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Optimizing pyrazinamide for the treatment of tuberculosis

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Abstract

Pyrazinamide is a potent sterilizing agent that shortens the treatment duration needed to cure tuberculosis. It is synergistic with novel and existing drugs for tuberculosis. The dose of pyrazinamide that optimizes efficacy while remaining safe is uncertain, as is its potential role in shortening treatment duration further.

Pharmacokinetic data, sputum culture, and safety laboratory results were compiled from TBTC Studies 27 and 28 and PanACEA MAMS-TB, multi-center Phase 2 trials in which participants received rifampicin (range 10-35 mg/kg), pyrazinamide (range 20-30 mg/kg), plus two companion drugs. Pyrazinamide pharmacokinetic-pharmacodynamic (PK/PD) and PK-toxicity analyses were performed.

In TBTC studies (n=77), higher pyrazinamide maximum concentration (C_{max}) was associated with shorter time to culture conversion (TTCC) and higher probability of two-month culture conversion (p-value < 0.001). Parametric survival analyses showed that relationships varied geographically, with steeper PK-PD relationships seen among non-African than African

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participants. In PanACEA MAMS-TB (n=363), TTCC decreased as pyrazinamide C_{max} increased and varied by rifampicin AUC (p-value < 0.01). Modeling and simulation suggested that very high doses of pyrazinamide (>4500 mg) or increasing both pyrazinamide and rifampicin would be required to reach targets associated with treatment shortening. Combining all trials, liver toxicity was rare (3.9% with Grade 3 or higher liver function tests, LFT), and no relationship was seen between pyrazinamide C_{max} and LFT levels.

Pyrazinamide's microbiologic efficacy increases with increasing drug concentrations. Optimizing pyrazinamide alone, though, is unlikely to be sufficient to allow tuberculosis treatment shortening; rather, rifampicin dose would need to be increased in parallel.

Summary

The activity of pyrazinamide, a critical drug for tuberculosis treatment, increases as drug concentrations go up, but optimizing this drug alone is unlikely to result in treatment shortening. Rather, rifampicin dosing must go up in parallel.

Keywords

Pyrazinamide; pharmacokinetics; pharmacodynamics; toxicity

INTRODUCTION

Pyrazinamide is a potent sterilizing agent against *Mycobacterium tuberculosis*. It is unique in its activity against semi-dormant bacilli in acidic environments and against bacilli that remain viable despite unfavorable local conditions and antibiotic pressure, so-called “persisters” that must be eliminated to cure tuberculosis disease (1).

Currently-recommended “short-course” treatment for drug-sensitive tuberculosis remains lengthy. Isoniazid, rifampicin, pyrazinamide, and ethambutol are given for two months (intensive phase), followed by isoniazid and rifampicin for four months (continuation phase). The addition of pyrazinamide to rifampicin and isoniazid during the intensive phase of therapy allows for treatment shortening from 9 to 6 months (2, 3). Whether or not optimization of pyrazinamide—giving it for longer, increasing the dose, or pairing it with synergistic drugs-- can contribute to a regimen that cures tuberculosis more quickly is unknown (4).

In the World Health Organization (WHO) 1984 treatment guidelines, the recommended daily dose of pyrazinamide was 35 mg/kg (in keeping with studies that showed its treatment shortening benefit). In 2003, WHO reduced the recommended daily dose to 25 mg/kg; the rationale was left unstated (5). In some studies, low pyrazinamide exposures have been associated with worse outcomes (6, 7). However, pyrazinamide can cause liver injury at high doses given for prolonged periods (8, 9). The relationship between pyrazinamide exposures and either efficacy or, on the flip side, hepatotoxicity is not firmly established (10).

The Tuberculosis Trials Consortium (TBTC) is a multinational trials network funded by the US Centers for Disease Control and Prevention (CDC). TBTC conducted two clinical

Pyrazinamide PK and Minimal Inhibitory Concentration (MIC) Assessments

Intensive PK sampling was performed in a subset of participants in S27 and S28 in Uganda, South Africa, or the United States (n=72). PK sampling was performed pre-dose, and 1, 2, 6, 12, and 24 hours post-dose, and so that PK values would reflect steady state measures, these were collected at least 10 days after beginning treatment. Pyrazinamide PK analysis was performed using a validated gas chromatography assay with mass selective detection (14). For MIC determinations, isolates were stored at baseline. The pyrazinamide MIC of participants' isolates were determined using the BD BACTEC MGIT 320 system. Pyrazinamide MIC testing was performed using the standard method described in the package insert, except that in addition to the standard test concentration of 100 mcg/mL, three additional concentrations were tested (25, 50, and 75 mcg/mL) (15).

In MAMS-TB, PK sampling was performed in a subset of participants (20/arm) four weeks after commencing therapy. Samples were collected pre-dose and 1, 2, 3, 4, 6, 8, 12, and 24h post-dose. pyrazinamide bioanalysis was performed using high performance liquid chromatography (13). MICs were not measured.

Population PK/PD Modeling

Pyrazinamide with standard-dose rifampicin—Using PK data from S27 and S28, a nonlinear mixed effects (NLME) model was previously developed to characterize pyrazinamide population PK (Supplemental Table 3) (14). The existing model was used to generate post-hoc Bayesian estimates of secondary PK parameter values (AUC_{0-24hr} ; and maximum concentration (C_{max})) in NONMEM (version 7.4.3, ICON, Gaithersburg, MD) for each participant, taking into account an individual's PK data and characteristics (e.g. dose, weight, sex, age). Pharmacodynamic indices (PD indices) were calculated using MIC data (e.g. AUC_{0-24hr}/MIC or C_{max}/MIC). Cox proportional hazards regression analysis in R program (version: 3.6.1; package: survival 2.44) was performed to assess the relationship between pyrazinamide PK and PD indices vs. outcomes (time to sputum culture conversion, probability of culture conversion by eight weeks of treatment). The PK parameter with the best fit was included in final models. Variables with p-values less than < 0.1 in univariate models or factors known to be associated with culture conversion were tested in multivariate models (e.g. sex, ethnicity, cavity status, regimen). After exploring relationships with Cox modeling, we then proceeded to parametric survival analysis, a more sophisticated modeling technique that allows for evaluation of predictors' influence on both the shape and scale of the survival curve (16) (Supplemental materials, Section A). Hazard function was defined by scale and shape parameters; covariates were tested on scale and shape parameters in analyses.

Pyrazinamide with standard vs. higher-dose rifampicin—A population PK model for pyrazinamide was developed based on PK data from MAMS-TB using NONMEM software to generate primary PK parameters (Supplemental materials, Section A, Supplemental Table 4, Supplemental Figure 1). The relationship between pyrazinamide C_{max} or AUC_{0-24hr} and treatment outcomes was assessed in similar fashion to S27 and S28 in R program (version). A number of covariates were evaluated for inclusion in multivariate models (baseline mycobacterial load, weight, HIV status, age, sex, radiographic findings,

rifampicin PK). Data were analyzed using parametric survival analyses (Supplemental materials, Section A). PK-PD assessments were restricted to twelve weeks.

Dosing and efficacy simulations—Final survival models with covariates for S27 and S28 and, separately, for MAMS-TB, were used to simulate scenarios to investigate the probability of 8- or 12-week culture conversion reaching certain targets (e.g. 90% and 95%) for different dosing strategies, assuming that high rates of early culture conversion are a prerequisite for a regimen that will effectively shorten treatment (Supplemental materials, Section A). Simulations of 500 trials were conducted for each scenario.

PK-toxicity Modeling

We evaluated the relationship between pyrazinamide PK parameters (e.g. C_{max} or AUC) and change in LFTs from baseline on the basis of their relevance. Linear regression was conducted to measure the association between pyrazinamide C_{max} and individual maximal LFT values in R program (version version: 3.6.1). Multiple R-squared and p-value were calculated individually for ALT, AST and total bilirubin.

RESULTS

Study Population

Table 1 shows demographic and dose information for the 72 participants in the PK substudies of S27 and S28. In MAMS-TB, data for 363 participants were available and used in safety assessments. 96 individuals participated in the PK substudy and had concentration data sufficient to produce PK estimates (characteristics in Table 2); data for 86 subjects had dose and time of dose recorded adequately for population PK analysis.

PK and MIC Results

444 plasma samples were used in PK analyses from S27 and S28. Post-hoc Bayesian estimates of pyrazinamide PK parameters and PD indices are in Table 3. Predicted PZA C_{max} ranged from 15 to 55 ug/mL; only 18 (25%) of participants had C_{max} above 35 ug/mL. Pyrazinamide MICs were 25, 50 and 75 µg/ml (27, 29 and 4 participants, respectively). 846 plasma PK samples from MAMS-TB were used in PK assessments (Supplemental Figure 1). Predicted pyrazinamide C_{max} ranged from 30 to 51 ug/mL; 55 (64.0%) of participants had C_{max} above 35 ug/mL (Table 3). Time to maximum concentration (T_{max}) median was 4h (range 3-6h). Observed rifampin C_{max} varied depending on the dose level. Parameter estimates for the final population PK models are in Supplemental Tables S3 and S4 (14).

PK-PD of pyrazinamide, with Standard-dose rifampicin (TBTC Trials)

Time to culture conversion.—In multivariate Cox regression analyses, the only significant predictors of time to culture conversion were pyrazinamide PK parameters (C_{max} p=0.046 or AUC p=0.015). In the more complex parametric survival analyses, pyrazinamide C_{max} and geographic site were the only covariates that improved the fit of the Weibull time-to-culture-conversion model significantly (pyrazinamide C_{max} influenced the shape parameter, and geographic site influenced both scale and shape parameters) (Figure

1, Supplemental Figure 2a, Supplemental Table 5). Efficacy improved over the full range of clinically-observed values of pyrazinamide C_{max}, without plateau (Figure 2, top).

Probability of culture conversion.—There was a positive relationship between pyrazinamide C_{max} and two-month culture conversion in non-African but not African participants; however, there was a positive relationship between C_{max} and probability of culture conversion by 3 months across groups (Supplemental Table 6). Simulations show that to achieve culture conversion by 2 months in 90% of participants, pyrazinamide C_{max} of 43 and 93 ug/mL would be needed for non-African and African participants, respectively. Average daily doses of 1800mg and 4600mg are required to achieve these targets (Table 4A).

PK-PD of pyrazinamide, with Higher-dose rifampicin (PanACEA MAMS-TB trial)

In Cox regression models, pyrazinamide C_{max} or AUC_{0-24hr} were associated positively with time to culture conversion ($p=0.0067$ and 0.73 , respectively). In parametric survival analysis, several factors were correlated with the scale parameter in the Weibull model (age, ethnicity, weight, baseline mycobacterial load, HIV status, pyrazinamide C_{max}, rifampicin AUC_{0-24hr}), and several factors correlated with the shape parameter (age, ethnicity, baseline mycobacterial load, pyrazinamide C_{max}). The final model, which included rifampicin AUC_{0-24hr} and pyrazinamide C_{max} on scale and pyrazinamide C_{max} on shape, demonstrated a significant exposure-response relationship for pyrazinamide that depended on rifampicin exposures (Supplemental Tables 7 and 8, Figure 3, Supplemental Figure 2b). Table 4B shows the doses of pyrazinamide and exposures of rifampicin needed to achieve 2-month or 3-month culture conversion proportions of 90, or 95% on liquid media. For context, in MAMS-TB, doses of 10, 20, and 35 mg/kg of rifampicin achieved median AUC_{0-24hr} values of 20.6, 61.7, and 164.2 ug*hr/mL (13).

PK-toxicity Analysis

One of 72 participants in the TBTC trials and 12 of 363 participants in MAMS-TB had LFT values greater than 3 times the upper limit of normal during TB treatment. In TBTC trials, no association could be shown between pyrazinamide C_{max} and AST or total bilirubin (Multiple R-squared = 0.023 and 0.00070, p -value = 0.19 and 0.82, respectively) (Figure 4A); Median C_{max} in those who had LFT>3x normal was 28.9 ug/mL versus 29.7 ug/mL in those who did not. Similarly, there was not an association between pyrazinamide C_{max} and ALT, or AST, in MAMS-TB (Multiple R-squared = 0.00063, 0.00026 and 0.019, p -value = 0.64, 0.76 and 0.16, respectively) (Figure 4B). Median C_{max} was 35.3 ug/mL in those who had LFT>3x normal and 37.2 ug/mL in those who did not.

DISCUSSION

Pyrazinamide is a standard component of first-line TB treatment, yet the ‘right dose’ is not established. In this study using data from three international Phase 2 clinical trials, higher concentrations of pyrazinamide were associated with higher culture conversion rates at 2 and 3 months of treatment. These analyses suggest that current dosing may be insufficient to maximize efficacy (17). In the trials that enrolled from geographically-diverse settings,

parametric survival modeling revealed that PK-PD relationships differed for participants from African vs. non-African sites. To achieve targets associated with treatment-shortening, drug doses that are beyond the range of tolerability would likely be needed in African patients in the absence of other new drugs. Modeling and simulation showed that increasing doses of both rifampicin and pyrazinamide appears to be a more promising strategy. The range of pyrazinamide concentrations was broad, yet elevations in liver enzymes were rare, and there was not an association between pyrazinamide levels and hepatotoxicity.

Pyrazinamide is an important sterilizing agent. Early on, it was highly effective in two-drug combinations with isoniazid, provided it was given at a high enough dose and for sufficient duration (8, 18-22). Following the discovery of rifampicin, adding pyrazinamide to rifampicin-containing regimens reduced relapses (2, 23-25), and pyrazinamide became an essential part of current “short-course” treatment. Giving it during the first eight weeks allows for the shortening of treatment from nine to six months. In the trials demonstrating its treatment shortening activity, though, the doses given were 30-40 mg/kg daily, not the currently-recommended 20-25 mg/kg for adults (26-28).

In our study, culture conversion rates increased with increasing pyrazinamide exposure both when pyrazinamide was combined with standard-dose rifampicin or higher-dose rifampicin, and the best activity was seen when exposures of both drugs were high, demonstrating that there was an observable exposure-response relationship even when the companion drug was a potent sterilizing agent given at a high dose. Interestingly, in the TBTC studies, PK-PD relationships were different for non-African and African participants. African participants tended to have higher baseline extent of disease (higher likelihood of 3+ sputum smear grade or large lung cavities) than non-Africans, but these factors were not significant in our multivariate models, and there are likely other unobserved factors contributing to lower treatment response. This same phenomenon of lower treatment response, even after adjusting for known risk factors, was seen in the larger S28 study population and in TBTC Studies 29 and 29X and remains unexplained (29, 30). While optimizing pyrazinamide in the context of first-line therapy is important, pyrazinamide also has a role in multidrug-resistant TB treatment; treatment outcomes are significantly worse if the MDR-TB strain is pyrazinamide-resistant (28). Pyrazinamide also enhances the activity of new and investigational drugs, namely bedaquiline, delamanid, and pretomanid, so optimization of pyrazinamide may be valuable in multiple contexts (31-33).

Our study was not the first modern study to find that pyrazinamide PK influenced treatment outcomes. In our study, C_{max} was the covariate identified in the final PK-PD model as the most informative. However, we note that C_{max} and AUC are highly correlated and each has a strong association with outcomes; depending on the sampling strategy of a given study, which influences how well that parameter is estimated, one might have a modestly stronger correlation or better precision. For example, single samples can sometimes fail to capture C_{max} well. In the TBTC Studies, CV% for C_{max} and AUC were similar at 23.1% and 26.1%, respectively, so variability in these estimates was similar. In a study in Botswana, after adjusting for HIV infection and CD4 cell count, patients with pyrazinamide C_{max} less than 35 ug/mL (a putative pyrazinamide PK target) had a 3.4-fold higher risk of poor treatment outcome (6). In South Africa, pyrazinamide AUC was a top

predictor of poor long-term outcomes (7). In children with and without HIV in India, low pyrazinamide and rifampicin C_{max} were associated with unfavourable outcomes (34). In a recent meta-analysis, low pyrazinamide concentrations were shown to increase the risk of poor outcomes with relative risk of 1.73 (35). At currently-recommended doses, a high proportion of patients do not have drug concentrations that reach 35 ug/mL, let alone a proposed alternative target of 58 ug/mL (10), and the evidence base for selection of a 20-25 mg/kg dose is limited. Likely the optimal dose for this drug lies somewhere between 35 and 45 mg/kg. Higher doses would produce exposures exceeding 5000 ug*h/mL (see below) in some patients (29, 36). At the current dose, we are undertreating many.

Pyrazinamide commonly causes arthralgias, but its most dreaded toxicity is liver injury. In early studies, doses of at least 40-50 mg/kg given for 24 weeks or longer caused unacceptable rates of liver toxicity (5-10%), while rates were substantially lower (2-5%) if the duration or dose was reduced (37). Currently, doses of 30-40 mg/kg daily are well-tolerated as part of MDR-TB treatment (38). In a meta-analysis involving 4490 individuals, risk of liver toxicity did not appear to increase as a function of drug exposure until exposures were quite high (weekly AUC of > 5000 ug*h/mL) (39). Pyrazinamide toxicity appears to be idiosyncratic up until a point, after which dose-related increases in liver toxicity are seen (10). The mechanism for and contributing factors to pyrazinamide-associated liver injury are not clearly understood and may differ for different companion drugs (40). Consistent with previous reports, in our study, median weekly AUC were 2100 (TBTC) and 2310 (PanACEA MAMS-TB) ug*h/mL, and liver injury was rare and not related to exposure in the ranges seen.

Our study has limitations. Because of differences in culture methodology and covariates collected between TBTC and PanACEA trials, we could not combine PK-efficacy data into a single model. Model-predicted doses assumed proportional dose effects at doses higher than those observed, which may not be the case. In our parametric survival analyses, we did not consider interval censoring. The concentration that increases risk of liver toxicity could not be determined, as liver toxicity was rare and the dose range limited. Adjusting pyrazinamide PK parameters for isolates' MIC values did not improve model fit; likely this is because the MIC range was narrow, MICs were not measured precisely, and many participants did not have MIC data. In one TBTC trial, some patients had intermittent dosing- sensitivity analyses suggested removing those patients did not change model parameters or fit. Lastly, predicting an effective treatment-shortening regimen using microbiologic data from Phase 2 trials is an uncertain science; while 90-95% culture conversion on solid media by 8 weeks of treatment has been proposed, there are no well-validated prediction models using liquid culture (41).

It is important that the dose of each drug in a TB treatment regimen be optimized. Higher-dose rifampicin and pyrazinamide have the potential to shorten TB treatment. Simply prolonging the duration over which pyrazinamide is given was not sufficient to reduce treatment duration from 6 to 4 months in historical trials (4). Using parametric survival modeling and trial simulations, we discovered that increasing just the pyrazinamide dose does not seem as though it will improve outcomes in the hardest-to-treat patients (29). Indeed, the predicted doses of pyrazinamide that would be required as part of the

standard regimen to produce rapid and sustained culture conversion (90% conversion by two months, for example) for all patients were high and would not be safe. Whether or not an enhanced multidrug regimen containing high-dose rifampicin (e.g. 35 mg/kg) and higher-dose pyrazinamide (e.g. 30-40 mg/kg) will be sufficient to meaningfully reduce TB treatment duration must be explored prospectively, with attention to safety and tolerability.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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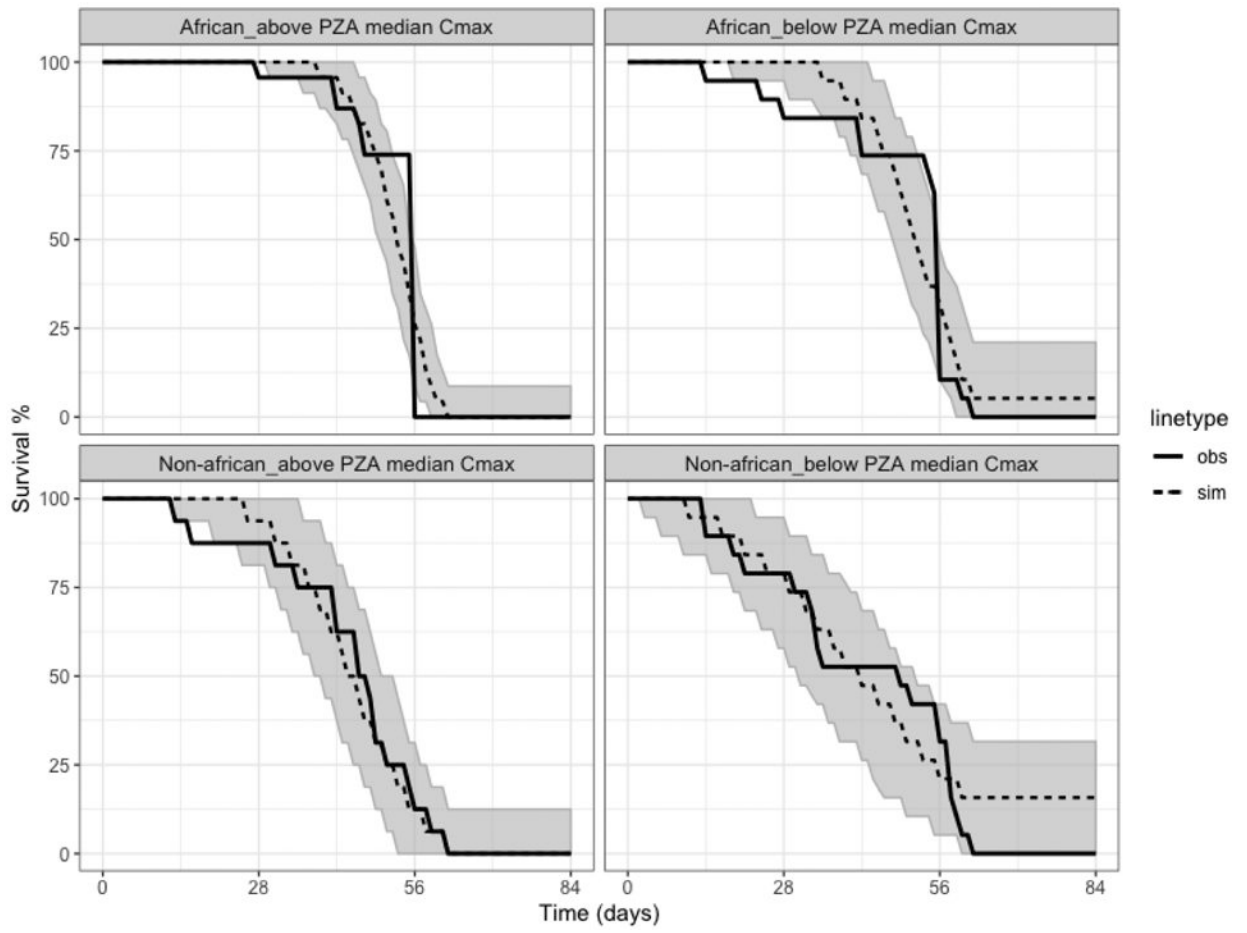
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REFERENCES

1. Zhang Y, Yew WW, Barer MR. Targeting persisters for tuberculosis control. *Antimicrobial Agents and Chemotherapy*. 2012;56(5):2223–30. [PubMed: 22391538]
2. Controlled trial of 6-month and 8-month regimens in the treatment of pulmonary tuberculosis. First report. *The American Review of Respiratory Disease*. 1978;118(2):219–28. [PubMed: 100029]
3. Controlled clinical trial of short-course (6-month) regimens of chemotherapy for treatment of pulmonary tuberculosis. *Lancet (London, England)*. 1972;1(7760):1079–85.
4. Clinical trial of six-month and four-month regimens of chemotherapy in the treatment of pulmonary tuberculosis: the results up to 30 months. *Tubercle*. 1981;62(2):95–102. [PubMed: 7029838]
5. World Health O. Treatment of tuberculosis. Guidelines for national programmes. WHO/CDS/TB/2003.313. 3 ed. Geneva, Switzerland 2003.
6. Chideya S, Winston CA, Peloquin CA, Bradford WZ, Hopewell PC, Wells CD, et al. Isoniazid, rifampin, ethambutol, and pyrazinamide pharmacokinetics and treatment outcomes among a predominantly HIV-infected cohort of adults with tuberculosis from Botswana. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America*. 2009;48(12):1685–94. [PubMed: 19432554]
7. Pasipanodya JG, McIlleron H, Burger A, Wash PA, Smith P, Gumbo T. Serum drug concentrations predictive of pulmonary tuberculosis outcomes. *The Journal of infectious diseases*. 2013;208(9):1464–73. [PubMed: 23901086]
8. Muschenheim C, Organick A, McCune RM Jr., Batten J, Deuschle K, Tompsett R, et al. Pyrazinamide-isoniazid in tuberculosis. III. Observations with reduced dosage of pyrazinamide. *American review of tuberculosis*. 1955;72(6):851–5. [PubMed: 13268857]
9. SEQUENTIAL use of paired combinations of isoniazid, streptomycin, para-aminosalicylic acid, and pyrazinamide. A United States Public Health Service tuberculosis therapy trial. *The American Review of Respiratory Disease*. 1959;80:627–40. [PubMed: 13856609]

10. Pasipanodya JG, Gumbo T. Clinical and toxicodynamic evidence that high-dose pyrazinamide is not more hepatotoxic than the low doses currently used. *Antimicrobial Agents and Chemotherapy*. 2010;54(7):2847–54. [PubMed: 20439617]
11. Burman WJ, Goldberg S, Johnson JL, Muzanye G, Engle M, Mosher AW, et al. Moxifloxacin versus ethambutol in the first 2 months of treatment for pulmonary tuberculosis. *American journal of respiratory and critical care medicine*. 2006;174(3):331–8. [PubMed: 16675781]
12. Dorman SE, Johnson JL, Goldberg S, Muzanye G, Padayatchi N, Bozeman L, et al. Substitution of moxifloxacin for isoniazid during intensive phase treatment of pulmonary tuberculosis. *American journal of respiratory and critical care medicine*. 2009;180(3):273–80. [PubMed: 19406981]
13. Boeree MJ, Heinrich N, Aarnoutse R, Diacon AH, Dawson R, Rehal S, et al. High-dose rifampicin, moxifloxacin, and SQ109 for treating tuberculosis: a multi-arm, multi-stage randomised controlled trial. *The Lancet Infectious diseases*. 2017;17(1):39–49.
14. Alsultan A, Savic R, Dooley KE, Weiner M, Whitworth W, Mac Kenzie WR, et al. Population Pharmacokinetics of Pyrazinamide in Patients with Tuberculosis. *Antimicrobial Agents and Chemotherapy*. 2017;61(6):10.1128/AAC.02625-16. Print 2017 Jun.
15. Whitfield MG, Warren RM, Streicher EM, Sampson SL, Sirgel FA, van Helden PD, et al. Mycobacterium tuberculosis pncA Polymorphisms That Do Not Confer Pyrazinamide Resistance at a Breakpoint Concentration of 100 Micrograms per Milliliter in MGIT. *J Clin Microbiol*. 2015;53(11):3633–5. [PubMed: 26292310]
16. Rudolph JE, Cole SR, Edwards JK. Parametric assumptions equate to hidden observations: comparing the efficiency of nonparametric and parametric models for estimating time to AIDS or death in a cohort of HIV-positive women. *BMC medical research methodology*. 2018;18(1):142–8. [PubMed: 30453971]
17. Phillips PP, Fielding K, Nunn AJ. An evaluation of culture results during treatment for tuberculosis as surrogate endpoints for treatment failure and relapse. *PLoS One*. 2013;8(5):e63840. [PubMed: 23667677]
18. Muschenheim C, McDermott W, McCune R, Deuschle K, Ormond L, Tompsett R. Pyrazinamide-isoniazid in tuberculosis. I. Results in 58 patients with pulmonary lesions one year after the start of therapy. *American review of tuberculosis*. 1954;70(4):743–7. [PubMed: 13197750]
19. Campagna M, Calix A, Hauser G. Observations on the combined use of pyrazinamide (Aldinamide) and isoniazid in the treatment of pulmonary tuberculosis; a clinical study. *American review of tuberculosis*. 1954;69(3):334–50. [PubMed: 13138868]
20. Campagna M, Hauser G, Greenberg HB. The eradication of Mycobacterium tuberculosis from the sputum of patients treated with pyrazinamide and isoniazid. A ten-year follow-up study. *The American Review of Respiratory Disease*. 1962;86:636–9. [PubMed: 14018084]
21. Schwartz WS, Moyer RE. The chemotherapy of pulmonary tuberculosis with pyrazinamide used alone and in combination with streptomycin, para-aminosalicylic acid, or isoniazid. *American review of tuberculosis*. 1954;70(3):413–22. [PubMed: 13189056]
22. McDermott W, Ormond L, Muschenheim C, Deuschle K, McCune RM, Tompsett R. Pyrazinamide-isoniazid in tuberculosis. *American review of tuberculosis*. 1954;69(3):319–33. [PubMed: 13138867]
23. Controlled trial of 6-month and 8-month regimens in the treatment of pulmonary tuberculosis: the results up to 24 months. *Tubercle*. 1979;60(4):201–10. [PubMed: 396701]
24. Controlled trial of four thrice-weekly regimens and a daily regimen all given for 6 months for pulmonary tuberculosis. *Lancet (London, England)*. 1981;1(8213):171–4.
25. Controlled trial of 4 three-times-weekly regimens and a daily regimen all given for 6 months for pulmonary tuberculosis. Second report: the results up to 24 months. *Hong Kong Chest Service/ British Medical Research Council. Tubercle*. 1982;63(2):89–98. [PubMed: 6758252]
26. Combs DL, O'Brien RJ, Geiter LJ. USPHS Tuberculosis Short-Course Chemotherapy Trial 21: effectiveness, toxicity, and acceptability. The report of final results. *Annals of Internal Medicine*. 1990;112(6):397–406. [PubMed: 2155569]
27. Nahid P, Dorman SE, Alipanah N, Barry PM, Brozek JL, Cattamanchi A, et al. Official American Thoracic Society/Centers for Disease Control and Prevention/Infectious Diseases Society of

- American Clinical Practice Guidelines: Treatment of Drug-Susceptible Tuberculosis. *Clin Infect Dis*. 2016;63(7):e147–e95. [PubMed: 27516382]
28. World Health O.WHO treatment guidelines for drug-resistant tuberculosis: 2016 update. 2016. Contract No.: Report.
 29. Mac Kenzie WR, Heilig CM, Bozeman L, Johnson JL, Muzanye G, Dunbar D, et al.Geographic differences in time to culture conversion in liquid media: Tuberculosis Trials Consortium study 28. Culture conversion is delayed in Africa. *PLoS one*. 2011;6(4):e18358. [PubMed: 21494548]
 30. Savic RM, Weiner M, MacKenzie WR, Engle M, Whitworth WC, Johnson JL, et al.Defining the optimal dose of rifapentine for pulmonary tuberculosis: Exposure-response relations from two phase II clinical trials. *Clinical pharmacology and therapeutics*. 2017;102(2):321–31. [PubMed: 28124478]
 31. Nuermberger E, Rosenthal I, Tyagi S, Williams KN, Almeida D, Peloquin CA, et al.Combination chemotherapy with the nitroimidazopyran PA-824 and first-line drugs in a murine model of tuberculosis. *Antimicrobial Agents and Chemotherapy*. 2006;50(8):2621–5. [PubMed: 16870750]
 32. Williams K, Minkowski A, Amoabeng O, Peloquin CA, Taylor D, Andries K, et al.Sterilizing activities of novel combinations lacking first- and second-line drugs in a murine model of tuberculosis. *Antimicrobial Agents and Chemotherapy*. 2012;56(6):3114–20. [PubMed: 22470112]
 33. Tasneen R, Li SY, Peloquin CA, Taylor D, Williams KN, Andries K, et al.Sterilizing activity of novel TMC207- and PA-824-containing regimens in a murine model of tuberculosis. *Antimicrobial Agents and Chemotherapy*. 2011;55(12):5485–92. [PubMed: 21930883]
 34. Ramachandran G, Kumar AK, Kannan T, Bhavani PK, Kumar SR, Gangadevi NP, et al.Low Serum Concentrations of Rifampicin and Pyrazinamide Associated with Poor Treatment Outcomes in Children with Tuberculosis Related to HIV Status. *The Pediatric infectious disease journal*. 2016;35(5):530–4. [PubMed: 26825153]
 35. Perumal R, Naidoo K, Naidoo A, Ramachandran G, Requena-Mendez A, Sekaggya-Wiltshire C, et al.A systematic review and meta-analysis of first-line tuberculosis drug concentrations and treatment outcomes. *Int J Tuberc Lung Dis*. 2020;24(1):48–64.
 36. Ahmad Z, Fraig MM, Bisson GP, Nuermberger EL, Grosset JH, Karakousis PC. Dose-dependent activity of pyrazinamide in animal models of intracellular and extracellular tuberculosis infections. *Antimicrobial Agents and Chemotherapy*. 2011;55(4):1527–32. [PubMed: 21282447]
 37. United States Public Health S. Hepatic toxicity of pyrazinamide used with isoniazid in tuberculous patients. 1959;3:371.
 38. Borisov S, Danila E, Maryandyshev A, Dalcolmo M, Miliauskas S, Kuksa L, et al.Surveillance of adverse events in the treatment of drug-resistant tuberculosis: first global report. *Eur Respir J*. 2019;54(6).
 39. Sahota T, Della Pasqua O. Feasibility of a fixed-dose regimen of pyrazinamide and its impact on systemic drug exposure and liver safety in patients with tuberculosis. *Antimicrobial Agents and Chemotherapy*. 2012;56(11):5442–9. [PubMed: 22777045]
 40. Tostmann A, Boeree MJ, Aarnoutse RE, de Lange WC, van der Ven AJ, Dekhuijzen R. Antituberculosis drug-induced hepatotoxicity: concise up-to-date review. *J Gastroenterol Hepatol*. 2008;23(2):192–202. [PubMed: 17995946]
 41. Bonnett LJ, Ken-Dror G, Koh GCKW, Davies GR. Comparing the Efficacy of Drug Regimens for Pulmonary Tuberculosis: Meta-analysis of Endpoints in Early-Phase Clinical Trials. *Clinical infectious diseases : an official publication of the Infectious Diseases Society of America*. 2017;65(1):46–54. [PubMed: 28402396]



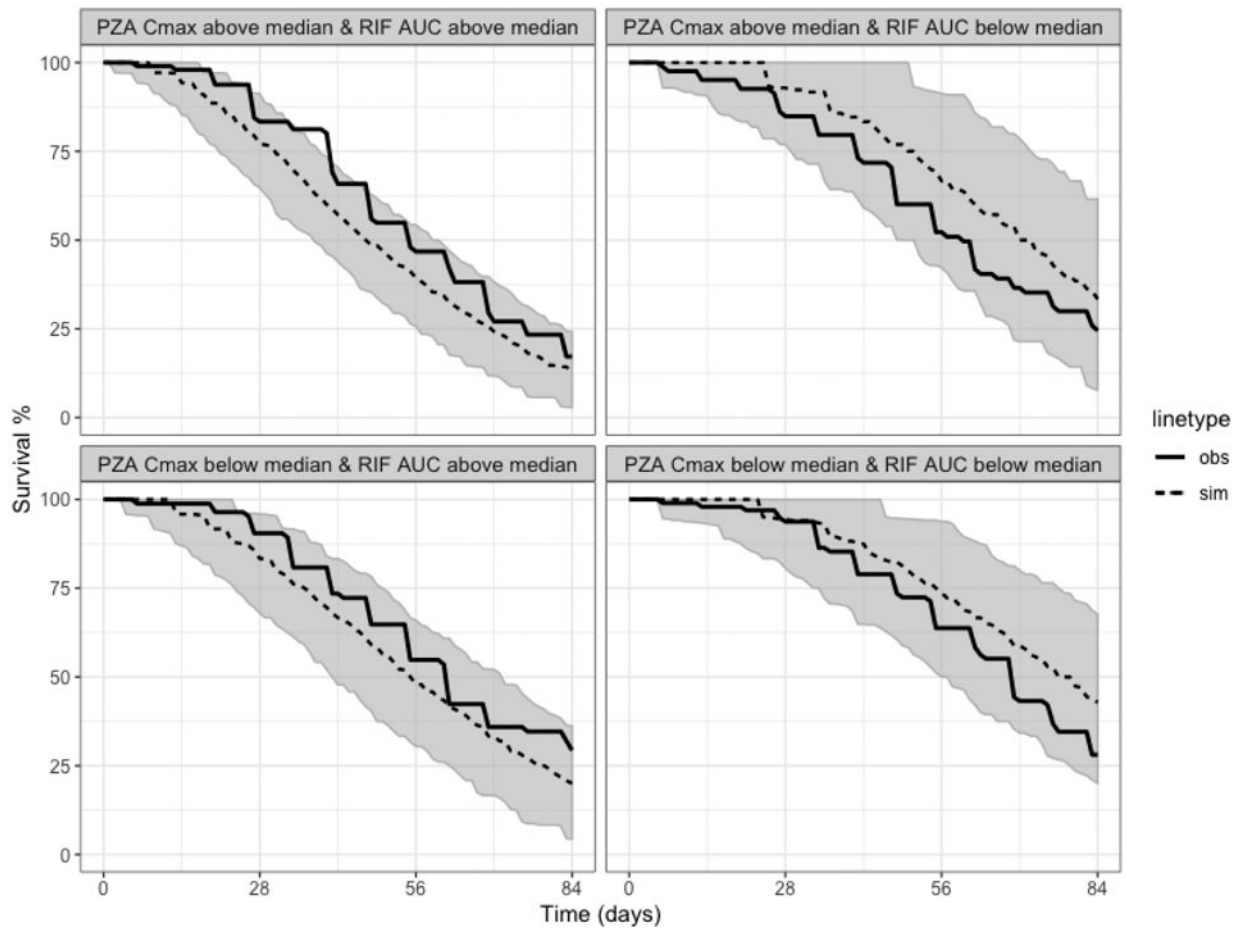


Figure 1. Visual predictive checks of the PK/outcome Weibull survival models for TBTC S27/28 (A) and PanANCEA MAMS trials stratified by covariates identified in the survival model (B).

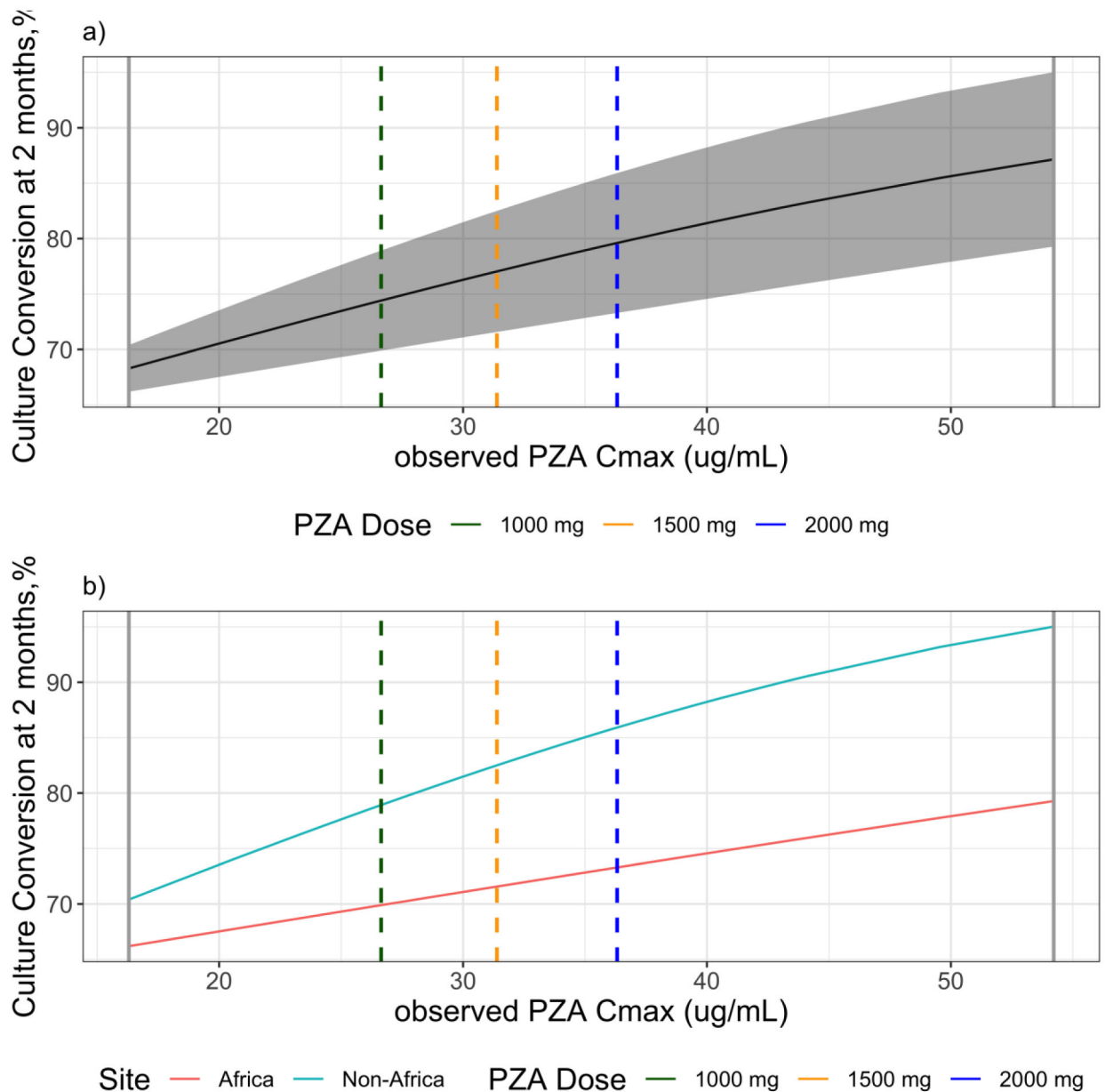


Figure 2.

Among participants taking combination treatment including pyrazinamide and standard-dose rifampicin in TBTC Studies 27 and 28, the relationship between maximum drug concentration (mcg/mL) and proportion with culture conversion to negative by 2 months of treatment. The median Cmax with drug doses of 1000mg, 1500mg, and 2000mg are shown in the vertical dash lines, and the observed range of Cmax values is contained within the vertical grey lines. In Panel a, the grey ribbon shows the 90% confidence interval of the proportion with culture conversion to negative with the black line as the median. In Panel b, the relationship between Cmax and two-month culture conversion is shown for African vs. non-African participants.

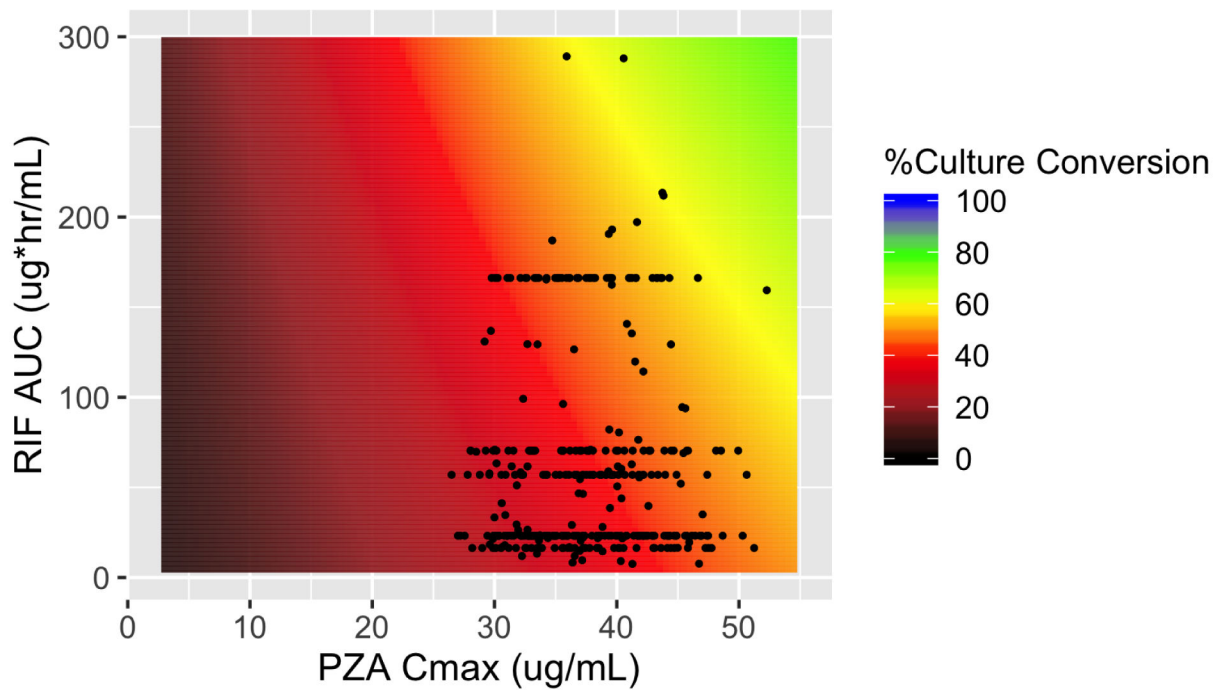


Figure 3.

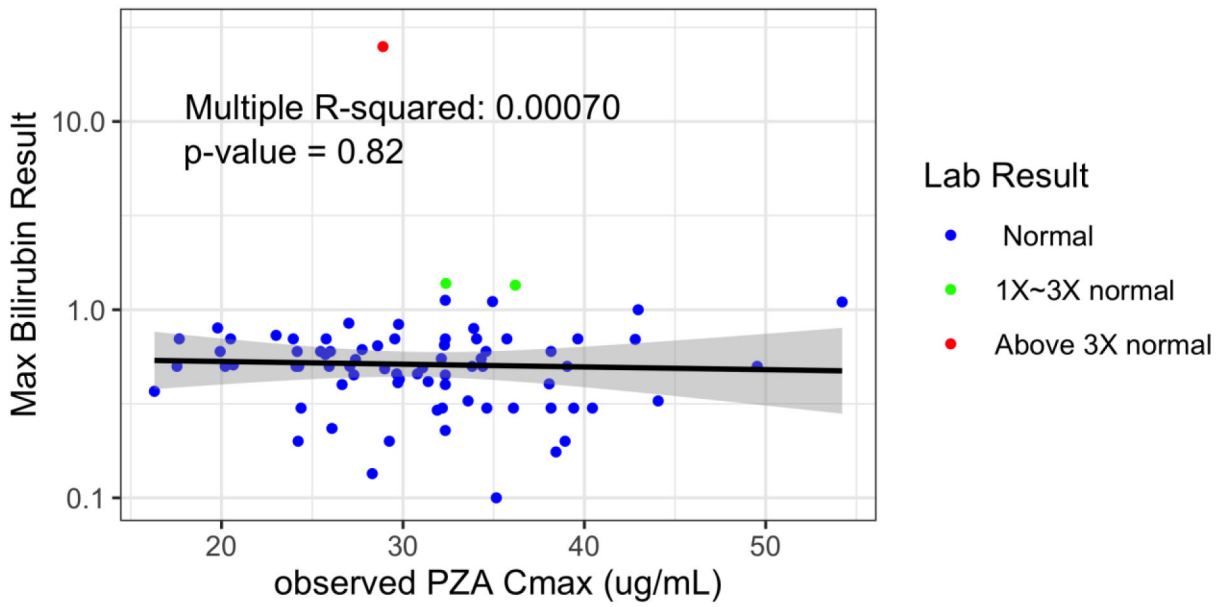
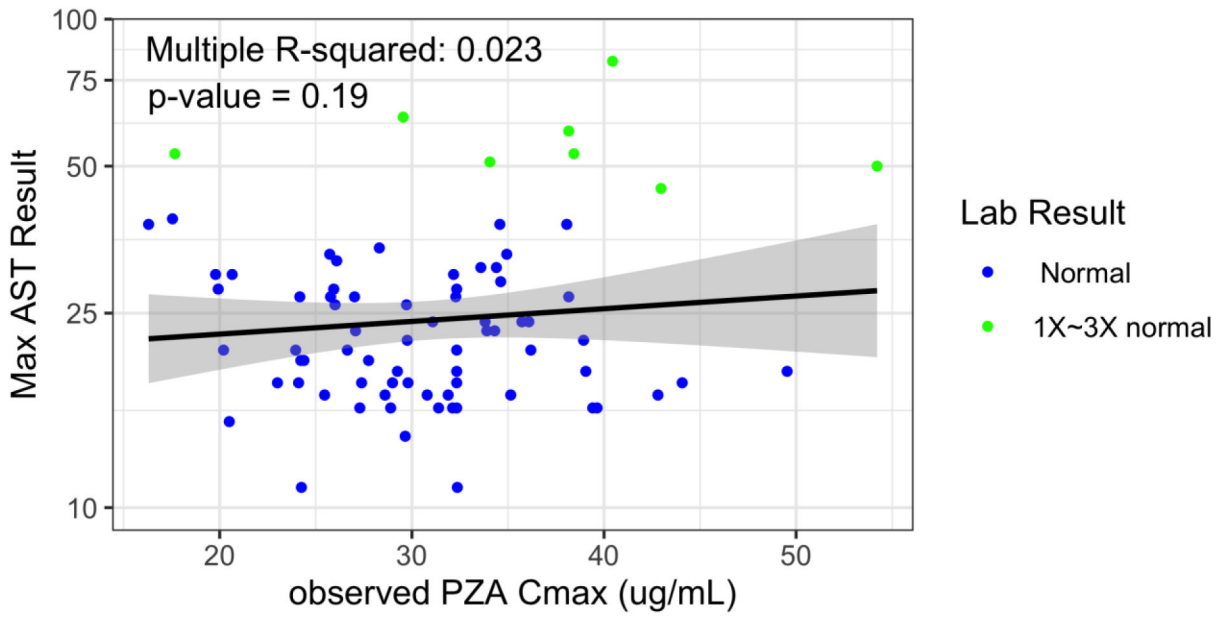
Simulated relationship between culture conversion on liquid medium by 2 months of treatment with maximum concentrations (C_{max}) of pyrazinamide and area under the curve (AUC_{0-24hr}) of rifampicin from PanACEA MAMS trial. Black dots are C_{max} values of pyrazinamide and rifampicin obtained by population PK models. Notes: for patients whose PK concentrations were not measured, pyrazinamide C_{max} values were imputed using the population PK model and rifampicin AUC values were imputed using geometric mean values of the regimen taken.

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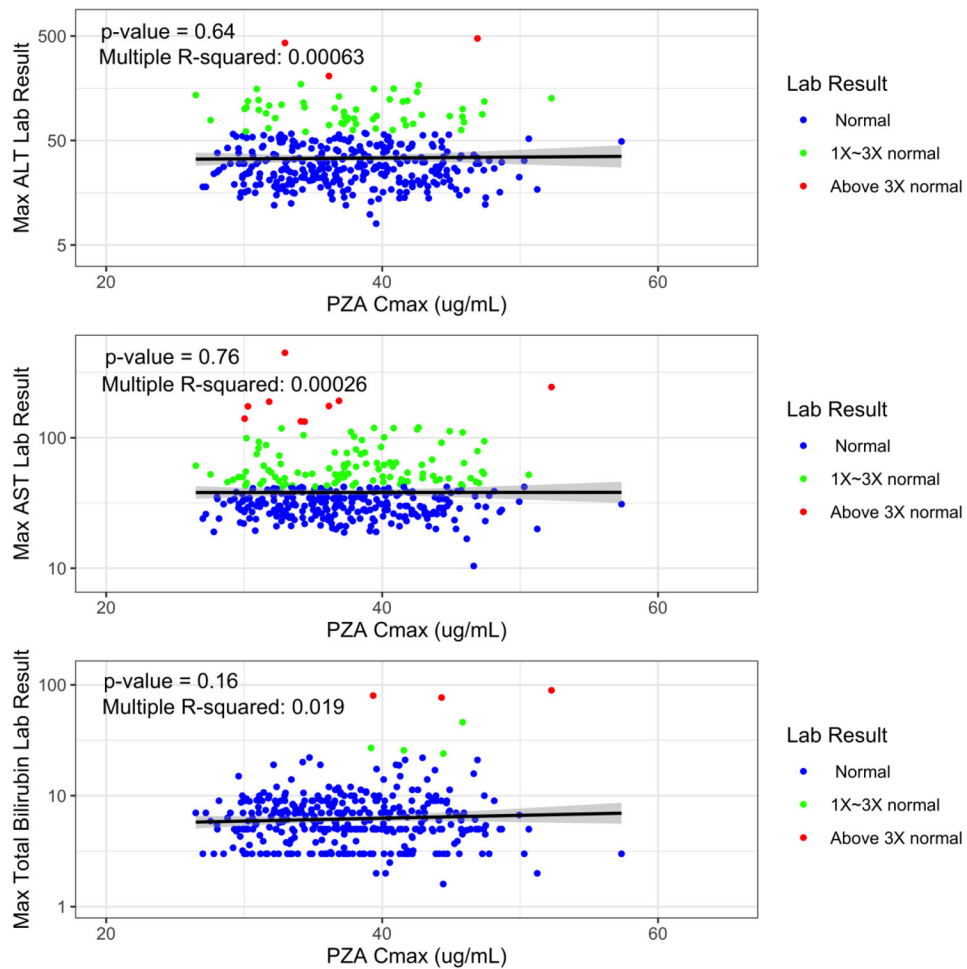


Figure 4. (A) Distribution and regression of individual maximal AST (left) and maximal total bilirubin (right) versus observed Cmax of pyrazinamide in TBTC 27 and 28 trials. (B) Distribution and regression of individual maximal ALT (left), individual maximal AST (middle), and individual total bilirubin (right) versus of pyrazinamide Cmax in PanANCEA MAMS trial.

Table 1.

Demographic, treatment, and clinical characteristics among participants enrolled in the pharmacokinetic sub-study of Tuberculosis Trials Consortium Studies 27 and 28

	Study 27 participants (n=9)	Study 28 participants (n=63)	All study participants (n=72)
Demographic Factors			
Age, years	50 (37-55)	33(26-38)	33 (27-42)
Female sex	1 (11%)	12 (19%)	13 (18%)
Enrollment from Africa *	0 (0%)	37 (59%)	37 (51%)
Race			
Black	2 (22%)	40 (63%)	42 (58%)
White	7 (78%)	22 (35%)	29 (40%)
Asian	0 (0%)	1 (1.6%)	1 (1.4%)
Hispanic Ethnicity	7 (78%)	18 (29%)	25 (35%)
Intensive Phase Treatment			
HRZE	7 (78%)	15 (24%)	22 (31%)
HRZM ^a	2 (22%)	---	2 (3%)
MRZE ^b	---	48 (76%)	48 (67%)
Thrice weekly therapy ^c	4 (44%)	---	4 (6%)
Pyrazinamide dose (mg)	1000 (1000-1500)	1500 (1000-1500)	1500 (1000-1500)
Pyrazinamide dose (mg/kg)	19.7 (18.6-23.9)	22.9 (20.2-25.4)	22.9 (19.9-25.3)
Clinical Factors			
Cavity on baseline chest X-ray	5 (56%)	50 (79%)	55 (76%)
HIV positive	1 (11%)	2 (3%)	3 (4%)
Weight (kg)	54.1 (53.3-74.0)	57.0 (51.1-63.0)	56.7 (51.3-63.3)

* Uganda or South Africa

Data presented are median (interquartile range) for age, weight, and dose and n (%) for all other factors.

^aHRZM not used in Study 28

^bMRZE not used in Study 27

^cIn Study 27, PK sampling was performed during the first two weeks of therapy, when dosing was daily; after that, some patients received thrice-weekly dosing

Abbreviations: HRZE: isoniazid-rifampicin-pyrazinamide-ethambutol intensive phase regimen; HRZM: isoniazid-rifampicin-pyrazinamide-moxifloxacin intensive phase regimen; MRZE: moxifloxacin-rifampicin-pyrazinamide-ethambutol intensive phase regimen; TB: tuberculosis; BMI: body mass index; HIV: human immunodeficiency virus

Table 2.

Demographic, treatment, and clinical characteristics among participants enrolled in the pharmacokinetic sub-study of PanACEA MAMS-TB

	All study participants (n=96)
Demographic Factors	
Age (years), median (IQR)	34.5 (28.9-39.2)
Male sex	67 (70%)
Race	
Black	83 (86%)
Mixed	13 (14%)
Intensive Phase Treatment	
HRZE	19
HR35ZE	20
HRZQ	19
HR20ZQ	19
HR20ZM	19
Pyrazinamide dose (mg), median (IQR)	1200 (1200, 1600)
Pyrazinamide dose (mg/kg), median (IQR)	25.7 (24.0, 28.3)
Clinical Factors	
Weight (kg), median (IQR)	54.0, (48.9, 56.5)
BMI, median (IQR)	19.2, (17.6, 20.5)*
BMI<18.0 kg/m ²	25 (26%)*
HIV positive	2 (2%)
Cavity on baseline chest X-ray	66 (68.8%)

* 1 patient had height missing

Table 3.

Post hoc Bayesian estimates of pyrazinamide PK parameters from TBTC Studies 27 and 28 and PanACEA MAMS-TB.

Parameter	Median	IQR	
TBTC Trials (68 participants with MIC data)			
Pyrazinamide Pharmacokinetic Parameters			
Predicted C _{MAX} (µg/ml)	29.2	(25.6, 35.0)	
Predicted AUC _{0-24hr} (µg*hr/ml)	306	(261, 357)	
Pharmacodynamic Parameters			
Predicted AUC _{0-24hr} /MIC	8.35	(5.36, 12.7)	
Predicted C _{MAX} /MIC	0.775	(0.549, 1.18)	
PanACEA MAMS Trial (86 participants)			
Pyrazinamide Pharmacokinetic Parameters			
Predicted C _{MAX} (µg/ml)	37.2	(33.4, 40.9)	
Predicted AUC _{0-24hr} (µg*hr/ml)	331	(278, 398)	
Rifampicin Pharmacokinetic Parameters			
Observed C _{MAX} (µg/ml)	Control: HR10ZE	5.56	(5.08, 7.26)
	Arm 1: HR35ZE	26.7	(23.6, 32.1)
	Arm 2: HR10ZQ	3.65	(2.49, 5.04)
	Arm 3: HR20ZQ	12.1	(9.81, 13.2)
	Arm 4: HR20ZM	12.1	(9.83, 14.4)
Observed AUC _{0-24hr} (µg*hr/ml)	Control: HR10ZE	23.4	(17.4, 29.3)
	Arm 1: HR35ZE	164	(131, 199)
	Arm 2: HR10ZQ	18.3	(10.8, 23.5)
	Arm 3: HR20ZQ	66.3	(56.7, 82.9)
	Arm 4: HR20ZM	61.7	(50.5, 78.7)

Table 4.

(A) Clinically-observed maximal concentration (C_{max}) and associated drug doses (90%CI) that would be required to achieve 90 and 95% culture conversion on solid media by 2 months of treatment in TBTC trials and (B) Doses (and clinically-observed C_{max}) of pyrazinamide and rifampicin that would be needed to achieve 90% or 95% culture conversion on liquid media by (i) two months or (ii) three months of treatment, using PanACEA MAMS-TB data.

A)				
Enrollment Site	90% Culture Conversion	95% Culture Conversion		
Non-Africa	43 µg/mL	54 µg/mL	Observed C_{max}	
Africa	93 µg/mL	120 µg/mL		
Non-Africa	1800 mg (2800, 6000)	2200 (1400, 4800)	Expected Dose Level	
Africa	4600 (3000, 8400)	5800 (3800, 8800)		

B)				
Pyrazinamide Dose (mg)*	Pyrazinamide C _{max} (µg/mL)	i. Rifampicin AUC (for conversion by 2 months) (µg*hr/mL)	ii. Rifampicin IF AUC (for conversion by 3 months) (µg*hr/mL)	%Culture Conversion
1500	42	>688	>450	95%
3000	83	>352	>=113	95%
1500	42	>588	>349	90%
3000	83	>251	>=13	90%

* for reference, median weight in MAMS-TB was 54 kg