Prenatal Exposure to Bisphenol A and Child Wheeze from Birth to 3 Years of Age

Adam J. Spanier,^{1,2} Robert S. Kahn,³ Allen R. Kunselman,² Richard Hornung,³ Yingying Xu,³ Antonia M. Calafat,⁴ and Bruce P. Lanphear^{3,5}

¹Department of Pediatrics, and ²Department of Public Health Sciences, Penn State University Hershey Medical Center, Hershey, Pennsylvania, USA; ³Department of Pediatrics, Cincinnati Children's Hospital Medical Center, Cincinnati, Ohio, USA; ⁴Division of Laboratory Sciences, National Center for Environmental Health, Centers for Disease Control and Prevention, Atlanta, Georgia, USA; ⁵Child and Family Research Institute, BC Children's Hospital and Faculty of Health Sciences, Simon Fraser University, Vancouver, Canada

BACKGROUND: Bisphenol A (BPA), an endocrine-disrupting chemical that is routinely detected in > 90% of Americans, promotes experimental asthma in mice. The association of prenatal BPA exposure and wheeze has not been evaluated in humans.

OBJECTIVE: We examined the relationship between prenatal BPA exposure and wheeze in early childhood.

METHODS: We measured BPA concentrations in serial maternal urine samples from a prospective birth cohort of 398 mother–infant pairs and assessed parent-reported child wheeze every 6 months for 3 years. We used generalized estimating equations with a logit link to evaluate the association of prenatal urinary BPA concentration with the dichotomous outcome wheeze (wheeze over the previous 6 months).

RESULTS: Data were available for 365 children; BPA was detected in 99% of maternal urine samples during pregnancy. In multivariable analysis, a one-unit increase in log-transformed creatinine-standardized mean prenatal urinary BPA concentration was not significantly associated with child wheeze from birth to 3 years of age, but there was an interaction of BPA concentration with time (p = 0.003). Mean prenatal BPA above versus below the median was positively associated with wheeze at 6 months of age [adjusted odds ratio (AOR) = 2.3; 95% confidence interval (CI): 1.3, 4.1] but not at 3 years (AOR = 0.6; 95% CI: 0.3, 1.1). In secondary analyses evaluating associations of each prenatal BPA concentration separately, urinary BPA concentrations measured at 16 weeks gestation were associated with wheeze (AOR = 1.2; 95% CI: 1.0, 1.5), but BPA concentrations at 26 weeks of gestation or at birth were not.

CONCLUSIONS: Mean prenatal BPA was associated with increased odds of wheeze in early life, and the effect diminished over time. Evaluating exposure at each prenatal time point demonstrated an association between wheeze from 6 months to 3 years and log-transformed BPA concentration at 16 weeks gestation only.

KEY WORDS: bisphenol A, BPA, child, cotinine, prenatal, tobacco, wheeze. *Environ Health Perspect* 120:916–920 (2012). http://dx.doi.org/10.1289/ehp.1104175 [Online 14 February 2012]

Asthma is one of the most common chronic and disabling conditions of childhood (Bloom et al. 2010). The prevalence of asthma in children has risen in the last few decades, currently affecting 1 in 10 children (Akinbami et al. 2011; Bloom et al. 2010). Several environmental risk factors for asthma have been identified, such as tobacco smoke and airborne pollutants (McConnell et al. 2010; Oberg et al. 2011). The reasons for the increase in childhood asthma are largely unknown, but other environmental exposures may play a role.

Recent studies have implicated exposure to plastic products in the development of childhood respiratory problems. Investigators have found an association of exposure to plastic wall materials (any plastic wall surfaces) with the development of bronchial obstruction, persistent wheeze, cough, and phlegm in children (Jaakkola et al. 1999, 2000). Jaakkola et al. (2006) found that exposure to plastic wall materials also was associated with more than two times the odds of asthma in adults. Indoor concentrations of common organic plasticizers (2,2,4-trimethyl-1,3-pentanediol monoisobutyrate and 2,2,4-trimethyl-1,3pentanediol diisobutyrate) in schools also have been associated with increased prevalence of asthmatic symptoms in students (Kim et al. 2007). Thus, the evidence for an association between plastic exposures and wheeze is growing, but it is unclear whether specific plastic constituents, such as bisphenol A (BPA) or phthalates, act as risk factors for childhood respiratory problems.

BPA is a high-production-volume chemical used in the manufacture of some plastics and epoxy resins that are found in many consumer products. The primary route of human exposure to BPA is through dietary ingestion (Wilson et al. 2007). Unfortunately, there is little information about the variability and routes of BPA exposure in pregnant women (Braun et al. 2011). BPA was detected in the urine of > 90% of the National Health and Nutrition Examination Survey (NHANES) 2003-2004 participants > 6 years of age (Calafat et al. 2008). Perinatal BPA exposure has been reported to promote experimental asthma in mice, but this relationship has not been evaluated in humans (Midoro-Horiuti et al. 2010). The objective of this analysis was to examine the relationship between prenatal BPA exposure and wheeze in early childhood.

Methods

We used data from the Health Outcomes and Measures of the Environment (HOME) Study for this project (Braun et al. 2009; Phelan et al. 2009). The HOME Study is a prospective birth cohort with the primary goal of investigating the effects of exposure to environmental toxicants on child growth, neurobehavioral development, and respiratory health. Between March 2003 and January 2006, the HOME Study enrolled English-speaking women at a mean ± SD of approximately 16 ± 2 weeks gestation who were \geq 18 years and lived in a home built before 1979. We tracked the women through pregnancy and are continuing to follow their children through 5 years of age. Women resided within five Ohio counties surrounding Cincinnati, received prenatal care from one of nine participating obstetrical clinics, and delivered at one of three participating hospitals. The obstetric clinics provided us with lists of new patients, and we prescreened women for eligibility criteria. We sent study information to eligible women and placed a follow up phone call to assess interest in the study and complete screening. Approximately 37% (468 of 1,263) of eligible women agreed to participate and provided written informed consent. We excluded potential participants if they were HIV positive, taking antiepileptic medication, or were diabetic (not gestational) or had bipolar disorder or schizophrenia.

Address correspondence to A. Spanier, Department of Pediatrics, Mail Code HS83, Penn State University, Hershey Medical Center, 500 University Dr., P.O. Box 850, Hershey, PA 17033-0850 USA. Telephone: (717) 531-8006. Fax: (717) 531-0869. E-mail: aspanier@hmc.psu.edu

We acknowledge the technical assistance of X. Ye, A. Bishop, X. Zhou, R. Hennings, T. Jia, P. Olive, and T. Bernert [Centers for Disease Control and Prevention (CDC), Atlanta, GA] for measuring the urinary bisphenol A and creatinine and serum cotinine.

This work was funded by a Flight Attendant Medical Research Foundation Young Clinical Scientist Award (062435_YCSA_Faculty) to A.J.S., R.S.K., A.R.K., R.H., Y.X., and B.P.L. and National Institute of Environmental Health Sciences grants 1K23ES016304 and P01ES11261.

The findings and conclusions in this report are those of the authors and do not necessarily represent the official position of the CDC.

The authors declare they have no actual or potential competing financial interests to disclose.

Received 6 July 2011; accepted 14 February 2012.

Infants of participating mothers were eligible for the longitudinal study. The longitudinal study included an embedded randomized control trial of a lead hazard reduction intervention and injury hazard reduction control.

We included only children with prenatal urinary BPA concentration and respiratory outcome data for this study. These data were available for 365 children (92% of HOME Study participants: 398 live-born infants, randomly excluding one child for twin births). The institutional review boards of Cincinnati Children's Hospital Medical Center, the Centers for Disease Control and Prevention (CDC), and the involved birth hospitals approved the HOME Study and this project.

Exposure assessment. We assessed prenatal BPA exposure by measuring the concentrations of BPA in serial maternal spot urine samples. We collected maternal urine and serum at enrollment (15.9 ± 1.9 weeks gestation), 26 weeks gestation, and birth. Urine and serum samples were sent to the Division of Laboratory Sciences at the CDC for analysis. Urinary BPA concentrations were quantified using online solid-phase extraction coupled with high-performance liquid chromatography/isotope-dilution tandem mass spectrometry (Ye et al. 2005a, 2005b). The limit of detection (LOD) was 0.4 µg/L. When BPA concentrations were below the LOD (< 13% of samples), we replaced these values with the LOD divided by $\sqrt{2}$ (Hornung and Reed 1990).

We assessed tobacco exposure using serum concentrations of cotinine, a metabolite of nicotine. We used serum cotinine, a biomarker of tobacco exposure, as a covariate in analyses because of the established association of prenatal tobacco exposure with wheeze in childhood (Sherriff et al. 2001; Spanier et al. 2011). Analyses of serum for cotinine were performed using high-performance liquid chromatography/atmospheric-pressure chemical ionization tandem mass spectrometry (Bernert et al. 1997, 2000). The LOD was 0.015 ng/mL. When serum cotinine values were below the LOD (~ 35% of samples), we imputed values by sampling randomly from the left tail of a lognormal distribution. We collected other environmental exposure information (i.e., pet ownership, cockroach exposure, and house characteristics) by survey as described below.

Outcome measure. After birth, we employed trained research assistants to survey parents every 6 months through the child's age 3 years to collect study data. The baseline, 12-, 24-, and 36-month surveys were conducted via home visits, and the 6-, 18-, and 30-month surveys were conducted over the phone. We designed these survey questions to parallel the NHANES wheeze question (National Center for Health Statistics 1994). We asked "Has [child's name] had wheezing or whistling in

his/her chest in the last 6 months?" We also asked about the number of wheeze attacks, and we dichotomized the number of wheeze attacks at each time point (no wheeze vs. any wheeze) to minimize effects of extreme values. We used this dichotomized value as our outcome measure. We also asked if wheezing usually occurred with or followed a cold or viral illness.

Covariates. Trained research assistants conducted extensive surveys at baseline and every 6 months after the child was born to collect data on potential covariates, including demographic characteristics, socioeconomic status, and health status. Demographic and socioeconomic characteristics such as maternal education, race/ethnicity, occupation, income, housing volume, and health insurance status were considered as possible covariates in all models. We also considered other factors potentially associated with wheeze by evaluating them as possible covariates [e.g., prenatal tobacco exposure (cotinine), season, history and duration of breast-feeding, family history of asthma, family history of allergy, child eczema, child allergy, neonatal characteristics, pet ownership, and cockroach exposure] (Ball et al. 2000; Fleming et al. 1987; Gold et al. 1999; Oddy et al. 2002; Simoes 2003; Taussig et al. 2003; Young et al. 2000).

We categorized residence location/type into three categories based on census tract information. Family residence was categorized as urban if their census tract was located in a city defined as a metropolitan area by U.S. Census 2000 (Cincinnati, OH; Newport, KY; and Covington, KY) (unpublished data), rural if their census tract had a 2000 Census population density < 500 persons/square mile, and suburban if their census tract did not fit into urban or rural categories.

Statistical analysis. We first calculated descriptive statistics for all demographic, exposure, and outcome data. We used a natural log transformation for all BPA data because urinary BPA concentrations were approximately log-normal, and to minimize the influence of outlying values. We used the geometric mean and 95% confidence interval (CI) as the primary descriptors of central tendency and dispersion for BPA data.

We used the mean of creatinine standardized prenatal urinary BPA concentrations as our primary independent variable to approximate exposure over pregnancy. To calculate this variable, we first standardized BPA concentrations for urinary creatinine (micrograms of BPA per gram of creatinine) at each time point. We then calculated the mean of all available standardized BPA concentrations and log transformed the mean of prenatal BPA concentrations for each participant. We analyzed the association of prenatal urinary BPA concentration with the dichotomous outcome variable wheeze (wheeze over the previous 6 months) using generalized estimating equations (GEEs) with a logit link. GEE is an extension of logistic regression that accounts for within-subject correlation resulting from repeated outcome measurements in longitudinal studies. Therefore, all data collected during the six time points (months 6, 12, 18, 24, 30, and 36) were used in the analysis. Interpretation of the association between the exposure variable and outcome (i.e., wheeze) using odds ratios (ORs) and 95% CIs from GEEs is similar to ordinary logistic regression but represents the association with wheeze over the entire 3-year period rather than at a single point in time. We first conducted bivariate analyses to evaluate the association of prenatal BPA concentration and potential covariates with wheeze. After bivariate analysis, we conducted a multivariable analysis using forward selection techniques. In the initial multivariable analysis, we included covariates that predicted wheeze with a *p*-value ≤ 0.2 in bivariate analysis. Covariates were retained if they were significantly associated with wheeze (p < 0.05)or if their addition caused a > 10% change in the estimate of the association between wheeze and prenatal BPA. In all analyses (including the bivariate analyses), we included a variable for survey time point (6, 12, 18, 24, 30, or 36 months) and a variable for intervention arm to account for any potential design effects of the embedded randomized trial. We also adjusted for log-transformed maternal serum cotinine (continuous) because previous studies have demonstrated an association of cotinine with wheeze (Spanier et al. 2011). After conducting the multivariable analysis, we tested for potential multiplicative interactions of prenatal BPA concentration with all covariates on the logit scale.

We conducted secondary analyses to explore potential windows of vulnerability by replacing the mean prenatal urinary BPA concentration variable with BPA concentration measured at each of the prenatal time points (i.e., 16 weeks, 26 weeks, and at birth) in three separate analyses, each standardized for same time urinary creatinine. GEE analyses were used for these secondary analyses. SAS (version 9.2; SAS Institute Inc., Cary, NC, USA) was used for all data analyses.

Results

More than half of the child participants were female (55.6%), most had a white mother (65.5%), and most were born full term (Table 1). Compared with the 33 participants without outcome data, those for whom outcome data were available were more likely to be white, have a mother with more than a high school education, live in nonurban environment, have married parents, have an older mother, and have a higher household income. Of 365 children with maternal prenatal BPA concentration and respiratory outcome data available, 99% were born to a mother with detectable urinary BPA concentrations at some point during pregnancy. The geometric mean of the mean prenatal urinary BPA concentration was 2.4 µg/g creatinine (95% CI: 2.3, 2.6 µg/g creatinine) (Table 2). During the study, the frequency of children with wheeze during a 6-month period ranged from a low of 15.3% to a high of 24% (Figure 1). Almost all wheezing episodes occurred in relation to a cold; the frequency of children who wheezed without a cold during any 6-month period ranged from a low of 0% to a high of 2.7% (data not shown).

The prenatal creatinine-standardized urinary BPA concentrations measured at 16 weeks of gestation, 26 weeks of gestation, and birth were only weakly correlated (Pearson's correlation coefficients of 0.07, 0.12, and 0.13 for the three pairs of measurements). In bivariate analysis, the OR for a one-unit change in logtransformed mean prenatal BPA concentration was 1.16 (95% CI: 0.90, 1.49). In multivariable analysis, adjusting for season, maternal allergy, mean prenatal serum cotinine concentration, time (categorical), and intervention group, log-transformed mean prenatal urinary BPA concentration was not associated with wheeze [adjusted OR (AOR) = 1.13; 95% CI: 0.86, 1.47].

There was a borderline interaction of mean prenatal urinary BPA concentration with time in the multivariable analysis (p = 0.06). We

dichotomized prenatal BPA at the median $(\geq 2.197 \text{ vs.} < 2.197 \text{ }\mu\text{g/g} \text{ creatinine})$ to permit exploration of the interaction of prenatal BPA concentration with time (Table 3). The interaction of dichotomized BPA and time was significant (p = 0.002). At 6 months of age, the adjusted odds of wheeze for children born to mothers with a BPA concentration above the median compared with those born to mothers with a BPA concentration below the median was 2.27 (95% CI: 1.28, 4.06); by 3 years, there was no longer a significant association of mean prenatal BPA concentration with wheeze (AOR = 0.57; 95% CI: 0.29, 1.10). There was no statistically significant interaction (p > 0.05) of BPA urinary concentrations with any of the other covariates or with sex.

Windows of vulnerability (timing). In secondary analyses, we used separate multivariable models to estimate associations of wheeze (from birth to 3 years of age) with creatininestandardized log-transformed maternal urinary BPA concentration at each time point during pregnancy. A one-unit change in logtransformed prenatal urinary BPA concentration at 16 weeks of gestation had a borderline association with wheeze (AOR = 1.21; 95% CI: 0.99, 1.49); wheeze was not associated with BPA at 26 weeks of gestation (AOR = 1.08; 95% CI: 0.87, 1.34) or at birth (AOR = 0.91; 95% CI: 0.73, 1. 15). Interactions of BPA with time (study time point) were not statistically significant in these analyses.

We also explored whether adjusting, rather than standardizing, for urinary creatinine altered our findings, but the results were similar. Specifically, wheeze was associated with a one-unit increase in log-transformed prenatal urinary BPA concentration at 16 weeks of gestation (AOR = 1.24; 95% CI: 1.00, 1.52) but not at 26 weeks of gestation (AOR = 1.07; 95% CI: 0.86, 1.33) or at birth (AOR = 0.89; 95% CI: 0.71, 1. 12).

Discussion

We evaluated the relationship of prenatal BPA exposure, estimated by measuring maternal urinary BPA concentration, with childhood wheeze in a prospective birth cohort. BPA was detected in the urine of virtually all of the pregnant women. We found that the mean urinary BPA concentration was associated with wheeze in children at 6 months of age, but there was no evidence of a persistent positive association by 3 years of age. In secondary analyses, we explored windows of vulnerability during pregnancy and found an association of wheeze with BPA exposure only during early pregnancy.

Previous investigators have noted an association of exposure to plastic wall materials with the development of bronchial obstruction, adult asthma, and respiratory symptoms typical of childhood asthma (Jaakkola et al. 1999, 2000, 2006; Kim et al. 2007). These studies focused on plastic or plasticizers as a broad category rather than evaluating specific chemicals in plastic. BPA, a chemical found in some plastics, promotes the development of experimental asthma in mouse pups, and perinatal BPA exposure enhanced allergic sensitization, airway hyperresponsiveness, and bronchial eosinophilic inflammation in mice (Midoro-Horiuti et al. 2010). Similarly,

Table 2. Prenatal creatinine-standardized urinary BPA concentrations (μ g/g).

Collection time	п	Geometric mean (95% CI)	Percent >LOD ^a
16 weeks gestation	363	1.9 (1.7, 2.1)	89.8
26 weeks gestation	353	2.2 (2.1, 2.4)	90.4
Birth	330	2.0 (1.9, 2.2)	87.6
Mean prenatal	365	2.4 (2.3, 2.6)	100

 $aLOD = 0.4 \ \mu g/L.$

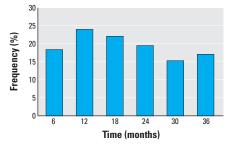


Figure 1. Percentage of children with wheeze over previous 6 months in the HOME Study (*n* = 365).

Table 1. Characteristics of study participants with available prenatal BPA c	concentration and wheeze
data [n (%)].	

Characteristic	Included	Excluded	<i>p</i> -Value ^a
No. of participants	365 (91.7)	33 (8.3)	Not applicable
Male	162 (44.4)	21 (63.6)	0.03
Maternal race			
White	239 (65.5)	5 (15.1)	
Black	103 (28.2)	18 (54.6)	< 0.0001
Other	23 (6.3)	4 (12.1)	
Unknown		6 (18.2)	
Maternal education			
< High school	77 (21.1)	18 (54.5)	< 0.0001
≥ Any college	288 (78.9)	9 (27.3)	
Unknown		6 (18.2)	
Maternal marital status			
Married	249 (68.2)	7 (21.2)	< 0.0001
Not married	116 (31.8)	20 (60.6)	
Unknown		6 (18.2)	
Household income			
< \$25,000	87 (23.8)	15 (45.4)	
\$25,000-50,000	78 (21.4)	6 (18.2)	0.003
\$50,000-80,000	91 (24.9)	1 (3.0)	
> \$80,000	102 (27.9)	3 (9.1)	
Unknown		8 (24.2)	
Residence			
Rural	12 (3.3)	1 (3.0)	
Suburban	133 (36.4)	6 (18.2)	0.04
Urban	220 (60.3)	26 (78.8)	
History of maternal allergy	157 (43.0)	11 (40.7)	0.82
Prenatal serum cotinine concentration, µg/L [geometric mean (95% CI)]	0.06 (0.04, 0.07)	1.15 (0.41, 3.23)	< 0.0001
Gestational age, weeks (mean \pm SD)	39.0 ± 1.8	38.0 ± 2.8	0.07

^aRepresents a test (chi-square or *t*-test) of the difference between the children in the included and excluded groups.

we found that prenatal exposure to BPA was associated with wheeze in infants.

Nearly all the children in this study were born to mothers exposed to BPA during pregnancy, based on urinary BPA concentrations. This finding is consistent with NHANES data: Urinary BPA was detected in nearly 93% of participants > 6 years of age (Calafat et al. 2008). The geometric mean urinary BPA concentration among women in the HOME Study cohort was 2.4 µg/g creatinine, compared with 2.6 µg/g creatinine reported in NHANES for 2003–2004 (Calafat et al. 2008).

There are limited human data about the bioavailability and metabolism of BPA during pregnancy. It has been demonstrated that BPA at low levels can cross the human placenta in an *ex vivo* model (Balakrishnan et al. 2010). It has also been noted that BPA levels in human amniotic fluid appear to be higher in early pregnancy than in later pregnancy (Ikezuki et al. 2002). Unfortunately, we did not have direct measures of fetal or amniotic BPA concentrations in this study.

We did not find an association between mean prenatal urinary BPA concentration and wheeze over the entire study period but did find evidence of an association with wheeze in infancy that decreased over time. This early association may be spurious, or it may be analogous to the transient early wheeze phenotype of pediatric asthma (Martinez et al. 1995).

Our primary measure of prenatal BPA exposure was the mean of up to three serial urinary BPA concentrations standardized for creatinine at each time point, but we also examined windows of vulnerability. Increased urinary BPA concentration at 16 weeks of gestation had a borderline association with increased odds of wheeze during the entire time period from 6 months to 3 years of age. This finding suggests that exposure to BPA in early pregnancy might be driving the observed association of mean prenatal BPA concentration and wheeze. There are no other human respiratory studies to compare with this result, but it is consistent with an evaluation of prenatal BPA exposure and child behavior in the same prospective cohort that found that 16-week-gestation urinary BPA concentration was significantly associated with externalizing behaviors (Braun et al. 2009).

We found that BPA concentrations from the three prenatal urinary measurements were weakly correlated. Other biological markers such as serum cotinine measured at similar intervals during pregnancy demonstrate higher correlations (correlation coefficients $r \ge 0.8$) (Spanier et al. 2011). Despite this weak correlation some investigators have suggested that a single urinary measure of BPA is predictive of long-term exposure (Mahalingaiah et al. 2008). Other investigators have noted that BPA concentrations for an individual vary both within and among days (Ye et al. 2011), and some suggest that studies may need to measure BPA concentration more than once during pregnancy (Braun et al. 2011). We chose to use mean prenatal urinary BPA concentration as the primary exposure measure to represent exposure averaged over pregnancy. However, using the mean prohibited us from adjusting for time-specific urinary creatinine, which was suggested by Barr et al. (2005). Rather than adjusting for urinary creatinine in our analyses, we standardized each BPA concentration before calculating the mean to account for differences in urinary concentration and kidney filtration. Our use of the mean of the maternal urinary BPA concentrations was a conservative approach that could have underestimated associations because it averaged several measures. Future work should evaluate the best approach for BPA exposure assessment throughout pregnancy.

There are several strengths and limitations to this study. First, although having serial measures of BPA is a strength, it forced us to standardize urinary BPA concentration for creatinine. Some investigators suggest that it may be better to adjust for creatinine in the analysis rather than standardize each time point. Adjustment requires inclusion of the creatinine and BPA as separate variables in the analysis. In this case we have three maternal measures, and it would not have been appropriate to place the three maternal BPA concentrations and the three creatinine concentrations in the same analysis. Second, because of the variability of urinary BPA concentrations over time, using this biomarker could result in exposure misclassification. Third, our outcome variable, wheeze, was based on parent report and could have

Table 3. Adjusted association of mean prenatal BPA urinary concentration (dichotomized at the median)^a with child wheeze.

Children reporting wheeze [n (%)]			
Child age	High-BPA group	Low-BPA group	AOR (95% CI)
6 months	46 (25.6)	22 (12.3)	2.27 (1.28, 4.06)
12 months	45 (27.3)	36 (20.6)	1.49 (0.89, 2.48)
18 months	38 (24.4)	31 (19.0)	1.20 (0.69, 2.09)
24 months	22 (15.4)	32 (22.5)	0.55 (0.31, 0.99)
30 months	25 (17.2)	18 (13.2)	1.23 (0.63, 2.40)
36 months	18 (13.4)	26 (20.3)	0.57 (0.29, 1.10)

^aMean prenatal BPA urinary concentration was dichotomized at the median (high BPA, ≥ 2.197 µg/g creatinine; low BPA < 2.197 µg/g creatinine).

been under- or overreported. Fourth, although the HOME Study is a representative sample of the population in the Cincinnati metropolitan area, it is not a random sample. Last, there was differential attrition for our cohort; the attrition rate for minority, low-income families was higher than for white, affluent families. Future studies should explore the association of BPA exposure with wheeze beyond 3 years of age, with objective measures of lung function, and with the diagnosis of asthma.

Conclusion

We found that mean prenatal BPA exposure, assessed from maternal urinary BPA concentration, was associated with increased odds of wheeze at 6 months of age, but the association diminished by 3 years of age. Evaluating BPA exposure at each prenatal measurement time demonstrated an association of wheeze with BPA at 16 weeks of gestation but not at 26 weeks of gestation or at birth, which suggests that early pregnancy may be the most sensitive time window of exposure. In the analyses of each separate prenatal measurement, there was no evidence that the effect diminished across the 3 years. If exposure to BPA is confirmed to increase the risk for developing wheeze or asthma, it may help explain the rising rates of childhood asthma and would have important policy implications for women of childbearing age and their children.

REFERENCES

- Akinbami L, Moorman J, Liu X. 2011. Asthma Prevalence, Health Care Use, and Mortality: United States, 2005–2009. National Health Statistics Report no. 32. PHS 2011-1250. Available: http://www.cdc.gov/nchs/data/nhsr/nhsr032.pdf [accessed 20 October 2011].
- Balakrishnan B, Henare K, Thorstensen EB, Ponnampalam AP, Mitchell MD. 2010. Transfer of bisphenol A across the human placenta. Am J Obstet Gynecol 202(4):393.e1–393.e7.
- Ball TM, Castro-Rodriguez JA, Griffith KA, Holberg CJ, Martinez FD, Wright AL. 2000. Siblings, day-care attendance, and the risk of asthma and wheezing during childhood. N Engl J Med 343(8):538–543.
- Barr DB, Wilder LC, Caudill SP, Gonzalez AJ, Needham LL, Pirkle JL. 2005. Urinary creatinine concentrations in the U.S. population: implications for urinary biologic monitoring measurements. Environ Health Perspect 113:192–200.
- Bernert JT Jr, McGuffey JE, Morrison MA, Pirkle JL. 2000. Comparison of serum and salivary cotinine measurements by a sensitive high-performance liquid chromatographytandem mass spectrometry method as an indicator of exposure to tobacco smoke among smokers and nonsmokers. J Anal Toxicol 24(5):333–339.
- Bernert JT Jr, Turner WE, Pirkle JL, Sosnoff CS, Akins JR, Waldrep MK, et al. 1997. Development and validation of sensitive method for determination of serum cotinine in smokers and nonsmokers by liquid chromatography/atmospheric pressure ionization tandem mass spectrometry. Clin Chem 43(12):2281–2291.
- Bloom B, Cohen R, Freeman G. 2010. Summary Health Statistics for U.S. Children: National Health Interview Survey, 2009. Vital and Health Statistics ser. 10, no. 247. PHS 2011-1575. Available: http://www.cdc.gov/nchs/data/series/sr_10/ sr10_247.pdf [accessed 20 October 2011].
- Braun JM, Kalkbrenner AE, Calafat AM, Bernert JT, Ye X, Silva MJ, et al. 2011. Variability and predictors of urinary bisphenol A concentrations during pregnancy. Environ Health Perspect 119:131–137.

Braun JM, Yolton K, Dietrich KN, Hornung R, Ye X, Calafat AM,

et al. 2009. Prenatal bisphenol A exposure and early childhood behavior. Environ Health Perspect 117:1945–1952.

- Calafat AM, Ye X, Wong LY, Reidy JA, Needham LL. 2008. Exposure of the U.S. population to bisphenol A and 4-*tertiary*-octylphenol: 2003–2004. Environ Health Perspect 116:39–44.
- Fleming DW, Cochi SL, Hightower AW, Broome CV. 1987. Childhood upper respiratory tract infections: to what degree is incidence affected by day-care attendance? Pediatrics 79(1):55–60.
- Gold DR, Burge HA, Carey V, Milton DK, Platts-Mills T, Weiss ST. 1999. Predictors of repeated wheeze in the first year of life: the relative roles of cockroach, birth weight, acute lower respiratory illness, and maternal smoking. Am J Respir Crit Care Med 160(1):227–236.
- Hornung R, Reed LD. 1990. Estimation of average concentration in the presence of nondetectable values. Appl Occup Environ Hyg 5(1):46–51.
- Ikezuki Y, Tsutsumi O, Takai Y, Kamei Y, Taketani Y. 2002. Determination of bisphenol A concentrations in human biological fluids reveals significant early prenatal exposure. Hum Reprod 17(11):2839–2841.
- Jaakkola JJ, Ieromnimon A, Jaakkola MS. 2006. Interior surface materials and asthma in adults: a population-based incident case-control study. Am J Epidemiol 164(8):742–749.
- Jaakkola JJ, Oie L, Nafstad P, Botten G, Samuelsen SO, Magnus P. 1999. Interior surface materials in the home and the development of bronchial obstruction in young children in Oslo, Norway. Am J Public Health 89(2):188–192.
- Jaakkola JJ, Verkasalo PK, Jaakkola N. 2000. Plastic wall materials in the home and respiratory health in young children. Am J Public Health 90(5):797–799.
- Kim JL, Elfman L, Mi Y, Wieslander G, Smedje G, Norback D. 2007. Indoor molds, bacteria, microbial volatile organic compounds and plasticizers in schools--associations with

asthma and respiratory symptoms in pupils. Indoor Air 17(2):153–163.

- Mahalingaiah S, Meeker JD, Pearson KR, Calafat AM, Ye X, Petrozza J, et al. 2008. Temporal variability and predictors of urinary bisphenol A concentrations in men and women. Environ Health Perspect 116:173–178.
- Martinez FD, Wright AL, Taussig LM, Holberg CJ, Halonen M, Morgan WJ. 1995. Asthma and wheezing in the first six years of life. The Group Health Medical Associates. N Engl J Med 332(3):133–138.
- McConnell R, Islam T, Shankardass K, Jerrett M, Lurmann F, Gilliland F, et al. 2010. Childhood incident asthma and traffic-related air pollution at home and school. Environ Health Perspect 118:1021–1026.
- Midoro-Horiuti T, Tiwari R, Watson CS, Goldblum RM. 2010. Maternal bisphenol A exposure promotes the development of experimental asthma in mouse pups. Environ Health Perspect 118:273–277.
- National Center for Health Statistics. 1994. Plan and Operation of the Third National Health and Nutrition Examination Survey, 1988–94. Vital and Health Statistics, ser. 1, Programs and Collections Procedures, no. 32. PHS 94-1308. Available: http://www.cdc.gov/nchs/data/series/ sr_01/sr01_032.pdf [accessed 20 October 2011].
- Oberg M, Jaakkola MS, Woodward A, Peruga A, Pruss-Ustun A. 2011. Worldwide burden of disease from exposure to second-hand smoke: a retrospective analysis of data from 192 countries. Lancet 377(9760):139–146.
- Oddy WH, Peat JK, de Klerk NH. 2002. Maternal asthma, infant feeding, and the risk of asthma in childhood. J Allergy Clin Immunol 110(1):65–67.
- Phelan KJ, Khoury J, Xu Y, Lanphear B. 2009. Validation of a HOME injury survey. Inj Prev 15(5):300–306.
- Sherriff A, Peters TJ, Henderson J, Strachan D. 2001. Risk factor associations with wheezing patterns in children

followed longitudinally from birth to 3(1/2) years. Int J Epidemiol 30(6):1473–1484.

- Simoes EA. 2003. Environmental and demographic risk factors for respiratory syncytial virus lower respiratory tract disease. J Pediatr 143(5 suppl):S118–S126.
- Spanier AJ, Kahn RS, Xu Y, Hornung R, Lanphear BP. 2011. Comparison of biomarkers and parent report of tobacco exposure to predict wheeze. J Pediatr 159(5):776–782.
- Taussig LM, Wright AL, Holberg CJ, Halonen M, Morgan WJ, Martinez FD. 2003. Tucson Children's Respiratory Study: 1980 to present. J Allergy Clin Immunol 111(4):661–675.
- Wilson NK, Chuang JC, Morgan MK, Lordo RA, Sheldon LS. 2007. An observational study of the potential exposures of preschool children to pentachlorophenol, bisphenol-A, and nonylphenol at home and daycare. Environ Res 103(1):9–20.
- Ye X, Kuklenyik Z, Needham LL, Calafat AM. 2005a. Automated on-line column-switching HPLC-MS/MS method with peak focusing for the determination of nine environmental phenols in urine. Anal Chem 77(16):5407–5413.
- Ye X, Kuklenyik Z, Needham LL, Calafat AM. 2005b. Quantification of urinary conjugates of bisphenol A, 2,5-dichlorophenol, and 2-hydroxy-4-methoxybenzophenone in humans by online solid phase extraction-high performance liquid chromatography-tandem mass spectrometry. Anal Bioanal Chem 383(4):638–644.
- Ye X, Wong LY, Bishop AM, Calafat AM. 2011. Variability of urinary concentrations of bisphenol A in spot samples, firstmorning voids, and 24-hour collections. Environ Health Perspect 119:983–988.
- Young S, Sherrill DL, Arnott J, Diepeveen D, LeSouef PN, Landau LI. 2000. Parental factors affecting respiratory function during the first year of life. Pediatr Pulmonol 29(5):331–340.