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Multivariate GWAS of Structural Dental Anomalies and Dental Caries in a Multi-Ethnic Cohort

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AUTHOR CONTRIBUTIONS

RNA generated the data set for the analyses. RNA analyzed data. RNA, BJH, LMU, MLM, JRS, SMW helped define outcomes to be studied. RNA wrote the first draft of the manuscript. MLM, JCM, JRS, SMW, BJH, LMU, and KN critically reviewed the manuscript. RNA & MLM designed the study. RNA, MLM, FWBD, KN, CP, IMO, CJB, JTH, GLW, SMW, and LMU collected and interpreted data. RNA & MLM generated the final draft of the manuscript. All authors read and approved the final manuscript.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by IRB. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin. Written informed consent was obtained from the individual(s), and minor(s)' legal guardian/next of kin.

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Abstract

Odontogenesis is a complex process, where disruption can result in dental anomalies and/or increase the risk of developing dental caries. Based on previous studies, certain dental anomalies tend to co-occur in patients, suggesting that these traits may share common genetic and etiological components. The main goal of this study was to implement a multivariate genome-wide association study approach to identify genetic variants shared between correlated structural dental anomalies and dental caries. Our cohort ($N = 3,579$) was derived from the Pittsburgh Orofacial Clefts Study, where multiple dental traits were assessed in both the unaffected relatives of orofacial cleft (OFC) cases ($n = 2,187$) and unaffected controls ($n = 1,392$). We identified four multivariate patterns of correlated traits in this data: tooth agenesis, impaction, and rotation (AIR); enamel hypoplasia, displacement, and rotation (HDR); displacement, rotation, and mamelon (DRM); and dental caries, tooth agenesis and enamel hypoplasia (CAH). We analyzed each of these four models using genome-wide multivariate tests of association. No genome-wide statistically significant results were found, but we identified multiple suggestive association signals ($P < 10^{-5}$) near genes with known biological roles during tooth development, including *ADAMTS9* and *PRICKLE2* associated with AIR; *GLIS3*, *WDR72*, and *ROR2* associated with HDR and DRM; *ROBO2* associated with DRM; *BMP7* associated with HDR; and *ROBO1*, *SMAD2*, and *MSX2* associated with CAH. This is the first study to investigate genetic associations for multivariate patterns of correlated dental anomalies and dental caries. Further studies are needed to replicate these results in independent cohorts.

Keywords

Multivariate GWAS; Dental Anomalies; Correlation; Dental Caries; Multi-ethnic

Introduction

Odontogenesis, the process of tooth development, is a complicated process that starts early in embryogenesis (1, 2). Mouse and human models indicate that interactions between the epithelium and mesenchymal cells during odontogenesis occur under the control of different families of signaling molecules and their receptors, including transforming growth factors (TGF), bone morphogenetic proteins (BMP), fibroblast growth factors (FGF), epidermal growth factor (EGF), and the hedgehog (Hh) and wntless (Wnt) families (3, 4). There are several known genes that regulate the communication and interaction between epithelium and mesenchymal cells, including *MSX1*, *MSX2*, *PAX9*, *RUNX2*, *AXIN2*, *EDA*, *GLI2*, and *GLI3*. Disturbances during the signaling process or changes in any of the regulating genes may result in dental anomalies, including changes in tooth development, structure, number, size, and morphology (5, 6).

Structural dental anomalies, such as tooth agenesis, enamel hypoplasia, impaction, rotation, displacement, mamelons, and supernumerary teeth (Supplementary Table 1 provides definitions for the dental anomalies used in this study) are presumed to be caused by interactions between genetic, epigenetic, and environmental factors during the process of tooth development (7). There is evidence that these dental defects tend to co-occur in the same patient, which could indicate a shared genetic etiology (8, 9). Although some dental anomalies might be asymptomatic, these anomalies can also lead to serious clinical problems, including delayed eruption or impaction of the teeth; temporomandibular joint pain and dysfunction; malocclusion; periodontal disease due to excessive occlusal force; and increased susceptibility to dental caries due to defects in tooth structure and/or crowding (10).

Different types of studies have taken advantage of next generation sequencing technologies to investigate the causes of genetic diseases and anomalies that affect the craniofacial complex. However, those studies to date have mostly focused on one disease or anomaly without taking into account that some genetic variants could cause multiple different anomalies. Therefore, the goal of this study was to better understand the inter-relationships between multiple structural dental anomalies and dental caries on a phenotypic level by identifying patterns of correlations among traits. We then conducted a multivariate genome wide association study (GWAS) to identify common variants that impact clusters of dental traits identified from these observed patterns of correlation.

METHODS AND MATERIAL

Sample

The cohort for this study is part of the Pittsburgh Orofacial Clefts Study (POFC) which studies risk factors for orofacial cleft birth defects (OFCs). Participants for POFC were recruited from multiple cleft centers in the United States, including Colorado, Iowa, Pennsylvania, Texas, and Puerto Rico, and internationally, from Argentina, the Philippines, Colombia, Guatemala, and Hungary. Institutional review board (IRB) approval was obtained at each site by the appropriate IRB process and committee, with a coordinating IRB at the University of Pittsburgh (IRB 0405013). The same data collection protocols were used for every site.

The total sample used for this study ($n = 3,579$, ages 8–82) included only OFC-unaffected individuals: 1,392 control individuals, with no personal nor family history of craniofacial anomalies, and 2,187 unaffected relatives of cases with OFCs, including case parents, siblings and spouses. Excluded were participants 7 years of age or younger; any case affected with an overt OFC, edentulous participants, or participants having any facial trauma or surgery. Table 1 summarizes the basic descriptive statistics for the participants involved in our study.

Data regarding the participant's dental history, including dental extractions and orthodontic treatment, were collected from all participants by self-report. In addition, each participant had an in-person dental exam and/or extensive intraoral photos taken. The reliability of data from intraoral photos was compared to in-person dental exams in a subset of individuals

with both dental exams and photos, prior to the start of data collection. Both the Intra-rater reliability and Inter-rater reliabilities were excellent (anomalies kappa =0.91–0.95; caries kappa =0.91- 0.99) (11).

Data Collection and Genotyping

All information regarding the data collection process, DNA collection, genotyping and quality control are summarized in the Supplementary material.

Structural Dental Anomalies and Dental Caries

For the current study we investigated enamel hypoplasia, microdontia, rotated and displaced teeth, supernumerary teeth, tooth agenesis, mamelons, and dental caries, all as binary (“yes/no”) traits.

Each of these traits was assigned a binary value of yes in each subject if there was at least one instance of a trait across the dentition. Then the prevalence of each trait was averaged over the total study for unaffected relatives and controls separately. We tested if there was a significant difference in the prevalence values between unaffected relatives and controls. Further, sex differences were evaluated for each dental anomaly and for dental caries. Lastly, we tested for correlations amongst the dental anomalies and dental caries.

Statistical Methods

All descriptive and comparative statistical analyses were performed using the R v3.4.1 statistical analysis environment (12). The prevalence of dental anomalies and dental caries, and comparisons between these traits were performed using χ^2 tests. The Spearman correlation coefficient was used to evaluate the associations between the traits, with a $P < 0.05$ was considered as significant correlation.

We utilized a multivariate GWAS approach in this study, the multivariate test of association implemented in PLINK (MV-PLINK) (13), with adjustment for age, age², sex, subject types (Control or unaffected relatives), site, and principal components (PCs) of genetic ancestry. Principle components (PCs) can be used to help clarify the differences among the sample participants in the genetic data (14). MV-PLINK uses canonical correlation analysis (CCA), a multivariate generalization of the Pearson product-moment correlation, in order to measure the association between sets of variables. CCA extracts the linear combination of traits that explain the largest possible amount of the covariation between the genetic variants and all traits. The CCA method implemented in MV- PLINK is equivalent to multivariate analysis of variance and has been shown to outperform other methods (15). Wilks' lambda was used to test the significance of the canonical correlations. Wilk's lambda (F) corresponds to the linear combination of traits with maximum correlation with the genetic variant; the correlation coefficients (Weights) for each individual trait indicate the contributions of each trait to the association result (13). Because our data comes from a large family study, we incorporated permutation testing implemented in MV-PLINK to correct for family structure.

Possible genomic inflation was assessed by calculating the genomic inflation factor, lambda (λ), visualized in a quantile-quantile (QQ) plot in R. In addition, we used R to create

Manhattan plots to visualize the association results. Single nucleotide polymorphisms (SNP variants) with minor allele frequencies lower than 5%, as well as SNPs with genotyping call rates less than 10% were filtered out. The threshold for genome-wide significance was set to $P = 5 \times 10^{-8}$ (Bonferroni correction for a million tests), and for suggestive significance to p -values between 10^{-5} and 5×10^{-8} . The top associated loci were then annotated.

We investigated all the genes within ± 500 kilobases (16) of the top association signals (index SNPs) for putative connections to dental development or specific dental anomalies. To visualize regions showing genome-wide significance and regions showing suggestive significance, we used the regional plots generated by LocusZoom. For investigation of functionally, genes were investigated using the resources such as the Gene and PubMed databases at the National Center for Biotechnology Information (NCBI). To examine the genes near the association signals, we searched these multiple databases including [OMIM](#), [UCSC](#), [Genome Browser](#), [EMAGE](#), [Ensembl](#), [ENCODE](#) and the Mouse Genome Informatics database by using terms including the name of the gene or corresponding protein, plus terms relevant to dental development and dental anomalies.

RESULTS

Summary of Dental Caries and Dental Traits in the Study Cohort

The study cohort included 1505 males (42.05%), and 2074 females (57.95%) with an age range of 8–82 years and a mean age of 31 years (Table 1), with 991 participants (27.69%) from U.S recruitment sites, and 2,588 participants (72.31%) from international sites. The distribution by age and recruitment site is summarized in Supplementary Table 2.

The distribution of the different dental anomalies is presented in Supplementary Figure 1. Rotation (70.80%) and Displacement (40.77%) were the most prevalent dental anomalies in the study cohorts. Females had significantly more rotation ($P = 6.478^{-09}$), and displacement ($P = 1.003^{-06}$) than males in the cohort (Table 2). As shown in Table 3, there were some differences in anomaly incidence between the OFC relatives and the unrelated controls, however these differences were not statistically significant after correcting for multiple testing.

Dental Caries

Previous studies indicated that relatives of individuals with clefts do not have an increased risk of dental caries when compared to the general population (17-19). Our study showed that there was a trend for higher rates of dental caries among unaffected relatives versus controls (DFT/*dft* percentage = 62.58% vs. 37.42%, $P = 0.0018$).

There was also a trend for an increased incidence for dental caries among females versus males (DFT/*dft* percentage = 1.13 vs. 1.54, $P = 0.002177$) across the different POFC sites. Higher dental caries prevalence among females when compared to males has been reported before (20, 21). Hypothesized reasons include a) earlier dental eruption in females, leading to a longer exposure to a cariogenic oral environment, b) hormonal changes during pregnancy, c) higher intake of snacks that might contain high sucrose amongst pregnant women, d) and behavioral factors such as poor oral hygiene (20, 21). In addition,

previous studies hypothesized that sex differences in dental caries could be explained by the differential effects of genes that influence dental caries risk (22, 23).

Dental Anomalies

We observed that unaffected relatives displayed more rotation ($P=0.0016$) than controls. We also observed a trend for an increased rate in unaffected relatives for impaction (80.49% vs. 19.51%, $P=0.01$), supernumerary teeth (84.62% vs. 15.38%, $P=0.014$), and dental caries (78.40% vs. 62.58%, $P=0.0018$) compared with controls.

Sex differences for all the dental anomalies plus dental caries as a binary trait, were tested. Females displayed significantly more rotation ($P=6.478^{-09}$) and displacement ($P=1.003^{-06}$) when compared to males. The tendency to develop more rotation and displacement in females could be explained by previous observations that females have different characteristics of the dentition (differences in the dentin, enamel, and mesio-distal teeth width) when compared to males (20, 24). A previous study by our group investigated the sex differences in certain dental anomalies within this cohort (POFC) and no significant sex difference were found, however note that tooth rotation and displacement were not included as separate traits in that previous study (11), they were both treated as one trait (malposition) and were excluded from the overall analysis.

Multivariate Patterns of Dental Traits

Based on the Spearman rank correlation coefficient, shown in Table 4, we found statistically significant evidence of correlation between the following groups of anomalies: tooth agenesis, impaction, and rotation (AIR); enamel hypoplasia, displacement, and rotation (HDR); and displacement, rotation, and mamelons (DRM). Further, dental caries was correlated with tooth agenesis and enamel hypoplasia (CAH). Notably, 84.4% of participants who had enamel hypoplasia had dental caries (Supplementary Table 3). However, note that the magnitude of most of the correlation coefficients indicate relatively weak correlation (i.e., We analyzed each of the four correlated groups of traits using MV-PLINK.

A total of 5,802,671 SNPs was available for analysis after applying quality control criteria. Although no SNPs reached genome-wide significance ($P < 5 \times 10^{-8}$) for any of the four correlated groups of traits (AIR, HDR, DRM and CAH) there were several regions of the genome that showed suggestive significance ($P \sim 10^{-5}$). Association results for the top SNPs for the four correlation groups are shown in Table 5 and Figure 1. The suggestive associations for each of the correlated groups are summarized in the following sections.

TOOTH AGENESIS, IMPACTION AND ROTATION—There were 100 SNPs showing suggestive associations with AIR, of which five were in regions with possible relevance to dental development, summarized below with regional plots shown in Figure 2.

Chr3:64541255 ($P=1.00E-06$) is located in an intron of *ADAMTS9*, which codes a protein located in the extracellular matrix (ECM) and interacting tightly with ECM proteins. *ADAMTS9* may modify the extracellular environment, influencing proliferation and survival of neural crest-derived cells(25). In mice, *Adamts9* expression is seen in developing craniofacial structures such as teeth and mandible(26). In addition, this variant

is also approximately 300kb upstream of *PRICKLE2*, a planar cell polarity protein involved in amelogenesis. In rats, *Prickle2* in rats is expressed in the differentiating inner enamel epithelial cells and early inner enamel-secretory ameloblasts (27).

Variant rs140220410 ($P = 3.00E-06$) is less than 100 kb downstream from *ARNT2*, a transcriptional regulator involved in several biological functions, including regulation of developmental genes. In mouse, *Arnt2* was expressed in the molar and incisor teeth, the odontoblasts, and both inner and outer enamel epithelium (28).

Variant rs9913511 ($P = 3.00E-06$) is intronic to *NTN1* which encodes a protein in the family of laminin-related secreted proteins. Although little is known about this gene with respect to dental development, it affects development of the craniofacial region in animal models (29), mediating a critical step in palatal fusion in mouse embryo (30).

Variant rs2251904 ($P = 4.00E-06$) is downstream of *TRPC4* which encodes a protein involved in multiple biological processes, including neurotransmitter release and cell proliferation. Notably, in rats *Tpc4* was highly expressed in the rat dental follicle and stellate reticulum cells during the early stage, and moderately expressed in odontoblasts (31).

Variant rs1838002 ($P = 6.00E-06$) is upstream of *WNT2B*, one of the WNT (wingless-type MMTV integration site) family of signaling factors that play an important role in human development, including the differentiation and proliferation of cementoblasts and odontoblasts (32).

ENAMEL HYPOPLASIA, DISPLACED, ROTATION—There were 105 SNPs showing suggestive associations with HDR of which four were in regions with possible relevance to dental development, summarized below (regional plots shown in Figure 3).

Variant rs12379966 ($P = 1.00E-06$) is less than 100 kb upstream from the zinc finger protein *GLIS*. The encoded protein of *GLIS3* regulates and improves osteoblast differentiation by acting interdependently with BMP2 and Shh. In addition, *GLIS3* promotes an increase in FGF18 expression during osteoblast differentiation (33). *GLIS3* has not been studied in tooth development.

Variant rs6479408 ($P = 1.00E-06$) is downstream from *ROR2*, an orphan tyrosine kinase that mediates Wnt5a-initiated noncanonical signaling and Wnt5a-inhibition. Wnt canonical signaling, required during the growth, patterning, and differentiation of teeth. *Ror2* expression has been observed during the development of teeth in mice, with tooth developmental retardation seen in *Ror2* mutant mice (34). This variant is also located upstream to the Osteomodulin gene (*OMD*), a member of the small leucine-rich proteoglycan family distributed in the extracellular matrix (ECM). *OMD* is expressed in the polarized odontoblasts and alveolar bone during early crown formation and plays an essential role in modulating the osteo/odontoblastic differentiation of human dental pulp stem cells (35).

Variant rs141429354 ($P = 6.00E-06$) is downstream of *WDR72*, a protein coding gene is essential during the maturation phase of amelogenesis for normal formation of the enamel (36). *Wdr72* expression was detected in the enamel organ of mouse incisors, and knockout mice (*Wdr72* $-/-$) have defects in enamel maturation (37). Mutations in this gene are associated with the hypomaturation seen in amelogenesis imperfecta (36).

Variant rs404727 ($P = 8.00E-06$), is upstream to *BMP7*, which encodes a secreted ligand of transforming growth factor-beta (TGF-beta) a superfamily of proteins that play a role in ectopic bone formation and odontogenesis. *Bmp7* is highly expressed during tooth development in mice; *Bmp7* knock-out mice had morphological and functional changes in teeth (38). In studies of human dental mesenchyme *BMP7* was expressed in the late bell-stage dental papilla and thus might play a role in inducing the odontogenic differentiation of human dental pulp stem cells (39).

DISPLACED, ROTATION, MAMELONS—There were 100 SNPs showing suggestive associations with DRM of which four were in regions with possible relevance to dental development, summarized below (regional plots shown in Figure 4).

Three of the variants were also suggestively associated with the HDR pattern, not surprisingly given that these two correlation groups share two traits (see the HDR section for detailed summaries). **Variant rs10511451** ($P = 1.00E-06$ for DRM), is upstream of the zinc finger protein *GLIS3*. One of the variants suggestively associated with HDR, variant rs12379966 ($P = 1.00E-06$), was also located upstream to *GLIS3* gene. **Variant rs141429354** ($P = 3.00E-06$ for DRM, $P = 6.00E-06$ for HDR) is downstream of *WDR72*; **Variant rs6479408** ($P = 5.00E-06$ for DRM and $P = 1.00E-06$ for HDR) is near *ROR2*.

Variant rs174814 ($P = 9.00E-06$ for association with DRM) is located approximately 100 kb upstream of *ROBO2* which encodes a protein that acts as a cell receptor for slit2 which plays a role in cell migration (40). In a gene set enrichment analysis study, *ROBO2* was listed as “potentially” associated with dental traits (41).

DENTAL CARIES, TOOTH AGENESIS, ENAMEL HYPOPLASIA—There were 149 SNPs showing suggestive associations with CAH of which six were in regions with possible relevance to dental development, summarized below (regional plots shown in Figure 4).

Variant rs79577009 ($P = 2.87E-07$) is located intronic to *SMAD2* which encodes a protein that mediate the signal of transforming growth factor (TGF)-beta, indirectly regulating multiple cellular processes, including cell proliferation, differentiation, and odontogenesis(4). In mice, *smad2* plays an essential role during early stages of tooth formation (42).

Variant rs11125855 ($P = 8.82E-07$) is upstream of *REL* which encodes a protein in the “Rel homology domain/immunoglobulin-like fold plexin, transcription factor” (RHD/IPT) family, with roles in biological processes including apoptosis, inflammation, and the immune response (e.g. survival and proliferation of B lymphocytes) which produce cytokines as a response to inflammation regulating the intensity and duration of the immune response

(43). In mice, *REL* (also known as c-rel) controls the development of the epidermis and associated appendages, such as teeth, during embryogenesis (44).

Variant rs6758898 ($P = 2.93E-06$) is located less than 100kb from *IL1A* and *IL1B* (part of the interleukin 1 cytokine family) whose encoded proteins have multiple functions within the immune system. *IL1A* (Interleukin-1 alpha) has a role in bone resorption, and positively affects the survival and differentiation of osteoclast and odontoclast(45). *IL1B* (Interleukin-1 beta) has an important function as a mediator in the inflammatory response (46). A case-control study in China found that a significant association between dental caries and *IL1B*(47). Interestingly, a systematic review of twenty-seven studies showed that genetic variation in both *IL-1a* and *IL-1b* could contribute to chronic periodontitis in whites (46).

Variant rs4868444 ($P = 3.09E-06$) is intronic to *MSX2* whose encoded protein provides the balance between survival and apoptosis of neural crest-derived cells, necessary for proper craniofacial morphogenesis. In addition, *msx2* protein is part of the bone morphogenic protein (BMP) signaling pathway, that regulates various processes including odontogenesis (48). A mutation in *MSX2* has been found in a family with amelogenesis imperfecta and impaired tooth eruption (49), and *Msx2* knockout mice showed a tooth abnormality resembling human amelogenesis imperfecta (50).

Variant rs5850440 ($P = 3.37 E-06$) is located intronic to *ROBO1*, with an encoded protein that functions as cell receptor for slit1. In mice, both slit1 and robo1 were expressed in the primary enamel knot and during the cap stage, plus robo1 expression in tooth germ and dental papilla was noted. Moreover, it was found that even before birth both of robo1 and robo2 were localized in preodontoblast (51). As detailed earlier, note that a variant in *ROBO2* was suggestively associated with the DRM pattern in this study.

DISCUSSION

In recent years there has been large number of new genomic approaches and new methods of analyzing and understanding the wealth of genomic data. The current study leveraged a novel multivariate approach to GWAS (MV-PLINK) (13) that can aid in understanding the inter-relationship between multiple traits, in this case dental anomalies, and to investigate genetic associations underlying these correlations. To do so, we utilized a large, multi-ethnic study cohort that was well-characterized for multiple dental characteristics including structural dental traits and dental caries.

Some studies have found that families with clefting have a significantly higher risk for developing dental anomalies than general population (52-54). However, there are also studies that found no significant differences (11, 55, 56). In our study, subject type, i.e. unaffected relatives vs. controls, was tested for all dental anomalies' variables, and we noticed that unaffected relatives had more rotation in their teeth ($P = 0.0016$) when compared to controls. In addition, a trend for an increased rate in unaffected relatives for impaction (80.49% vs. 19.51%, $P = 0.01$), supernumerary teeth (84.62% vs. 15.38%, $P = 0.014$), and for dental caries (78.40% vs. 62.58%, $P = 0.0018$) compared with controls. However, we do not consider these differences significant in our study after correcting for

multiple testing, which is also consistent with the findings from a previous study in this same cohort (11). Rotation and displacement were the most prevalent anomalies in our study. This could be due to the different patterns of racial/ethnic admixture in our study cohort, and that potentially could induce bias (57).

Based on Spearman rank correlation coefficients, we found multivariate correlations between different structural dental anomalies and between some of these anomalies and dental caries. Note that other studies have shown similar correlations. For example, one study has found a significant association between impacted maxillary canine and the dental anomalies of tooth number, tooth size and rotations which suggest a shared common genetic background (58). Another study also discovered associations among different dental anomalies, including an association between tooth agenesis and displacement of maxillary canines, and between tooth agenesis and tooth transposition (59). We hypothesized that these correlations are due to similar genetic backgrounds, emphasizing the importance of investigating these anomalies on a genetic level. Therefore, we used a multivariate GWAS approach to investigate genetic factors underlying these patterns, as recommended when genetic correlations between traits are relatively weak, as observed in the current analyses.

This study is the first to apply multivariate GWAS to identify possible genetic loci associated with the presence of patterns of correlated dental anomalies (AIR, DRM, HDR, or CAH). Although none of our multivariate GWAS results reached the strict genome-wide significance level, there were a large number of variants with suggestive genome-wide significance relevant to odontogenesis and dental caries. The suggestive variants were annotated to help generate hypotheses and nominate these variants for further investigations using information from different bioinformatics databases. We focused our search on genes with known biological functions related to tooth development process (odontogenesis) and oral/dental health, either in human or in rodents. However, there were other suggestive variants near genes with biological roles in the body that could be of interest for further investigation in future studies.

The strongest association signal was in the CAH model (dental caries, tooth agenesis, enamel hypoplasia). The lead variant, rs79577009 ($P = 2.87E-07$) is located intronic to the *SMAD2* gene, which has an important function in mediating the signal of the transforming growth factor (TGF)-beta and helps indirectly (TGFb) in regulating multiple cellular processes, including odontogenesis (4). In addition, *smad2* was found to be involved during the early stages of tooth development in mice, so we hypothesize that variants in *SMAD2* might increase the chance of having any of the correlated dental anomalies: dental caries, tooth agenesis, and/or enamel hypoplasia.

An overlap seen in results of the HDR and DRM GWAS could be due to the fact that the two patterns share two dental anomalies (D and R). The results of these overlapped regions of significance are presented in Supplementary Table 4, and include suggestive association signals near the *WDR72* gene, which has an essential role in the mineralization and maturation of tooth enamel (36). Further investigations needed to replicate and understand the extent of these associations.

There are no previously reported GWAS studies directed at identifying risk loci associated with multiple correlated dental anomalies. There have been however, GWAS's of third molar tooth agenesis in cohorts of European ethnicity (60, 61). Further, in another GWAS of tooth agenesis (*excluding* third molars) in European ancestry that used a sample from the POFC study for replication, several risk variants were identified, near *ASCL5/CACNA1S*, *ARHGAP15*, *FOXI3*, *EDAR*, and *WNT10A* (62). Sequencing studies of tooth agenesis found that nonsynonymous, nonsense, and missense mutations in *WNT10A* are strongly associated with tooth agenesis of 1-3 teeth and also for 4 or more teeth (63). A previous study also identified several genes that have been associated with non-syndromic forms of tooth agenesis, including *MSX1*, *PAX9*, *AXIN2*, and *EDA*(64). Each of these genes play an important role during tooth development.

We examined the candidate genes from previous sequencing and GWASs of tooth agenesis in our AIR and CAH GWAS modules results to see if they had been replicated. GWAS of tooth agenesis was the only dental anomaly that was investigated before in a single ancestry and in a sample that has no other syndromes. Only limited evidence of suggestive association was seen near the previously reported tooth agenesis genes (results shown in Supplementary Table 5).

The use of binary traits in GWAS's is very common and has multiple advantages. However, the limitation of this approach is that the phenotype is less precise which could negatively affect the power of a GWAS study (65), that is, there might be false negative associations but not false positives. The major limitation of this study was the lack of replication cohorts available for testing the variants nominated with the patterns of correlated dental characteristics. Additional studies in larger and more diverse cohorts are warranted to assess the effects of the potential variants identified in this study.

In summary, we nominate genes with known biological roles during tooth development, including *ADAMTS9* and *PRICKLE2* from the AIR pattern; *GLIS3*, *WDR72*, and *ROR2* from HDR and DRM patterns; *ROBO2* from the DRM pattern; *BMP7* from the HDR pattern; and *ROBO1*, *SMAD2* and *MSX2* from the CAH pattern. In addition, genes were identified with plausible roles in tooth development such as: *ARNT2* and *WNT2B* from the AIR pattern; *OMD* from the HDR pattern; and *REL*, *IL1A* and *IL1B* from the CAH pattern.

CONCLUSION

Evidence of correlation between multiple different structural dental anomalies, including the correlation between dental caries and enamel hypoplasia, reflect support for the hypothesis that similar genetic background or other etiologic factors may underlie multiple dental characteristics and dental disease. This is the first study to perform multivariate GWAS for patterns of associated dental anomalies and dental caries, and we were able to identify suggestive genetic loci for four correlated dental anomaly patterns that play plausible biological roles during tooth development. Further studies are needed to replicate the analyses in independent cohorts.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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DATA AVAILABILITY STATEMENT

The datasets analyzed in this paper are available in the dbGaP repository, https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000774.v2.p1, further queries can be forwarded to the corresponding author/s.

References:

1. Thesleff I Epithelial-mesenchymal signalling regulating tooth morphogenesis. *Journal of Cell Science*. 2003.
2. Thesleff I, Nieminen P. Tooth morphogenesis and cell differentiation. *Current Opinion in Cell Biology*. 1996.
3. Hu X, Zhang S, Chen G, Lin C, Huang Z, Chen Y, et al. Expression of SHH signaling molecules in the developing human primary dentition. *BMC Developmental Biology*. 2013.
4. Sarkar L, Cobourne M, Naylor S, Smalley M, Dale T, Sharpe PT. Wnt/Shh interactions regulate ectodermal boundary formation during mammalian tooth development. *Proceedings of the National Academy of Sciences of the United States of America*. 2000.
5. Bailleul-Forestier I, Molla M, Verloes A, Berdal A. The genetic basis of inherited anomalies of the teeth. Part 1: Clinical and molecular aspects of non-syndromic dental disorders. 2008.
6. Thesleff I, editor *The genetic of tooth development and dental defects* 2006 2006.
7. Brook AH, Griffin RC, Townsend G, Levisianos Y, Russell J, Smith RN. Variability and patterning in permanent tooth size of four human ethnic groups. *Archives of Oral Biology*. 2009.
8. Markovic M Hypodontia in twins. *Swedish dental journal Supplement*. 1982.
9. Mossey PA. The heritability of malocclusion: part 2. The influence of genetics in malocclusion. 1999.
10. Temilola DO, Folayan MO, Fatusi O, Chukwumah NM, Onyejaka N, Oziegbe E, et al. The prevalence, pattern and clinical presentation of developmental dental hard-tissue anomalies in children with primary and mix dentition from Ile-Ife, Nigeria. *BMC Oral Health*. 2014.
11. Howe BJ, Cooper ME, Vieira AR, Weinberg SM, Resick JM, Nidey NL, et al. Spectrum of dental phenotypes in nonsyndromic orofacial clefting. *Journal of Dental Research*. 2015.
12. R Development Core Team. *R: A language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing; 2019.
13. Ferreira MAR, Purcell SM. A multivariate test of association. *Bioinformatics*. 2009.

14. Price AL, Zaitlen NA, Reich D, Patterson N. New approaches to population stratification in genome-wide association studies. *Nat Rev Genet.* 2010;11(7):459–63. [PubMed: 20548291]
15. Galesloot TE, Van Steen K, Kiemeny LALM, Janss LL, Vermeulen SH. A comparison of multivariate genome-wide association methods. *PLoS ONE.* 2014.
16. Tang W, Teichert M, Chasman DI, Heit JA, Morange P-E, Li G, et al. A genome-wide association study for venous thromboembolism: the extended cohorts for heart and aging research in genomic epidemiology (CHARGE) consortium. *Genetic epidemiology.* 2013;37(5):512–21. [PubMed: 23650146]
17. Al-Dajani M Comparison of dental caries prevalence in patients with cleft lip and/or palate and their sibling controls. *Cleft Palate-Craniofacial Journal.* 2009.
18. De Castilho ARF, Das Neves LT, De Carvalho Carrara CF. Evaluation of oral health knowledge and oral health status in mothers and their children with cleft lip and palate. *Cleft Palate-Craniofacial Journal.* 2006.
19. Howe BJ, Cooper ME, Wehby GL, Resick JM, Nidey NL, Valencia-Ramirez LC, et al. Dental Decay Phenotype in Nonsyndromic Orofacial Clefting. *Journal of Dental Research.* 2017.
20. Martinez-Mier EA, Zandona AF. The impact of gender on caries prevalence and risk assessment. 2013.
21. Shaffer JR, Leslie EJ, Feingold E, Govil M, McNeil DW, Crout RJ, et al. Caries Experience Differs between Females and Males across Age Groups in Northern Appalachia. *International Journal of Dentistry.* 2015.
22. Ferraro M, Vieira AR. Explaining Gender Differences in Caries: A Multifactorial Approach to a Multifactorial Disease. *International Journal of Dentistry.* 2010.
23. Vieira AR, Marazita ML, Goldstein-McHenry T. Genome-wide scan finds suggestive caries loci. *Journal of Dental Research.* 2008.
24. Lukacs JR, Largaespada LL. Explaining sex differences in dental caries prevalence: Saliva, hormones, and "life history" etiologies. *American Journal of Human Biology.* 2006.
25. Christian L, Bahudhanapati H, Wei S. Extracellular metalloproteinases in neural crest development and craniofacial morphogenesis. 2013.
26. Jungers KA, Le Goff C, Somerville RPT, Apte SS. Adamts9 is widely expressed during mouse embryo development. *Gene Expression Patterns.* 2005.
27. Nishikawa S, Kawamoto T. Localization of core planar cell polarity proteins, PRICKLES, in ameloblasts of rat incisors: Possible regulation of enamel rod decussation. *Acta Histochemica et Cytochemica.* 2015.
28. Aitola MH, Pelto-Huikko MT. Expression of Arnt and Arnt2 mRNA in developing murine tissues. *J Histochem Cytochem.* 2003;51(1):41–54. [PubMed: 12502753]
29. Salminen M, Meyer BI, Bober E, Gruss P. netrin 1 is required for semicircular canal formation in the mouse inner ear. *Development.* 2000.
30. Serafini T, Colamarino SA, Leonardo ED, Wang H, Beddington R, Skarnes WC, et al. Netrin-1 is required for commissural axon guidance in the developing vertebrate nervous system. *Cell.* 1996.
31. Yang J, Cai W, Lu X, Liu S, Zhao S. RNA-sequencing analyses demonstrate the involvement of canonical transient receptor potential channels in rat tooth germ development. *Frontiers in Physiology.* 2017.
32. Yi Q, Liu O, Yan F, Lin X, Diao S, Wang L, et al. Analysis of senescence-related differentiation potentials and gene expression profiles in human dental pulp stem cells. *Cells Tissues Organs.* 2017.
33. Beak JY, Kang HS, Kim YS, Jetten AM. Krüppel-like zinc finger protein Glis3 promotes osteoblast differentiation by regulating FGF18 expression. *Journal of Bone and Mineral Research.* 2007.
34. Lin M, Li L, Liu C, Liu H, He F, Yan F, et al. Wnt5a regulates growth, patterning, and odontoblast differentiation of developing mouse tooth. *Developmental Dynamics.* 2011.
35. Lin W, Gao L, Jiang W, Niu C, Yuan K, Hu X, et al. The role of osteomodulin on osteo/odontogenic differentiation in human dental pulp stem cells. *BMC Oral Health.* 2019;19(1):22-. [PubMed: 30670012]

36. El-Sayed W, Parry DA, Shore RC, Ahmed M, Jafri H, Rashid Y, et al. Mutations in the Beta Propeller WDR72 Cause Autosomal-Recessive Hypomaturation Amelogenesis Imperfecta. *American Journal of Human Genetics*. 2009.
37. Katsura KA, Horst JA, Chandra D, Le TQ, Nakano Y, Zhang Y, et al. WDR72 models of structure and function: A stage-specific regulator of enamel mineralization. *Matrix Biology*. 2014.
38. Zurowski C, Jamniczky H, Graf D, Theodor J. Deletion/loss of bone morphogenetic protein 7 changes tooth morphology and function in *Mus musculus*: Implications for dental evolution in mammals. *Royal Society Open Science*. 2018.
39. Gao B, Zhou X, Zhou X, Pi C, Xu R, Wan M, et al. BMP7 and EREG contribute to the inductive potential of dental mesenchyme. *Scientific Reports*. 2015.
40. Nguyen-Ba-Charvet KT, Plump AS, Tessier-Lavigne M, Chédotal A. Slit1 and slit2 proteins control the development of the lateral olfactory tract. *Journal of Neuroscience*. 2002.
41. Wang Q, Jia P, Cuenco KT, Zeng Z, Feingold E, Marazita ML, et al. Association Signals Unveiled by a Comprehensive Gene Set Enrichment Analysis of Dental Caries Genome-Wide Association Studies. *PLoS ONE*. 2013.
42. Ito Y, Zhao J, Mogharei A, Shuler CF, Weinstein M, Deng C, et al. Antagonistic Effects of Smad2 Versus Smad7 are Sensitive to Their Expression Level during Tooth Development. *Journal of Biological Chemistry*. 2001.
43. Jontell M, Okiji T, Dahlgren U, Bergenholtz G. Immune defense mechanisms of the dental pulp. 1998.
44. Gugasyan R, Voss A, Varigos G, Thomas T, Grumont RJ, Kaur P, et al. The Transcription Factors c-rel and RelA Control Epidermal Development and Homeostasis in Embryonic and Adult Skin via Distinct Mechanisms. *Molecular and Cellular Biology*. 2004.
45. Song JS, Hwang DH, Kim SO, Jeon M, Choi BJ, Jung HS, et al. Comparative Gene Expression Analysis of the Human Periodontal Ligament in Deciduous and Permanent Teeth. *PLoS ONE*. 2013.
46. Karimbux NY, Saraiya VM, Elangovan S, Allareddy V, Kinnunen T, Kornman KS, et al. Interleukin-1 Gene Polymorphisms and Chronic Periodontitis in Adult Whites: A Systematic Review and Meta-Analysis. *Journal of Periodontology*. 2012.
47. Hu XP, Song TZ, Zhu YY, Wu LL, Zhang X, Zhou JY, et al. Association of ENAM, TUFT1, MMP13, IL1B, IL10 and IL1RN gene polymorphism and dental caries susceptibility in Chinese children. *Journal of International Medical Research*. 2019.
48. Davidson D The function and evolution of Msx genes: pointers and paradoxes. 1995.
49. Suda N, Kitahara Y, Ohyama K. A case of amelogenesis imperfecta, cleft lip and palate and polycystic kidney disease. *Orthodontics and Craniofacial Research*. 2006.
50. Satokata I, Ma L, Ohshima H, Bei M, Ian W, Nishizawa K, et al. Msx2 deficiency in mice causes pleiotropic defects in bone growth and ectodermal organ formation. *Nature Genetics*. 2000.
51. Loes S, Luukko K, Kvinnsland IH, Kettunen P. Slit1 is specifically expressed in the primary and secondary enamel knots during molar tooth cusp formation. *Mechanisms of Development*. 2001.
52. Aspinall A, Raj S, Jugessur A, Marazita M, Savarirayan R, Kilpatrick N. Expanding the cleft phenotype: the dental characteristics of unaffected parents of Australian children with non-syndromic cleft lip and palate. *Int J Paediatr Dent*. 2014;24(4):286–92. [PubMed: 24237197]
53. Eerens K, Vlietinck R, Heidbuchel K, Van Olmen A, Derom C, Willems G, et al. Hypodontia and tooth formation in groups of children with cleft, siblings without cleft, and nonrelated controls. *Cleft Palate Craniofac J*. 2001;38(4):374–8. [PubMed: 11420017]
54. Kuchler EC, da Motta LG, Vieira AR, Granjeiro JM. Side of dental anomalies and taurodontism as potential clinical markers for cleft subphenotypes. *Cleft Palate Craniofac J*. 2011;48(1):103–8. [PubMed: 20507240]
55. Anderson PJ, Moss AL. Dental findings in parents of children with cleft lip and palate. *Cleft Palate Craniofac J*. 1996;33(5):436–9. [PubMed: 8891376]
56. Haria S, Noar JH, Sanders R. An investigation of the dentition of parents of children with cleft lip and palate. *Cleft Palate Craniofac J*. 2000;37(4):395–405. [PubMed: 10912719]

57. Li Q, Yu K. Improved correction for population stratification in genome-wide association studies by identifying hidden population structures. *Genet Epidemiol.* 2008;32(3):215–26. [PubMed: 18161052]
58. Nagpal A, Pai KM, Sharma G. Palatal and labially impacted maxillary canine-associated dental anomalies: a comparative study. *J Contemp Dent Pract.* 2009;10(4):67–74.
59. Lagana G, Venza N, Borzabadi-Farahani A, Fabi F, Danesi C, Cozza P. Dental anomalies: prevalence and associations between them in a large sample of non-orthodontic subjects, a cross-sectional study. *BMC Oral Health.* 2017;17(1):62. [PubMed: 28284207]
60. Haga S, Nakaoka H, Yamaguchi T, Yamamoto K, Kim YI, Samoto H, et al. A genome-wide association study of third molar agenesis in Japanese and Korean populations. *Journal of Human Genetics.* 2013.
61. Vukelic A, Cohen JA, Sullivan AP, Perry GH. Extending genome-wide association study results to test classic anthropological hypotheses: Human third molar agenesis and the “probable mutation effect”. 2017.
62. Jonsson L, Magnusson TE, Thordarson A, Jonsson T, Geller F, Feenstra B, et al. Rare and Common Variants Conferring Risk of Tooth Agenesis. *Journal of Dental Research.* 2018.
63. Arzoo PS, Klar J, Bergendal B, Norderyd J, Dahl N. WNT10A mutations account for 1/4 of population-based isolated oligodontia and show phenotypic correlations. *American Journal of Medical Genetics, Part A.* 2014.
64. Matalova E, Fleischmannova J, Sharpe PT, Tucker AS. Tooth agenesis: From molecular genetics to molecular dentistry. 2008.
65. Rekaya R, Smith S, Hay EH, Farhat N, Aggrey SE. Analysis of binary responses with outcome-specific misclassification probability in genome-wide association studies. *Appl Clin Genet.* 2016;9:169–77. [PubMed: 27942229]

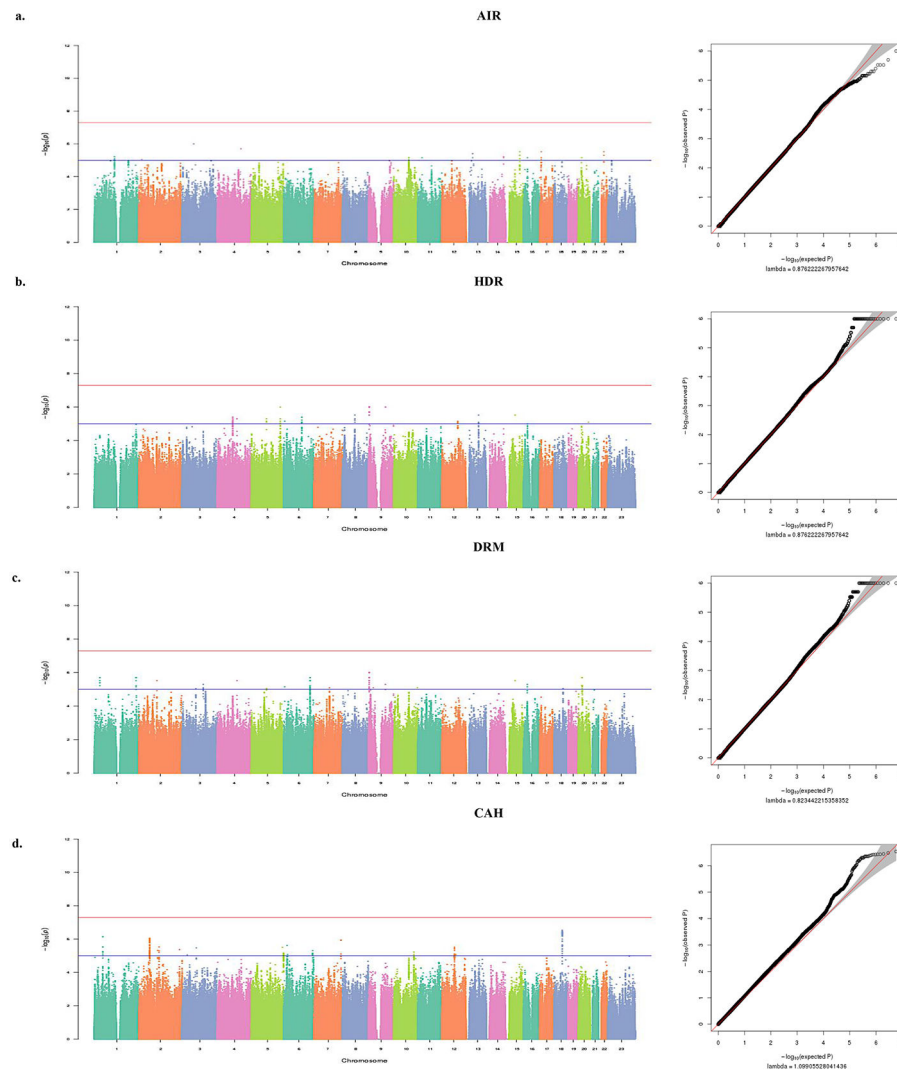


Figure.1. Manhattan plots and (Q-Q) plots showing multivariate GWAS results for the correlated dental anomalies patterns and dental caries. In the Manhattan plot the red lines represent thresholds for genome-wide significance (p value $< 5 \times 10^{-8}$). Blue lines represent thresholds for suggestive significance (p value $< 5 \times 10^{-6}$) and it shows negative log₁₀-transformed p values (y-axis) across the whole genome (x-axis). Genotyped and imputed SNPs are plotted together. **a.** on the left is the Manhattan plot for Tooth Agensia, impaction and Rotation (AIR), on the right is the quantile–quantile plot (Q-Q), and the genomic inflation factor (λ) is 0.87. **b.** on the left is the Manhattan plot for Hypoplasia, displacement and rotation (HDR), on the right is the quantile–quantile plot (Q-Q), and the genomic inflation factor (λ) is 0.87. **c.** on the left is the Manhattan plot for Displacement, Rotation and Mamelon (DRM), on the right is the quantile–quantile plot (Q-Q), and the genomic inflation factor (λ) is 0.82. **d.** on the left is the Manhattan plot for Dental caries, Agensia and Hypoplasia (CAH), on the right is the quantile–quantile plot (Q-Q), and the genomic inflation factor (λ) is 1.09.

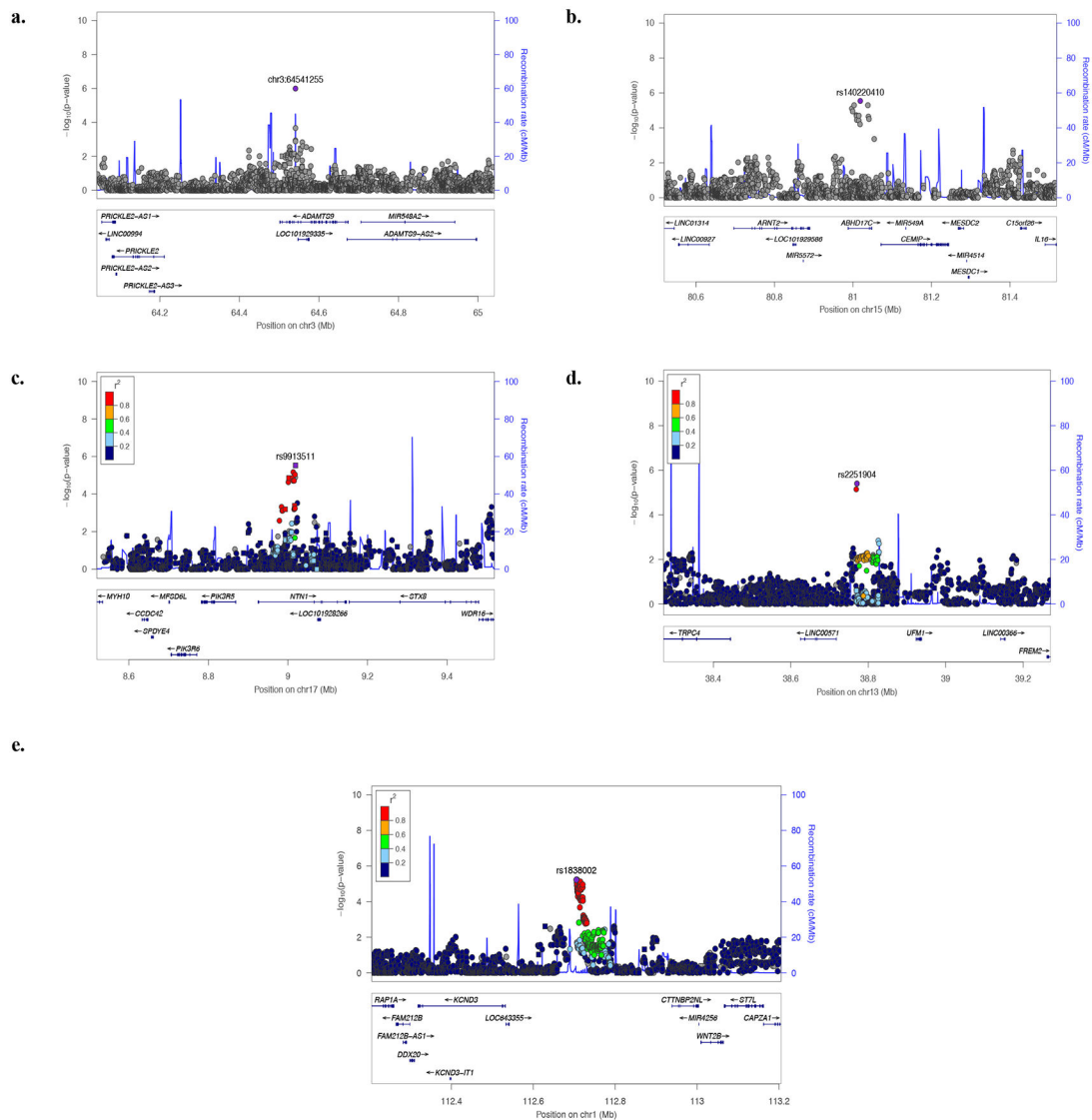


Figure.2.

Regional Association Plots from (AIR) results. LocusZoom were used to generate plots for regions of Interest. **a.** suggestive locus near *ADAMTS9* and *PRICKLE2* on chromosome 3. **b.** suggestive locus near *ARNT2* gene on chromosome 15. **c.** suggestive locus near *NTN1* gene on chromosome 17. **d.** suggestive locus near *TRPC4* on chromosome 13. **e.** suggestive locus near *WNT2B* gene on chromosome 1. The genome build used for the recombination rate was based on 1000 Genomes November 2014 EUR data. All of the gene positions and directions of transcription are annotated on the plots.

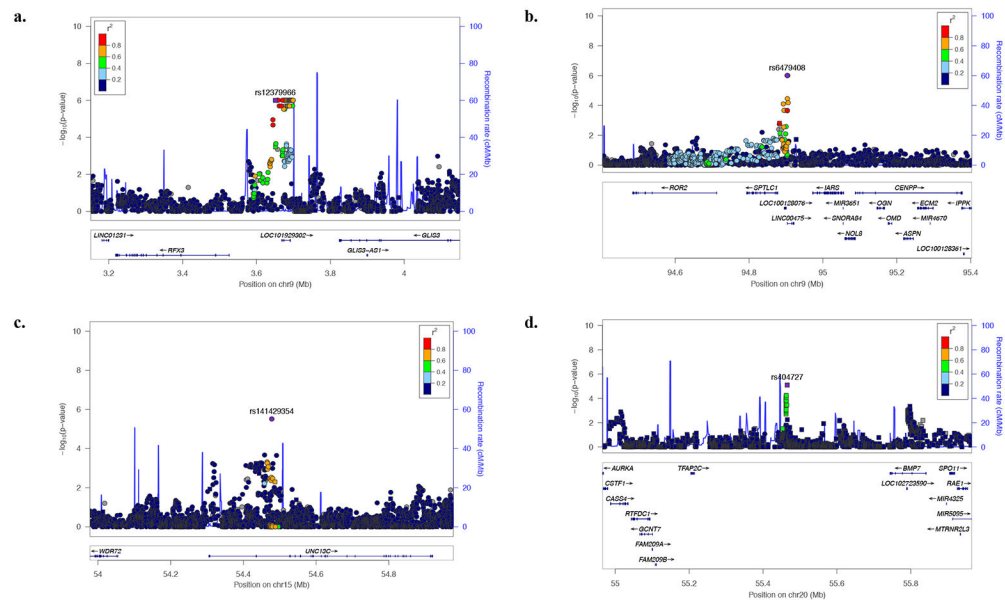


Figure.3.

Regional Association Plots from (HDR) results. LocusZoom were used to generate plots for regions of Interest. **a.** suggestive locus near *GLIS3* on chromosome 9. **b.** suggestive locus near *ROR2* and *OMD* genes on chromosome 9. **c.** suggestive locus near *WDR72* gene on chromosome 15. **d.** suggestive locus near *BMP7* on chromosome 20. The genome build used for the recombination rate was based on 1000 Genomes November 2014 EUR data. All of the gene positions and directions of transcription are annotated on the plots.

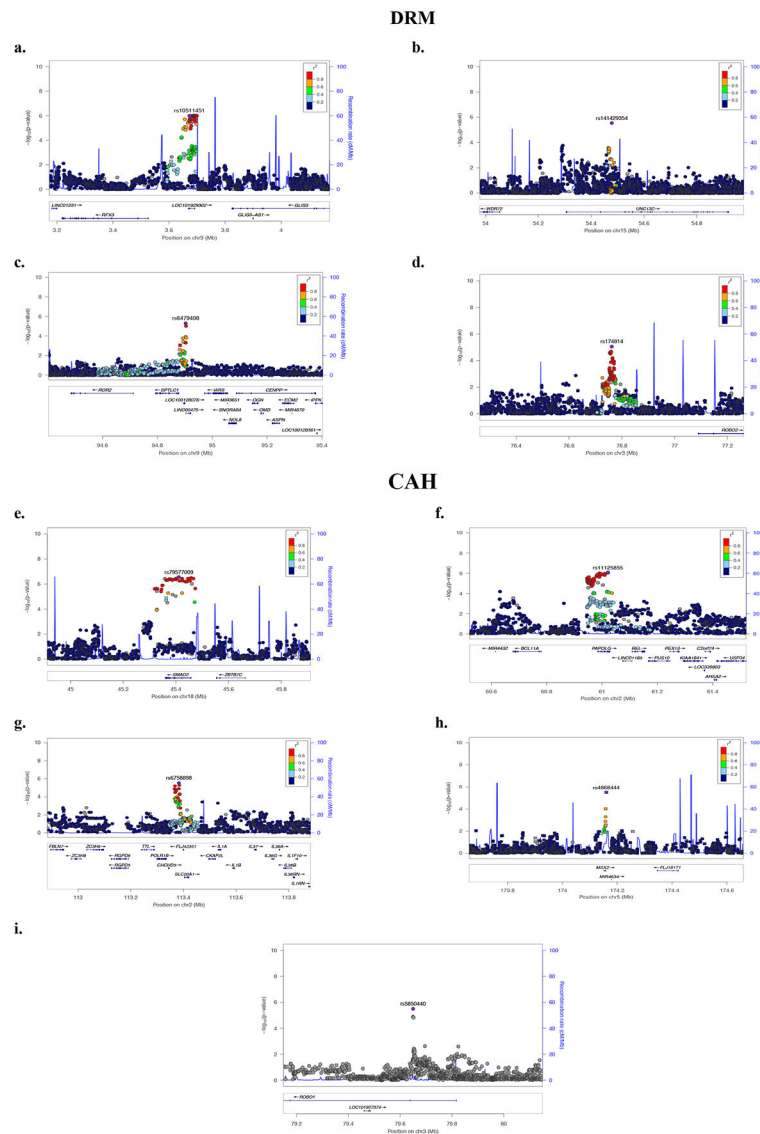


Figure.4. Regional Association Plots from (DRM) and (CAH) results. LocusZoom were used to generate plots for regions of Interest. **a.** suggestive locus near *GLIS3* on chromosome 9. **b.** suggestive locus near *WDR72* gene on chromosome 15. **c.** suggestive locus near *ROR2* and *OMD* on chromosome 9. **d.** suggestive locus near *ROBO2* on chromosome 3. **e.** suggestive locus near *SMAD2* on chromosome 18. **f.** suggestive locus near *REL* on chromosome 2. **g.** suggestive locus near *IL1A* and *IL1B* on chromosome 2. **h.** suggestive locus near *MSX2* on chromosome 5. **i.** suggestive locus near *ROBO1* on chromosome 3. The genome build used for the recombination rate was based on 1000 Genomes November 2014 EUR data. All of the gene positions and directions of transcription are annotated on the plots.

Table 1:

POFC Cohort (1,505 male, 2,074 female)

SITE	Subject Type				Total (%)
	<i>Unaffected relatives</i>		<i>Control</i>		
	Sex		Sex		
	M	F	M	F	
USA-COLORADO	15	20	--	--	35 (0.98)
USA-IOWA	90	162	87	146	485 (13.55)
USA-PITTSBURGH	31	42	45	65	190 (5.31)
USA-TEXAS	119	161	--	1	281 (7.85)
COLOMBIA	129	136	90	89	444 (12.4)
GUATEMALA	55	94	90	185	424 (11.85)
HUNGARY	142	163	205	231	741 (20.70)
ARGENTINA	82	185	16	30	313 (8.75)
PHILIPPINES	246	276	49	49	620 (17.32)
PUERTO RICO	11	21	3	11	46 (1.28)
	920	1267	585	807	
TOTAL					3,579
	2,187		1,392		

Table 2:

Distribution of dental anomalies by sex

Dental Anomaly	n (%)	Sex (%)		P-value*
		M (%)	F (%)	
Tooth Agenesis	170 (4.7)	80 (2.2)	90 (2.5)	0.175
Enamel Hypoplasia	282 (7.8)	136 (3.8)	146 (4)	0.029
Impaction	41 (1.14)	15 (0.42)	26 (0.73)	0.476
Rotation	2534 (70.80)	1144 (32)	1390 (38.8)	6.478e-09
Displacement	1459 (40.77)	685 (19.14)	774 (21.63)	1.003e-06
Mamelons	354 (9.89)	155 (4.33)	199 (5.56)	0.486
Supernumerary teeth	26 (0.73)	16 (0.45)	10 (0.28)	0.043
Dental Caries	2806 (78.40)	1,169 (41.66)	1,637 (58.34)	0.368

Theses *P*-values are based on χ^2 tests

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Table 3:

Distribution of dental anomalies by subject type

Dental Anomaly	n (%)	Subject Type (%)		P-value
		Unaffected relatives	Control	
Tooth Agenesis	170 (4.7)	116 (68.24)	54 (31.76)	0.051
Enamel Hypoplasia	282 (7.8)	177 (62.77)	105 (32.23)	0.551
Impaction	41 (1.14)	33 (80.49)	8 (19.51)	0.010
Rotation	2534 (70.80)	1,599 (63.10)	935 (36.90)	0.0016
Displacement	1459 (40.77)	893 (61.21)	566 (38.79)	0.919
Mamelon	354 (9.89)	233 (65.82)	121 (34.18)	0.055
Supernumerary teeth	26(0.73)	22 (84.62)	4 (15.38)	0.014
Dental Caries	2,806 (78.40)	1,756 (62.58%)	1,050 (37.42)	0.0018

* These *P*-values are based on χ^2 tests

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Table 4:

Correlation results between the different dental anomalies

	Tooth Agenesis	Enamel Hypoplasia	Impaction	Displaced	Rotation	Mamelons	Supernumerary Teeth
Tooth Agenesis		-0.002	0.038 *	0.007	0.045 **	-0.026	-0.004
Enamel Hypoplasia	-0.002		0.017	0.057 ***	0.049 **	-0.007	0.036
Impaction	0.038 *	0.017		0.028	0.029	-0.000	-0.009
Displaced	0.007	0.057 ***	0.028		0.459 ***	0.112 ***	0.009
Rotation	0.045 **	0.049 **	0.029	0.459 ***		0.192 ***	-0.003
Mamelon	-0.026	-0.007	-0.000	0.112 ***	0.192 ***		-0.006
Supernumerary	-0.004	0.036 *	-0.009	0.009	-0.003	-0.006	
Dental Caries	0.059 **	0.043 *	-0.007	0.019	0.029	-0.137	0.005

* $P < 0.05$ ** $P < 0.01$ *** $P < 0.001$.

Table 5:

Top GWAS hits for the Correlated Patterns of Dental anomalies and Dental Caries

SNP	CHR	BP	F*	Weights*	P	Effect Allele	MAF	Type
<i>AIR</i>								
chr3:64541255	3	64541255	5.914	0.536,0.758,0.413	1.00E-06	G	0.0693	Imputed
rs140220410	15	81018892	2.508	-0.474, -0.736,0.524	3.00E-06	A	0.05183	Imputed
rs9913511	17	9019184	11.08	0.340, -0.322,0.883	3.00E-06	T	0.0898	Genotyped
rs2251904	13	38770991	7.978	0.422, -0.027,0.906	4.00E-06	A	0.1618	Imputed
rs1838002	1	112705328	8.079	0.274, -0.635,0.721	6.00E-06	G	0.3036	Imputed
<i>HDR</i>								
rs12379966	9	3650939	9.534	0.135,0.825, -0.455	1.00E-06	G	0.1802	Genotyped
rs6479408	9	94903215	11.37	-0.01,0.792, -0.505	1.00E-06	G	0.4517	Imputed
rs141429354	15	54478011	8.714	-0.335,0.921, -0.087	3.00E-06	T	0.05562	Imputed
rs404727	20	55465434	10.12	0.407, -0.623,0.577	8.00E-06	A	0.251	Imputed
<i>DRM</i>								
rs10511451	9	3672822	11.13	-0.787,0.522,0.0586	1.00E-06	C	0.1631	Imputed
rs141429354	15	54478011	8.516	0.941, -0.089,0.290	3.00E-06	T	0.05562	Imputed
rs6479408	9	94903215	11.26	0.790, -0.508,0.029	5.00E-06	G	0.4517	Imputed
rs174814	3	76761788	9.488	0.196,0.769,0.680	9.00E-06	C	0.4874	Imputed
<i>CAH</i>								
rs79577009	18	45411221	11.13	-0.205, -0.003,0.981	2.87E-07	T	0.1441	Imputed
rs11125855	2	61021349	10.35	0.857,0.518, -0.197	8.82E-07	G	0.4123	Imputed
rs6758898	2	113382684	9.52	0.038,0.967,0.236	2.93E-06	G	0.4478	Imputed
rs4868444	5	174160113	9.484	0.048,0.956, -0.296	3.09E-06	T	0.07077	Imputed
rs5850440	3	79649799	9.422	-0.788, -0.545,0.361	3.37E-06	AT	0.3316	Imputed

* F-Statistic tests the significance of the canonical correlations

* Weights reflect the correlation coefficients for each individual trait

Tooth Agenesis, impaction and Rotation (AIR); Hypoplasia, displacement and rotation (HDR); Displacement, Rotation and Mamelon (DRM); and Dental caries, Agenesis and Hypoplasia (CAH).