**Supplemental materials and methods**

**Preparation of 10% stool suspension**. Stool specimens were weighed in 50 ml tubes with 6 mm glass beads and then the appropriate volume of PBS was added to make a 10% (w/v) suspension. The stool suspension was vortexed until a slurry was created and then centrifuged at 10,000 rpm for 20 min at 4oC. The supernatant was transferred to a new 50 ml centrifuge tube and spin at 10,000 rpm for 20 min at 4oC. Aliquots of the 10% stool suspension were frozen at −80oC until use.

**Rhesus macaque-specific immune response gene array**. Synthesis of cDNA was carried out with 500 ng of extracted total RNA using RT2 First Strand Kit (QIAGEN, Carlsbad, CA).

**Determination of HEV RNA titer by real-time PCR.**

200 μl of sample were used for extraction and isolated RNA was eluted in 70 μl of elution buffer. Aliquots of extracted RNA were frozen at -80oC until use. Synthesis of cDNA was conducted with 10 μl of extracted total RNA using SuperScript VILO cDNA Synthesis Kit (Life Technologies, Carlsbad, CA). The PCR reaction was performed using primer pairs targeting the ORF2/3 region (23). Primers and TaqMan probes were used at concentrations of 250 nM and 100 nM, respectively. The real-time PCR assay (detection limit: 20 WHO IU/ml) was performed according to the manufacturer’s instructions (Life Technologies, Carlsbad, CA). Briefly, real-time PCR was performed in a 50 μl reaction volume consisting of 10 μl of cDNA and 40 μl of master mix containing HEV-specific primers and probe, and Taqman DNA polymerase master mix (Life Technologies) using a ViiA7 PCR instrument (Life Technologies, Carlsbad, CA) in a 96-well format. The following conditions were used: 2 min at 50oC and 10 min at 95oC for initial denaturation and then followed by 55 cycles of amplification with denaturation at 95oC for 15 s, at 50oC for 1 min, and at 72oC for 1 min.