A vs D; NS: D vs E	^e <i>p</i> = 0.002 A vs C; ^f <i>p</i> <0.001 B vs C
Poliovirus shedding at 19 weeks: <i>n</i> (%)	12/203	5.9%	119/196	60.7%	89/156	57.1%	99/151	65.6%	122/211	57.8%	NS: C vs D	
Type 3												

	A tOPV	B bOPV	C iPV*	D f-IPV*	E f-IPV/ bOPV	Fisher's exact test (a priori)	Fisher's exact test (post hoc)**
Seroconversion by 14 weeks: <i>n</i> (%)	174/205 84.9% ^{b,c,e}	181/203 89.2% ^{a,f}	54/161 33.5% ^{e,f}	22/155 14.2% ^{c,d}	153/211 72.5% ^{a,b,d}	^a <i>p</i> < 0.001 B vs E; ^b <i>p</i> = 0.003 A vs E; ^c <i>p</i> < 0.001 A vs D; ^d <i>p</i> < 0.001 D vs E; NS: A vs B	^e <i>p</i> < 0.001 A vs C; ^f <i>p</i> < 0.001 B vs C
Priming response by 15 weeks: <i>n</i> (%) [†]	-	-	84/87 96.6%	94/107 87.9%	-		
Cumulative effect of one dose (seroconversion and priming): <i>n</i> (%) [†]	-	-	129/132 97.7%	115/128 89.8%	-		
Seroconversion by 18 weeks: <i>n</i> (%)	192/203 94.6%	188/200 94.0%	152/156 97.4%	135/152 88.8%	198/211 93.8%	NS: B vs E; A vs B; A vs E; A vs D; D vs E	NS: A vs C; B vs C
Poliovirus shedding at 19 weeks: <i>n</i> (%)	16/203 7.9%	12/196 6.1% ^a	50/156 32.1%	64/151 42.4%	29/211 13.7% ^a	^a <i>p</i> = 0.013 B vs E; NS: C vs D	

NS, not significant; tOPV, trivalent oral poliovirus vaccine (OPV); bOPV, bivalent OPV; iPV, inactivated poliovirus vaccine; f-IPV, fractional IPV.

* Test comparison of IPV (Arm C) and f-IPV (Arm D) presented in Fig. 2.

** Bonferroni correction. Significance at *p* < 0.0125.

[†] Analysis restricted to those with serological results at 6, 14, 15 and 18 weeks.